

Studies on Character Improvement in Tomato (*Solanum lycopersicum* L.) by Heterosis

ABSTRACT

Most of the tomato varieties in Bangladesh are of inbred type and produced low yield indicating need to develop high yielding variety through the hybridization. Heterosis breeding is used to improve yield and quality of tomato because traditional methods cannot be used to achieve this goal. A half diallel design was employed to develop F_1 s from seven parents of winter tomato. 21 F_1 s along with their parents were evaluated for yield and quality traits. Heterosis analysis revealed that heterotic vigor was present for growth and yield characters among hybrids. Heterosis for better parent was negative for days to flowering, days to harvest, harvest duration, number of locules, and number of seeds per fruit but positive for fruit set, number of fruits per plant, yield per plant, pericarp thickness and TSS. None of the hybrid was heterotic for all characters simultaneously. The hybrids G5, G13, G16, G17, G18, and G20 had 25.73, 19.92, 39.20, 36.49, 53.77, and 50.31% higher heterosis compared to the better parent, respectively, for fruit yield per plant as well as for many other yield contributing traits. High heterosis for yield appears to be the consequence of heterosis of yield attributing traits; therefore, these hybrids offer scope of developing improved commercial lines through heterosis breeding.

Keywords: Heterosis breeding, quantitative trait, tomato, yield

1. INTRODUCTION

Tomato (*Solanum lycopersicum* L.) is one of the most popular and extensively consumed vegetable over the world. Currently, tomato is grown around the globe for either fresh market or processing [1] and considered as a high value crop. As a cash crop, it has a great demand in local as well as the international market. Unfortunately, the production of tomato in Bangladesh is limited due to the scarcity of high yielding varieties. As a result, a huge quantity of tomato is imported every year from the neighboring countries to meet up the local market demand. Recently, the crop has received more attention to the policy makers and researchers. As the development of hybrid varieties with higher yield has been thought to be an effective strategy increasing tomato production, a number of projects have been implemented recent years developing new hybrids in Bangladesh. On the other hand, heterosis breeding is predicted to be the most powerful genetic approach developing hybrids with higher yield [2]. Heterosis, which is the superiority in performance of hybrid individuals compared with their parents [3], has been reported for a wide range of crop species including both self and cross-pollinated crops. Therefore, the estimation of heterosis is one of the goals to assess the hybrid vigor selecting promising hybrids.

37 Heterosis was first observed by Hedrick and Booth [4] in tomato for higher yield. Afterwards a numerous
38 studies have been done in relation to heterosis for yield, its components and quality traits [3,5,6,7,8].
39 However, the exploitation of heterosis is a quick and an effective way of selecting hybrids for high yield
40 potential, earliness and quality attributes. Unfortunately, a very few attempts in this regard has been taken
41 in the past in Bangladesh. The present study was therefore, executed to estimate the level of percent
42 better and mid parent heterosis among F_1 hybrids of tomato. This information would be useful to
43 investigate the performance and relationship of F_1 hybrids with their parents and to select suitable parents
44 and/or population for designing an effective breeding programme.

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2. MATERIALS AND METHODS

48 **2.1 Planting materials:** Seven inbred lines of tomato namely VRT001 (P1), VRT007 (P2), VRT008 (P3),
49 C11 (P4), C41 (P5), LE02 (P6) and TLB133 (P7) were used in the hybridization. A half diallel mating
50 fashion was followed in developing F_1 s in winter 2009-10 (Table 1). Twenty one F_1 s along with the seven
51 parents were evaluated in winter 2010-11. Parental genotype denoting VRT is virus tolerance, LE is
52 *Lycopersicon esculentum*, TLB is tolerance to late blight and C is heat tolerance.

53 **2.2 Experimental site:** The experiment was conducted at the Vegetable Research Field of Horticulture
54 Research Centre (HRC), Bangladesh Agricultural Research Institute (BARI) Bangladesh from October
55 2010 to March 2011. The climate of the experimental site is subtropical characterized by heavy rainfall
56 from May to September and scanty rainfall rest of the year. The soil of the experimental site was sandy
57 loam in texture and acidic in nature with a pH around 6.0. This area belongs to the "Shallow red-brown
58 terrace" soil of Madhupur tract as reported by Haider *et al.* [9]. The land was prepared and fertilized as
59 described by Salim *et al.* [10].

60 **2.3 Seedling raising and transplanting:** Seeds were sown thinly in a raised seed bed on October 15,
61 2010. Seed bed was shaded partially with black net after sowing the seeds. Young seedlings were also
62 covered by a fine mesh white net to protect them from insect attack. 7-days old seedlings were
63 transplanted to a second seed bed at the spacing of 5 x 5 cm for hardening. Thirty days old seedlings
64 were transplanted in the main field on November 15, 2010. Light irrigation was given to each seedling
65 immediately after transplanting for their better establishment.

66 **2.4 Experimental design and plot layout:** Tomato seedlings were grown in a raised seed bed and 30-
67 days old seedlings were transplanted in the main field following randomized complete block design with
68 three replications. Each genotype with spacing of 60 cm x 40 cm represented double row having 12
69 plants per row accommodating in total 24 plants per plot. The unit plot was separated by 50 cm irrigation
70 drain, while blocks were separated by 75 cm drain. Recommended cultural practices as well as plant
71 protection measures were followed.

