

# An Evaluation of the Chemical Compositions and Antifungal Activity of *Ocimum gratissimum* (Nchuanwu) leaves against some Plant Pathogens

## Abstract

**Aim:** This work was carried out to determine the chemical compositions of *Ocimum gratissimum* leaves using GC-MS and its antifungal potential against some plant pathogenic fungi.

**Study Design:** The study was designed to determine its chemical compositions by GC-MS and to test the inhibitory ability of the plant extract on plant pathogens.

**Place and Duration of Study:** Department of Chemistry, Alvan Ikoku Federal College of Education, Owerri and Department of Plant Science and Biotechnology, Imo State University, Owerri, Nigeria, between February to July 2017.

**Methodology:** The leaf ethanol extract of *Ocimum gratissimum* was evaluated using GC-MS to determine the chemical compositions of the plant. The identification of compounds was done by comparing spectrum of the unknown component with the spectrum of the known components stored in the NIST library. The essential oil of the plant was used to analyze the antifungal potential of the plant. This was done against some plant pathogenic fungi using disc diffusion method and MIC using broth micro dilution method.

### Results:

The GC-MS analysis revealed eight compounds with n- Hexadecanoic acid constituting the bulk of the oil (37.21 %), followed by Oleic acid (25.38 %) and Octadecanoic acid (16.19 %). Other compounds present in the plant are Glycyl alcohol (2.47 %), Methyl alpha -D- Glucopyranoside (8.33 %), Tetradecanoic acid (5.77 %), Palmitic amide (2.72 %) and d-Glucose, 2,3- diethyl-4,5- dithioacetyl (1.93 %). *Ocimum gratissimum* exhibited different degrees of antifungal activity against the mycelial growth of *Aspergillus niger*, *Botryodiplodia theobromae*, *Rhizopus stolonifer*, *Penicillium expansum* and *Colletotrichum spp* and *Fusarium oxysporium*. The maximum percentage degree inhibition of *Ocimum gratissimum* oil was observed on *A.niger* at different concentrations while the least inhibition was observed in *Colletotrichum spp* at different concentrations.

Analysis of some of the compounds found in *Ocimum gratissimum* such as Methyl alpha.-d-glucopyranoside, Oleic acid etc, reveals the rich pharmacological potential of this medicinal plant and the inhibitory potential of the plant against fungi justify the use of *Ocimums gratissimum* as a medicine traditionally.

**Key words:** *Ocimum gratissimum*, pharmacological activities, fungal growth

## Introduction

Nigeria is blessed with several medicinal plants and *Ocimum gratissimum* plant is one of the medicinal plants used widely in herbal medicine and as spice in many delicacies. *Ocimum gratissimum* also called nchuanwu or scent leaves hails from Africa and is found throughout Hawaii and other tropical regions, it has many health benefits. It belongs to *Lamiaceae* family

and is widely known as clove basil or African basil, this plant is used by herbalists to treat a variety of diseases, from bacterial infections and diabetes to pain and liver damage [1]. *Ocimum gratissimum* is a herb used in making anti-bacterial medicines. It is a home grown plant and is also commercially cultivated .

The plant is commonly used in folk medicine to treat different diseases such as upper respiratory tract infection, diarrhoea, headache, ophthalmic, skin diseases, pneumonia, cough fever and conjunctivitis [2]., *O. gratissimum* has been reported to be active against several species of bacteria and fungi [3].

Several studies have confirmed the efficacy of essential oil from *Ocimum gratissimum* in treating various diseases. This is largely credited to the plant's high concentrations of a phenylpropene compound called eugenol. Eugenol, an isolate from *O. gratissimum* has been reported to possess insecticidal properties, nematicidal and antihelminthic properties [4]

The antibacterial qualities of *Ocimum gratissimum* are perhaps the most studied and verified. Several studies have been performed that lend credence to herbalist use of this plant for treating diarrhea and other gastrointestinal infections. It was found that the leaf extract provided relief from diarrhea in lab rats and guinea pigs. It was found that the essential oil relaxed the small intestine in lab rats, furthering claims that the plant is beneficial in relieving gastrointestinal ailments.

