

*Original Research Article*

*Evaluation of Microbial Contamination of Combs and Brushes in Beauty salons within the University of Port Harcourt, Rivers State, Nigeria*

**ABSTRACT**

Beauty salons may provide a suitable medium for the growth and transfer of pathogenic microorganisms which may be of public health significance. This study was aimed at investigating the microbial contamination of beauty salon tools within the University of Port Harcourt, Rivers State, Nigeria. Nutrient agar was used for the determination of total culturable heterotrophic bacterial counts and Potato dextrose agar was used for the determination of total spore counts. Bacterial isolates were subjected to different biochemical tests while the fungal cultures were identified by macroscopy and microscopy. Results revealed bacterial load obtained from combs and brushes across the three campuses studied ranged from  $6.3 \times 10^5$  to  $2.8 \times 10^6$  CFU/swab area and  $5.8 \times 10^5$  to  $1.8 \times 10^6$  CFU/swab area respectively. Total spore counts obtained from combs and brushes across the three campuses ranged from  $1.8 \times 10^5$  to  $1.0 \times 10^6$  CFU/swab area and  $4.2 \times 10^5$  to  $9.3 \times 10^5$  CFU/swab area respectively. The bacterial isolates obtained from the salon tools include *Staphylococcus aureus*, *Bacillus* spp., *Micrococcus* spp., *Serratia* spp., *Citrobacter* spp., *Proteus* spp. and *Shigella* spp., while the fungal isolates include *Aspergillus flavus*, *Penicillium* spp., *Trichophyton* spp. and *Microsporium* spp. *Staphylococcus aureus* (27.7%) and *Bacillus* spp. (22.2%) were the predominant bacterial isolates in the study while *Aspergillus flavus* (36.3%) and *Penicillium* spp. (27.3%) were the most occurring fungi. The study showed that fomites used in beauty salons harbour significantly high microbial load including microorganisms of possible public health significance.

Key words: Beauty salons; pathogenic microorganisms

**1.0 INTRODUCTION**

Besides the day to day interactions of people which constitute one way of spreading disease, the major source and spread of infections in communities are fomites [1]. The environment plays an important role in transmission of microbial agents to humans, with many environmental materials serving as vehicles [2]. Tools used in Beauty salons can become contaminated with pathogenic microorganisms and can be potential reservoirs of such pathogenic microorganisms. Any service with the potential to break the skin's surface can be associated with infections, and these infections can be transmitted to and between clients if proper infection control procedures are not implemented.

Beauty salon services may pose potential health concerns to their clients, including risk of infection and injury. These health risks will vary depending on the nature of the service, the tools used, the health status of the clients and service providers, as well as the infection control procedures implemented. While invasive procedures, such as piercing and tattooing, are clearly associated with bacterial and viral infection risks, even non-invasive procedures, such as hair dressing, pedicure and manicure can result in infections [3].

Beauty salons play an important role in possible transfer of skin and eye infections due to the use and reuse of beauty salon tools and equipment [4]. Items such as razors, scissors, combs, clippers and hairpins can accidentally pierce the skin. Nail and cuticle clippers, nail files, and callus removers used in beauty salons have also been implicated in disease establishment among beauty salon users [5].

Beauty salons are considered high-exposure environments for transmission of human pathogens [6]. Ruddy *et al.* [7] reported a case of Methicillin-resistant *Staphylococcus aureus* (MRSA) infection in patient previously tested negative for MRSA, after a visit to the hospital hairdresser. Improperly sanitized cuticle cutters had been attributed to cause varying serious complications,

ranging from an inflamed cuticle to hepatitis [5]. Pelenita [8] stated that dirty instruments also contribute to infection by blood borne diseases such as HIV or hepatitis. Other infections that can be spread in hairdressing premises include skin infections on the scalp, face and neck such as impetigo and fungal infections such as *Tinea capitis* and *Tinea barbae* [9-11].

Commonly isolated bacteria from hair dressing and beauty salons include *Staphylococcus aureus*, *Staphylococcus epidermis*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Bacillus* spp. *Micrococcus* spp. *Enterococcus* spp. and *Enterobacter* spp. while fungal isolates include *Aspergillus flavus*, *Aspergillus fumigates*, *Alternaria* spp., *Cladosporium* spp., *Geotrichum candidum*, *Rhizopus nigricans*., *Cladosporium*, *Trichophyton* spp., *Mucor* spp., *Rhizopus arrhizus*, *Candida albicans*, *Penicillium* spp. [4,5,12,13].

