(Comparative Studies on Effectiveness Of
Bra	anded And Unbranded Disinfectants On on
	E. coli and Staphylococcus species.
ABST	TRACT (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)
clinica Study disinfe Staph Place Depar Lahore Metho analyz Staph both b micro Resul gram unbran from 0 for Sta rangeo 21mm While positiv from 1 did no Concl	to compare the antimicrobial potential of branded and unbranded disinfectants on a bacterial isolates. design: the study was for a period of two months (June-July, 2018). Ten- bectants of which five were branded and five unbranded were used against <i>E. coli</i> and <i>ylococcus aureus.</i> and Duration of Study: Sample: Department of Medicine (Medical Unit IV) and tment of Radiology, Services Institute of Medical Sciences (SIMS), Services Hospital e, between June 2009 and July 2010. bdology: faecal samples were collected from the University Medical centre and was teed in the Microbiology Laboratory for the isolation of <i>Escherichia coli</i> and <i>ylococcus aureus</i> using standard microbiological method. The antimicrobial potential of the technique and the well in agar technique ts: revealed that both branded and unbranded disinfectants were effective on both negative and positive organisms with varying zones of inhibition. However, the negative and positive organisms with varying zones of inhibition. However, the finded were only effective at high concentrations. <i>E. coli had</i> zone of inhibition ranging to 22mm when tested with the unbranded disinfectant, while 0 to 17mm was recorded aphylococcus aureus. The zones of inhibition of the branded disinfectant on <i>E. coli</i> drom 0 to 28mm, while zone diameter of <i>Staphylococcus aureus</i> ranged from 0 to 28mm, while zone diameter of <i>Staphylococcus aureus</i> ranged from 0 to 2. Among the unbranded disinfectants, Lysol produced the highest zone of inhibition. The re control was effective against all tested organisms with zones of inhibition ranging 7-26 mm. On the other hand, as expected, the negative control (sterile distilled water) t show any zone of inhibition.

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18 **1. INTRODUCTION**

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20 Antimicrobials are substances that have the ability to kill or inhibit the growth or proliferation 21 of microorganisms [17]. This implies that these substances when introduced in objects or 22 other materials or consumed could either be bacteriostatic or bactericidal in action. 23 According to Douglas and Braide [6], antimicrobial substances that when introduced on 24 inanimate objects kills or inhibits the growth of microbes. Thus, a good disinfectant should be able to offer complete and full microbiological sterilization, without harming humans and 25 26 useful form of life, be inexpensive and noncorrosive. However, most disinfectants are also, 27 by nature, potentially harmful to humans and animals. The choice of disinfectant to be used 28 may depend on the demanding situation. According to Van et al [15], the idea of using 29 disinfectants and antiseptics is to control or reduce the presence of microorganisms. In order to prevent infections as it regards injury, the most vital measure is to kill or inhibit the growth 30 31 of microorganisms on the skin, wounds and in human body cavity [3]. The antimicrobial 32 potentials of these disinfectants could be influenced by their formulation properties, concentration of organic components, temperature, synergy, rate of dilution and 33 experimental procedures, mode of application, water solubility and pH [5 and 6]. Application 34 factors include the type of surface to be applied, the type of (organic) soil, the temperature 35 and contact time as well as humidity and the method of application (with or without 36 mechanical action) [8]. A disinfectant could be branded or unbranded (Maillard, 2005). 37 38 These unbranded disinfectants are hawked from place to place and also sold in the local 39 markets [9]. They could be good alternative disinfecting agents if their effectiveness against 40 some clinical isolates is known [10]. Unbranded disinfectants are produced locally by people 41 that are taught how to make different household washing, cleaning and disinfecting agents. 42 When these disinfectants are made by these persons, they are normally packaged in 43 containers (usually liable plastic bottles). There are two different ways by which disinfectants 44 can act on microorganisms: growth inhibition (bacteriostasis and fungistasis) or lethal action (bactericidal, fungicidal or viricidal effects) [2]. Thus, this study is aimed at comparing the 45 antimicrobial potential of branded and unbranded disinfectants on clinical bacterial isolates. 46

B 2. METHODOLOGY

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50 2.1 Collection of clinical Samples

51 Faecal samples were collected from the Rivers State University Medical Center, Port 52 Harcourt in specimen bottle and transported to the Microbiology laboratory of Rivers State 53 University, Port Harcourt.

