

Original Research Article

ANTIBIOTIC SUSCEPTIBILITY OF BACTERIA ISOLATED FROM ABATTOIR EFFLUENT-IMPACTED TAGANGU RIVER, ALEIRO, KEBBI STATE, NORTH- WESTERN NIGERIA

ABSTRACT

This study aimed to evaluate the impact of abattoir effluent on microbiological quality of the receiving Tagangu River and the susceptibility of isolates to commonly-used antibiotics. The total heterotrophic population as well as *Escherichia coli* O157:H7 numbers in a total of 30 water samples collected over a period of three months at three strategic points of the river indicated that the river has been heavily polluted with the effluent discharges and did not meet any of the WHO guidelines for natural water sources fit for irrigation or other domestic purposes.

In accordance with CLSI guidelines, four of the eight bacteria (*Enterobacter* sp., *Pseudomonas aeruginosa*, *Proteus vulgaris* and *Citrobacter* sp.) isolated, demonstrated multiple antibiotic resistance (MAR) against at least three of septrin, chloramphenicol, amoxicillin, augmentin, gentamicin, tarivid and streptomycin. All the isolates (*Escherichia coli*, *Klebsiella pneumoniae*, *Enterobacter* sp., *Pseudomonas aeruginosa*, *Proteus vulgaris*, *Citrobacter* sp., *Serratia marcescens* and *Aerobacter aerogenes*) showed either high or intermediate susceptibility to sparfloxacin, ciprofloxacin and pefloxacin. Indiscriminate discharge of abattoir effluent could impact on the microbiological quality and promote increased incidence of multiple antibiotic resistant bacteria in a receiving river.

Keywords: Abattoir, effluent, Tagangu River, Microbiological quality, antibiotic susceptibility Test.

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25 **1.0 INTRODUCTION**

26 Abattoir waste disposal in many developing countries including Nigeria has been a major
27 challenge for years [1]. In most cases, waste materials are disposed of without regard to sound
28 environmental management practices, thus making them harmful to humans and other terrestrial
29 and aquatic life [2]. Studies from Nigeria and Ghana show that many abattoirs in the respective
30 countries either deposit waste materials in the immediate environs or dispose of them directly
31 into water bodies, some of which serve as sources of water for the abattoirs [3].

32 The major known sources of water pollution are municipal, industrial and agricultural. The most
33 polluting of them are sewage and industrial waste discharges into rivers. Industrial effluents
34 mostly contained microbes, heavy metals, acids, hydrocarbons and atmospheric depositions [4].

Comment [TKC3]: rephrase it

35 In Nigeria, Meat processing activities are generally carried out in unsuitable buildings and by
36 untrained personnel or butchers who are most of the time unaware of sanitary principles [5]. The
37 major activities involved in the operations of an abattoir are: receiving and holding of livestock;
38 slaughter and carcass dressing of animals; chilling of carcass products; carcass boning and
39 packaging; freezing of finished carcass and cartooned product; rendering processes; drying of
40 skins; treatment of wastes and transport of processed materials [5].

41 **1.1 Abattoir Effluent as a Pollutant**

42 In Nigeria, available reports cite gross contamination of most major river bodies across the
43 nation by discharge of industrial effluents, sewage and agricultural wastes among others [6].

44 Abattoir activities may be another source of water pollution since human activities such as
45 animal production and meat processing have been reported to impact negatively on soil and
46 natural water composition leading to pollution of soil, natural water resources and the entire
47 environment [7].

48 Yahaya *et al.*, [8], reported that animals which graze on contaminated plants and drink from
49 polluted waters, as well as marine lives that breed in heavy metal polluted waters also
50 accumulate such metals in their tissues and milk if lactating. When such animals are killed, these
51 metals are released in the soil as natural sink but subsequently leached out into nearby streams or
52 water bodies.