Beauty salons around the University environment have been observed to receive a lot of patronage, and a better percentage of this population is made up of students who make direct or indirect contact with each other. The study therefore sets out to determine the microbial population and diversity in selected beauty salon tools across the different campuses of the University of Port Harcourt.

## **2.0 MATERIALS AND METHODS**

### **2.1 Collection of Samples**

Twelve composite samples were collected from the combs and brushes in the selected public salons within the three university campuses (Choba, Abuja and Delta Park), within the study period (May-June, 2018) using sterile swab sticks. The sterile swab sticks were moistened in normal saline first before they were used to swab the combs and brushes. They were replaced into

the container, labeled appropriately and were immediately transported to the laboratory for microbiological analysis.

## **2.2 Isolation and enumeration of bacterial and fungal isolates**

This was done to estimate the number of organisms in different samples. Swab samples were diluted in 10 ml sterile normal saline to make a stock solution and shaken mechanically for 10 minutes. Exactly 1ml from the sample stock solution was pipette aseptically into a test tube containing 9ml of normal saline to make  $10^{-1}$  dilution and from this dilution,  $10^{-2}$ ,  $10^{-3}$ ,  $10^{-4}$ , and  $10^{-5}$  dilutions were made.

Nutrient agar and potato dextrose agar were prepared for plating out the diluted samples. Duplicate plates were set for the plating of the dilution of the different samples, 0.1ml of  $10^{-3}$  dilution was collected and dropped on the surface of the agar using a sterile pipette and spreading was done using a sterilized hockey stick. Bacterial plates were incubator at  $37^{\circ}\text{C}$  for 24 hours while fungal plates were incubated at  $27^{\circ}\text{C}$  for 48-72 hours. The number of colonies that developed from each plate ranging between 30 and 300 after incubation was counted and recorded.

The bacterial isolates were identified based on their cultural and biochemical characteristics with reference to Holt *et al.* [14]. Morphological characteristics such as shape, colour, arrangement of spores, structure of the mycelium, and structure of hyphae and arrangement of sporangiophores were used in identifying the fungal isolates as described in Ellis *et al.* [15].

## **2.3 Ethical Consideration**

Ethical approval was sought from the department in the University of Port Harcourt. A letter of consent was presented to the salon owners before the commencement of the study.

## RESULTS

The results of bacterial load obtained from combs and brushes from salons across the three campuses was shown in Table 1, which ranged from  $1.8 \times 10^6$  to  $3.7 \times 10^6$  CFU/swab area and  $1.6 \times 10^6$  to  $3.4 \times 10^6$  CFU/swab area respectively. In Table 2, the total spore counts obtained from combs and brushes from salons across the three campuses were recorded and it ranged from  $6.9 \times 10^5$  to  $1.9 \times 10^6$  CFU/swab area and  $1.2 \times 10^6$  to  $1.6 \times 10^6$  CFU/swab area respectively.

The bacterial isolates obtained from combs and brushes are *Staphylococcus aureus*, *Bacillus* spp., *Micrococcus* spp., *Serratia* spp., *Citrobacter* spp., *Proteus* spp. and *Shigella* spp. while the fungal isolates include *Aspergillus flavus*, *Penicillium* spp., *Tricophyton* spp. and *Microsporium* spp. (Table 3).

The percentage occurrence of bacterial isolates was shown in Figure 1. The organisms and their level of occurrence include *Staphylococcus aureus* (27.7%),

*Bacillus* spp. (22.2%), *Micrococcus* spp. (11.1%), *Serratia* spp (16.7%), *Citrobacter* spp. (5.6%) *Proteus* spp. (11.1%) and *Shigella* spp. (5.6%).

The percentage occurrence of fungal isolates is shown in Figure 2. The organisms and their level of occurrence include *Aspergillus flavus* (36.3%), *Penicillium* spp. (27.3%), *Tricophyton* spp. (18.2%) and *Microsporium* spp. (18.2%).