72 **2.5 Data collection and statistical analysis:** Data for different characters (Table 2) were recorded from
73 10 randomly selected plants of parents and F_1 s. Analysis of variance (ANOVA) was performed as
74 suggested by Gomez and Gomez [11]. Heterosis was estimated using basic formula described by

75 Falconer [12]. Usually, the magnitude of heterosis depends on the accumulation of favorable dominant
 76 alleles in the F₁ population. If the parental populations differ from each other for favorable dominant
 77 alleles, the magnitude of heterosis supposed to be proportionally higher. This relationship was estimated
 78 by the basic formula 1. Where; d = magnitude of dominance, y = difference between the parental
 79 population for allelic frequencies at the locus.

$$\text{Heterosis in F}_1 = \sum dy^2 \dots\dots\dots (1)$$

80
 81 For estimation of heterosis in each character the mean values of the 21 F₁'s have been compared with
 82 better parent (BP) for heterobeltiosis and with mid parent (MP) for heterosis over mid parental value.
 83 Percent heterosis was calculated by the formula 2 and 3.

$$\text{Heterosis (BP)} = \frac{(F_1 - BP)}{BP} \times 100 \dots\dots\dots (2)$$

$$\text{Heterosis (MP)} = \frac{(F_1 - MP)}{MP} \times 100 \dots\dots\dots (3)$$

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 86 Where, F₁ = mean performance of F₁ hybrid, BP = mean performance of better parent and MP = mean
 87 performance of mid parent.

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 89 The test of significance for heterosis was done by using standard error of the value of better parent and
 90 mid parent as suggested by Turner [13]. Mean error variance from the combined analysis of variance of
 91 parents and F₁'s were used for calculating the standard error (SE) of difference. The mean values over
 92 replications were used for the comparison. Finally, critical difference (CD) was calculated by the formula 4
 93 and 5 for heterosis over better and mid parent respectively. Note that the difference between F₁ and the
 94 parent used for the estimation of heterosis were taken into account cross wise. While the difference
 95 between F₁ and the parent was greater than CD it was considered significant and vice versa.
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$$CD (BP) = \sqrt{\frac{2}{r}} EMS \times t \dots\dots\dots (4)$$

$$CD (MP) = \sqrt{\frac{3}{2r}} EMS \times t \dots\dots\dots (5)$$

97
 98
 99 Where, EMS = error mean square from ANOVA table, r = number of replications and t = tabulated value
 100 either at 5% or 1% level of probability.

101 **3. RESULTS AND DISCUSSION**

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103 **3.1 Analysis of variance:** Analysis of variance (ANOVA) for the genotypes *i.e.* parents and F_1 s showed
104 highly significant differences ($P = 0.05$ or $P = 0.01$) for the maximum characters studied except fruit set
105 percentage (Table 2). The estimation of percent heterosis observed in F_1 s over mid and better parent was
106 presented in Table 3 to Table 5.

107 **3.2 Days to 1st flowering:** All the F_1 s showed highly significant differences ($P = 0.05$ or $P = 0.01$)
108 heterosis for flowering time, ranging from -9.89 to -0.09% over mid parent and -11.59 to -2.22% over
109 better parent (Table 3). Out of 21 F_1 combinations, the highest heterobeltiotic effect of -11.59% was found
110 in cross G4 followed by G15 (-11.50%), and G20 (-11.44). The entire cross combinations produced
111 negative heterosis indicating early flowering in hybrids when compared with their parents. Earliness
112 actually leads to the early production and early supply in the market, resulting good price for the
113 producers. Thus the heterosis for flowering time is considered to be an economic parameter for this
114 study. The negative heterosis for flowering time was also reported in earlier studies [5,6,14,15].

115 **3.3 Days to 50% flowering:** The significant differences ($P = 0.05$ or $P = 0.01$) were also observed among
116 the F_1 crosses for the heterosis over mid and better parent (Table 3). Positive heterosis was shown for
117 mid parent whereas negative heterosis ranging from -4.45 to -14.82% was shown for better parent.
118 Negative heterosis showed in flowering indicating earliness by the hybrids as compared to their parents.
119 As the farmers prefer to get a high price from the early supply, therefore, negative heterosis for this trait is
120 preferable. This study is in accordance with the findings of Patwary *et al.* [16], Islam *et al.* [17] and
121 Baishya *et al.* [18], those who reported negative heterosis for this trait over better parent in their studies.

122 **3.4 Days to 1st harvest:** Out of 21 cross combinations, 20 exhibited significant different ($P = 0.05$ or $P =$
123 0.01) negative heterosis over better parent ranging from -3.05 to -11.92% whereas 18 combinations
124 showed negative heterosis over mid parent (Table 3). The results were very similar to Sharma *et al.* [19]
125 who reported heterosis ranged of -2.90 to -11.20% over better parent in tomato. More than 10% negative
126 heterosis over better parent was observed from three F_1 s *viz.* G5 (-11.92%), G1 (-10.38%), and G12 (-
127 10.18%), which was superior to the previous study -7.14% of heterosis over better parent, reported by
128 Sharma *et al.* [20]. Negative heterosis here is suggesting early harvest of tomato fruits. Therefore, those
129 genotypes can further be utilized to develop inbred lines toward a variety development program.

130 **3.5 Harvest duration (days):** Harvest duration showed significant negative better parent heterosis in
131 fourteen F_1 s whereas negative mid parent heterosis was showed in thirteen F_1 s (Table 3). The highest
132 significant negative heterosis over better parent was estimated from the cross combination G1 (-6.77%)
133 followed by G12 (-6.68%). On the other hand, the highest negative heterosis over mid parent was also
134 estimated from the cross G1 (-6.50%). In contrast, four crosses produced significant positive heterosis
135 over better parent *viz.* G18 (5.58%), G16 (4.72%), G8 (3.87%), and G17 (3.05%), which also showed
136 positive heterosis over their mid parent (Table 3). Positive heterosis suggests longer harvest period
137 whereas negative heterosis suggests shorter harvest period. Generally, longer and shorter harvest
138 duration is preferred by the homestead and commercial growers, respectively. Positive heterosis for the
139 trait was also reported by Kumari and Sharma [14] and Khan and Jindal [21]. Therefore, these genotypes

140 would be the effective combination in exploiting heterosis for the homestead and commercial growers as
141 their desire.