53 **1.2 Impact of Untreated Abattoir Effluent**

54 The continuous drive to increase meat production for the protein needs of the ever increasing
55 world population has some problems attached [9]. In developing countries like Nigeria, water
56 pollution from abattoir frequently arises from activities in meat production as a result of failure
57 in adhering to good manufacturing practices and good hygienic practices [10].

58 Discharge of abattoir wastewater to surface waters affects the water quality. One of the
59 environmental effects of discharging slaughterhouse wastewater causes de-oxygenation of rivers
60 and the contamination of groundwater [11, 12, 13]. Moreover, discharge of high levels of
61 biodegradable organics into receiving streams results in increased microbial activity associated
62 with excessive nutrient loadings which requires greater amounts of oxygen than natural aeration
63 processes. This decreases the available dissolved oxygen which negatively affects aquatic
64 organisms [14].

65 A specific example of what happen is logging of contaminated water in the soil. In that situation,
66 oxygen become less, available as an electron acceptor, promoting denitrifying bacteria to reduce
67 available nitrate into gaseous nitrogen which enters the atmosphere with resultant negative
68 effects [13]. Also, anaerobic archaea (methanogens) may produce excessive methane at a high
69 rate than aerobic methane oxidizing bacteria (methanotrophs) could cope with, there for

70 contributing to greenhouse effect and global warming [15]. Increasing in methane is of concern
71 because it is five times more effective as a greenhouse gas than carbon dioxide (CO₂).

72 Wrongful discharge of blood and animal faeces into streams may cause oxygen-depletion as well
73 as nutrient over enrichment of the receiving system which could cause increased rate of toxin
74 accumulation [16]. Humans may also be affected through outbreak of water borne diseases and
75 other respiratory and chest diseases [17].

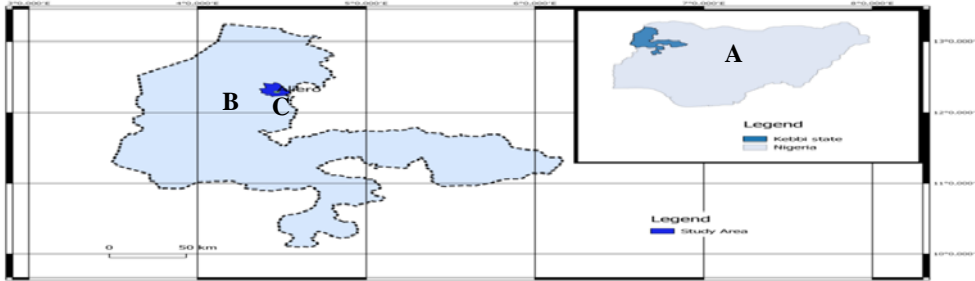
76 **1.3 Antibiotic Susceptibility Testing**

77 Antibiotic susceptibility testing can be used for drug discovery, epidemiology and prediction of
78 therapeutic outcome. After the revolution in the “golden era”, when almost all groups of
79 important antibiotics (Tetracycline, Cephalosporin, Aminoglycosides and Macrolides) were
80 discovered and the main problems of chemotherapy were solved in the 1960s, the history repeats
81 itself nowadays and these exciting compounds are in danger of losing their efficacy because of
82 the increase in microbial resistance [18]. Currently, its impact is considerable with treatment
83 failures associated with multidrug-resistant bacteria and it has become a global concern to public
84 health [19].

85 **2.0 MATERIALS AND METHODS**

86 **2.1 Study Area**

87 The study area was a section along the Tagangu seasonal River at old Kasuwa (Market) area
88 located in Sarkin Fada 1 Ward Aleiro, Kebbi State, Nigeria. Kebbi State was created on 27th
89 August in 1991 from the old Sokoto State. It is located in the North Western part of Nigeria
90 between the latitude 11.6781⁰N and longitude 4.0695⁰ E. According to the 2011 National
91 Population Census (NPC) estimate, the total population of Kebbi State is 3,802,500. Its capital
92 city is Birnin Kebbi.



