

4
5 ABSTRACT

6 **Aims:** To investigate the effect of food blends (plantain, soybean and ginger) on the blood
7 glucose, lipid profile and haematological indices on *streptozotocin* induced diabetic rats

8 **Methodology:** A total of 35 rats of mean body weight 219.07g separated into 7 groups (5 per
9 group) where induced by a single intraperitoneal (I.P) injection of *streptozotocin* (0.1g dissolved
10 in 5ml of freshly prepared sodium citrate buffer 0.1M, pH 4.5) at a dose of 40 mg/kg body
11 weight after fasting for 12 hours and fed with flours/blends. The flours were produced from plant
12 materials for different treatments/blends (blend A=100% unripe plantain, B=80% unripe plantain,
13 14% soybean, 6% ginger, C=70% unripe plantain, 26% soybean, 4% ginger, D= 60% unripe
14 plantain, 38% soybean, 2% ginger, E= 50% unripe plantain, 50% soybean) and the
15 phytochemicals and minerals content were determined. Blood glucose was determined at 5 days
16 interval for 25 days. Diabetes was confirmed in rats with blood glucose concentrations ≥ 200
17 mg/dl. After 25 days rats were anaesthetized with chloroform vapour and blood samples
18 collected by cardiac puncture for haematology and lipid profile determination.

19 **Results:** The results showed that unripe plantain, soya beans and ginger in adequate
20 proportion (C=70% unripe plantain, 26% soybean, 4% ginger or D= 60% unripe plantain, 38%
21 soybean, 2% ginger) could help to reduce blood glucose, improve haematological parameters and
22 lipid profile. Significant reduction was observed in the blood glucose level of rats fed blends C
23 and D from 286 to 85mg/dl and 307 to 90mg/dl respectively at the end of experiment. These
24 results also demonstrated that the inclusion of ginger at 6% causes rise in blood glucose
25 level. Total cholesterol (TC) increased in all the blends. However, the lowest concentration of TC
26 was observed in blends C and D. The highest packed cell volume (60%) and Haemoglobin
27 (20g/dl) level observed in rats fed blend C was significantly higher than the normal control fed
28 conventional feeds. The increase in packed cell volume (PCV) (50%) and Hb (17g/dl) in diabetic
29 rats demonstrated that the formulated blend C was able to raise PCV and Hb above 50% and
30 17g/dl (Normal control NC) respectively. Significant increase ($P < 0.05$) in low density
31 lipoprotein cholesterol (LDLc) was also observed in all the blends with blend C having the
32 least (4.0mg/dl) close to NC (2.0mg/dl).

33 **Conclusion:** From the results it is evident that blend C will manage and improve the health
34 status of diabetic patients.

35 **Key words:** Diabetes mellitus, streptozotocin (STZ), haematology, lipid profile, plant materials

36 **INTRODUCTION**

37
38 Diabetes mellitus has become a major global problem in our world today. It is a common
39 disorder associated with increased morbidity and mortality and can be defined as a group of
40 metabolic diseases characterized by chronic hyperglycemia due to defective insulin secretion,
41 insulin action, or both, resulting in impaired carbohydrate, lipid, and protein metabolism [1].

42 The combat against diabetes mellitus must be made a matter of top priority by all due to
43 the continual increase in the global prevalence of this social ill. Globally the prevalence was
44 estimated to increase in year 2000 to 2010 from 14.2 million to 17.5 million in North America,

45 15.6 million to 22.5 million in South America, 26.5 million to 32.9 million in Europe, 9.4 million
46 to 14.1 million in Africa, 84.5 million to 132.2 million in Asia and 1.0 million to 1.3 million in
47 Australia giving a total global increase in prevalence from 151 million people in 2000 to 221
48 million people in 2010 [2]. This was projected to 324 million by 2025 by Zimmet *et al.* [3] and 366
49 million 2030 [4]. In 2013, 382 million people had diabetes mellitus worldwide and this is
50 expected to rise to 592 million by 2035 [5]

51 The increasing interest in herbal medicine for the treatment of diabetes and many
52 prevailing diseases is not surprising. This may be attributed to the upsurge in cases of drug
53 resistance, cost and several side effects associated with most orthodox medicines. The use of
54 plant materials as spices, condiments and for medicinal purposes has therefore become more
55 popular and as such more plants materials such as plantain and soybean that have low
56 carbohydrate content with high mineral values are being exploited.