**Table 1: Total bacterial counts obtained from Salon tools**

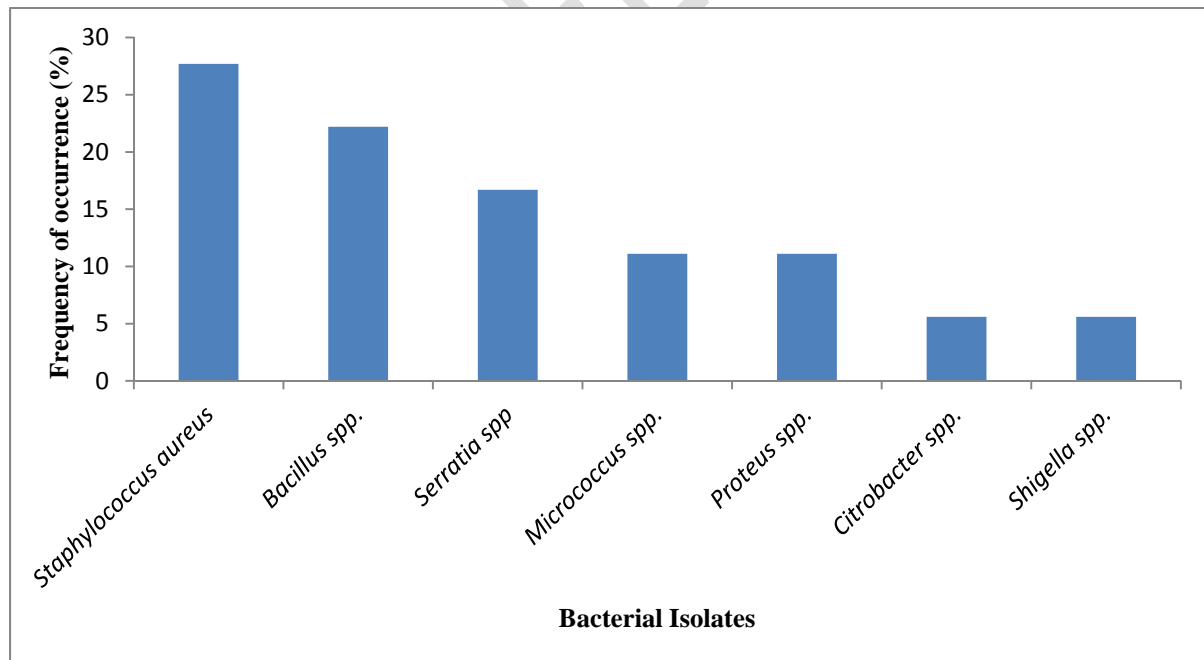
Salon location	Bacterial counts (CFU/swab area)	
	Comb	Brush
Abuja campus	$2.5 \times 10^6$	$3.2 \times 10^6$
Delta campus	$1.8 \times 10^6$	$3.4 \times 10^6$
Choba campus	$3.7 \times 10^6$	$1.6 \times 10^6$

**Table 2: Total fungal counts obtained from Salon tools**

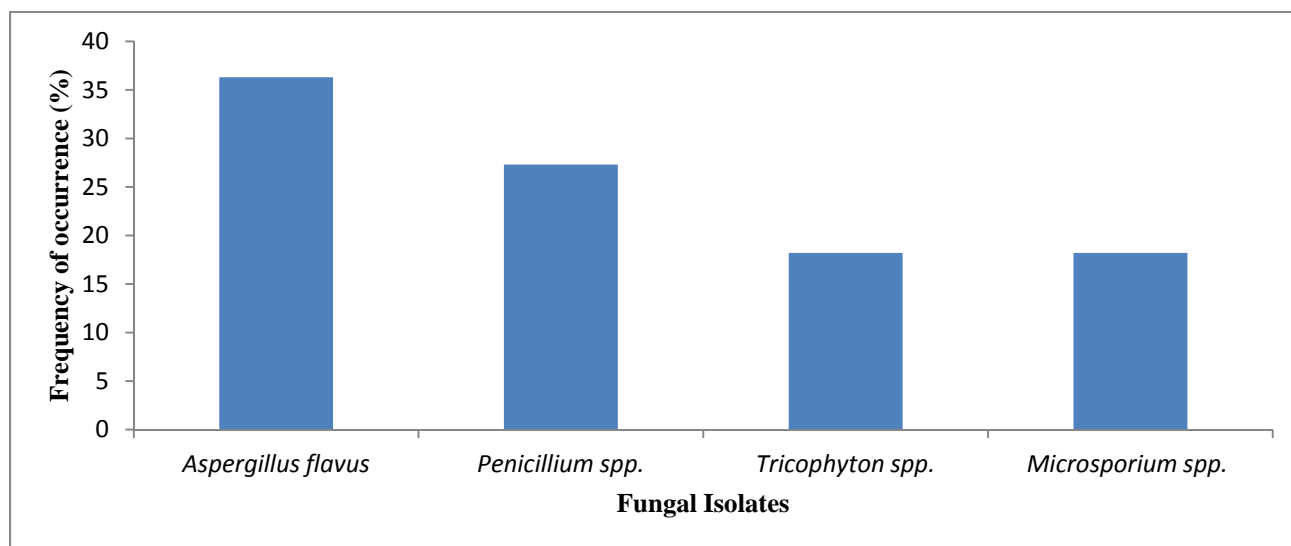
Salon location	Spore counts (CFU/swab area)	
	Comb	Brush
Abuja campus	$1.9 \times 10^6$	$1.3 \times 10^6$
Delta campus	$9.9 \times 10^5$	$1.6 \times 10^6$
Choba campus	$6.9 \times 10^5$	$1.2 \times 10^6$