93
94 **Fig. 1: Map of Nigeria (A) Kebbi State map (B) Aliero map (C)**

95 **2.2 Samples Collection and Preparation**

96 A total of thirty (30) water samples were collected, ten (10) each from the three sections named
97 as downstream, upstream and irrigation site denoted as A, B and C respectively, along the
98 Tagangu seasonal River receiving the abattoir effluent. The water samples were collected as
99 described by Cheesbrough, [19], the water samples were collected at the point's representative of
100 the sampling sites (A, B and C) and transported to the laboratory in an ice jacket box and
101 subsequently processed within 4 hours of sampling.

102 **2.3 Media Preparation**

103 All the media used under this study were prepared as described by [20].

104 **2.4 Bacteriological Analyses**

105 **2.4.1 Isolation and Identification of Bacterial Isolates**

106 The organisms were isolated and identified based on colonial morphology, cultural
107 characteristics and biochemical tests as described by [20, 21, 22].

108
109
110 **2.5 Antibiotic Susceptibility Test Profile of the Isolates**

Comment [TKC5]: 1989 right?

Comment [TKC6]: [20]

Comment [TKC7]: What are the specific media used?

Comment [TKC8]: I was wondering how the identification done for all the isolates just based on morphology, cultural characteristic and biochemical tests?

111 The antibiotic susceptibility testing (Agar disk diffusion method) of the isolated organisms was
112 carried out in accordance with the standard approved by the Clinical and Laboratory Standards
113 Institute (CLSI) [23].

114 **2.6 Statistical Analyses of the Results**

115 ANOVA system of analysis was carried out using SPSS computer application. The results were
116 typed, analyzed and interpreted.

117 **3.0 RESULTS AND DISCUSSION**

118 **3.1 Bacteriological Analyses**

119 **3.1.1 Total heterotrophic bacteria plate count**

120 Table 1 represents the number of the heterotrophic bacterial count (cfu/ml). Sample A had the
121 highest count of $1.64 \pm 1.94 \times 10^7$ cfu/ml, followed by sample C with the count of $1.62 \pm 1.69 \times 10^7$
122 (cfu/ml), while the least count of $1.57 \pm 1.64 \times 10^7$ (cfu/ml) was observed in sample B.

123 The total heterotrophic bacterial plate count recorded was highest in samples A ($1.64 \pm 1.94 \times 10^7$
124 cfu/ml) followed by samples C ($1.62 \pm 1.69 \times 10^7$ cfu/ml), while the lowest number of $1.57 \pm 1.64 \times$
125 10^7 cfu/ml was observed in samples B. This is so because samples A were obtained from
126 upstream, where the incoming substances including microbes do reside before getting to other
127 portions of the river, it's also a point at which abattoir effluent directly find their way into the
128 river body without treatment, and that must contained high level of contamination.

129 Samples B were also collected from downstream where the effluent has to travel far away to get
130 to the site, while samples C were also obtained from a place called irrigation space; where the
131 farmers use the water for growing crops, and therefore was expected to have a fair number of
132 microbial count, but much physicochemical contaminations. This was in agreement with what

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133 UNESCO [24] reported that agricultural run-off is another major water pollutant as it contains
134 nitrogen compound and phosphorus from fertilizers, pesticides, salts, poultry wastes and washes
135 down from abattoirs. Contaminants are usually of varied composition ranging from simple
136 organic substances to complex organic compounds with varying degrees of toxicity.

Comment [TKC10]: Nitrogen and phosphorus compounds

137 **Table 1:** Total Heterotrophic Bacterial (THB) Plate Count

Samples	Total heterotrophic bacterial count (cfu/ml)
A	$1.64 \pm 1.94 \times 10^7$
B	$1.57 \pm 1.64 \times 10^7$
C	$1.62 \pm 1.69 \times 10^7$

138 **Keys:** cfu/100ml= Colony forming unit/100ml.