142 **3.6 Plant height at 1st harvest:** Significant negative heterosis for better parent was manifested by five
143 **F₁s** viz. G11 (-15.32%), G20 (-10.56%), G13 (-10.25%), G1 (-9.76%) and G19 (-7.74%). Only two **F₁s** viz.
144 G11 (-11.85%), and G20 (-6.76%) produced significant negative heterosis for their mid parent (**Table 3**).
145 Significant positive heterosis for better parent was also found from the crosses G14 (16.60%) and G17
146 (8.87%). **This result is similar to that of** Baishya *et al.* [18] and Padma *et al.* [22]. Patwary *et al.* [16]
147 reported both positive and negative heterosis for their study whereas Fageria *et al.* [23] reported only
148 positive heterosis. So, these genotypes can further be used to develop inbred lines toward developing of
149 both taller and dwarf varieties.

150 **3.7 Fruit set (%):** Seventeen out of 21 **F₁s** produced significant different ($P = 0.05$ or $P = 0.01$) positive
151 heterosis over their better parent whereas 16 produced significant positive heterosis over their mid parent
152 (**Table 4**). Ten cross combinations viz. G20 (25.57%), G8 (17.00%), G18 (14.82%), G9 (10.29%), G19
153 (4.71%), G16 (3.72%) and G11 (2.04%) produced significant positive heterosis either their mid or better
154 parent indicating potential increment of fruit set. On the other hand, seven **F₁s** performed negative
155 heterosis ranging from -1.68 to -22.11% indicating a reduction in fruit setting. Both positive and negative
156 heterosis in respect of fruit setting was reported by El-Ahmadi and Stevens [24].

157 **3.8 Number of fruits per plant:** About 50% of the **F₁s** showed significant different ($P = 0.05$ or $P = 0.01$)
158 positive heterosis over their better parent ranging from 7.86 to 45.99% (**Table 4**). More than 40%
159 heterosis over their better parent was produced by four crosses viz. G3, G10, G13, G18. On the other
160 hand, about 76% of the **F₁s** produced significant positive heterosis over their mid parent ranging from
161 12.05 to 63.55% (**Table 4**). This result suggested a potential increment of fruits number in the tomato
162 plant. This study showed a bit higher amount of heterosis for fruits number than the previous study by
163 Patwary *et al.* [16]. It could be due to the variation of the parents used in the study. Our study also had an
164 agreement with the previous research [6,18,19,20,23].

165 **3.9 Fruit length (cm):** Fourteen hybrids showed positive heterosis, of which 5 hybrids exhibited positive
166 significant heterosis over better parents (**Table 4**). More than 10% heterosis was estimated from four
167 **crosses viz.** G6, G4, G14, and G20. Only one hybrid G18 (-12.93%) produced the significant negative
168 heterosis over better parent. Since, only a genotype out of twenty one showed significant negative
169 heterosis over better parent, indicating character is mainly governed by non-additive gene effects. Islam
170 *et al.* [18] also reported similar results for fruit length. Significant positive heterosis has been reported by
171 Ahmad *et al.* [6], and Sharma *et al.* [20]. These findings of significant positive heterosis over mid and
172 better parent are in line with the findings of Singh *et al.* [5] and Kumar and Singh [25] as well.

173 **3.10 Fruit diameter (cm):** About 62% **hybrids** exhibited with significant positive heterosis over better
174 parent, whereas 76% produced significant positive heterosis over mid parent (**Table 4**). The highest value
175 of positive heterotic effect was exhibited by the **cross** G4 (53.70 %) followed by G2 (48.46 %), G13 (46.54
176 %), G7 (42.50 %) and G14 (40.00 %). One-third of the hybrids produced significant negative heterosis for

177 either mid or better parent, which suggested that the character is possibly governed by non-additive gene
178 action. Heterosis for fruit diameter in tomato was also reported by Ahmad *et al.* [6], Padma *et al.* [23], and
179 Sharma *et al.* [20].

180 **3.11 Average fruit weight (g):** The entire cross combinations except G18 and G4 exhibited with negative
181 heterosis over mid and better parent, whereas two hybrids G18 (12.09%) and G4 (12.01%) showed
182 significant positive heterosis over mid parent (Table 4). The best hybrid was G18, which showed the
183 highest per se performance with the highest heterosis (12.09%) over mid parent. Positive heterosis for
184 fruit weight has been reported by Sharma *et al.* [19,20], whereas both positive and negative heterosis
185 over better parent reported by Patwary *et al.* [16] and Ahmad *et al.* [6] in their studies. These findings of
186 positive heterosis over mid parent and check co-relate with the findings of Kumari and Sharma [14] and
187 Marbal *et al.* [26].

188 **3.12 Total soluble solid (TSS):** Significant positive heterosis over mid and better parent was observed in
189 all the F_1 s confirming additive gene effect for the trait (Table 4). The highest positive heterosis was
190 observed in cross G20 (141.67%) followed by G17 (84.76%), and G16 (80.83%). Similar range of
191 heterosis was also noted by the previous studies [8,17,19,20,22,27]. Total soluble solid is responsible for
192 the sweetness of tomato hereafter high TSS is a preferable character in processing tomatoes. So, these
193 genotypes can further be advanced toward developing a processing variety.