Studies have shown that essential oil obtained from the leaf of *Ocimum gratissimum* has shown marked antibacterial activity [5]

These range from *Shigella* and *Salmonella* to *Escherichia* and *Proteus* strains. The oil is aromatic, yet deadly, it is used as mosquito repellant [6]; [7]. A polyherbal preparation of a water extract obtained from the leaves of *Ocimum gratissimum* showed analgesic activity [8]. Extracts of the leaves are documented to possess antidiabetic properties [9], anti-hyperlipidemic effect and recently, it was shown to improve hematological variables in experimental diabetes mellitus and it has antioxidant property [10].

In spite of the rich pharmacological potential of *Ocimum gratissimum*, so far the chemical constituents of the plant have not been fully documented, hence this study.

## Materials and methods

**Sample Collection / Preparation of Plants material.** Fresh leaves of *Ocimum gratissimum* were collected from farm in Owerri Municipal council. The plant was identified and authenticated by Prof F.N Mbagwu, Department of Plant science and biotechnology, Imo State University, Owerri, Nigeria. The leaves were washed, allowed to drain, then pounded with mortar and pestle. The pounded leaves were soaked in ethanol for 48 hours and concentrated, 1ml of the extract was subjected to GC/MS analysis.

### Experimental Procedure of Gas Chromatography – Mass Spectrometry (GC-MS).

The GC analysis were carried out in SHIMADZU JAPAN gas chromatography 5890-11 with a fused GC column (OV- 101) coated with polymethyl silicon (0.25mm x 50m) and the conditions were as follows: Temperature programming from 80- 200°C held at 80°C for 1 minute, rate 5°C/min and at 200°C for 20 min. FID temperature 300°C, injection temperature of 250°C and carrier gas nitrogen at a flow of 1ml /min, split ratio 1:75. GC- MS analysis was conducted using GCMS- QP 2010 PLUS SHIMADZU JAPAN with injector temperature of 230°C and carrier gas pressure of 100 Kpa. The column length was 30m with a diameter of 0.25mm and the flow rate of 50ml/min. the elutes were automatically passed into a mass spectrometer with a detector voltage set at 1.5kv and sampling rate of 0.2 sec. The mass spectrometer was also equipped with a computer fed mass spectra data bank. HERMLE Z 233 M-Z centrifuge Germany was used. Reagents and solvents like ethanol, chloroform, diethyl ether, hexane were all analytical grade and were procured from MERCK, GERMANY [11].

**Component Identification:** Oil components were identified by matching the peaks with Computer Wiley MS libraries and confirmed by comparing mass spectra of the peaks with those from literature [11].

### Experimental Procedure of Antifungal Activity

**Isolation of Essential oils.** Fresh leaves of *Ocimum gratissimum* were subjected to hydro distillation using clevenger's apparatus for 8 hours. The distillate was extracted using diethyl ether and dried over anhydrous sodium sulphate. Antifungal activity of the essential oil was

performed using disc diffusion method as described by [12] the oil was added acetone and serial dilution was made to obtain a concentrations 1000, 750, 500, 250µg/ml. respectively.

### Isolation and Culturing of the Pathogenic Fungi.

Following the procedures of [13], the fungi isolates were obtained from dried and sterized rotted yam discs (2x2mm) and cultured on potato dextrose agar (PDA) and incubated at 30°C for 5days. About 3mm of each fungal culture were placed on the centre of sterilized Petri dish containing PDA. Then 10ml of each concentration of *Ocimum gratissimum* oil was placed inside each sterile paper disc (6mm diameter) and then placed on the PDA containing the fungi culture. Synthetic antifungal chemical, mancozeb acted as control. All the Petri dishes in 3 replications were incubated at 30°C for 5days and monitor for growth inhibition.