57 There is therefore no doubt that orthodox medicine itself appears to be strongly anchored
58 on traditional medicine [6]. The fact that the tropics into which majority of Africa lies is host to
59 about 2/3 of the world's flora and fauna means that a lot of medicinal plants can be found here
60 for both curative and management of diseases [7].

61 Plantain (*M. paradisiaca*) is a staple food crop in West Africa where its starchy fruits are
62 generally cooked or fried before consumption. During unripe plantain ripening, the starch is
63 changed to reducing sugars and sucrose. The medicinal value of plants have assumed a more
64 important dimension in the past decades owing largely to the discovery that their extracts contain
65 not only minerals but also a diverse array of secondary metabolites with antioxidant potentials
66 [8,9]. These antioxidants have been implicated in the therapeutic effects of several plants and
67 vegetables that are used in traditional medicine [10, 11]. Plantain contains a high fiber content,
68 and thus is capable of lowering cholesterol and helps to relieve constipation and hence
69 prevention of colon cancer. Besides, its high potassium content is found to be useful in the
70 prevention of rising blood pressure and muscle cramp [12]. Various parts of the plant such as the
71 leaves, root, fruit stalk, bract and fruit have been used for medicinal and domestic purposes.

72 Soybean is known as the "Golden bean" or the super legume of the twentieth century,
73 because it contains a good proportion of oil more than 20 percent. Soybean is also categorized as
74 oilseed, represents an excellent source of unsaturated fatty acids, high quality proteins and fiber.
75 Soybean contains very small amount of saturated fatty acid but do not contain any Trans fatty
76 acid. Both omega-6 and omega-3 fatty acids such as linoleic acid (56 % of total fat) and alpha
77 linolenic acid (7-8 % of total fat) are present in soybean. Cooked Soybeans are rich in iron,
78 phosphorous, magnesium, vitamin B2 (riboflavin) and folate. Kadamet *et al.* [13] stated that
79 legumes have been known as "a poor man's meat". They supply protein, complex carbohydrates,
80 fiber and essential vitamins and minerals to the diet, which are low in fat and sodium and contain
81 no cholesterol.

82 Spices are food adjunct commonly added to food to improve the sensory properties but
83 many spices have been observed to exert medicinal effects. Some spices which have been
84 reported to exert hypoglycemic effect both in laboratory animals and human subjects are:
85 Fenugreek seeds (*Trigonella foenum*), garlic (*Allium sativum*), Onion (*Allium cepa*), turmeric
86 (*Curcuma longa*), cumin seeds (*Curminum cuminum*), ginger (*Zingiber officinale*), mustard
87 (*Brassica nigra*), curry leaves (*Murrayakoenigi*) and coriander (*Coriandumsativum*)
88 [14].

89 Ginger is a perennial plant with narrow, bright green, grass-like leaves. It is cultivated in
90 the tropics for its edible rhizomes and has been found to be useful for both culinary and medicinal
91 purposes [15, 16]. Fresh ginger contains 80.9% moisture, 2.3% protein, 0.9% fat, 1.2% minerals,
92 2.4% fiber and 12.3% carbohydrates. The minerals presented in ginger are iron, calcium and
93 phosphorous. It also contains vitamins such as thiamine, riboflavin, niacin and vitamin C. The
94 composition varies with the type, variety, agronomic conditions, curing methods, drying and
95 storage conditions [17].