54 2.2 Collection of Disinfectant Samples

55 The branded disinfectants used were; Purit, Dettol, Ivy's, Salvon and Robert. While the 56 unbranded disinfectants were; Lysol, Pine oil, Morigade, Nigertol, Chlonoxynol. The 57 disinfectants were purchased from different markets within Port Harcourt Metropolis, Rivers 58 State.

59 **2.3 Isolation of Test Organisms**

Isolation of the test organisms was carried out as described by Cheesbrough [4]. A thick
suspension of the faecal sample was emulsified in 1ml sterile peptone water. Afterwards a
loop full of the emulsified sample was inoculated on Mannitol salt agar plates (MSA) and
Eosin methylene blue agar plates (EMB). Plates were then incubated at 37°C for 24 hours.

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67 2.3.1 Confirmation of Test Organisms

Ensuing colonies on the MSA and EMB plates were carefully picked using a sterile wire loop and subcutured on fresh plates of MSA and EMB agar. Pure isolates were then stored in nutrient agar slants and stored in the refrigerator for further use.

47 48 The respective pure isolates were identified using conventional methods as described by Cheesbrough [4]. Further confirmation of isolates was done by comparing their biochemical results with those presented in Bergey's manual of determinative bacteriology [18]. The conventional methods include; microscopy, motility, coagulase, catalase, oxidase, indole production, methyl red, citrate utilization, vogues Proskauer test and sugar fermentation [4].

76 **2.4 Standardization of Test Inoculum**

Test isolates were standardized using the 0.5 McFarland. The test isolates were placed in sterile test tubes containing 4ml distilled water. The turbidity was ascertained using the already prepared McFarland standard [4]. The standardized isolates were carefully spread on prepared sterile Mueller-Hinton agar plates as described by Wemedo and Robinson [16]. Plates were allowed to dry before 4 wells using a 6mm well borer were made on the dried seeded plates.

83 2.4.1 Antimicrobial Assay (Well-in-agar method)

The antimicrobial activity of each disinfectants with different concentration was tested in vitro against *E. coli* and *Staphylococcus aureus*. Aliquots (0.1ml) of 10%, 25%, 50% and 100% concentration of the different disinfectants were transferred using sterile Pasteur pipette in to the four wells. The plates were then incubated at $37 \,^{\circ}$ C for 18 to 24hours in an upright position. Autoclaved distilled water was used as negative control while ofloxacin was used as a positive control. After incubation, the plates were observed and the zones of inhibition that developed were read and interpreted [16].

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92 2.4.2 Broth Dilution Method

93 The minimum inhibitory concentrations of the different disinfectants were done using the 94 broth dilution method as described by Prescott et al (2011). Different concentrations of the disinfectants were prepared (10, 25, 50, 75 and 100) mg/ml [1]. One milliliter (1ml) of the 95 96 standardized inoculum and the various concentrations of the disinfectants were put into the 97 sterile tubes of nutrient broth respectively. Tubes containing nutrient broth and organisms 98 without the disinfectant served as negative control while the tube containing only the broth 99 and disinfectant without organism served as positive control. These tubes were incubated at 100 37 °C for 18 to 24 hours. Thereafter, the tubes were examined for visible growth or turbidity and recorded. The MIC is the concentration at which no visible growth was observed when 101 102 compared with the control [9].

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105 3. RESULTS AND DISCUSSION

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107 The result in Table1 showed the characteristics of the two bacterial isolates to some 108 biochemical tests as well as their morphology. The result showed that the isolates were 109 Escherichia coli and Staphylococcus aureus. In this current study, the antimicrobial activities 110 of both the branded disinfectants and unbranded disinfectants on Escherichia coli and Staphylococcus aureus using the agar well diffusion showed some level of inhibition. In 111 Table2, the effect of the unbranded disinfectants on Escherichia coli showed that the 112 effectiveness of the unbranded disinfectants occurred at the 50% and 100% concentration 113 114 and at 50% concentration only Lysol, morigade and pine oil were able to produce a clear zone of inhibition while Chlonoxynol and Nigertol showed no antimicrobial effect. 115 Furthermore, all the unbranded disinfectants were able to exert some antimicrobial 116 117 properties thereby leading to the formation of zones of inhibition at 100% concentration 118 (Table2.). Lyaol and Morigade showed the highest zones of inhibition of 22mm and 20 mm 119 respectively thereby making them the most effective unbraded disinfectants on E. coli.