Percentage inhibition =  $100 \times [(1 - \text{radial growth of treatment (mm)})]$

---

Radial growth of control (mm)

### Determination of minimum inhibitory concentration (MIC)

This is described as the lowest concentration of the oil that reduced the growth of fungus. It was done by broth dilution technique by following the procedure of [14].

The essential oil was added acetone to make 1000µg/ml. Serial dilution was made to obtain concentrations of 125µg/ml, 250µg/ml, 500µg/ml, 750µg/ml, 1000µg/ml. Then 1ml of the essential oil and 10µl spore suspension (80 spores /ml) of each fungus was inoculated in the test tubes in potato dextrose broth medium and incubated for 5days at 30°C. The control tubes contained PDA medium that were separately added 0.3g/ml mancozeb. Each was inoculated with different fungal spore suspensions (80 spores/ml).

The data collected were subjected to statistical analysis using analysis of variance (ANOVA) method according to Duncan multiple range test (DMRT) and treatment means were separated

121 using fishers least significant difference (LSD) at 5% level of propability, using statistical  
122 package for social science (SPSS) software, version 11.5, Chicago. IL. USA.

123

## 124 Results and Discussion

125 **Table1; Percentage inhibitions of fungal pathogens, 5 days after inoculation with *Ocimum***  
126 ***gratissimum* oil and Mancozeb, incubated at 30<sup>0</sup> C and their MIC values.**

Fungal Pathogens	Concs. of <i>Ocimum</i> oil (µg/ml)				Mancozeb 0.3g/100ml	MIC µg/ml
	250	500	750	1000		
<i>Aspergillus niger</i>	60±2.01	84 ± 1.01	98 ± 0.01	100 ± 0.02	100 ± 0.23	34 ± 0.03
<i>B. theobromae</i>	40 ± 0.40	60 ± 0.31	75 ± 0.31	100 ± 0.07	100 ± 0.01	41.20± 0.01
<i>R.stolonifer</i>	37 ± 0.71	54 ± 0.4	68 ± 0.05	100 ± 0.01	95 ± 0.21	55 ± 0.25
<i>Penicillium expansum</i>	38 ± 1.01	50 ± 0.02	60 ± 0.11	98 ± 0.41	100 ± 0.31	37 ± 0.02
<i>Colletotrichum spp.</i>	23 ± 0.01	37±0.51	44 ± 0.41	70 ± 0.21	100 ± 0.04	70 ± 0.01
<i>F.oxysporium</i>	48 ± 0.01	56 ± 1.01	60 ± 0.01	100 ± 0.61	100 ± 0.31	38 ± 0.04

127 N.B: Values in brackets are the standard errors of treatments

The ethanol extracts of *Ocimum gratissimum* leaves contain rich phytochemical constituents which resulted in the identification of eight different compounds by GC/MS analysis. The chromatogram of the GC/MS analysis is given in figure 1. The individual names of compounds identified. Compounds revealed include n- Hexadecanoic acid constituting the bulk of the oil (37.21 %), followed by Oleic acid (25.38 %) and Octadecanoic acid (16.19 %). Other compounds present in the plant are Glycyl alcohol (2.47 %), Methyl alpha –D- Glucopyranoside (8.33 %), Tetradecanoic acid (5.77 %), Palmitic amide (2.72 %) and d-Glucose, 2,3- diethyl-4,5-dithioacetyl (1.93 %).

Oleic acid is used as emollients, small amount of oleic acid is used as an excipient in pharmacy, and consumption of oleate in olive oil has been associated with a decreased risk of breast cancer and reduction of blood pressure [15] in [13].

n-Hexadecanoic acid was also found to be present in *Ocimum gratissimum*. In India, medicated oils rich in n-Hexadecanoic acid are used in the treatment of rheumatism and inflammation [16]. Ethyl alpha.-d-glucopyranoside has antituberculous activity, antioxidant alpha amylase inhibitory activity, Hypolipemic activity, Anticonvulsant [17].