**Table 3: Microbial isolates obtained from combs and brushes**

Microbial isolate	Salon tool	
	Comb	Brush
<b>Bacteria</b>	<i>Staphylococcus aureus</i> , <i>Bacillus</i> spp., <i>Micrococcus</i> spp., <i>Serratia</i> spp., <i>Citrobacter</i> spp., <i>Proteus</i> spp. and <i>Shigella</i> spp	<i>Staphylococcus aureus</i> , <i>Bacillus</i> spp., <i>Micrococcus</i> spp. and <i>Citrobacter</i> spp. and <i>Proteus</i> spp.
<b>Fungi</b>	<i>Aspergillus flavus</i> , <i>Penicillium</i> spp., <i>Trichophyton</i> spp. and <i>Microsporium</i> spp.	<i>Aspergillus flavus</i> , <i>Penicillium</i> spp., <i>Trichophyton</i> spp. and <i>Microsporium</i> spp.



**Figure 1: Percentage occurrence of bacterial isolates**



**Figure 2: Percentage occurrence of fungal isolates**

#### 4.1 Discussion

The microbial population and diversity of combs and brushes used in public salons were determined. Results revealed that bacterial load obtained from combs and brushes from salons across the three campuses studied to range from  $1.8 \times 10^6$  to  $3.7 \times 10^6$  CFU/swab area and  $1.6 \times 10^6$  to  $3.4 \times 10^6$  CFU/swab area respectively. This is similar to the study carried out by Mbajiuka *et al.* [16] who reported total bacterial counts obtained from brushes, combs and dryer in beauty salons to be between  $1.4 \times 10^6$  to  $1.6 \times 10^6$  cfu/swab area. The total spore count ranged from  $6.9 \times 10^5$  to  $1.9 \times 10^6$  CFU/swab area and  $1.2 \times 10^6$  to  $1.6 \times 10^6$  CFU/swab area.



The high microbial load obtained from the beauty salon tools can be attributed to the public services these tools are subjected to. A survey carried out in the course of this study revealed that no form of cleaning or sterilization was carried out for these tools and this will definitely lead to a build-up of microorganisms, hence putting customers of these salons at risk of infections should these organisms be pathogenic. Inevitably, salon workers handle these tools with their hands and this can contribute to the spread of infections if these hands are not thoroughly washed. Many studies have demonstrated the beneficial impact of hand washing and/or use of alcohol-based hand rubs for reducing pathogenic bacteria on hands and/or reducing infection rates in various institutional settings [17,18]. Since hands are important for intrapersonal and interpersonal transfer of microorganisms, as well as environmental transfer, the dynamics of hand microbial communities and factors impacting them are of considerable importance [19].

The bacterial isolates obtained from the salon tools in this study were *Staphylococcus aureus*, *Bacillus* spp., *Micrococcus* spp., *Serratia* spp., *Citrobacter* spp., *Proteus* spp. and *Shigella* spp., while the fungal isolates were *Aspergillus flavus*, *Penicillium* spp., *Trichophyton* spp. and *Microsporium* spp. In a similar study by Hassan *et al.*, [20], bacterial isolates including *Micrococcus*, *Bacillus* and *Staphylococcus* were obtained from salon tools, while the fungal isolates including *Cladosporium*, *Trichophyton*, *Mucor*, *Candida* and *Penicillium* were obtained.

