

# Impact of Aluminum and Silicon Nanocomposite on Foodborne Pathogens in Chicken Fillets

**Running head:** Nanoparticles and improving quality of chicken fillets

## ABSTRACT

Nanotechnology is an innovative technology for improving food quality and safety.

**Aims:** The aim of this study was to evaluate the efficacy of hydroxy propyl methyl cellulose (HPMC) films containing nanoparticles against **three** foodborne pathogens.

**Study design:** All data in this study were collect by different results we have got about them by this study.

**Place and Duration:** All experiments were done in Food Technology Department, Benha University, Egypt; Nanomaterial Laboratory, Beni-Suef University, Egypt and Agricultural Research Center, Egypt and were done within **three** months.

**Methodology:** All results had obtained by different experiments in different labs as preparation edible film, antimicrobial activity, mode of action, challenge study and the scanning electron microscope (SEM) as well mechanical properties of HPMC films were tested.

**Results:** The results obtained from this study showed that, the nanoparticles (~80 nm) at 80 ppm were active against *Bacillus cereus*, *Staphylococcus aureus*, and *Salmonella* Typhimurium compared with 20 and 40 ppm. The HPMC films including Al<sub>2</sub>O<sub>3</sub>-NPs were active against *B. cereus* than *S. aureus* and *S. Typhimurium*, while the SiO<sub>2</sub>-NPs were more effective against *S. Typhimurium* and *B. cereus* compared with *S. aureus*. In challenge studies, HPMC films including Al<sub>2</sub>O<sub>3</sub>-NPs and SiO<sub>2</sub>-NPs at 80 ppm decreased the viability of the **three**-foodborne pathogens associated with chicken fillets stored at 4±1°C for 15 days, as compared with the control sample. HPMC films incorporated with nanoparticles inhibited the microbial population ~ 2-3 **log<sub>10</sub>** CFU/cm<sup>2</sup> over the chicken fillet during storage period.

**Conclusion:** This work indicated that HPMC films incorporated with nanoparticles (~ 80 nm) at 80 ppm could enhance the safety of refrigerated chicken fillets.

**Keywords:** antimicrobial activity, HPMC edible film, nanoparticles, chicken fillets, cold storage.

## INTRODUCTION

Foodborne pathogen **are** one of the important biological hazards which causes a lot of **diseases**, harmful in food product leading to lose much money (16). According to Center for Disease Control and Prevention report, food-borne diseases account for approximately 48 million illnesses, 128000 hospitalizations and 3000 deaths cases, as well costed 15.6 billion \$ each year in the United States (9). Five foodborne pathogens record about (88%) of food poisons: *Norovirus* (26%), *Salmonella*

39 *nontyphoidal* (35%), *Campylobacter* (15%), *E. coli* (STEC) O157 (4%), and *Toxoplasma gondii* (8%).  
40 Moreover, twenty food products recalled in which exposure occurred in one state such as apple cider,  
41 bread, chicken, drink mix, ground beef, muffins, pork, raw tuna, and roast beef. (9).

42 Recently, nanotechnology have many applications in food sector particularly food industry, quality and  
43 safety (3). These applications used to improve food safety and extend shelf-life of food products (6).  
44 Nanoparticles one of the most types utilized in food safety as antimicrobial and supplementation. As  
45 well, inorganic nanoparticles as antimicrobial have taken more attention against food-borne  
46 pathogens i.e. aluminum oxide nanoparticles ( $Al_2O_3$ -NPs) and silica oxide nanoparticles ( $SiO_2$ -NPs)  
47 (15).

48  $Al_2O_3$ -NPs food grade are non-toxic, active against food-borne pathogens and permitted by FDA.  
49  $Al_2O_3$  NPs at  $1000\text{ mg ml}^{-1}$  significantly inhibits *Escherichia coli* growth in ready to eat foods (20). One  
50 study demonstrated  $Al_2O_3$ -NPs incorporated with polyvinylidene fluoride films reduced the *E. coli*  
51 growth (31). A study conducted by the author (27) reported that aluminum oxide nanoparticles were  
52 active against *Salmonella* Typhimurium, *Listeria monocytogenes*, *Fusarium oxysporum*,  
53 *Chromobacterium violaceum*, and *Aspergillus flavus*.

54 Food grade  $SiO_2$ -NPs are non- toxic, anticaking, has been used as food additive and permitted by  
55 FDA (7). Oregano silane containing  $SiO_2$ -NPs has been reported to prevented biofilm formation of  
56 food-borne pathogens (14).  $SiO_2$ -NPs reduce food-borne pathogens growth and make significate  
57 changes in cell morphology such as *Salmonella enterica* (30).