This result differs from the result of the analysis carried out by [1] and [18]. According to them, the Phytochemical screening of the aqueous extract of *Ocimum gratissimum* revealed the presence of many active ingredients, such as flavonoids, triterpenes, alkaloids, citral, saponins, eugenol, linalol, methyl cinnamate, camphor, and thymol.

The results of antifungal activity of *Ocimum gratissimum* is shown in Table 1. Different concentrations of the essential oil from *O.gratissimum* exhibited different degrees of antifungal activity against the mycelial growth of *Aspergillus niger*, *Botryodiplodia theobromae* *Rhizopus stolonifer*, *Fusarium oxysporium*, *Penicillium expansum* and *Colletotrichum spp*. The maximum percentage degree inhibition of *Ocimum gratissimum* oil was observed on *A.niger* at different concentrations while the least inhibition was observed in *Colletotrichum spp* at different concentrations. *A. niger* exhibited least MIC value (34 µg/ml), this is followed by *Fusarium oxysporium* (38 µg/ml) while the highest MIC value was seen in *Colletotrichum spp* (70 µg/ml).



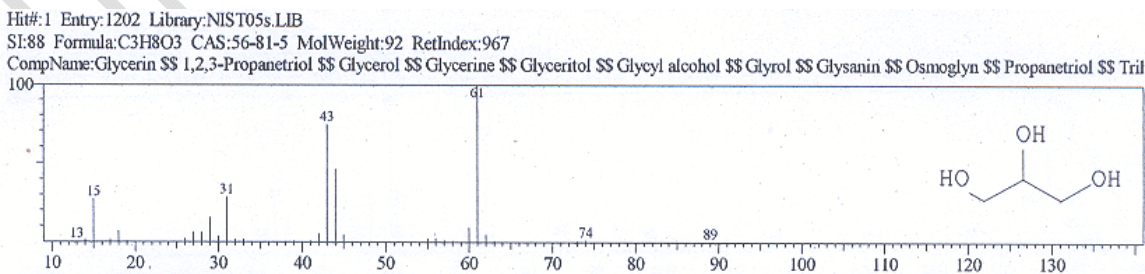
Synthetic antifungal chemical (Mancozeb) compared favourably with *O.gratissimum* oil in inhibiting the mycelial growth of all the fungal plant pathogens.

This result agrees with the report of [18] who reported that *O. gratissimum* is among important plants whose extracts are capable of checking the spread of many fungal diseases of food crops such as *R. stolonifer*, *F. culmorum*, *S. Sclerotiarum* and *P. expanum* associated with the post harvest decay of carrots, in vitro., *Ocimum gratissimum* was reported to inhibit the growth of *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi* and *Salmonella typhimurium*, pathogenic bacteria that cause diarrhea. and the minimum inhibitory concentration (MIC) ranged from 0.1% for *S. aureus* to 0.01% for *E. coli* and *S. typhimurium*, and 0.001% for *S. typhi*. [20].

*O. gratissimum* leaf extract effectively protected maize seeds from seed borne infection of *Fusarium moniliforme* and completely inhibited conidial germination of *Mycosphaerella fijiensis* that cause sigatoka disease of banana [21]. Also [19] reported that crude extracts of *O. gratissimum* effectively exhibited antifungal activity on *Cercospora arachidicola*, the causal organism of leaf spot disease of groundnut.