*Staphylococcus aureus* (27.7%) and *Bacillus* spp. (22.2%) were the predominant isolates in the study. In the studies Enemuor *et al.* [4] and Hassan *et al.* [20] *Staphylococcus* spp. was also identified as the predominant isolate from all salons investigated. Similarly, Naz *et al.* [21] also reported the presence of *Staphylococcus* in unpreserved beauty salon tools after use. *Staphylococcus* spp. are able to cause various diseases in humans such as skin abscess, impetigo

contagiosa, scalded-skin syndrome, and it is the most commonly identified agent that is responsible for skin and soft tissue infection [4,22]

In the study carried out by Mbajiuka *et al.* [16] *Aspergillus Spp*, *Mucor Spp* and *Rhizopus Spp* were isolated from beauty salon tools. Infections that can be spread in salon premises include skin infections on the scalp, face and neck such as impetigo and fungal infections such as ring worm or dermatophytosis [16,23]. Contamination of hairdressing salons is used as an indicator of the burden of *Tinea capitis* in society, particularly where the fungi are prevalent and occur in epidemics [24]. Salons are exposed to many irritants and allergens that may cause occupational diseases. It has been estimated that 10–20% of beauty salon customers are affected by skin disorders [25].

It is also noteworthy that these **microorganisms** can be transferred from the salon tools to the hands and from one surface to another. Several factors have been identified to affect the transfer rate of bacteria from surface to another surface. These include bacteria type, source, and type of surface and moisture level [26]. Since hands are important for intrapersonal and interpersonal transfer of microorganisms, as well as environmental transfer, the dynamics of hand microbial communities and factors impacting them are of considerable importance [19]. This makes hand washing an important part of infection control from fomites such as salon tools. Several hygienic measures were reported to prevent cross-contamination from surface to another surface. Hand hygiene is one of the imperative tools to reduce and prevent surface-to-surface cross-contamination [27]. **It is advisable that salons use single use products such as razor blades, disposable gloves, paper toweling where possible and all equipment must either be discarded or cleaned in hot water and detergent and allowed to dry before re-used on another client.**

## Conclusion

The study showed that fomites (combs and brushes) used in beauty salons harbor significantly high microbial load, which have the potential of causing epidemic if the organisms are pathogenic. The beauty salon tools can serve as reservoirs and carriers of microorganisms which are transmissible from person to person. High level of hygiene practice should be adopted in all salons within the university campuses to prevent the spread of infections via salons.

## REFERENCES

1. Kramer A, Schwebke I, Kampf G. How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. *BioMed Central (BMC) Infectious Diseases*. 2006; 6:130-8.
2. World Health Organization (WHO). The Determinants of Health. Geneva. (Accessed 12 May 2018).
3. Stout JE, Gadkowski LB, Rath S, Alspaugh JA, Miller MB, Cox GM. Pedicure-associated rapidly growing mycobacterial infection: an endemic disease. *Clinical Infectious Diseases*. 2011; 53(8):787-92.
4. Enemuor SC, Ojih MI, Isah S, Oguntibeju OO. Evaluation of bacterial and fungal contamination in hairdressing and beauty salons. *African Journal of Microbiology Research*. 2013; 7(14): 1222-1225.
5. Onajobi IB, Okerentugba PO, Adeyemi SA, Laba SA. Microbial Evaluation of Manicure and Pedicure Shops along Adewole Estate, Ilorin Kwara, Nigeria. *Nigerian Journal of Microbiology*. 2015; 28: 2939-2945.
6. Bures S, Fishbain JT, Uyehara CF, Parker JM, Berg BW. Computer keyboards and faucet handles as reservoirs of nosocomial Pathogens in the Intensive Care Unit. *American Journal of Infection Control*. 2000; 28(6): 465- 471.
7. Ruddy M, Cummins M, Drabu Y. Hospital hairdresser as a potential source of cross-infection with Methicillin Resistant *Staphylococcus aureus* (MRSA). *Journal of Hospital Infection*. 2001; 49(3):225-227.
8. Pelenita TM. Acrylic Nail and Native Bacteria. *Saint Martin's University Biology Journal*. 2006; 1:67-92.
9. Brown NJ. Guideline for public health standards of practice for hairdressing 2nd ed. Australia. 2006; pp. 1-4.