96 Several studies have reported the hypoglycemic effect of different forms of ginger in both
97 animals and human subjects. Among the fairly recent reports are: Arablouet *al.*[18]; Mozaffari-
98 Khosravi *et al.*[19] and Mahluji *et al.*[20] used ginger powder in Type 2 diabetic patients; Son *et*
99 *al.*[21] used 6-gingerol isolated from ginger in obese diabetic mice; Sukalingam *et al.*[22] used 6-
100 gingerol in STZ-induced diabetic rats; Abdulrazaq *et al.* [23] used aqueous ginger extract STZ-
101 induced diabetic rats; while Jafri *et al.* [24] used aqueous extract in alloxan-induced diabetic rats.
102 Very limited studies have reported the hypoglycemic effect of ginger juice while there is abject
103 scarcity of scientific findings on hypoglycemic effect of cooked ginger extract, which is highly
104 needed since the spice is mostly consumed in cooked forms in various cuisines. Hence, the
105 objective of this study is to determine the effect of food blends (plantain, soybean and ginger) on
106 the blood glucose, lipid profile and haematological indices on streptozotocin induced diabetic rats

107 MATERIALS AND METHODS

108 Materials

109 Unripe plantain and ginger roots were bought from Jattu market in Auchi, Edo State;
110 defatted soy bean flour (Variety TGX 1448-2E) was purchased from Benin City in Edo State.
111 Streptozotocin (STZ) Sigma NO SO130 was a product of Sigma-Aldrich chemical company,
112 UK. Every other chemical used were bought from Promise laboratory in Ekpoma, Edo State.

113 Processing of plantain flour:

114 Fresh unripe plantain was peeled, sliced using slicer and dried in an oven at 60°C for
115 48 hours. Dried sample was ground into powder (plantain flour).

116 Processing of soybean to defatted flour:

117 Soybean seeds were cleaned and sorted manually to remove dirt, leaves and stones. The
118 clean soybean seeds were coarsely milled to separate the coat from the cotyledon. The dehulled
119 seeds were milled to fine soybean flour using an attrition mill. The fine soybean flour was then
120 defatted using cold extraction with n-hexane. The defatted flour was then air-dried and the
121 clumps broken into fine flour, then sieved through a mesh screen.

122 Processing of ginger powder

123 Fresh ginger roots were sorted and washed to remove soil and other foreign materials
124 then sliced to thin layers and dried in an oven at 60°C for 24 hours before milling to powder.

125 Formulation of unripe plantain, soybeans and ginger flour blends:

126 Five samples were prepared from the combinations of unripe plantain, defatted soybean
127 and ginger as blends:
128
129

- 130 A=100% unripe plantain
 131 B=80% unripe plantain, 14% soybean, 6% ginger
 132 C=70% unripe plantain, 26% soybean, 4% ginger
 133 D= 60% unripe plantain, 38% soybean, 2% ginger
 134 E= 50% unripe plantain, 50% soybean

135 **Induction of Diabetes in Wister rats**

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 137 A total of 35 adult male albino rats with mean body weight of 219.07g were obtained
 138 from the disease free stock of the animal house, attached to Ambrose Alli University. The rats
 139 were separated into seven groups with five rats per group including NC and DC as follows in
 140 table 1.

141 **Table 1: Rat Groups and Treatments**

Groups	Number of rats	Treatments
A	5	STZ-induced diabetic rats fed with 100% unripe plantain),
B	5	STZ-induced diabetic rats fed with 80% unripe plantain, 14% soybean, 6% ginger),
C	5	STZ-induced diabetic rats fed with 70% unripe plantain, 26% soybean, 4% ginger) and
D	5	STZ-induced diabetic rats fed with 60% unripe plantain, 38% soybean, 2% ginger)
E	5	STZ-induced diabetic rats fed with 50% unripe plantain and 50% soybean),
NC	5	Not induced and fed with rat pellet
DC	5	Induced and fed with rat pellet).

142
 143 Prior to experimentation, the rats were acclimatized to laboratory condition and fed with
 144 rat pellet and water ad libitum for a week. Diabetes was induced in rats by a single intraperitoneal
 145 (I.P) injection of freshly prepared solution of streptozotocin (0.1g dissolved in 5ml of freshly
 146 prepared sodium citrate buffer 0.1M, pH 4.5) at a dose of 40 mg/kg body weight after fasting for
 147 12 hours. Good hygiene was maintained by constantly cleaning and removal of faeces and spilled
 148 feeds from cages daily. Fasting blood glucose (FBG) was determined using Accucheck Active
 149 glucometer, Roche Germany, with blood obtained from the tail vein of the rats. This test was
 150 repeated on day 5, 10, 15, 20 and 25. Diabetes was confirmed in STZ treated rats with blood
 151 glucose concentrations ≥ 200 mg/dl.