## Conclusion

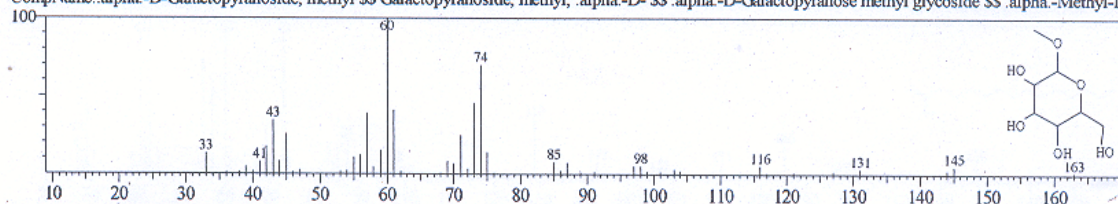
This study revealed that ethanol extract of *Ocimum gratissimum* contains compounds that can be used to treat different diseases. It exhibited different degrees of antifungal activity against some plant pathogenic fungi. Hence the oil might be used as natural antifungal agents replacing synthetic fungicides for the control of some fungal plant pathogens.



Hit#:1 Entry:15252 Library:NIST05s.LIB

SI:95 Formula:C7H14O6 CAS:3396-99-4 MolWeight:194 RetIndex:1714

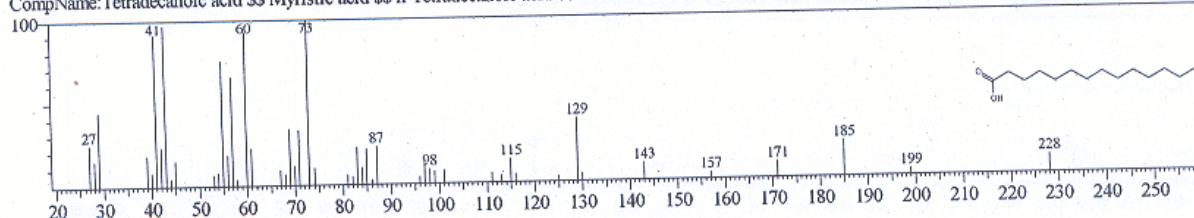
CompName:alpha-D-Galactopyranoside, methyl SS Galactopyranoside, methyl, alpha-D- SS alpha-D-Galactopyranose methyl glycoside SS alpha-Methyl-D



Hit#:1 Entry:19250 Library:NIST05s.LIB

SI:85 Formula:C14H28O2 CAS:544-63-8 MolWeight:228 RetIndex:1769

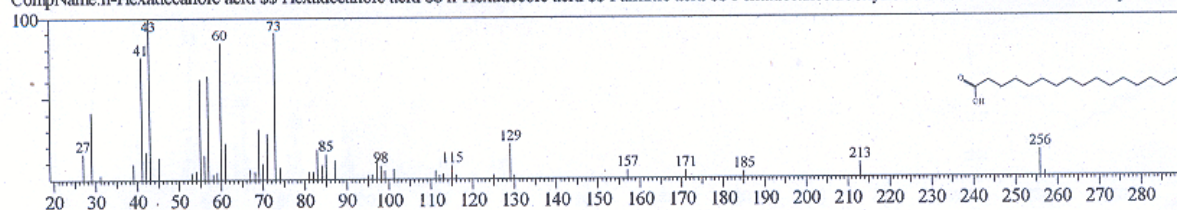
CompName:Tetradecanoic acid SS Myristic acid SS n-Tetradecanoic acid SS n-Tetradecoic acid SS Neo-Fat 14 SS Univol U 316S SS 1-Tridecanecarboxylic acid



Hit#:1 Entry:74999 Library:NIST05s.LIB

SI:95 Formula:C16H32O2 CAS:57-10-3 MolWeight:256 RetIndex:1968

CompName:n-Hexadecanoic acid SS Hexadecanoic acid SS Palmitic acid SS Pentadecanecarboxylic acid SS 1-Pentadecanecarboxylic acid



Hit#:1 Entry:22869 Library:NIST05s.LIB

SI:94 Formula:C18H34O2 CAS:112-80-1 MolWeight:282 RetIndex:2175

CompName:Oleic Acid SS 9-Octadecenoic acid (Z)- SS delta.(Sup9)-cis-Oleic acid SS cis-delta.(Sup9)-Octadecenoic acid SS cis-Oleic Acid SS cis-9-Octadec