58 Hydroxy propyl methylcellulose (HPMC) edible film is approved by the by FDA for food packaging (21  
59 CFR 172.8741). It has a good characters such as tasteless and odorless, transparent, and barrier  
60 (29). As well, HPMC films including poly lactic acid and incorporated with green tea extract  
61 nanoparticles improved shelf-life of fatty foods (32). Additionally, HPMC films contained  $TiO_2$   
62 nanoparticles was reported to inhibit *E. coli* and *S. aureus* growth (24)

63 In Egypt, chicken product consuming growing up nowadays for many reasons, in my opinion that is  
64 due to highly nutrition value, easily absorption in human body, cheaply price, availability, and easy  
65 cooking.

66 The aims of this study were (a) Improve the quality and safety of chicken fillets; (b) development the  
67 packaging systems; (c) extending the shelf-life of chicken fillets; and (d) discovering a new  
68 antimicrobial

69 .

## 70 2. MATERIALS AND METHODS

### 71 2.1. Bacterial strains

72 Three bacterial strains utilized in this work were purchased from American Type Culture Collection  
73 (ATCC) *Bacillus cereus* (ATCC 10876), *Staphylococcus aureus* (ATCC 11988), and *S. Typhimurium*  
74 (ATCC 14028). The strains activated at Food Technology Department, Benha University, Egypt. All  
75 strains were cultivated twice on Tryptic Soy Agar (TSB; Bio-life company, Italy) at  $37\text{ }^\circ\text{C}$  for 24 h, and  
76 kept at  $4\text{ }^\circ\text{C}$  till using (17).

### 77 2.2. Antimicrobials agents

78 Food-grade aluminum oxide nanoparticles ( $\text{Al}_2\text{O}_3$ -NPs), and silica oxide nanoparticles ( $\text{SiO}_2$ -NPs) at  
79 (~80 nm) were obtained from Nanomaterial Laboratory, Beni-Suef University, Egypt.

### 80 2.3. Preparation of Hydroxy Propyl Methyl Cellulose (HPMC) films

81 Hydroxy propyl methyl cellulose films (HPMC) were prepared according to follow. Briefly, 4 % of  
82 HPMC was dissolved in 100 mL distilled water at 70 °C with stirring at 1000 rpm/min for 2 h. A 1 mL  
83 of glycerol 30% was added with stirring at 1000 rpm for 30 min. The nanoparticles were added and  
84 stirred at 1000 rpm/min for 15 min. The solution was sterilized at (121°C/15 min). Then, casted and  
85 dried, as well kept under cold storage till utilized (24).

### 86 2.4. Antimicrobial activity of nanoparticles against food-borne pathogens

87 Antimicrobial activity of nanoparticles was evaluated by disk diffusion method on tryptic soy agar  
88 media (TSA). In briefly, different concentration of nanoparticles i.e. 20, 40 and 80 ppm against food-  
89 borne pathogens. Add 10µl from bacterial strains. Then, 100µl from nanoparticles agent were added.  
90 Afterward, the dishes put in incubator at 37°C for 48 h. At the end of incubation time clear zones were  
91 appeared and measured by ruler (22).

### 92 2.5. Mode of nanoparticles action against bacterial strains

93 The mode of action was done according to (14) with slightly modification. Briefly, 2 ml of sterilized  
94 Tryptic Soy Broth (TSB) were added. 1 ml of bacterial strain and 1 mL of antimicrobial were added.  
95 After that, the tube was incubated at 37°C for 24 h. Then, the pellets were collected by centrifuge at  
96 2500 rpm for 10 min. Finally, all glass slides were prepared by washing by acetone and methanol,  
97 then spread the cells onto slides with drying at 37°C for 15 min and examining by scanning electron  
98 microscope.

### 99 2.6. Challenge study

100 Raw chicken fillets were purchased from local Cairo, Egypt. The fillets were transferred in ice box to  
101 laboratory, and freshly used. The fillets were cut down (5 × 5 cm) sections under sterilized conditions.  
102 Then, the samples treated with ultraviolet light (UV) at 260 nm for 15 min to decrease bacterial  
103 population. Chicken fillets were inoculated for 24 h by aseptically diluted cultures of *S. Typhimurium*,  
104 *S. aureus* and *B. cereus* approximately 5 log<sub>10</sub> CFU/cm<sup>2</sup> on the surface. After impregnation, the  
105 samples were kept at 25 ± 1 °C for 20 min to allow cell attachment. Then, raw chicken fillets were  
106 coated with HPMC films (5 × 5 cm) incorporated with nanoparticles. Control samples covered by  
107 control HPMC films. After 0, 3, 6, 9,12 and 15 days, the samples were tested to determine remain  
108 microbial colonies. 1mL was spread plated in duplicate onto brilliant green agar for *S. Typhimurium*,  
109 paired parker (M043) for *S. aureus*, *Bacillus cereus* agar base (M833) for *B. cereus* to demonstrate  
110 microbial growth. Resulting colonies were counted after 24:48 h incubation at 37°C, populations  
111 measured by log<sub>10</sub>, and expressed as log<sub>10</sub> CFU/cm<sup>2</sup> (19).