120 The antimicrobial activities of the unbranded disinfectant on *Staphylococcus aureus* showed 121 that the unbranded disinfectants were not effective at 10 and 25 % concentrations. Also, at 122 50% concentration only pine oil was able to inhibit the staphylococcal isolates at a zone of 123 10mm. whereas at 100% concentration, all unbranded disinfectants except Pine oil exerted 124 some level of antimicrobial activities showing visible zones (Table 3).

The result of the antimicrobial activities of the branded disinfectant on Escherichia coli is 125 presented in Table 4. The result showed that only Robert and Salvon were able to inhibit the 126 isolates of E. coli at 10% with zones of inhibition observed to be 12mm and 7mm 127 128 respectively. While at 25, 50 and 100%, all branded disinfectants produced visible zones of 129 inhibition on the isolates. At 25% concentration, Robert was the most effective having zone of 15mm while lvy's and Salvon were the most effective disinfectants at the 50% 130 131 concentration with zones observed around 18mm and 20mm respectively. At 100% only Purit produced the least zone of inhibition of 18mm on the isolates while other disinfectants 132 had greater zones of inhibitions. 133

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135 Table1. Colonial morphology and biochemical characteristic of the bacterial isolates

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Isolate	Morphology	microscopy	G	L	S	М	F	Cat	Coa.	Ind.	MR.	Cit.	Mot	Identity
												X.		
А	Metallic-silver	-ve bacilli	+	+	+	-	-	+	-	+	+		+	Escherichia coli
	small round											Ŵ		
	flat									X				
В	Golden yellow	+ve clustered	+	+	+	+	+	+	+		+	+	-	Staphylococcus
	round smooth	cocci								•				aureus

136 Key: G; glucose, L; Lactose, S; Sucrose, M; Maltose, Cat. ;Catalase, Coa.; Coagulase, Ind.;Indole;

- 137 MR; Methyl red, Mot.; Motility, Oxi.; Oxidase,
- 138 SSA; Salmonella and Shigella Agar, MSA; Manitol salt Agar

139 Table2 effect of unbranded disinfectants on Escherichia coli

Control					Disinfectants								
Con.		itive oxacin	Negative (Sterile	Chlo	onoxynol	Lyso)I	Mor e	igad	Nige	ertol	Pine	oil
) P1	P2	water) P1 P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2
10%	8	10	00 00	00	00	00	00	00	00	00	00	00	00
25%	12	11	00 00	00	00	00	00	00	00	00	00	00	00
50%	16	18	00 00	00	00	12	12	10	10	00	00	6	6
100%	10	22	00 00	00	00	22	22	20	20	12	10	10	14
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- 151 **Table3. effect of unbranded disinfectants on** *Staphylococcus aureus*

		С	ontrol					Disi	infecta	ants				
Con.	on. Positive (Ofloxacin)		Negative (Sterile water)		Chlonoxynol		Lysol		Morigade		Nigertol		Pine oil	
	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2
10%	12	10	00	00	00	00	00	00	00	00	00	00	00	00
25%	17	16	00	00	00	00	00	00	00	00	00	00	00	00
50%	21	20	00	00	00	00	00	00	00	00	00	00	10	10
100%	28	28	00	00	00	00	17	19	12	10	14	14	00	00

155 Table 4. effect of branded disinfectants on Escherichia coli

		С	ontrol			Disinfectants									
Con.	Posi (Oflo	itive oxacin)	Nega (Ster	itive ile water)		Dettol	I	vy's	Purit	Ro	bert	Sa	vlon		
	P1	P2	P1	P2	P1	P2	P1	P2	P1 P2	P1	P2	P1	P2		
10%	8	10	00	00	00	00	00	00	00 00	12	10	5	7		
25%	12	11	00	00	12	10	12	14	11 9	13	15	12	14		
50%	16	18	00	00	14	14	18	16	12 14	15	17	20	18		
100%	10	22	00	00	22	20	26	25	16 18	24	26	26	28		
156															