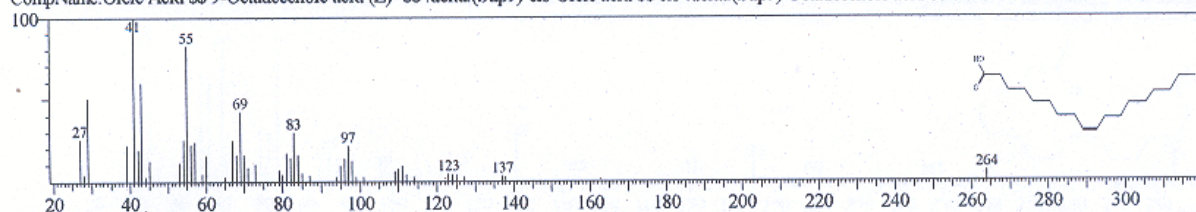


Fig 1: Structures of some of the compounds obtained from GC-MS Analysis of *Ocimum gratissimum*



## 189    **References**

- 190[1] Ofem, O.E, Ani, E.J and Eno, A.E (2012).Effect of aqueous leaves extract of *Ocimum*  
 191        *gratissimum* on hematological parameters in rats. *Int J Appl Basic Med Res.* 2(1): 38–42.
- 192    [2] Onajobi F.D (1986). Smooth muscle contracting lipidic soluble principle in chromatographic  
 193        functions of *Ocimum gratissimum*. *J. Ethnopharmacol.* 18: 3-11.
- 194    [3] Nakaruma CV, Nakaruma TU, Bando E, Melo AFN, Cortez DAG, Diaz Filho BP (1999).  
 195        Antibacterial activity of *Ocimum gratissimum* L.essential Oil. *Mem. Inst. Oswaldo*  
 196        *Cruz.* 94: 675-578.
- 197    [4] Pessoa LM, Morais SM, Bevilaque CM, Luciano JH (2002). Antihelmintic activity of  
 198        essential oil of *Ocimum gratissimum* Linn, and eugenol against *Heamonchus contortus*.  
 199        *Veterinary Parasitolog.* 10:59–63.
- 200    [5] Fern,K.Database,tropical.theferns.info.  
 201        <tropical.theferns.info/viewtropical.php?id=Ocimum+gratissimum>
- 202    [6] Lamiaceae- Silva L.L., Heldwein C.G., Reetz L.G.B., Hörner R., Mallmann C.A.,  
 203        Heinzmann B.M. (2010). Chemical composition, antibacterial activity in vitro and  
 204        brine-shrimp toxicity of the essential oil from inflorescences of *Ocimum gratissimum*  
 205        L. *Brazilian Journal of Pharmacognosy* 20:5 pg 700-705.
- 206    [7] Nweze E.I. and Eze E.E. (2009). Justification for the use of *Ocimum gratissimum* L in herbal  
 207        medicine and its interaction with disc antibiotics *Complementary and Alternative*  
 208        *Medicine* 9 (37) 1472.
- 209    [8] Oboh F.O.J., Madsodje H.I., Enabulele S.A. (2009). Nutritional and antimicrobial properties  
 210        of *Ocimum gratissimum* leaves. *Journal of Biological Sciences* 9:4, 377-380
- 211    [9] Aguiyi J.C, Obi C.I, Gang S.S, Igweh A.C (2000). Hypoglycaemic activity of *Ocimum*  
 212        *gratissimum* in rats. *Fitoterapia*; 71:444-446.
- 213    [10] Egesie U.G, Adelaiye A.B, Ibu J.O, Egesie O.J (2006). Safety and hypoglycaemic  
 214        properties of aqueous leaf extract of *Ocimum gratissimum* in streptozotocin induced  
 215        diabetic rats. *Niger J Physiol Sci* 21:31-5