194 **3.13 Fruit yield per plant (kg):** Of 21 crosses, six produced significant different ($P = 0.05$ or $P = 0.01$)
195 positive heterosis over better parent, whereas 15 produced significant positive heterosis over mid parent
196 (Table 5). More than 20% heterosis over better parent was observed in five F_1 s viz. G18 (53.77%), G20
197 (50.31%), G16 (39.20%), G17 (36.49%), and G5 (25.70%). The cross combinations G18 (70.00%), G16
198 (58.74%) and G20 (55.63%) showed higher positive heterosis over mid parent. This result suggested a
199 potential yield increment by the heterosis, and is predicted to be the reason of high yielding parents used
200 in the hybridization [28]. Eight genotypes exhibited with significant negative heterosis over either mid or
201 better parent. Positive better parent heterosis ranging from 13.58 to 282.63% was reported in heat
202 tolerant tomato [16], which was higher than this study. Bhatt *et al.* [8,27] observed 2.92 to 54.17% better
203 parent heterosis for yield per plant in tomato, which is very identical to our findings. Similarly,
204 heterobeltiosis in tomato hybrids was also reported in many studies [3,6,14,25,26,29,30]. Therefore,
205 these genotypes may be selected as heterotic hybrids for yield and can further be advanced toward
206 developing a high yielding variety.

207 **3.14 Number of locules per fruit:** Seven cross combinations out of 21 showed positive heterobeltiosis
208 but only two was significant. Positive heterosis for this trait ranged from 1.94 to 56.66% (Table 5). On the
209 other hand, nine cross combinations produced significant negative heterosis over better parent ranging
210 from -18.15 to -51.38%. More than 35 % negative heterosis was manifested by five F_1 s namely G8 (-
211 51.38 %), G16 (- 46.03 %), G18 (- 46.03 %), G17 (- 40.02 %) and G15 (- 36.29 %). Similarly, eight F_1 s
212 showed significant positive heterosis over mid parent and five F_1 s showed significant negative heterosis
213 over mid parent. The hybrid G20 showed no heterosis regarding locule number in fruit (Table 5).

214 However, the estimation of negative heterobeltiosis from -4.50 to -51.39% was observed from the study,
215 indicating the importance of non-additive gene action for the trait. As a result, heterosis breeding can be
216 exploited very well to reduce the locule number in tomato fruits. This result supported by Duhan *et al.*
217 [31], Kurian *et al.* [7] and Dod *et al.* [32] in where identified heterotic hybrids for lower locule number in
218 tomato. On the other hand, Ahmad *et al.* [6] reported significant positive heterosis for this trait. From the
219 quality point of view, less locule is desirable in tomato. This study is predicted the potential genotypes for
220 future breeding in reducing locule as we have seen negative estimation of heterosis.

221 **3.15 Pericarp thickness:** The highly significant different ($P = 0.05$ or $P = 0.01$) heterosis was estimated
222 by the majority of the hybrids towards positive heterosis over mid parent, whereas 12 hybrids produced
223 significant positive heterosis for better parent ranging from 26.67 to 109.06% (Table 5). More than 25%
224 heterosis exhibited by the 57% hybrids, indicating possibility of the enhancement of fruit quality by
225 improving pericarp thickness. Only a single hybrid G12 produced significant negative heterosis for both
226 mid and better parent. The results of the study in relation to pericarp thickness were agreed by the
227 previous studies [14,16,19,21,33,34]. Pericarp thickness usually contributes much for long storability.
228 Positive heterosis is the indicator of additive gene action for the trait, and is predicted to increase pericarp
229 thickness of tomato using these genotypes in a variety development program.

230 **3.16 Number of seeds per fruit:** Significant negative heterosis was manifested by 19 hybrids varying
231 from -10.30 to -67.56% for both mid and better parent (Table 5). The highest negative heterotic value was
232 achieved by the hybrid G8 (-67.56) followed by G3 (-65.41), G21 (-59.51) and G9 (-59.39) whereas the
233 lowest negative heterosis was provided by the hybrid G4. Ahmad *et al.* [6] and El-Ahmadi and Stevens
234 [24] reported higher degree of heterosis for this trait. Negative heterosis is an indication of the reduction
235 of seeds in tomato as the consumers expect. So, these cross combinations can be further used toward
236 developing less seeded tomato varieties.

237 **3.17 1000-seed weight:** The highly significant different ($P = 0.05$ or $P = 0.01$) positive heterosis was
238 observed by 48% of the hybrids over better parent (Table 5) indicating seed quality can be improved
239 through the hybridization. More than 10% positive heterosis was manifested by five hybrids *viz.* G4
240 (18.11%), G10 (15.81%), G9 (14.45%), G19 (11.39%), and G21 (12.13%). Nine hybrids provided
241 significant negative heterosis ranging from -4.30 to -26.94%. This result is in accordance with the findings
242 of Subburamu *et al.* [35].

243

244 4. CONCLUSION

245 None of the cross combinations was heterotic for all characters simultaneously. In this study, promising
246 hybrids for yield per plant with significant over better parent in desirable direction and also revealed for
247 other traits *viz.* days to flowering and harvesting, number of fruits per plant, fruit length, fruit diameter,
248 pericarp thickness, number of locules per fruit, plant height, TSS, 1000-seed weight (Table 6). As a result,
249 high heterosis for yield appears to be the consequence of heterosis of the yield attributing traits. Among
250 the hybrids G5, G13, G16, G17, G18 and G20 were promising for yield per plant as well as for many

251 other yield contributing traits. Therefore, these hybrids can be used to develop high yielding varieties
252 along with other quality traits.

