

1 **EFFECT OF SAGO EFFLUENT ON THE GROWTH HORMONE LEVELS IN**
2 ***CLARIAS BATRACHUS* BLOOD SAMPLE**

3

4 **Abstract:** The aim of the study was to determine the effect of Sago effluent on the levels of
5 growth hormone in the blood samples of the fresh water fish *Clarias batrachus*. The fish were
6 exposed to control and different concentrations of treated sago effluents. The concentrations
7 chosen were 25%, 50% and 75% of treated sago effluent. The levels of the growth hormone were
8 increased in the blood sample of the experimental fish *Clarias batrachus*, when compared with
9 that of controls.

10

11 **Keywords:** Growth hormone, Sago effluent, fish, *Clarias batrachus*.

12

13 **Introduction**

14 The aquatic environment is the ultimate sink for all the environment pollutants any chemical
15 pollutant either natural or synthetic is most likely to reach the aquatic environment sooner or
16 later. The toxicity may be either acute or chronic to all forms of biota in aquatic system and also
17 varies to different aquatic organisms. The toxic effects may include both lethal and sublethal
18 concentrations, which may change the growth rate, development, reproduction, histopathology,
19 biochemistry, physiology and behavior [1]. Alterations in the physiological and biochemical
20 parameters of toxicant treated fish have recently emerged as an important tool for the water
21 quality assessment and to know the pathological status of fish in the field of environmental
22 toxicology [2, 3]. The alteration in various physiological and biochemical parameters of an
23 aquatic animal due to exposure of different toxicant has been shown to be directly or indirectly
24 related to the behaviour, immune system, neurotransmission, energy metabolism and
25 reproduction [4, 5]. Accumulation of the environmental pollutants and toxicants has been shown
26 to cause alteration in the activity of many enzymes concerning to cellular energy metabolism [6,
27 7, 8, 9]. Alteration in enzyme activities of the fish is one of the major biomarker indicating the
28 level of changes consequent of pollutants in the tissues, organs and body fluid of the fish that can
29 be recognized and associated with established health impairment process [10]. Moreover,
30 Gabriel and Akinrotimi [11] noted that enzymes can be used to confirm and asses fish exposure
31 to toxicants, providing a link between external and internal structure and degree of responses to
32 toxicant exposure observed between different individuals. However, the applications of enzyme
33 determinations in fish, as an indicator of chemical intoxication seem to be promising. It is most
34 relevant and appropriate in sublethal exposure which spans over many days [12]. Toxicants also
35 can inhibit the activity or synthesis of enzymes[13], resulting in decreased activities in the
36 organs.

37 Growth hormone is a major participant in control of several complex physiologic processes,
38 including growth and metabolism. Growth hormone is also of considerable interest as a drug
39 used in both human and animals. Growth is a very complex process and requires the coordinated
40 action of several hormones. The major role of growth hormones in stimulating body growth is

41 to stimulate the liver and other tissues to secrete IGF – 1. IGF – 1 stimulates proliferation of
42 chondrocytes (cartilage cells), resulting in bone growth. Growth hormone has important effects
43 on protein, lipid and carbohydrate metabolism. Growth hormone is the primary hormone
44 responsible for stimulating tissue repair, cell replacement, brain function and enzyme production
45 [14].

46 Fish are sensitive indicators of pollutants present in water. These pollutants cause various
47 physiological and physical alterations in fishes. In the present work an attempt has been taken to
48 study the alterations in the levels of Growth hormone in the blood sample of the fresh water fish
49 *Clarias batrachus*.

50 **Materials and Methods**

51 The Sago industry effluents were collected from a private Sago industry, situated at Ponnachi
52 near Ammapet of Erode District, Tamil Nadu, India. The effluent from the industry was
53 collected and transported to the laboratory and used for further experiments following standard
54 method. Fingerlings of healthy *Clarias batrachus* were brought to the laboratory and
55 acclimatized for 15 days. The fish were well fed during the acclimatized period. Then fish were
56 exposed to control and 25%, 50%, 75% concentrations of treated sago effluents for period of 28
57 days. Feeding was stopped one day before commencement of the experiment.