### 112 2.7. Scanning electron microscope (SEM) of HPMC films

113 Hitachi S-4700 scanning electron microscope (Hitachi, Toronto, Ontario, Canada) was used to study  
114 the morphology of nanoparticles and films. The samples were deposited onto aluminum specimen  
115 stubs using double-stick carbon tabs (Ted Pella Inc., Redding, CA, USA) and coated with  
116 gold/palladium on an ion sputter coated (Denton Vacuum Inc., Moorestown, NJ, USA) for 45 s at 20

117 mA. All samples were examined using an accelerating beam at a voltage of 1.5 kV. Magnifications of  
118 40,000x ;and 60,000x were used (10).

#### 119 2.8. Film solubility and thickness characterization

120 The solubility of films in water were studied. Thickness was determined by using digital micrometer  
121 model 7326 (Mitutoyo Manufacturing, Tokyo, Japan) at 6 different positions on the film according to  
122 (28).

#### 123 2.9. Tensile of HPMC films determination

124 The tensile of films were determined by Texture Analyzer TA.XT2 (Stable Micro System, Surrey, UK),  
125 according to the ASTM Standard Method D 88283 (initial grip separation = 50 mm and cross head  
126 speed = 100 mm/min) according to (10).

#### 127 2.10. Water vapor permeability

128 Water vapor permeability was evaluated by ASTM E96-92 gravimetric method with some  
129 modifications to measure the relative humidity (RH) of HPMC films according to (11). Water vapor  
130 permeability was calculated according to follow relation:  $WVP = \frac{WVTR}{(P_2 - P_3)} y$

131 Where WVTR was obtained from the slope of the weight loss rate through the film surface and  $p^2$  was  
132 the water vapor partial pressure on the film underside.  $p^3$  was water vapor partial pressure at the film  
133 underside,  $y$  the average film thickness. Water vapor permeability of each film was measured as the  
134 mean and standard deviations of 5 replications.

#### 135 2.11. Gases vapor permeability (O<sub>2</sub> and CO<sub>2</sub>)

136 The gas vapor permeability was determined at 30°C in a designed stainless cell by gas testing  
137 instrument, model Witt Oxybaby headspace gas analyzer (O<sub>2</sub>/CO<sub>2</sub>) following the method described by

138 following equation:  $P = \frac{Q \cdot X}{A \cdot t \cdot \Delta p}$

139 The gas permeability (P) was calculated according to (11).

140 Where, P is the permeability of gas, (m<sup>3</sup>/m. day. mmHg), Q is the quantity of gas diffused m<sup>3</sup>, X is the  
141 thickness of film, A an area of the film, m<sup>2</sup>, t is the time, day and  $\Delta p$  is the pressure difference across  
142 the film.

143

#### 144 2.12. Statistical analysis

145 The challenge study, statistical analyses for bacterial growth were carried out utilizing one-way  
146 ANOVA with a significance value of  $P \leq 0.05$  by using SPSS software, var. 18 (IBM; Armonk, N.Y.,  
147 U.S.A.). Results were analyzed as a completely randomized design according to (26). All challenge  
148 experiments were performed in triplicate, using 3 samples per treatment. Multiple comparisons were  
149 carried out applying least significant difference and Tukey's test.

### 150 3. RESULTS AND DISCUSSION

#### 151 3.1 Antimicrobial activity of nanoparticles against food-borne pathogens

152 As shown in **Table 1 and 2**. The antibacterial activity of inorganic nanoparticles i.e. aluminum oxide  
153 nanoparticles (Al<sub>2</sub>O<sub>3</sub>-NPs) and silica oxide nanoparticles (SiO<sub>2</sub>-NPs) against food-borne pathogens

154 such as *Bacillus cereus*, *Salmonella* Typhimurium and *Staphylococcus aureus* were evaluated. The  
155 result showed that Al<sub>2</sub>O<sub>3</sub>-NPs and SiO<sub>2</sub>-NPs (~80 nm) at 80 ppm were effective against food-borne  
156 pathogens i.e. *B. cereus*, *S. Typhimurium* and *S. aureus*, than 20 and 40 ppm respectively, the results  
157 were partially agreement with (8). Moreover, Al<sub>2</sub>O<sub>3</sub>-NPs were more active against *B. cereus* and *S.*  
158 *aureus* than *S. Typhimurium*, the results were partially agreement with author (12). In addition, SiO<sub>2</sub>-  
159 NPs were more active against *B. cereus*, and *S. Typhimurium* compared *S. aureus* that is not it at all  
160 by (13). the results indicated that, the Al<sub>2</sub>O<sub>3</sub>-NPs were more active against spores and gram positive  
161 than gram negative bacteria, while SiO<sub>2</sub>-NPs more effective against gram negative and spores  
162 compared with gram positive bacteria. The results are agreement with data reported by (4).