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COMPETING INTERESTS

256 Authors have declared that there was no competing interests exist
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REFERENCES

- 259 1. Nowicki M, Kozik EU, Foolad MR. Late blight of tomato. In: Varshney RK, Tuberosa R, editors.
260 Translational genomics for crop breeding, 1st edn. Wiley-Blackwell, Hoboken, New Jersey, USA;
261 2013.
- 262 2. Choudhary B, Punia RS, Sangha HS. Manifestation of hybrid vigour in F₁ and its correlation in F₂
263 generation of tomato (*Lycopersicon esculentum* Mill). Indian J. Hort. 1965; 22: 52-59.
- 264 3. Tamta S, Singh JP. Heterosis in Tomato for Growth and Yield Traits. International Journal of
265 Vegetable Science, 2018; 24(2): 169-179.
- 266 4. Hedric UP, Booth NP. Mendelian characters in tomato. Proc. Am. Soc. Hort. 1907; 5: 19-24.
- 267 5. Singh NB, Paul A, Wani SH, Laishram JM. Heterosis studies for yield and its components in
268 tomato (*Solanum lycopersicum* L.) under valley conditions of Manipur. Int. J. Life Sci. 2012; 1:
269 224-232.
- 270 6. Ahmad S, Quamruzzaman AKM, Islam MR. Estimate of heterosis in tomato (*solanum*
271 *lycopersicum* L.) Bangladesh J. Agril. Res. 2011; 36: 521-527.
- 272 7. Kurian A, Peter KV, Rajan S. Heterosis for quality traits in tomato. J. Tropical Agric. 2001. 39: 5-8.
- 273 8. Bhatt RP, Biswas VR, Kumar N. Heterosis, combining ability, genetics for vitamin C, total soluble
274 solids and yield in tomato (*Lycopersicon esculentum* Mill) at 1700 m altitude. J.Agric. Sci. 2001;
275 137: 71-75.
- 276 9. Haider J, Marumoto T, Azad AK. Estimation of microbial biomass, carbon and nitrogen in
277 Bangladesh soils. Sci. Plant Nutr. 1991; 37: 591-599.
- 278 10. Salim MMR, Rashid MH, Hossain MM, Zakaria M. Morphological characterization of tomato
279 (*Solanum lycopersicum* L.) genotypes. Journal of the Saudi Society of Agricultural Sciences
280 2018; (In press). <https://doi.org/10.1016/j.jssas.2018.11.001>
- 281 11. Gomez KA, Gomez AA. Statistical Procedures for Agricultural Research. 2nd edition. John Wiley
282 and Sons, New York; 1983.
- 283 12. Falconer DS. Introduction to Quantitative Genetics, Longman Inc. Ltd. New York; 1981.
- 284 13. Turner JH. A study of heterosis in upland cotton, combining ability and inbreeding effects. Agron
285 J. 1953; 43: 478-490.
- 286 14. Kumari S, Sharma MK. Exploitation for yield and its contributing traits in tomato (*Solanum*
287 *lycopersicum* L.). International Journal of Farm Sciences 2011; 1: 45-55.

- 288 15. Dod VN, Kale PB. Heterosis for certain quality traits in tomato (*Lycopersicon esculentum* Mill).
289 Crop Res. 1992; 5: 303-308.
- 290 16. Patwary MMA, Rahman MM, Ahmad S, Miah MAK, Barua H. Study of heterosis in heat tolerant
291 tomato during summer. Bangladesh J. Agril. Res. 2013; 38: 531-544.
- 292 17. Islam MR, Ahmad S, Rahman MM. Heterosis and qualitative attributes in winter tomato (*Solanum*
293 *lycopersicum* L.) hybrids. Bangladesh J. Agril. Res. 2012; 37: 39-48.
- 294 18. Baishya KC, Syamal MM, Singh KP. Heterosis studies in tomato (*Lycopersicon esculentum* Mill.).
295 Veg. Sci. 2001; 28: 168-169.
- 296 19. Sharma DK, Chaudhary DR, Pandey DP. Studies on hybrid vigour in tomato (*Lycopersicon*
297 *esculentum* Mill.). Haryana J. Hort. Sci. 2001; 30: 236-238.
- 298 20. Sharma HR, Sharma D, Thakur AK. Studies on analysis of genetic divergence in tomato
299 (*Lycopersicon esculentum* Mill.). J. Hort. Sci. 2006; 1: 52-54.
- 300 21. Khan A, Jindal SK. Exploiting yield potential in tomato (*Solanum lycopersicum* L.) through
301 heterosis breeding. Plant Gene and Trait 2016; 7: 1-7.
- 302 22. Padma E, Senkar CR, Rao BV. Heterosis and combining ability in tomato (*Lycopersicon*
303 *esculentum* Mill.). The Andhra Agric. J. 2002; 49: 285-292.
- 304 23. Fageria MS, Kohli UK, Dhaka RS. Studies on heterobeltiosis for fruit yield and yield attributing
305 traits in tomato. Haryana J. Hort. Sci. 2001; 30: 131-133.
- 306 24. El-Ahmadi AB, Stevens MA. Genetics of high temperature fruit set in the tomato. J. Amer. Soc.
307 Hort. Sci. 1979; 104: 691-696.
- 308 25. Kumar C, Singh SP. Heterosis and inbreeding depression to identify superior F₁ hybrids in tomato
309 (*Solanum lycopersicum* L.) for the yield and its contributing traits. J. App. and Nat. Sci. 2016; 8:
310 290 – 296.
- 311 26. Marbhal SK, Ranpise SA, Kshirsagar DB. Heterosis study in cherry tomato for quantitative traits.
312 Int. Res. J. Multidiscip. Stud. 2016; 2: 1-6.
- 313 27. Bhatt RP, Adhekari RS, Narendra K. Genetical analysis for quantitative and qualitative traits in
314 tomato (*Lycopersicon esculentum* Mill.) under open and protected environment. The Indian J.
315 Genet. Pl. Breed. 2004; 64: 125-129.
- 316 28. Courtney WH, Peirce LC. Parent selection in tomato based on morpho-physiological traits.
317 HortScience 1979; 14: 458.
- 318 29. Hassan AA, Moustafa SES, Abdel AK, Mohammad AA. Development and release of some new
319 tomato (*Lycopersicon esculentum* Mill.) hybrids. Egyptian J. Hort. 2000; 27: 210-218.
- 320 30. Dharmatti PR, Madalgeri BB, Mannikeri IM, Patil RV, Patil G. Genetic divergence studies in
321 summer tomatoes. Karnataka Journal of Agricultural Sciences 2001; 14: 407-411.
- 322 31. Duhan D, Partap PS, Rana MK, Basawana KS. Study of heterosis for growth and yield characters
323 in tomato. Haryana J. Hort. Sci. 2005; 34: 366-370.