58 After the experimental period the fish exposed to sago effluent were sacrificed. Blood samples
59 were collected from the caudal vein by using the hypodermic micro syringes pre-rinsed with
60 heparin. Blood was centrifuged at 3200 rpm for 15 min and plasma was stored at -26°C until it
61 was used for the estimation of plasma cortisol and growth hormone. The growth hormone level
62 was estimated by ELISA method.

63 **Results**

64 The growth hormone level in the muscle of *Clarias batrachus* was increased with increase in the
65 concentrations of treated sago effluent. The control fish were able to record 0.20ng/ml and the
66 fish treated with the effluents recorded 0.32 ng/ml for 25%, 0.35 ng/ml for 50% and 0.46 ng/ml
67 for 75% respectively.

68

69

70

71

72

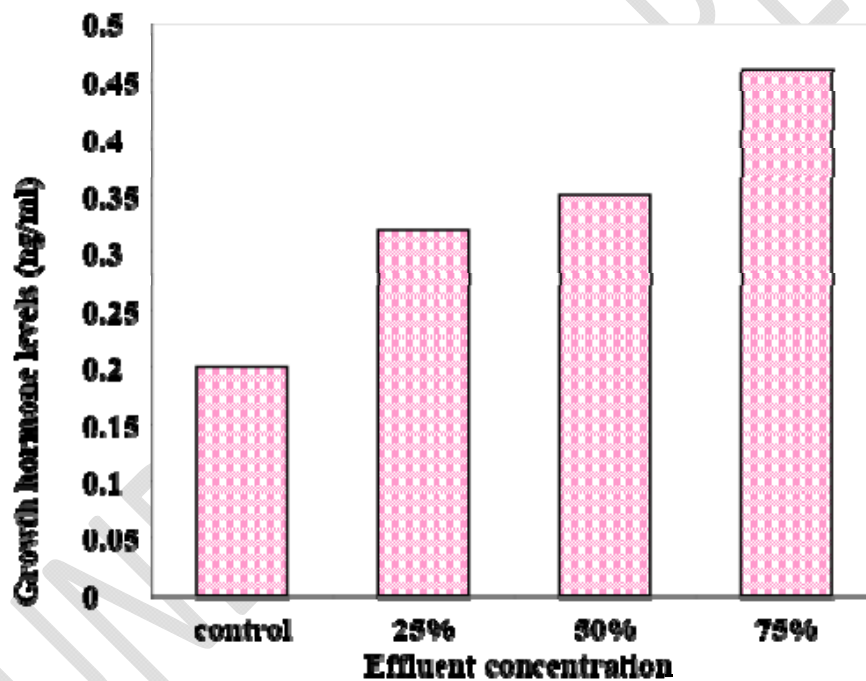
73

74

75 **Table.1. Levels of Growth Hormone in the blood sample of *Clarias batrachus* exposed to**
76 **control and different concentrations of sago effluent.**

Effluent Concentration	Growth Hormone level ng/ml
Control	0.20 ng/ml
25%	0.32 ng/ml
50%	0.35 ng/ml
75%	0.46 ng/ml

Fig.1. Growth Hormone levels in the blood sample of *Clarias batrachus* on exposure to control and different concentrations of treated sago effluent.



82 Growth hormone affects almost all body tissues. Growth Hormone is considered as a master
83 hormone which controls many organ and body function. It also regulates itself. The rejuvenating
84 effects of Growth Hormone are all encompassing, acting on both the mind and body.

85 Growth hormone is the primary hormone responsible for stimulating tissue repair, cell
86 replacement, and brain function and enzyme production. Growth hormone is the ultimate anti-
87 aging therapy and affects almost every cell in the body, rejuvenating the skin and bones,
88 regenerating the heart, liver, lungs and kidneys, bringing back organ and tissue function to more
89 youthful level.