152
 153
 154 **Collection and analysis of blood**

155 The rats were anaesthetized with chloroform vapour, twelve hours(12 h) after last day of feed
156 administration, and blood samples were collected by cardiac puncture into a set of plain and
157 fluoride oxalate sample bottles.

158

159 **Hematological parameters**

160 The packed cell volume (PCV) was measured by the micro hematocrit centrifuge.
161 Hemoglobin (Hb) concentration was determined by the cyanomethemoglobin technique [25].
162 The white blood cell components were also determined.

163

164 **Lipid Profile Studies**

165 Blood sample was centrifuged to collect plasma which was used to estimate total
166 cholesterol, high density lipoprotein cholesterol (HDL-C), low density lipoprotein cholesterol
167 (LDL-C), and triglycerides (TG) using commercial kits obtained from Randox Laboratories, UK.

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RESULTS AND DISCUSSION

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Mineral Composition of Formulated Food

171 Table 2 depicts the composition of the studied minerals. Food blend E had the highest
172 potassium content (1099.42ppm), this was followed by D (944.79ppm) while the lowest
173 potassium content was observed in A (704.80ppm). The highest potassium observed in food blend
174 E could be attributed to its high inclusion of soybean (50%) which is known to be a rich source
175 of potassium. Potassium is an important mineral in the body that regulates fluid balance, muscle
176 contraction and nerve signals. High potassium may reduce blood pressure and water retention,
177 protect against stroke and prevent osteoporosis and kidney stones.

178 Food blend A had the highest sodium content (75.65ppm), this was followed by B
179 (67.19ppm) while the lowest content (47.80ppm) was observed in E. sodium is essential for life. It
180 helps to control the body's fluid balance. It send nerve impulses and affects muscle function.

181 Food blend E had the highest calcium content (804.02ppm), this was followed by D
182 (626.91ppm) and C (435.71ppm) while the lowest calcium content (236.16ppm) was observed in
183 food blends A. calcium plays an important role in muscle contraction, transmitting messages
184 through the nerves and the release of hormones. Calcium is also important mineral in the
185 formation of teeth and bones

186 Food blend E had the highest iron content (141.49ppm), this was followed by D
187 (121.42ppm) and C (114.64ppm) while the lowest content (28.60ppm) was observed in food
188 blends A. Iron is an important component of haemoglobin, the substance in red blood cell,
189 responsible for carrying oxygen and transports it throughout the body.

190 The mineral content (potassium, calcium and iron except sodium) of the blends, increased
191 with increasing soybean inclusion level (Table 2), depicting that soybean is rich in these minerals.

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193

194

Table 2:- Mineral composition of formulated food blends

Blends	Minerals (ppm)				195
	Potassium	Sodium	Calcium	Iron	
A	704.80 ^a	75.65 ^a	236.16 ^c	28.60 ^e	
B	931.82 ^b	67.19 ^b	430.77 ^d	92.89 ^d	
C	942.17 ^c	66.00 ^b	435.71 ^c	114.64 ^c	
D	944.79 ^b	62.08 ^c	626.91 ^b	121.42 ^b	
E	1099.42 ^a	47.80 ^d	804.02 ^a	141.49 ^a	
SEM	0.05	0.54	0.06	0.05	

196 Means with the same letters down the column are not significantly different (P>0.05)

197 A=100% unripe plantain

198 B=80% unripe plantain, 14% soybean, 6% ginger

199 C=70% unripe plantain, 26% soybean, 4% ginger

200 D= 60% unripe plantain, 38% soybean, 2% ginger

201 E= 50% unripe plantain, 50% soybean

202 SEM= Standard error of mean

203

204 **Phytochemical Properties**

205 Table 3 shows the phytochemical compositions of the blends. The lowest tannin content
 206 (tannin 0.27mg/100g) was observed in food blend A and was followed by blend B (0.55
 207 mg/100g). Blends C, D and E had the same tannin content (0.61mg/100g).