- 324 32. Dod VN, Kale PB, Wankhade RV. Heterosis and combining ability in tomato (*Lycopersicon*
 325 *esculentum* Mill.). PKV Res. J. 1995; 19: 125-129.
- 326 33. Makesh S, Puddan M, Ashok S, Banu MR. Heterosis studies for quality and yield in tomato
 327 (*Lycopersicon esculentum* Mill.). Advances in Plant Science 2002; 15: 597-601.
- 328 34. Dhaliwal MS, Surjan S, Badha BS, Cheema DS, Singh S. Diallel analysis for total soluble solids
 329 content, pericarp thickness and locule number in tomato. Veg. Sci. 1999; 26: 120-122.
- 330 35. Subburamu K, Jayapragasam M, Thandapani V. Heterosis for seed and seedling characters in
 331 tomato (*Solanum lycopersicum*). Seed-Res. 1999; 26: 187-190.

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333 **Table 1 Developed F₁ hybrids by a half diallel fashion**

Parent (P)	P1	P2	P3	P4	P5	P6	P7
P1 (WP10)	-	P1 × P2 (G1)	P1 × P3 (G2)	P1 × P4 (G3)	P1 × P5 (G4)	P1 × P6 (G5)	P1 × P7 (G6)
P2 (VRT003)			P2 × P3 (G7)	P2 × P4 (G8)	P2 × P5 (G9)	P2 × P6 (G10)	P2 × P7 (G11)
P3 (VRT004)				P3 × P4 (G12)	P3 × P5 (G13)	P3 × P6 (G14)	P3 × P7 (G15)
P4 (LE009)					P4 × P5 (G16)	P4 × P6 (G17)	P4 × P7 (G18)
P5 (TLB182)						P5 × P6 (G19)	P5 × P7 (G20)
P6 (WP02)							P6 × P7 (G21)
P7 (TLB111)							-

334 ⁴Hybrid

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337 **Table 2 ANOVA for various traits of 21 F₁s and seven parents of tomato**

Characters	Mean Squares		
	Replications (df = 2)	Genotypes (df = 27)	Error (df = 54)
Days to 1 st flowering	0.94	8.47**	0.77
Days to 50% flowering	3.62	24.29**	0.58
Days to 1 st harvesting	9.33	41.37**	10.54
Harvest duration	52.27	95.44*	14.33
Plant height at 1 st harvest (cm)	60.69	174.81**	11.253
Fruit set percentage (%)	130.47	107.71	67.81
Number of fruits per plant	7.20	291.75**	9.65
Fruit length (cm)	1.48	1.44**	0.21
Fruit diameter (cm)	0.65	4.33**	0.13
Average fruit weight (g)	38.56	1829.54**	29.67
Yield per plant (kg)	0.11	0.47**	0.07
Total soluble solid (%)	0.35	6.09**	0.26
Locules per fruit	0.81	5.55**	0.76
Pericarp thickness (mm)	4.51	4.53**	0.44
Seeds per fruit	1505.23	1063.47**	5.04
1000-seed weight (g)	2.99	0.27**	0.001

338 ^YDegree of freedom; *, ** = Significant difference at P = 0.05 and P = 0.01 respectively

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Table 3 Percent heterosis over mid parent (MP) and better parent (BP) for days to 1st flowering, days to 50 % flowering, days to 1st harvest, harvest duration and plant height at 1st harvest in winter tomato.

Genotypes	Days to 1 st flowering		Days to 50% flowering		Days to 1 st harvest		Harvest duration		Plant height at 1 st harvest (cm)	
	MP	BP	MP	BP	MP	BP	MP	BP	MP	BP
G1	-9.890**	-10.87**	14.129**	-12.10**	-9.979**	-10.38**	-6.495**	-6.77**	-1.087	-9.76*
G2	-0.090**	-9.35**	0.268**	-7.64**	-0.086**	-9.89**	-0.056**	-6.49**	0.146**	6.12
G3	-2.571**	-4.32**	30.297**	-7.64**	-4.573*	-9.02**	-2.953**	-5.92**	-0.531	-6.394
G4	-9.290**	-11.59**	39.753**	-12.10**	-8.299**	-8.98**	-5.378**	-5.84**	5.172*	-4.18
G5	-8.834**	-10.15**	21.191**	-10.97**	-10.619**	-11.92**	-4.046**	-4.96**	10.810**	7.64
G6	-8.644**	-9.30**	28.177**	-14.82**	-5.660**	-7.78**	0.888	-0.58	8.778**	3.08
G7	-5.836**	-7.19**	11.644**	-12.10**	-6.899**	-7.85**	1.441*	0.76	13.370**	-3.47
G8	-2.222**	-2.22*	2.923	-8.91**	-2.877	-5.04*	5.400**	3.87**	0.971	-2.31
G9	-3.011**	-4.44**	14.573**	-9.56**	-5.703**	-6.82**	1.561*	0.78	-1.098	-1.25
G10	-3.357**	-3.71**	6.805**	-4.45**	-5.436**	-6.40**	-3.556**	-4.20**	9.213**	-2.94
G11	-3.284**	-5.01**	13.311**	-8.65**	-3.348	-5.93**	-2.161**	-3.87**	-11.851**	-15.32**
G12	-10.216**	-11.50**	1.038	-14.01**	-7.207**	-10.18**	-4.678**	-6.68**	8.902**	-4.61
G13	-5.933**	-8.63**	18.200**	-12.10**	-7.578**	-9.59**	-4.937**	-6.30**	5.535*	-10.25**
G14	-4.769**	-6.48**	19.816**	-5.73**	-6.977**	-6.98**	-4.775**	-4.78**	22.472**	16.60**
G15	-6.817**	-7.16**	21.154**	-12.35**	-3.774*	-7.27**	-0.686	-3.06**	9.953**	-3.18
G16	-3.755**	-5.18**	11.696**	-12.10**	-2.000*	-3.05*	5.442**	4.72**	1.355	-2.09
G17	-4.093**	-4.44**	3.398	-12.73**	-2.703**	-5.82**	5.263**	3.05*	18.877**	8.87*
G18	-1.102	-2.87**	7.447*	-4.94**	0.786	0.32	5.898**	5.58**	5.484*	4.72
G19	-5.660**	-6.72**	13.927**	-13.38**	-7.284**	-9.31**	-4.745**	-6.11**	3.951	-7.74*
G20	-8.501**	-11.44**	23.821**	-16.67**	-4.324**	-5.78**	-2.185**	-3.15**	-6.762**	-10.56**
G21	-8.036**	-10.01**	11.162**	-15.43**	-6.181**	-9.59**	-3.419**	-5.73**	7.221**	-1.15
SE	0.620	0.72	0.539	0.62	2.296	2.65	2.677	3.09	2.372	2.74
CD at 5%	0.507	0.83	0.442	0.72	1.879	3.07	2.192	3.60	4.766	7.78
CD at 1%	0.675	1.10	0.588	0.96	2.503	4.09	2.919	4.82	6.347	10.36