163 Furthermore, according to **Table 3**, the effect of hydroxy propyl methyl cellulose (HPMC) edible films  
164 incorporated with nanoparticles were decreased *B. cereus*, *S. aureus* and *S. Typhimurium* population  
165 growth. The results showed that Al<sub>2</sub>O<sub>3</sub>-NPs were inhibited *B. cereus* and *S. aureus* growth than *S.*  
166 *Typhimurium*. Although, SiO<sub>2</sub>-NPs less effective against *S. aureus* than *B. cereus*, and *S.*  
167 *Typhimurium*. the results were similar to the results obtained by (5).

### 168 3.2 Mode of action nanoparticles against foodborne pathogens

169 The mode of action it seems necessary because it presented all changesets in bacterial cells. **Fig. 1**,  
170 illustrated that Al<sub>2</sub>O<sub>3</sub>-NPs were highly effective against gram positive than gram negative bacteria, this  
171 is reverting to the Al<sub>2</sub>O<sub>3</sub>-NPs action as follows, Al<sub>2</sub>O<sub>3</sub>-NPs interact with bacteria membrane and made  
172 changes in cell morphology such as (a) the formation of 'pits' in their cell wall. Moreover, made  
173 disruption and drastic in cell wall. (b) As well, it produces reactive oxygen species (ROS) which allow  
174 to penetrate the cell membrane and led the cell to death. (c) Moreover, causes cell oxidative stress  
175 and formed free-radical scavenging that is led the bacteria to die that is reported by (18).

176 In addition to, SiO<sub>2</sub>-NPs more effective against gram negative and spores than gram positive bacteria.  
177 That is due to (a) the ability of SiO<sub>2</sub>-NPs to make morphological changes, lose the cell to preform it in  
178 function role. (b) As well, reactive oxygen spices (ROS) generation, and lose the DNA function and  
179 led to damage. (c) Additionally, cause the oxidative stress regulation in gens according to (15)

### 180 3.3 Challenge study

181 Based on the results of antimicrobial activity of HPMC films incorporated with nanoparticles, the films  
182 were utilized to cover raw chicken fillets at 4±1°C up to 15 days. **Fig. 2, 3, and 4**, reported that the  
183 bacterial population was gradually grew during the storage period over 15 days, when used control  
184 films compared with the nanoparticles films. HPMC films including nanoparticles reduced the food-  
185 borne pathogens growth approximately 2:3 log<sub>10</sub> during the challenge study.  
186 HPMC films include SiO<sub>2</sub>-NPs were stronger antimicrobial against *B. cereus*, *S. Typhimurium* and *S.*  
187 *aureus* than Al<sub>2</sub>O<sub>3</sub>-NPs on raw chicken fillets, these results are agreement with (18), (27) and (23).

### 188 3.4 Scanning electron microscope of HPMC films including nanoparticles agent

189 **Fig. 5** showed that, the cross sections and surface appearance of the control film, which appear to be  
190 homogeneous, smooth, colorless and free of any dimples or crevices. The HPMC films incorporated  
191 with nanoparticles were completely dispersion. Al<sub>2</sub>O<sub>3</sub>-NPs and SiO<sub>2</sub>-NPs loaded films show no pores  
192 with smooth surface. The presence of these pores is likely due to the flocculation and coalescence of

193 small drops during film preparation. Also, the nanoparticles distribution were found to be  
194 homogeneous in all films according to (1).

### 195 3.5 Mechanical properties of films

196 As shown in **Table. 4**, the tensile, water vapor permeability oxygen vapor permeability and carbon  
197 dioxide vapor permeability were evaluated, HPMC films containing SiO<sub>2</sub>-NPs were the highest values  
198 compared with HPMC films control and Al<sub>2</sub>O<sub>3</sub>-NPs films in mechanical properties. Additionally, SiO<sub>2</sub>-  
199 NPs increased the films water vapor permeability, carbon dioxide vapor permeability, tensile, oxygen  
200 vapor permeability and formed strong structure of films. That is due to (a) the ability of SiO<sub>2</sub>-NPs to fill  
201 the pores between the HPMC films structure (b) HPMC diffusion with SiO<sub>2</sub>-NPs and form  
202 homogenized structure (c) the ration of glycerol and it is ability to prevent water evaporation. As well,  
203 Al<sub>2</sub>O<sub>3</sub>-NPs were the lowest values and formed a weak structure, that is revert to the Al<sub>2</sub>O<sub>3</sub>-NPs can  
204 not interference with HPMC films and there is heterogenous distribution. In the control HPMC films,  
205 the transparence and thickness, was the lowest values than Al<sub>2</sub>O<sub>3</sub>-NPs and SiO<sub>2</sub>-NPs films. **That is**  
206 **refers to the colour** of nanoparticles and nanoparticles doses in films solution. Regarding solubility,  
207 there are non-significant results between HPMC films control and HPMC films including  
208 nanoparticles, **there are no pervious works in this point, but these results were similar to the results**  
209 **were obtained by other authors (2), (21) and (25).** Moreover, these results we had got from  
210 **experimental**