90 Growth hormone (GH) has multiple targets and diverse effects in vertebrates. It is a principal
91 promoter of growth, and also influences the metabolism. During the past years, it has become
92 clear that GH alters the behaviour of fish as it increases appetite, swimming activity, aggression,
93 and reduces anti-predator behavior [15].

94 Lescroart [16] have reported that the several neurotransmitters and intraperitoneal injections
95 induce the secretion of growth hormone and increase in plasma Growth hormone levels in the
96 African Cat fish (*Clarias gariepinus*) by sensitive radio immuno assay.

97 Peterson et al. [17] have studied the effect of recombinant bovine growth hormone (rbGH) on
98 growth rate, feed efficiency, body composition and insulin-like growth factor binding proteins
99 (IGFBPs) in Norris.

100 The scientists have discovered few synthetic growth hormones like methyl testosterone and ethyl
101 estradiol, which evidences that the synthetic growth hormone promotes weight in several fishes.

102 The fish *Betta splendens* were given 17 α methyl testosterone at different dietary levels under
103 laboratory conditions for 15 days. The maximum growth was found in methyl testosterone
104 treated fish than the control fish [18]. Higher dose of methyl testosterone induced growth in
105 different fish species was reported by various studies [19, 20, 21].

106 Sumera et al. [22] have studied the changes in growth hormone and cortisol profile due to lead
107 induced toxicity in *Labeo rohita* and according to their study; Pb acts as endocrine disruptor and
108 has profound influence on the hormonal profiles and specific growth rate of carp. El-Shebly [23]
109 reported that exposing fish to Pb significantly interferes with the activity of serum GH.

110 Moreover, exposure to toxicants disrupts hormone signaling cellular pathways favoring the
111 findings of present study [24].

112 **Conclusion**

113 The above findings and the results of the present study indicates that the growth hormone levels
114 in the fish has increased with increasing the concentrations of the effluent. This could be due to
115 some toxicants which is present in effluent could have acted as endocrine disruptor and had
116 profound influence on the hormone levels.

117

118 **References:**

- 119 1. Rand GM, Petrocelli SR, In: 'Fundamentals of Aquatic Toxicology Methods and
120 Applications'. (Eds.) Hemisphere Publishing Corporation, Washington, U.S.A. (1985):
121 1-28.
122
- 123 2. Racicot YC, Gander M, Leay C, Blood and liver enzymes in rainbow trout
124 (*Salmo gairdneri* Rich.) with emphasis in their diagnostic study of CCl₄ toxicity and a
125 case of aeromonas infection J Fish Biol 7 (1975): 825.
126
- 127 3. Wieser W, Hinterleintner S, Serum enzymes in rainbow trout as tools in the diagnosis of
128 water quality. Bull Environ contam Toxicol 25 (1980): 188-193.
129
- 130 4. Ekweozor IKE, Bobmanuel NOK, Gabriel UU, Sublethal effect of ammonial fertilizer
131 effluents on the three commercial species from Niger Delta area. J app Sci Environ
132 Mange 5 (2001): 63-68.
133
- 134 5. Adeyemo OK, Hematological and histopathological effects of cassava mill effluent in
135 *Clarias gariepinus*. Afr J Biomed Res 8 (2005) : 179-183.
136
- 137 6. Niwelinski J, Zamorska L, Kaczarski F, Pawlicki R, Enzyme histochemistry and
138 microstructure of the human placenta as indicators of environmental pollution
139 Archiwumochrony Srodowiska 3 (1990): 53-59.
140
- 141 7. Claireaux G, Dutil JD, Physiological responses of Atlantic cod (*Gadus morrhna*) to
142 hypoxia at various environmental salinities J Ep Biol 163 (1992): 97-118.
143
- 144 8. Sebert P, Simon B, Barthelemy, Hydrostatic pressure induces a state resembling
145 histotoxic hypoxia in *Anguilla Anguilla* Comp Biochem Physiol 105 B (1993) : 255-258.
146
- 147 9. Almeida-Val VME, Farias IP, Silva MNP, Duncan WP, Val AL, Biochemical
148 adjustments to hypoxia by Amazon cichids *Braz J Med Biol Res* 28 (1995) : 1257-1263.
149
- 150 10. Akinrotimi OA, Abu OMG, Ansa EJ, Edun OM, George OS, Haematological responses
151 of *Tilapia guineensis* to acute stress. J Nat Appi Sci 5 (2009) : 338 - 343.
152
- 153 11. Gabriel UU, Akinrotimi OA, Management of stress in fish for aquaculture development,
154 Researcher 3(4) (2011): 28- 38.
155
- 156 12. Cengiz EJ, Vnlu E, Sublethal effects of commercial deltamethrin on the structure of the
157 gill, liver and gut tissues of mosquito fish, *Gambusia affis* microscopic study Environ
158 Toxicol Pharmacol 21 (2006) : 246-253.
159
- 160 13. Jung SH, Sim DS, Kim Y, Effects of formalin on haematological and blood chemistry in
161 olive flounder *Paralichthys olivaceus* Aquat Res 34 (2000) : 1269-1275.
162
- 163 14. Wilmore D W. Growth hormone and growth factors in catabolic illness. *Endocrinal*
164 *metab*, 2 (Supp B) (1995) : 77-84.