139 3.1.2 The frequency and percentage occurrence of identified organisms

140 Figure 2 represents the frequency and percentage occurrence of the identified bacteria from the
141 water samples. *Escherichia coli* have the highest percentage occurrence of 56.7% while
142 *Aerobacter aerogenes* has the least of 20%.

143 The frequency and percentage of isolates reported in this study indicates that *Escherichia coli*
144 have the highest occurrence of 17 and a percentage of 56.7% while *Aerobacter aerogenes* have
145 the lowest occurrence of 6 and the percentage of 20%. This was contraindicated with the
146 statement of International Reference Center for Community water supply and sanitation, which
147 stipulated that, the level of coliforms which should be presence in any giving water body should
148 be less than 10/100ml of a sample, and the number of *E. coli* should be less than 2.5/100ml of a
149 sample.

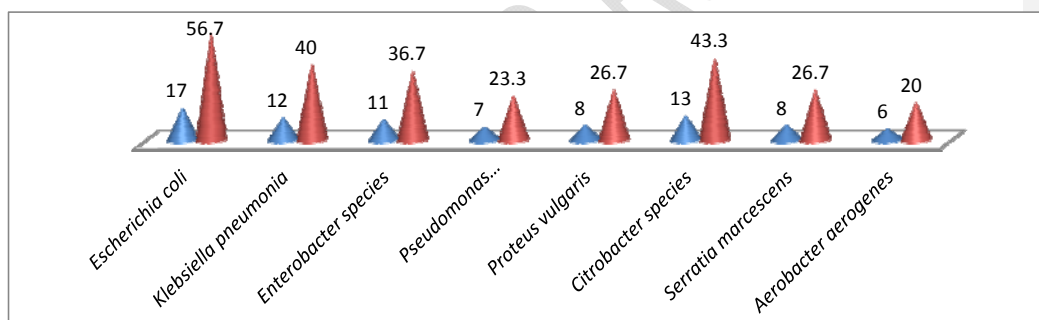
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150 The bacteria isolated from the River Tagangu were enterobacteriaceae. The presence of enteric
151 bacteria like *Serratia marcescens*, *Salmonella species*, *Shigella species*, *Klebshialla species* and

Comment [TKC12]: *Klebsiella*

152 *Escherichia coli* O157:H7 can be attributed to high level of faecal, municipal and abattoir waste
 153 contaminations which may constitute health hazard to the people drinking or using the water for
 154 domestic activities or both. The high incidence of Enterobacteriaceae recorded in this study
 155 could be due to the virulent factors present within these organisms which gives them the ability
 156 to be resistant to antibiotics.

157 The result of this study also agreed perfectly with the similar result carried out by Olayemi and
 158 Oyadege, [25], were as high as 45.3% incidence of Enterobacteriaceae among other organisms
 159 were recorded in Gombe state, Nigeria. Similarly *Escherichia coli* was also incriminated as the
 160 highest organism (36.6%) that was isolated from the gastrointestinal tract of fresh water fish as
 161 reported by Trust [26].



162
 163 **Figure 2:** Frequency and percentage (%) occurrence of identified bacteria

164 3.2 Antibiotic Susceptibility Test Profile

165 Table 2: represents antibiotic susceptibility profile test of each of the identified organisms in
 166 each of the antibiotic disc tested. *Escherichia coli* indicate the highest zone of inhibition of
 167 $18.6 \pm 0.06 \text{ mm}$ with Sparfloxacin, Amoxicillin and Tarivid respectively, and the least of
 168 $16.6 \pm 0.04 \text{ mm}$ with Septrin. *Klebsiella pneumoniae* demonstrates the highest zone of inhibition
 169 of $19.3 \pm 0.07 \text{ mm}$ with Tarivid, and the least of $15 \pm 0.03 \text{ mm}$ with Gentamycin.

Comment [TKC13]: Spacing between 0.06 and mm

Comment [TKC14]: Spacing between 0.04 and mm, please apply it to the rests...