10. Amodio E, Benedetto MA, Gennaro L, Maida CM, Romano N. Knowledge, attitudes and risk of HIV, HBV and HCV infections in hairdressings of Palermo City (South Italy). *European Journal of Public Health*. 2010; 20:433-437.
11. Barn P, Chen T. Infections associated with personal service establishments: aesthetics. National Collaborating Centre for Environment Health ISBN: 978-1-926933-29-0. 2011; pp. 1-10.
12. Janmohammadi F, Ghodous F, Daem R, Kayvan F. Evaluation of Bacterial and fungal Contaminations in Barbershops in Kamyaran city, Iran-Summer 2015. *International Journal of Medical Research and Health Sciences*. 2016; 5 (9):368-371.
13. Sekula SA, Havel A, Otilar LJ. Nail salons can be risky business. *Archives of Dermatological Research*. 2002; 138(3):414-415.
14. Holt JG, Krieg NR, Sneath PHA. (Ed.). *Bergey's Manual of Determinative Bacteriology* (9<sup>th</sup> Ed.). Lippincott Williams & Wilkins. 1994.
15. Ellis D, Davis S, Alexiou H, Handke R, Bartley R. Descriptions of Medical Fungi. Mycology Unit Women's and Children's Hospital School of Molecular and Biomedical Science University of Adelaide. 2007; pp 1-204.
16. Mbajiuka CS, Obeagu EI, Ochei KC, Iheke SO. Evaluation of Microbial Contamination of Tools Used in Hair Dressing Salons in Michael Okpara University of Agriculture, Umudike, Abia State. International Organization of Scientific Research (IOSR). *Journal of Dental and Medical Sciences*. 2014; 13 (17): 22-27.
17. Hammond B, Ali Y, Fendler E, Dolan M, Donovan S. effect of hand sanitizer use on elementary school absenteeism. *Am J Infect Control*. 2000; 28(5):340-6.
18. White C, Kolble R. and Carlson, R. The effect of hand hygiene on illness rate among students in university residence halls. *American Journal of Infection Control*. 2003; 31(6); 364-370.
19. Lax S, Smith P. Longitudinal analysis of microbial interactions between humans and the indoor environment. *Science Magazine*. 2014; 345, 1048.
20. Hassan SM, Hamad AK, Shallal AF, Abdullah SM. Isolation of Pathogenic Microbes from Beauty Salons in Ranya, Iraq. *Galen Medical Journal*. 2018; 29: 104-106.
21. Naz S, Iqtedar M, Ul Ain Q, Aftab K. Incidence of Human Skin Pathogens from Cosmetic Tools used in Beauty Salons in Different Areas of Lahore, Pakistan, *Journal of Scientific Research*. 2012; 4:523.

22. Helaskoski E, Suojalehto H, Virtanen H, Airaksinen L, Kuuliala O, Aalto-Korte K, Pesonen M. Occupational asthma, rhinitis, and contact urticaria caused by oxidative hair dyes in hairdressers. *Annals of Allergy, Asthma and Immunology*. 2014; 11:2462-52.
23. Moore JE, Miller BC. Skin, hair, and other infections associated with visits to barber's shops and hairdressing salons. *American Journal of Infection Control*. 2007; 35:203-204.
24. Coulibaly O, Thera MA, Piarroux R, Doumbo OK, Ranque S. High dermatophyte contamination levels in hairdressing salons of a West African suburban community. *Mycoses*. 2015; 58: 65–68.
25. Dadashi L, Dehghanzadeh R. Investigating incidence of bacterial and fungal contamination in shared cosmetic kits available in the women beauty salons. *Health promotion perspectives*. 2016; 6:159.
26. Rusin P, Maxwell S, Gerba C. Comparative surface-to-hand and fingertip-to-mouth transfer efficiency of gram-positive bacteria, gram-negative bacteria, and phage. *Journal of Applied Microbiology*. 2002; 93(4): 585-592.
27. Kendall D, Viator C, Karns S, Durocher B. Modeling the effects of food handling practices on the incidence of foodborne illness. Washington, DC, USA. 2003; pp. 1-118.