208 The highest alkaloid content (6.43%) was observed in blend A and was followed by B
 209 (6.23%), C (5.99%), D (5.75%), and E (4.84%) in that decreasing order.

210 Blends B and C had the same flavonoid content(0.42 mg/100g) which was higher than
 211 the other blends. The lowest flavonoid content (0.11mg/100g) was observed in blend A; this was
 212 followed by E(0.31mg/100g) and D (0.35mg/100g).

213 Blend A (0.16 mg/100g) had the lowest saponin content and was followed by B
 214 (2.39mg/100g), C (3.99mg/100g), D (4.22mg/100g), and E (6.33mg/100g) in that decreasing
 215 order.

216 Saponins are known to possess both beneficial(cholesterol lowering) and deleterious
 217 (cytotoxicpermeabilization of the intestine and paralysis of the sensory system) properties [26].
 218 Flavonoids, alkaloids and tannins are polyphenolic compounds with antioxidant properties. In
 219 addition, phenolic compounds existing in plants are also responsible for their contribution to
 220 colour, sensory and antioxidant properties of food [27].

221 The low phytochemical values (Table 3) recorded in this study are significantly lower than
 222 (P<0.05) the results of Elezuet *al.*[28] who recorded significant values (saponin 1.827, flavonoid
 223 0.981 and tannin 1.577) in unripe plantain flour. However, he further reported that the levels of
 224 saponin in the flour are quite too low to cause any deleterious effects.

225 **Table 3:- Phytochemical Properties of formulated food blends**

Blends	Phytochemicals			
	Tannin (mg/100g)	Alkaloids (%)	Flavonoids (mg/100g)	Saponin (mg/100g)

A	0.27 ^c	6.43 ^a	0.11 ^d	0.16 ^c
B	0.55 ^b	6.23 ^b	0.42 ^a	2.39 ^d
C	0.61 ^a	5.99 ^c	0.42 ^a	3.99 ^c
D	0.61 ^a	5.75 ^d	0.35 ^b	4.22 ^b
E	0.61 ^a	4.84 ^e	0.31 ^c	6.33 ^a
SEM	0.008	0.014	0.005	0.008

226 Means with the same letters down the column are not significantly different (P>0.05)

227 A=100% unripe plantain

228 B=80% unripe plantain, 14% soybean, 6% ginger

229 C=70% unripe plantain, 26% soybean, 4% ginger

230 D= 60% unripe plantain, 38% soybean, 2% ginger

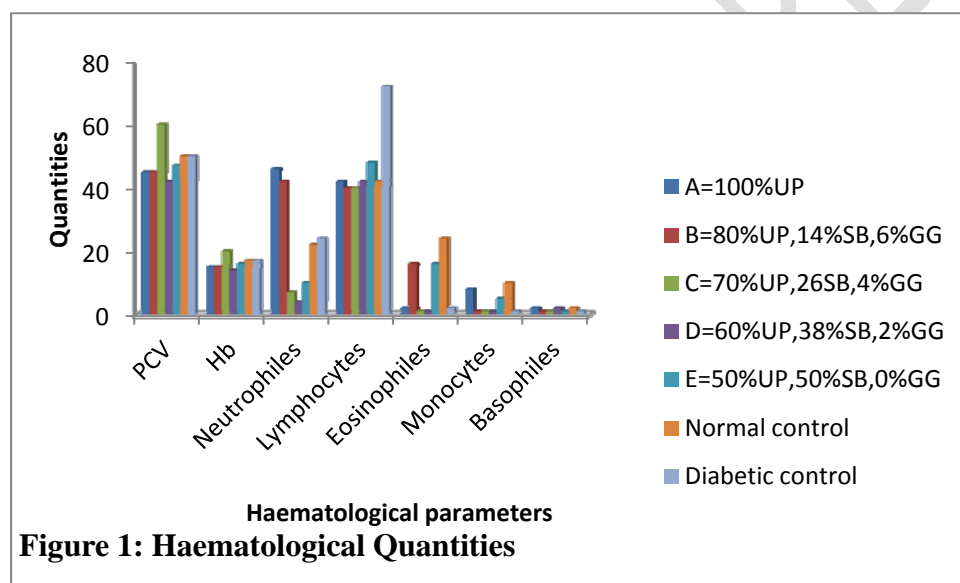
231 E= 50% unripe plantain, 50% soybean

232 SEM= Standard error of mean

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246 Food and nutrients play vital role in the normal functioning of the body. In this study,
 247 plant materials such as unripe plantain, soybean and ginger were used to formulate food blends
 248 with the aim of studying its effect on the haematological parameters, lipid profile and blood
 249 glucose level of diabetic rats.