341 *, ** = Significant difference at $P = 0.05$ and $P = 0.01$ respectively

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Table 4 Percent heterosis over mid parent (MP) and better parent (BP) for fruit set (%), number of fruits, fruit length, fruit diameter, average fruit weight, and TSS% in winter tomato.

Genotypes	Fruit set (%)		Number of fruits per plant		Fruit length (cm)		Fruit diameter (cm)		Average fruit weight (g)		Total soluble solid (TSS %)	
	MP	BP	MP	BP	MP	BP	MP	BP	MP	BP	MP	BP
G1	17.873**	8.48**	16.130**	-0.79	14.129**	-5.10	18.182**	17.50**	-18.374**	-23.37**	43.992**	40.78**
G2	-0.209**	-22.1083**	-0.024	-14.73*	0.268**	3.47	0.493**	48.46**	-0.153**	-30.77**	0.487**	45.07**
G3	-14.062**	-22.1083**	4.847	40.47**	30.297**	6.03	0.242	-14.35**	1.402	-49.55**	60.432**	50.81**
G4	2.278	-11.3983**	14.251**	10.08	39.753**	26.91**	79.342**	53.70**	12.014**	-1.87	75.949**	41.26**
G5	-5.431**	-9.92**	45.655**	27.97**	21.191**	3.93	25.897**	25.53**	-15.462**	-26.28**	34.054**	26.02**
G6	-3.437*	-14.20**	23.217**	-3.43	28.177**	27.47**	37.422**	6.81*	-13.165**	-38.18**	89.394**	52.44**
G7	-1.727	-8.21**	-1.680	-24.83**	11.644**	9.03*	42.500**	42.50**	-14.049**	-33.07**	34.302**	34.04**
G8	18.968**	17.00**	18.833**	10.36*	2.923	0.17	-14.538**	-26.62**	-11.228**	-26.96**	51.266**	39.22**
G9	17.851**	10.29**	52.504**	26.34**	14.573**	3.83	31.680**	12.31**	-13.790**	-19.95**	74.170**	37.48**
G10	11.079**	7.12**	46.113**	41.42**	6.805**	2.91	24.397**	24.04**	-18.484**	-24.68**	58.228**	45.63**
G11	6.043**	2.04	38.380**	24.60**	13.311**	-6.19	17.764**	-8.85**	-18.163**	-39.32**	71.779**	35.92**
G12	1.652	-6.51**	30.239*	-5.34	1.038	0.69	8.434**	-6.90**	-23.550**	-48.02**	50.947**	38.69**
G13	1.689	-10.69**	58.967**	43.51**	18.200**	4.86	71.815**	46.54**	-9.009**	-32.77**	84.049**	45.07**
G14	-9.178**	-12.14**	0.562	-21.29**	19.816**	12.85**	40.405**	40.00**	-1.410	-27.41**	47.368**	35.40**
G15	-3.204*	-12.76**	-13.762	-38.61**	21.154**	-1.56	56.522**	21.15**	-1.887	-37.74**	83.599**	45.07**
G16	9.104**	3.72	63.554**	27.84**	11.696**	-1.21	0.366	-24.41**	-7.057**	-18.52**	114.227**	80.83**
G17	-2.192	-7.18**	39.286**	25.52**	3.398	-2.93	-11.594**	-24.28**	-1.117	-12.91**	84.758**	84.76**
G18	17.400**	14.82**	50.968**	45.99**	7.447*	-12.93**	-2.376	-32.00**	12.090**	-2.04	97.817**	67.44**
G19	15.743**	4.71	12.053**	-4.65	13.927**	6.88	38.009**	17.99**	-13.097**	-13.56**	103.010**	71.36**
G20	29.269**	25.57**	41.351**	7.86*	23.821**	11.88**	51.534**	34.61**	0.955	-21.08**	142.475**	141.67**
G21	5.802**	-1.68	22.581**	7.23	11.162**	-5.11	21.945**	-5.42	-6.990*	-27.01**	86.357**	57.74**
SE	5.823	6.72	2.197	2.54	0.326	0.38	0.255	0.29	3.851	4.45	0.254	0.29
CD at 5%	1.941	3.17	1.798	2.94	0.267	0.44	0.209	0.34	3.152	5.15	0.208	0.34
CD at 1%	2.586	4.22	2.394	3.91	0.356	0.58	0.278	0.45	4.198	6.86	0.277	0.45

345 *, ** = Significant difference at $P = 0.05$ and $P = 0.01$ respectively

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Table 5 Percent heterosis over mid parent (MP) and better parent (BP) for yield, number of locules, pericarp thickness, number of seeds and 1000-seed weight in winter tomato.