### 211 4. CONCLUSION

212 **The results of this investigation had demonstrated that HPMC films including Al<sub>2</sub>O<sub>3</sub>-NPs and SiO<sub>2</sub>-**  
213 **NPs were active against food-borne pathogens such as *S. Typhimurium*, *B. cereus* and *S. aureus* in**  
214 **chicken fillets. Additionally, nanoparticles (~80 nm) at 80 ppm showed a significant inhibition**  
215 **compared with 20 and 40 ppm respectively. Moreover, SiO<sub>2</sub>-NPs has a stronger antimicrobial activity**  
216 **against food-borne pathogens than Al<sub>2</sub>O<sub>3</sub>-NPs. However. HPMC films incorporated with SiO<sub>2</sub>-NPs **has****  
217 **improve** mechanical property than **HPMC films combined with Al<sub>2</sub>O<sub>3</sub> – NPs. HPMC films containing**  
218 **nanoparticles have the potentials to increase the shelf – life property and improve** chicken fillets  
219 safety and quality.

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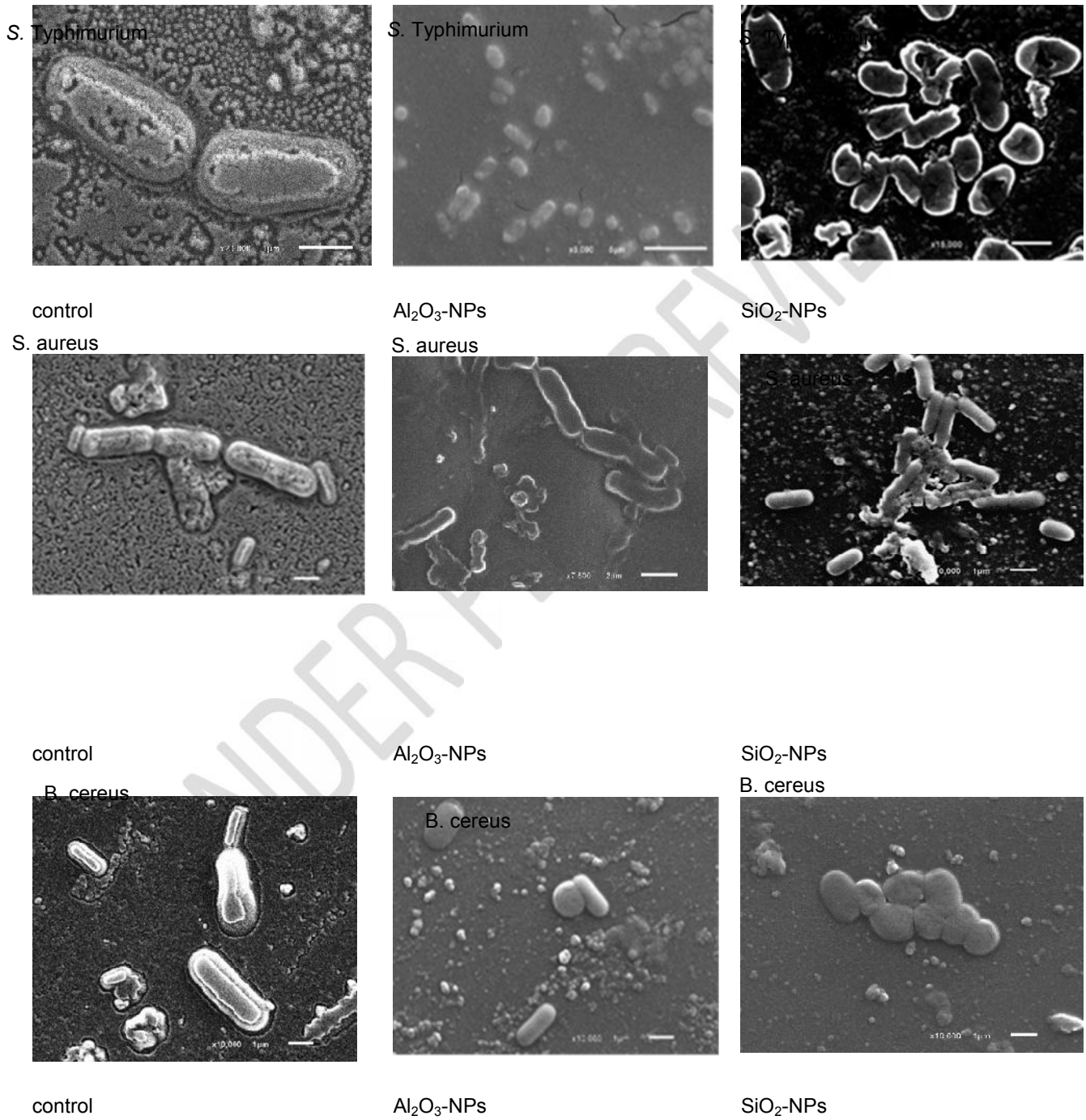
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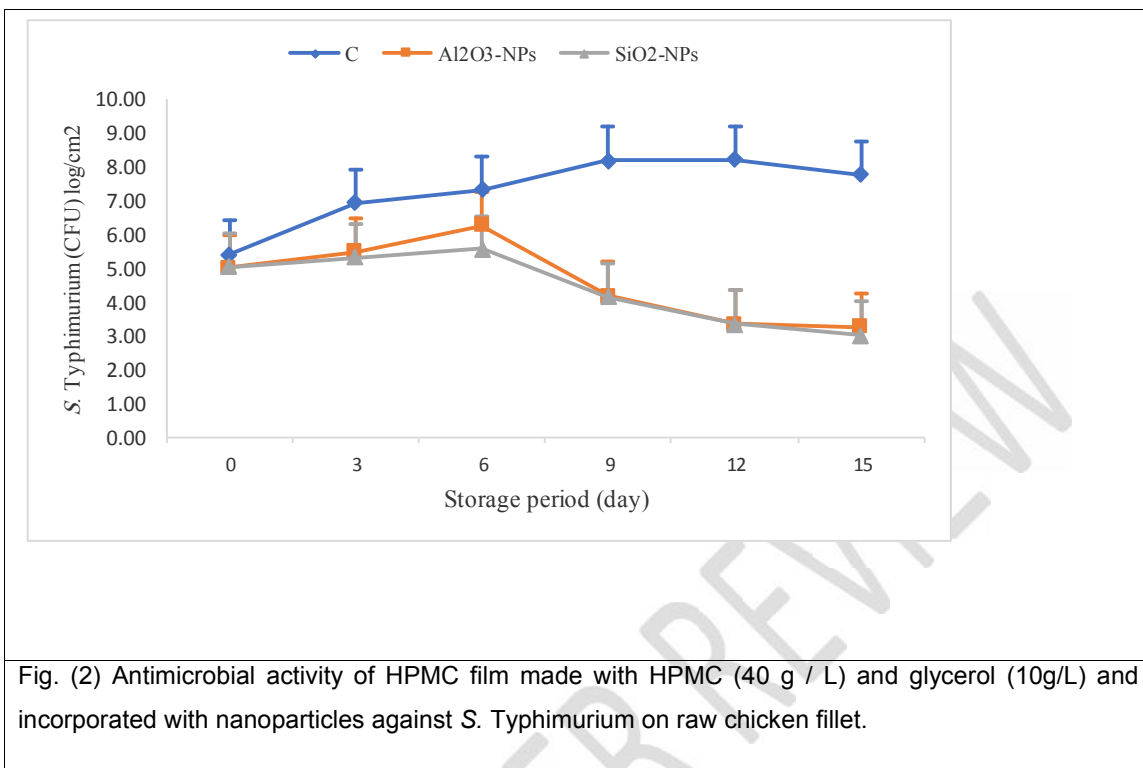


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337 Fig.1 The mode of action of nanoparticles against foodborne pathogens using SEM.