170 *Enterobacter species* points the highest zone of inhibition of 21 ± 0.09 mm with Augmentin, and
171 the least of 12.3 ± 0.01 mm with Gentamycin. *Pseudomonas aeruginosa* indicates the highest zone
172 of inhibition of 18.3 ± 0.06 mm with Ciprofloxacin, and the least of 13.3 ± 0.01 mm with Tarivid.
173 *Proteus vulgaris* counts the highest zone of inhibition of 19.6 ± 0.07 mm with Amoxicillin, and the
174 least of 14.3 ± 0.02 mm with Augmentin. *Citrobacter species* happens to have the highest zone of
175 inhibition of 20.6 ± 0.08 mm with Tarivid, and the least of 15.6 ± 0.03 mm with Amoxicillin.

176 *Serratia marcescens* reveals the highest zone of inhibition of 17 ± 0.05 mm with Ciprofloxacin and
177 Pefloxacin respectively, and the least of 12 ± 0.01 mm with Amoxicillin. *Aerobacter aerogenes*
178 indicates the highest zone of inhibition of 19 ± 0.07 mm with Amoxicillin, and the least of
179 14 ± 0.02 mm with Augmentin and Septrin respectively.

180 The antibiotic susceptibility profile of all the identified bacteria tested, *Enterobacter species*
181 revealed the highest zone of inhibition of 21 ± 0.09 mm with Augmentin, followed by *Citrobacter*
182 *species* with the zone of inhibition of 20.6 ± 0.08 , while the least zone of inhibition of 12 ± 0.01
183 was observed with *Serratia marcescens*. This finding is similar to the some previous
184 investigations in other regions carried out in non-domestic environment [27, 28], the findings
185 stated that *Serratia marcescens*, *Citrobacter* and *Enterobacter species* were investigated to have
186 the highest resistant with most antibiotics in non-domestic environment in Portugal.

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188

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Table 2: Antibiotic susceptibility test profile of the identified organisms from the water samples

Antibiotics	Potency	<i>Escherichia coli</i>	<i>Klebsiella pneumonia</i>	<i>Enterobacter species</i>	<i>Pseudomonas aeruginosa</i>	<i>Proteus vulgaris</i>	<i>Citrobacter species</i>	<i>Serratia marcescens</i>	<i>Aerobacter aerogenes</i>
	Zones of inhibition (mm) measured								
SXT	30µg	18.3±0.06	17.3±0.05	00.0±0.00	15.3±0.03	17±0.05	19.6±0.07	16.6±0.04	16±0.04
CH	30µg	17.3±0.05	17.3±0.05	00.0±0.00	15±0.03	18±0.06	00.0±0.00	16.3±0.04	14.6±0.02
SP	10µg	18.6±0.06	19±0.07	19.3±0.07	17.3±0.05	18±0.06	17.3±0.05	16.6±0.04	16±0.04
CPX	10µg	17.3±0.05	18±0.06	19±0.07	18.3±0.06	18.6±0.06	17.3±0.05	17±0.05	16.3±0.04
AM	30µg	18.6±0.06	18±0.06	16.6±0.04	17±0.05	19.6±0.07	15.6±0.03	12±0.01	19±0.07
AU	30µg	00.0±0.00	16.3±0.04	21±0.09	00.0±0.00	14.3±0.02	00.0±0.00	13.5±0.1	14±0.02
CN	10µg	00.0±0.00	15±0.03	12.3±0.01	00.0±0.00	15±0.03	00.0±0.00	14±0.02	15±0.03
PEF	30µg	17.3±0.05	17.3±0.05	15.6±0.03	16.3±0.04	17.3±0.05	17.6±0.05	17±0.05	15.6±0.03
OFX	10µg	18.6±0.06	19.3±0.07	00.0±0.00	13.3±0.01	19±0.07	20.6±0.08	16.3±0.04	17.3±0.05
S	30µg	16.6±0.04	00.0±0.00	13.3±0.01	00.0±0.00	15.3±0.03	16.3±0.04	14±0.2	14±0.02

Comment [TKC17]: s not separated..for overall result please show some evidence pictures of significant result (zone inhibition)

Keys: SXT= Septrin, CH= Chloramphenicol, SP= Sparfloxacin, CPX= Ciproflaxacin, AM= Amoxicillin, AU= Augmentin, CN=

Gentamycin, PEF= Pefloxacin, OFX= Tarivid, S= Streptomycin.