- 216 [11] Uchegbu, R.I.; Ngozi – Olehi , L.C.; Mbadiugha, C.N.; Ahuchogu, A.A; Ogbuagu, O.E.. ( 217 2015). Phytochemical Evaluation by GC-MS Analysis of the seeds of *Mucuna* 218 *flagellipes* Extract. *Journal of Natural Sciences Research* Vol.5, No.12, 103 – 109.
- 219 [12] Murray P.R, Baron E.J, Pfaller M.A, Tenover, F.C, and Tenover F.C, and Tenover R.H,(1995). Manual of 220 clinical microbiology, 6<sup>th</sup> ed. ASM, Washington.
- 221 [13] Uchegbu, R.I., Akalazu, J.N., Ukpai, K.U. and Iwu, I.C. (2017). Antimicrobial Assessment of 222 *Annona muricata* Fruits and Its Chemical Compositions. *Asian Journal of Medicine and* 223 *Health* 3(1): 1-7.
- 224 [14] Gulluce M, Sokmen M, Salun F. Sokmen A. Adiquzel A and Ozer H (2004); Biol. 225 Activities of the essential oil methanotic extract of micromeria frullicosa (L) Druce 226 ssp. Serpyllifolia (Bleb) Ph Davis plants from the eastern Anatolia region of Turket .J. 227 *Sc Food Agric* . 84, 735-741.
- 228 [15] Teres S., Barcelo- Coblijn, G, Benet, M., Alvarez, R., Bressani, R., Halver, J.E. and Escriba, 229 P.V. (2008). Oleic acid content is responsible for the reduction in blood pressure induced 230 by olive oil. *Proceedings of the National Academy of Science*. 105 (37): 13811 – 13816.
- 231 [16] Aparna, V., Dileep, K.V., Mandal, P.K., Karthe, P., Sadasivan, C. and Haridas, M. (2012). 232 Anti-inflammatory property of n- hexadecanoic acid : structural evidence and kinetic 233 assessment. John Wiley & Sons A/S. Pp 1-2
- 234 [17] Rane Zab, Anish Kumar P, Anusha Bhaskar (2012). Phytochemical evaluation by GC-MS 235 and *in vitro* antioxidant activity of *Punica granatum* fruit rind extract. *Journal of* 236 *Chemical and Pharmaceutical Research*, 4(6):2869-2873.
- 237 [18] Lemos Jde A, Passos XS, Fernande Ode F, Paula JR, Ferri PH, Souza LK, (2005). Antifungal 238 activity from *Ocimum gratissimum* L.towards *Cryptococcus neoformans*. *Mem Inst* 239 *Oswaldo Cruz* 100: 55- 58.
- 240 [19] Okoi, A. I. and Afuo C. O. (2009), Effect of leaf extracts of three plant species on 241 *Cercospora arachidicola* Hori, the causal fungus of leaf spot disease of groundnut 242 (*Arachis hypogea* L) Nig. Jour. *Plt. Protect* 22. (Special edition) 132-139. 243

- [20] Adebolu T. T. and Salau A O, (2005). Antimicrobial activity of leaf extracts of *Ocimum gratissimum* on selected diarrhoea causing bacteria in southwestern Nigeria  
Antimicrobial activity of leaf extracts of *Ocimum gratissimum* on selected diarrhoea causing bacteria in southwestern Nigeria. *African Journal of Biotechnology* Vol. 4 (7), pp. 682-684.
- [21] Okigbo R.N and Emoghene A.O (2004). Antifungal activity of leaf extracts of some plant species on *Mycosphaerella fijiensis* Morelet, the causal organism of black sigatoka disease of Banana (*Musa acuminata*) KMITL Sci. J. 4: 20-31.

#### **Authors' contributions**

This work was carried out in collaboration between all authors.

Author RIU designed the study, performed the chemical composition, wrote the protocol, and wrote the first draft of the manuscript.

Author JNA determined the fungi associated with deterioration and wrote the second draft of the manuscript

Author CES did the analysis

#### **The work is an Original Research Article**