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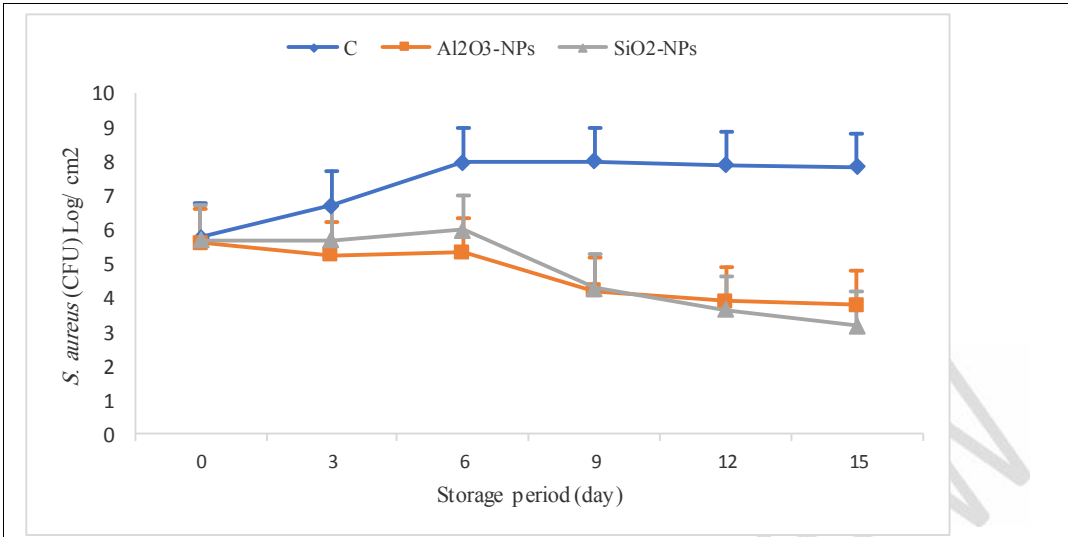


Fig. (3) Antimicrobial activity of HPMC film made with HPMC (40 g / L) and glycerol (10g/L) and incorporated with nanoparticles against *S. aureus* on raw chicken fillet.

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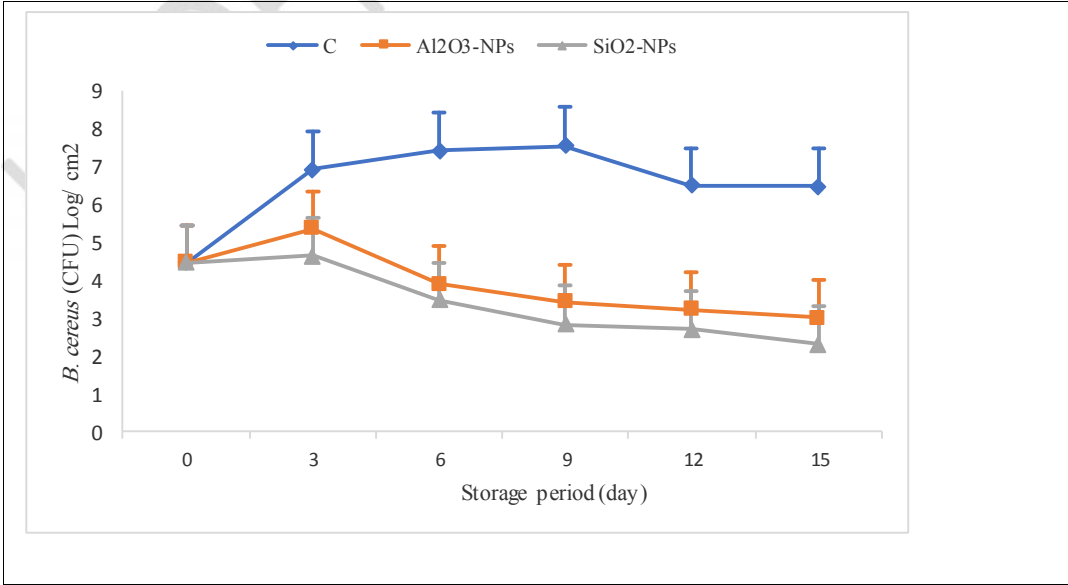
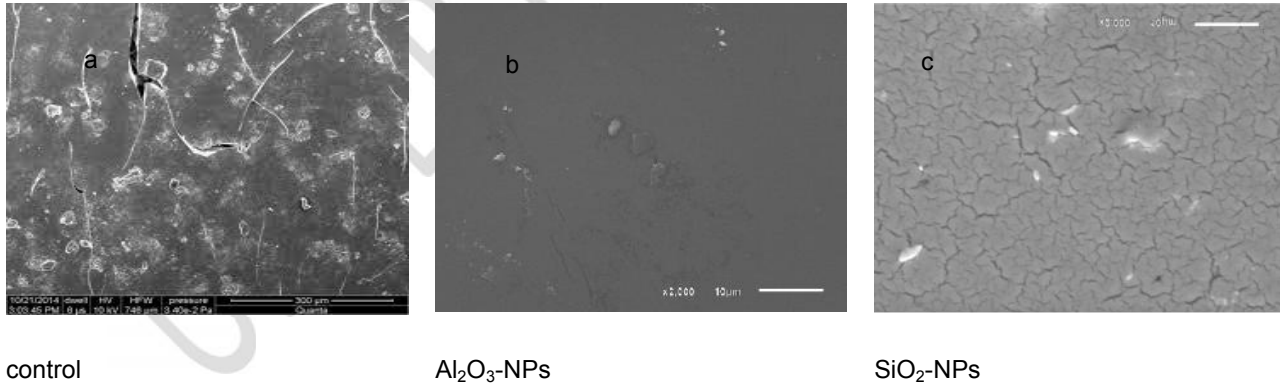


Fig. (4) Antimicrobial activity of HPMC film made with HPMC (40 g / L) and glycerol (10g/L) and incorporated nanoparticles against *B. cereus* on raw chicken fillet.