4.0 CONCLUSION

The high level of enteric pathogens demonstrated in Tagangu seasonal River located at Shiyar Fada 1, Aleiro Local Government, Kebbi State Nigeria, which always receives a tremendous amount of Aleiro abattoir effluent, and their multiple resistance to commonly used antibiotics, further confirmed the dangers associated with discharging municipal waste, organic waste and untreated wastewater to the river, which have a fatal impact on the river and its users. Therefore, it has been concluded that the water from the river is microbiologically unhygienic and unsafe for domestic (washing of clothes, animal products and feeding of animal) and agricultural purposes (growing of crops) without bacteriological treatment.

CONFLICT OF INTEREST

There was no conflict of interest exist

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REFERENCES

- [1] Adelegan, J. A. Environmental policy and slaughterhouse waste in Nigeria. 228th Water, Engineering and Development Conference Report: Calcutta, India. 2002.
- [2] Osibanjo, O. and Adie, G. U. Impact of effluent from Bodija abattoir on the physicochemical parameters of Oshunkaye stream in Ibadan City, Nigeria: *African Journal of Biotechnology*. 2007; **6** (15): 1806-1811.
- [3] Weobong, C. A. A. and Adinyira, E. Y. Operational impacts of the Tamale abattoir on the environment. *Journal of Public Health and Epidemiology*: 2011; **3** (9): 386-393.

- [4] Alam, M. J., Islam, M. R., Muyen Z., Mamun M. and Islam S. Water quality parameters along rivers. *International Journal of Environmental Science & Technology*: 2007; **4** (1): 159-167.
- [5] Olanike, K. A. Unhygienic operation of city abattoir in South Western Nigeria. Environmental implication AJEAM/RAGEE: 2002; **4** (1): 23- 28.
- [6] World Health Organization. Guidelines for Drinking Water Quality Recommendation, World Health Organization, Geneva: 2008; 1: 130-135.
- [7] Adesemoye, A. O., Opere, B. O. and Makinde, S. C. O. Microbial content of abattoir wastewater and its contaminated soil in Lagos, Nigeria: *African Journal of Biotechnology*: 2006; **5** (20): 1963-1968.
- [8] Yahaya, M. I., Mohammed, S. and Abdullahi B. K. Seasonal Variations of Heavy Metals Concentration in abattoir Dumping Site Soil in Nigeria: *Journal of Applied Sciences and Environmental Management*: 2009; **13** (4): 9-13.
- [9] Hinton, M. H., Mead, G. C. and Rowlings, C. Microbiology control in the meat industry. Flair flow Europe technical manual: 2002; **2** (339): 4-12.
- [10] Amisu, K. O., Coker, A. O., Onanmi, S. L. W. and Isokpehi, R. D. Arcobacter butzleri strains from poultry abattoir effluent in Nigeria. *East African medical journal*: 2003; **80** (4): 218-222.
- [11] Quinn, J. M. and McFarlane, P. N. Effects of slaughterhouse and dairy factory wastewaters on epilithon: A comparison in laboratory streams. *Water Research*: 1989; **23** (10): 1267-1273.
- [12] Ejaz, N. Investigation of the Characteristics of Effluent Mixing in Streams. A Doctoral dissertation, University of Engineering and Technology Taxila-Pakistan: 2009.