250 The analysis of variance showed significant difference (P<0.05) in the packed cell
 251 volume (PCV) and haemoglobin (Hb) level of the diabetic rats (Figure 1). The highest PCV and
 252 Hb level (60%, 20g/dl respectively) were observed in rat fed blend C that contains 70% unripe
 253 plantain, 26% soybean and 4% ginger. This was significantly higher (P<0.05) than the
 254 normal control (not induced) (50%, 17g/dl) fed conventional feeds. The increase in PCV and

255 Hbin diabetic ratsshowed that the formulated blends were able to raise the PCV and Hb above
 256 50% and 17g/dl.

257 The degree of anemia in diabetic patients can be associated with a number of factors such
 258 as glomerular filtration rate andglycated h (HbA1c) level. Thomas *et al.*[29] reported that anemia
 259 is due to diminished erythropoietin production by failing kidneys and increased non enzymatic
 260 glycosylation of red blood cell (RBC) membrane protein. In this study, increase in PCV and Hb
 261 level of some of the diabetic rats does not depict occurrence of anemia rather shows its potency
 262 in the management of the ailment (diabetes). This could be attributed to the phytochemicals and
 263 mineral present in the blends. The antioxidant properties of these phytochemicals especially
 264 flavonoids have been reported in several studies. Onatet *al.*[30] reported the anti-sickling
 265 properties. This according to Palaciouset *al.* [31] it prevents oxidation of RBC and Hb that often
 266 lead to haemolysis. According to Egunyomiet *al.* [32] it may also stimulate formation or
 267 secretionof erythroprotein in the stem cells of the animals as evidenced by the increased level of
 268 PCV and Hb. There is no significant difference ($P < 0.05$) in the lymphocytes of the formulated
 269 blends (A and D) from the normal control. The diabetic control rat had lymphocytes (72%)
 270 significantly higher ($P < 0.05$) than every otherrat. The high lymphocytes level could be attributed
 271 to unknown infection.The values of Neutrophiles, Ecsinophiles, Basophiles and
 272 Monocytesobtained in rats fed with blends C, D and E were significantly lower ($P < 0.05$) than the
 273 normal control rats.

274 **Changes in Blood Glucose and Body Weight of Streptozotocininduced Wister Rats**

275 **Table 4: Blood glucose of Streptozotocin rats**

Food blends	Days						SEM
	0	5	10	15	20	25	
A	104.40 ^{mno}	272.00 ^h	334.00 ^e	504.00 ^b	409.00 ^c	413.20 ^c	
B		93.00 ^{nop}	245.00 ⁱ	301.00 ^f	562.00 ^a	559.00 ^a	506.00 ^b
C	107.00 ^{lmn}	286.00 ^g	73.00 ^q	99.00 ^{lmno}	101.00 ^{mno}	85.00 ^p	
D		103.00 ^{lmno}	307.00 ^f	99.00 ^{mno}	114.00 ^{lm}	103.00 ^{lmno}	90.00 ^{op} 4.15
E	109.00 ^{lmn}	247.00 ⁱ	370.00 ^e	392.00 ^d	375.00 ^e	402.00 ^{cd}	
NC	108.00 ^{lmn}	110.00 ^{lmn}	133.00 ^k	109.00 ^{lmn}	106.00 ^{mno}	103.00 ^{lmno}	
DC	120.00 ^l	229.0 ^j	184.00 ^k	214.00 ^k	225.00 ^{jk}	283.00 ^{gh}	