- 165
166
167
168
169
170
171
172
173
174
175
176
177
178
179
180
181
182
183
184
185
186
187
188
189
190
191
192
193
194
195
196
197
198
199
200
201
202
15. Elisabeth J, Björn TB. Physiological functions of growth hormone in fish with special reference to its influence on behavior. *Fisheries science* 68 Issue sup1 (2002): 742-748
 16. Lescroart O, Roelats I, Mikolajczyk T, Bosma PT, Schulz RW, Kuhn R and Ollevier F. A Radio immuno assay for African cat fish growth hormone: Validation and effects of substance modulating the release of Growth Hormone. *General and comparative Endocrinology* 104 (1996): 147-155.
 17. Peterson BC, Small BC, Bosworth BG. Effects of Bovine Growth hormone (Polisac) on growth performance, body composition and IGFBPs in two strains of channel cat fist. *Aquaculture* 232 (2004): 651-663.
 18. Adsul AD, Singh H. fect of 17α - methyl testosterone on growth and maturation of the fish *Betta splendens*. *J Ecobiol*, 15(1) (2003):23-27
 19. Guerrero RD. Use of androgens for the production of male *Tilapia aurea* (Staindanchner) *Trans. Am. Fish Soc*, 4(2) (1975):342-348.
 20. Nirmala ARC, Pandian TJ. The effect of steroid injection on the food utilization in *Channa striatus*. *Proc.Indian Acad.Sci*, 92 (3) (1983): 221-229.
 21. Sindhu S, Pandian TJ. Effect of administration of different doses of 17α methyl testosterone in *Heteropneustes fossilis* (Bloch). *Proc. Indian Acad.Sci*, 93(6) (1984): 511-516.
 22. Sumera S, Husna M, Laiba S, Aqsa C. Changes in Growth Hormone and Cortisol Profile due to Lead Induced Toxicity in *Labeo rohita*. *Turkish Journal of Fisheries and Aquatic Sciences* 18 (2018): 921-926
 23. El-Shebly AA. Protection of Nile Tilapia (*Oreochromis niloticus*) from lead pollution and enhancement of its growth by α - tocopherol vitamin E. *Research Journal of Fisheries and Hydrobiology*, 4(1) (2009): 17-21.
 24. Gagnon A, Jumarie C, Hontela A. Effects of Cu on plasma cortisol and cortisol secretion by adrenocortical cells of rainbow trout (*Oncorhynchus mykiss*). *Aquatic Toxicology*, 78(1) (2006): 59–65. <https://doi.org/10.1016/j.aquatox.2006.02.004>