- [13] Kanu, I., and Achi, O. K. Industrial effluents and their impact on water quality of receiving rivers in Nigeria. *Journal of applied technology in environmental sanitation*: 2011; 1 (1): 75-86.
- [14] USEPA. Technical development document for the final effluent limitations guidelines and standards for the meat and poultry products point source category: EPA 821-R- 04- 011, Washington, DC. Available at <http://www.epa.gov/ogwdw000/ccl/pdfs/regdetermine1/supportcclmagnesedwreort.pdf> 2004.
- [15] Semrau J., D. Current knowledge of microbial community structures in landfills and its cover soils. *Applied microbiology and biotechnology*: 2011; 89 (4): 961-9.
- [16] Nwachukwu, M. I., Akinde, S. B., Udujih, O. S. and Nwachukwu, I. O. Effect of abattoir wastes on the population of proteolytic and lipolytic bacteria in a recipient water body (Otamiri River): *Global Research Journal of Science*: 2011; (1): 40-42.
- [17] Mohammed, S. and Musa, J. J. Impact of Abattoir Effluent on River Landzu, Bida, Nigeria. *Journal of Chemical, Biological and Physical Sciences (JCBPS)*: 2011; 2 (1): 132-139
- [18] Mayers, D. L., Sobel, J. D., Ouellette, M., Kaye, K. S. and Marchaim, D. Antimicrobial drug resistance: clinical and epidemiological aspects, Springer: 2017; 2.
- [19] Guschin, A., Ryzhikh, P., Rummyantseva, T., Gomberg, M. and Unemo, M. Treatment efficacy, treatment failures and selection of macrolide resistance in patients with high load of *Mycoplasma genitalium* during treatment of male urethritis with josamycin. *BMC infectious diseases*: 2015; 15 (1): 40-53.
- [20] Cheesbrough, M. Water and sanitation decade. *Medical Laboratory Manual for Tropical Countries*: Butter Worth and co-Publisher: 1989; 206-219.
- [21] Dubey, R.C. and Maheshwari, D.K. Microbial ecology. A text book of microbiology; Multicolor Illustrative Edition: S. Chand And Company Ltd. 2006; 785-797.

- [22] Manga, B. S. and Oyeleke, S. B. Biochemical tests. *Essentials of laboratory practicals in microbiology*: Tobest publisher, Minna, Niger state: 2008; 20-67.
- [23] Clinical and Laboratory Standard Institute. Performance Standards for Antimicrobial Disk Susceptibility Tests, Approved Standard, 7th ed., CLSI document M02-A11: CLSI, 950 West Valley Road, Suite 2500, Wayne, Pennsylvania 19087, USA. 2012.
- [24] UNESCO. Water a Shared Responsibility .The United Nations World Water Development Report. Retrieved from <[http:// www.unesco.org/water/wwap](http://www.unesco.org/water/wwap)>: 2006.
- [25] Olayemi, A. B. and Oyagede, J. S. O. Incidence of antibiotic resistance among Escherichia coli isolated from clinical and river water Nigeria. *Med. Journal*: 1997; **17** (4): 207-209.
- [26] Trust, T. J. and Sparrow, R. A. H. The bacterial flora in the alimentary tract of freshwater salmonid fishes: *Canadian Journal of Microbiology*: 1974; **20** (9): 1219- 1228.
- [27] Fluckey, W. M., Loneragan, G. H., Warner, R. and Brashears, M. M. Antimicrobial drug resistance of Salmonella and Escherichia coli isolates from cattle feces, hides, and carcasses. *Journal of food protection*: 2007; **70** (3): 551-556.
- [28] Miranda, J. M., Guarddon, M., Vázquez, B. I., Fente, C. A., Barros-Velazquez, J., Cepeda, A. and Franco, C. M. Antimicrobial resistance in enterobacteriaceae strains isolated from organic chicken, conventional chicken and conventional turkey meat: A comparative survey. *Food Control*: 2008; **19** (4): 412-416.