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384 Fig. (5) The SEM of (a) HPMC films incorporation (b) Al<sub>2</sub>O<sub>3</sub>-NPs and (c) SiO<sub>2</sub>-NPs.

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402 Table (1). Antibacterial activity of Al<sub>2</sub>O<sub>3</sub>-NPs and SiO<sub>2</sub>-NPs nanoparticles (~80 nm) at different  
403 concentration against foodborne pathogens.

Bacterial strains	Al <sub>2</sub> O <sub>3</sub> -NPs			SiO <sub>2</sub> -NPs		
	20 ppm	40 ppm	80 ppm	20 ppm	40 ppm	80 ppm
S. Typhimurium	9±0.3	11±0.3	13±0.2	11±0.3	15±0.2	18±0.3
S. aureus	8±0.3	12±0.3	14±0.3	12±0.3	13±0.3	16±0.3
B. cereus	ND	12±0.3	15±0.3	13±0.3	15±0.3	18±0.3

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ND: Not Detect

Al<sub>2</sub>O<sub>3</sub>-NPs: Aluminum oxide nanoparticles

SiO<sub>2</sub>-NPs: Silica oxide nanoparticles

410 Table (2). Antibacterial activity of Al<sub>2</sub>O<sub>3</sub>-NPs and SiO<sub>2</sub>-NPs nanoparticles (~80 nm) at 80 ppm against  
411 foodborne pathogens.

Bacterial strains	Nanoparticles agents	
	Al <sub>2</sub> O <sub>3</sub> -NPs	SiO <sub>2</sub> -NPs
S. Typhimurium	13±0.2	18±0.3
S. aureus	14±0.3	16±0.3
B. cereus	15±0.3	18±0.3

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413 Al<sub>2</sub>O<sub>3</sub>-NPs: Aluminum oxide nanoparticles

414 SiO<sub>2</sub>-NPs: Silica oxide nanoparticles

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416 Table (3). Antibacterial activity of HPMC film incorporation with nanoparticles (~80 nm) at 80 ppm

417 against foodborne pathogens.

Bacterial strains	HPMC films incorporation nanoparticles	
	Al <sub>2</sub> O <sub>3</sub> -NPs	SiO <sub>2</sub> -NPs
S. Typhimurium	16±0.2	22±0.4
S. aureus	17±0.3	20±0.3
B. cereus	18±0.3	22±0.4

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419 HPMC: Hydroxy propyl methyl cellulose

420 Al<sub>2</sub>O<sub>3</sub>-NPs: Aluminum oxide nanoparticles

421 SiO<sub>2</sub>-NPs: Silica oxide nanoparticles

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434 Table (4). Physical and mechanical properties of HPMC films incorporated with Al<sub>2</sub>O<sub>3</sub>-NPs and SiO<sub>2</sub>-NPs

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samples	Properties (tests results)						
	Tensile (MPa)	Water vapor permeability (g mm K <sup>-1</sup> Pa <sup>-1</sup> h <sup>-1</sup> m <sup>-2</sup> )	O <sub>2</sub> vapor permeability P (ml mm cm <sup>-2</sup> s <sup>-1</sup> cm Hg <sup>-1</sup> )	Co <sub>2</sub> vapor permeability P (ml mm cm <sup>-2</sup> s <sup>-1</sup> cm Hg <sup>-1</sup> )	Transparence	Thickness	Solubility
control	38.1	0.108	0.188×10 <sup>-8</sup>	2.25×10 <sup>-9</sup>	0.065	0.5 mm	100%
HPMC- Al <sub>2</sub> O <sub>3</sub> -NPs	31.6	0.056	1.074×10 <sup>-8</sup>	1.44×10 <sup>-9</sup>	0.079	0.5mm	100%
HPM -SiO <sub>2</sub> -NPs	43.17	0.541	2.17×10 <sup>-8</sup>	14.4×10 <sup>-9</sup>	0.082	0.51 mm	100%

436 HPMC: Hydroxy propyl methyl cellulose

437 Al<sub>2</sub>O<sub>3</sub>-NPs: Aluminum oxide nanoparticles

438 SiO<sub>2</sub>-NPs: Silica oxide nanoparticles

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UNDER PEER REVIEW