Genotypes	Fruit yield per plant (kg)		Number of locules per fruit		Pericarp thickness (mm)		Number of seeds per fruit		1000-seed weight (g)	
	MP	BP	MP	BP	MP	BP	MP	BP	MP	BP
G1	-3.967	-13.21**	52.624**	48.04**	100.667**	89.91**	-23.822**	-29.60**	10.891**	9.375**
G2	-0.144**	-20.72**	0.254**	-4.50	0.254**	-4.75	0.066**	2.91*	0.017**	-9.907**
G3	14.286**	-5.98	0.000	-18.96**	59.627**	26.67**	-65.341**	-65.41**	13.238**	-13.932**
G4	26.923**	7.94	67.598**	56.66**	80.317**	79.18**	19.874**	-10.30**	19.284**	18.110**
G5	26.588**	25.70**	22.549**	15.47	78.650**	77.81**	-30.740**	-34.59**	7.486**	2.941**
G6	20.533**	5.61	20.950**	13.06	38.436**	34.07*	-14.287**	-23.42**	1.581**	0.000**
G7	-7.364*	-9.81*	28.088**	-4.50	37.066**	0.33	-12.614**	-16.51**	-18.480**	-26.935**
G8	7.759*	-5.66	-38.588**	-51.38**	66.221**	26.67**	-67.249**	-67.56**	-1.606**	-4.297**
G9	29.639**	1.51	5.916	1.94	105.034**	95.21**	-42.850**	-59.39**	14.902**	14.453**
G10	18.908**	6.79	26.103**	15.47	121.891**	109.06**	-37.866**	-38.93**	19.318**	15.809**
G11	18.310**	-4.91	5.916	1.94	58.621**	54.88**	-41.260**	-43.38**	-13.450**	-13.619**
G12	21.778**	9.16	3.704	-4.50	-23.130**	-27.54**	-32.397**	-34.45**	7.257**	-6.192**
G13	50.125**	19.92***	12.570*	-18.15**	18.960*	-10.00	33.333**	-2.50	2.600**	-8.359**
G14	10.390**	1.59	-8.576	-27.29**	37.849**	5.08	-27.156**	-28.82**	9.916**	1.238**
G15	9.223*	-10.36*	-12.383*	-36.29**	10.915*	-17.54	-18.898**	-25.18**	-2.414**	-12.384**
G16	58.739**	39.20**	-29.895**	-46.03**	41.149**	11.48	14.594**	-17.81**	-14.516**	-16.535**
G17	40.488**	36.49**	-30.095**	-40.52**	19.070**	-5.19	-28.994**	-29.55**	7.782**	1.838**
G18	70.000**	53.77**	-29.895**	-46.03**	9.677*	-15.00	-52.369**	-54.76**	5.812**	2.724**
G19	-2.493	-16.59**	4.439	-7.62	97.788**	95.66**	-20.550**	-42.74**	15.209**	11.397**
G20	55.627**	50.31**	0.000	0.00	86.230**	81.47**	-31.146**	-52.12**	0.587	0.000
G21	18.280**	4.27	-13.055	-23.10*	45.543**	40.31**	-57.053**	-59.51**	15.312**	12.132**
SE	0.184	4.27	0.616	0.71	0.470	0.54	1.587	1.83	0.022	0.03
CD at 5%	0.151	0.25	0.505	0.82	0.385	0.63	1.299	2.12	0.018	0.03
CD at 1%	0.201	0.33	0.672	1.10	0.512	0.84	1.730	2.83	0.024	0.04

350 *, ** = Significant difference at $P = 0.05$ and $P = 0.01$ respectively

351 **Table 6 Promising F_1 hybrids showing higher per se performance and better-parent heterosis**
 352 **(BPH) for yield per plant and significant BPH for other characters**

Genotypes	Yield per plant (kg)	BPH (%) for yield	BPH for other characters
G18	3.06	53.77**	# of locule, # of seeds per fruit, 1000-seed weight, harvest duration, TSS
G20	2.42	50.31**	Days to 1 st flowering, days to 50% flowering, days to 1 st harvest, harvest duration, fruit length, fruit diameter, TSS, pericarp thickness, # of seeds per fruit, # of fruits per plant
G16	2.77	39.20**	Days to 1 st flowering, days to 50% flowering, days to 1 st harvest, harvest duration, fruit diameter, TSS, # of seeds per fruit, # of fruits per plant
G17	2.88	36.49**	Days to 1 st flowering, days to 50% flowering, days to 1 st harvest, harvest duration, plant height, fruit diameter, TSS, # of seeds per fruit, # of fruits per plant, 1000-seed weight
G5	2.67	25.70**	Days to 1 st flowering, days to 50% flowering, days to 1 st harvest, harvest duration, fruit diameter, TSS, # of seeds per fruit, # of fruits per plant, 1000-seed weight
G13	3.02	19.92**	Days to 1 st flowering, days to 50% flowering, days to 1 st harvest, harvest duration, fruit diameter, TSS, # of fruits per plant, # of locule

353 ** = Significant difference at $P = 0.05$; # refers to number
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