Table 5. effect of branded disinfectants on Staphylococcus aureus

		С	ontrol		Ψ.		Dis	infecta	ints				
Con.	Posi (Oflo	tive oxacin)	Negative (Sterile water)		Dettol	ľ	vy's	Ρι	urit	Rol	bert	Sav	/lon
	P1	P2	P1 P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2
10%	12	10	00 00	12	10	00	00	9	11	00	00	8	8
25%	17	16	00 00	16	18	00	00	21	17	16	14	16	14
50%	21	20	00 00	20	22	10	8	23	21	17	18	22	20
100%	28	28	00 00	22	24	14	16	24	26	21	23	22	24

Table6. Concentration of Activity of Branded Disinfectants (MIC)

Organisms	Branded Disinfectants	Concentration (%)
Staphylococcus aureus	Purit	75
	Dettol	50
	lvy's	75
	Salvon	75
	Robert	75
Escherichia coli	Purit	50
	Dettol	75
	lvy's	75
	Salvon	50
	Robert	75

MIC: minimal inhibitory concentration

167		Activity of Unbranded Disinfect	
	Organisms	Unbranded Disinfectants	Concentration (%)
	Staphylococcus aureus	Lysol	75
		Pine oil	75
		Morigade	75
		Nigertol	75
		Chlonoxynol	75
	Escherichia coli	Lysol	50
		Pine oil	75
		Morigade	50
		Nigertol	75
		Chlonoxynol	75
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168 MIC: minimal inhibitory concentration

The bacterial isolates in this current study have shown some level of resistance and 169 susceptibility to the various form of disinfectants. 170

171 Staphylococcus aureus is a known cause of various form of infections ranging minor skin 172 infections, such as pimples, impetigo, boils, cellulitis, folliculitis, carbuncles, scalded skin 173 syndrome, and abscesses, to life-threatening diseases such as pneumonia, meningitis, 174 osteomyelitis, endocarditis, toxic shock syndrome, gastrointestinal diseases, bacteremia and 175 sepsis [13 and 20]. While some strains of Escherichia coli are virulent and are responsible 176 for diarrheal infections worldwide as well as neonatal meningitis, septicemia, and urinary 177 tract infections (UTIs) [14].

178 The result in this study showed that the branded disinfectants are very much effective than the unbranded disinfectants. There is also a dearth of information on the effectiveness as 179 well as the composition of unbranded disinfectants. However, Douglas and Braide (2015) in 180 a study of the effectiveness of Locally Formulated Unbranded disinfectants on clinical 181 182 bacterial isolates reported that unbranded (locally formulated) disinfectants are more potent 183 when not diluted and that the differences in the activities of the unbranded and branded disinfectants may be due to the different substances used in formulations, as well as the 184 185 structure and nature of the cell wall of the microbes. The disinfectants in this current study 186 showed some level of activity on both Gram negative and positive bacterial isolates indicating that they have broad spectrum of activity. This is in agreement with Douglas and 187 188 Braide [6] who also reported that disinfectants show a broad spectrum of activity against 189 different bacterial isolates.

Effectiveness of Dettol and Savlon has been reported by [9] who carried out a study on the 190 efficacy of some disinfectants on clinical isolates including Escherichia coli and 191 192 Staphylococcus aureus in their study, Dettol was more active against the isolates compared 193 with Savlon and other tested disinfectants. Other studies carried out by Olowe [12] and 194 Olasehinde et al [11] also reported Dettol to be a strong disinfectant. Furthermore, El-Mahmood and Doughari [7] in a study of Bacteriological examination of some diluted 195 disinfectants routinely used in the specialist hospital Yola, Nigeria reported that Purit has a 196 higher activity on E. coli than S. aureus whereas in this study Purit was more effective 197 against S. aureus than E. coli. 198

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4. CONCLUSION 201

The antimicrobial effectiveness of five unbranded disinfectants and five branded 203 disinfectants Staphylococcus aureus, and Escherichia coli was evaluated. Despite some 204

level of antimicrobial actions observed in the unbranded disinfectants, the findings in this study has shown that the branded disinfectants are more effective than the unbranded disinfectants. Also, since the unbranded disinfectants have shown some level of antimicrobial actions, increasing the formulation or the quantity for disinfection would be necessary.

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211 Competing interests

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213 Authors have declared that no competing interests exist.

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6 ETHICAL APPROVAL (WHERE EVER APPLICABLE)

The permission to undertake this study was obtained from the Rivers State Health Research Ethical Committee.

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