276 Means with same superscript down the column and along the row are not significantly different
 277 ($P > 0.05$)

- 278 A=100% unripe plantain
- 279 B=80% unripe plantain, 14% soybean, 6% ginger
- 280 C=70% unripe plantain, 26% soybean, 4% ginger
- 281 D= 60% unripe plantain, 38% soybean, 2% ginger
- 282 E= 50% unripe plantain, 50% soybean
- 283 NC= Normal control, fed conventional feed (not induced)
- 284 DC= Diabetic control, fed conventional feed (induced)

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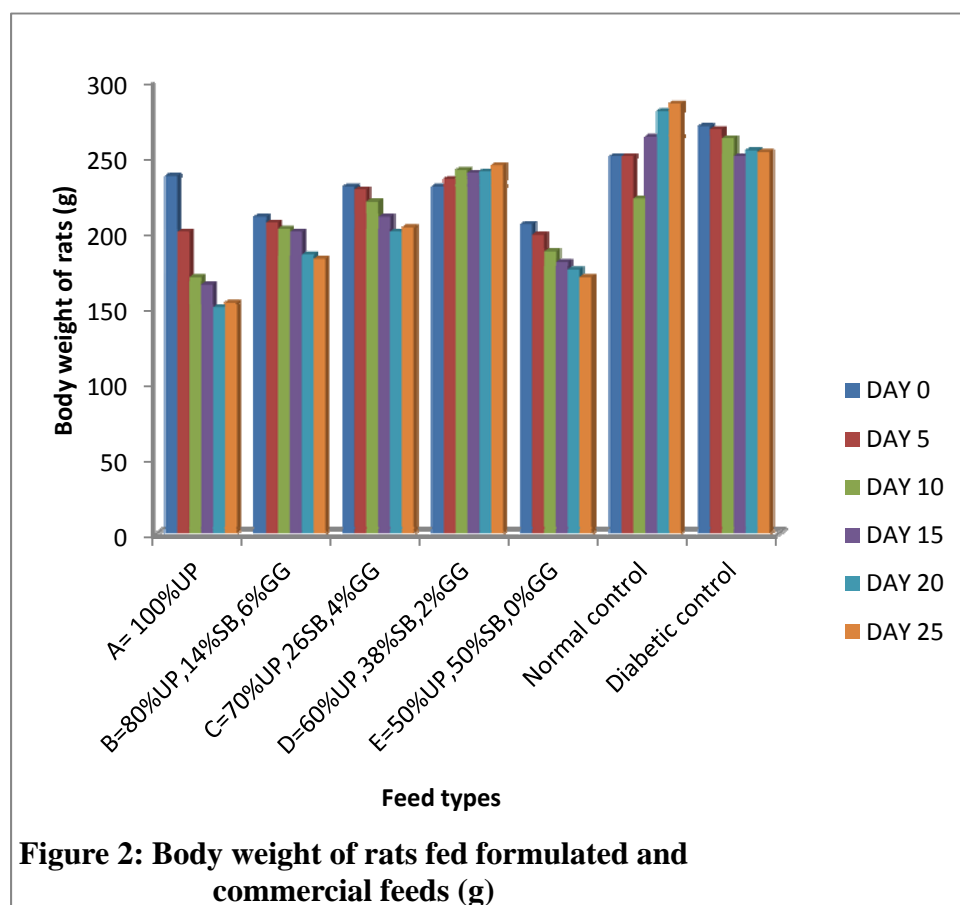


Figure 2: Body weight of rats fed formulated and commercial feeds (g)

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299 Blood glucose and body weight were monitored for total duration of 25 days. At 5 days
 300 interval blood glucose level and body weight were determined. The initial measurements were
 301 taken before induction at day 0 for glucose level and body weight. The various rat groups had
 302 blood glucose level between 93-120mg/dl and body weight between 205-270g (day 0). They were
 303 induced and fed formulated food blends and water ad libitum.

304 On the 5th day, all the induced groups had significant increase ($P < 0.05$) in glucose
 305 level > 200 mg/dl (Table 4) with corresponding decrease in body weight (Figure 2). Thus the rats
 306 were considered diabetic at ≥ 200 mg/dl. The rat fed blend D had the highest blood glucose level
 307 307mg/dl. NC rats had the lowest blood glucose level (110.0 mg/dl) and showed no significant
 308 ($P > 0.05$) change throughout the period of experiment.

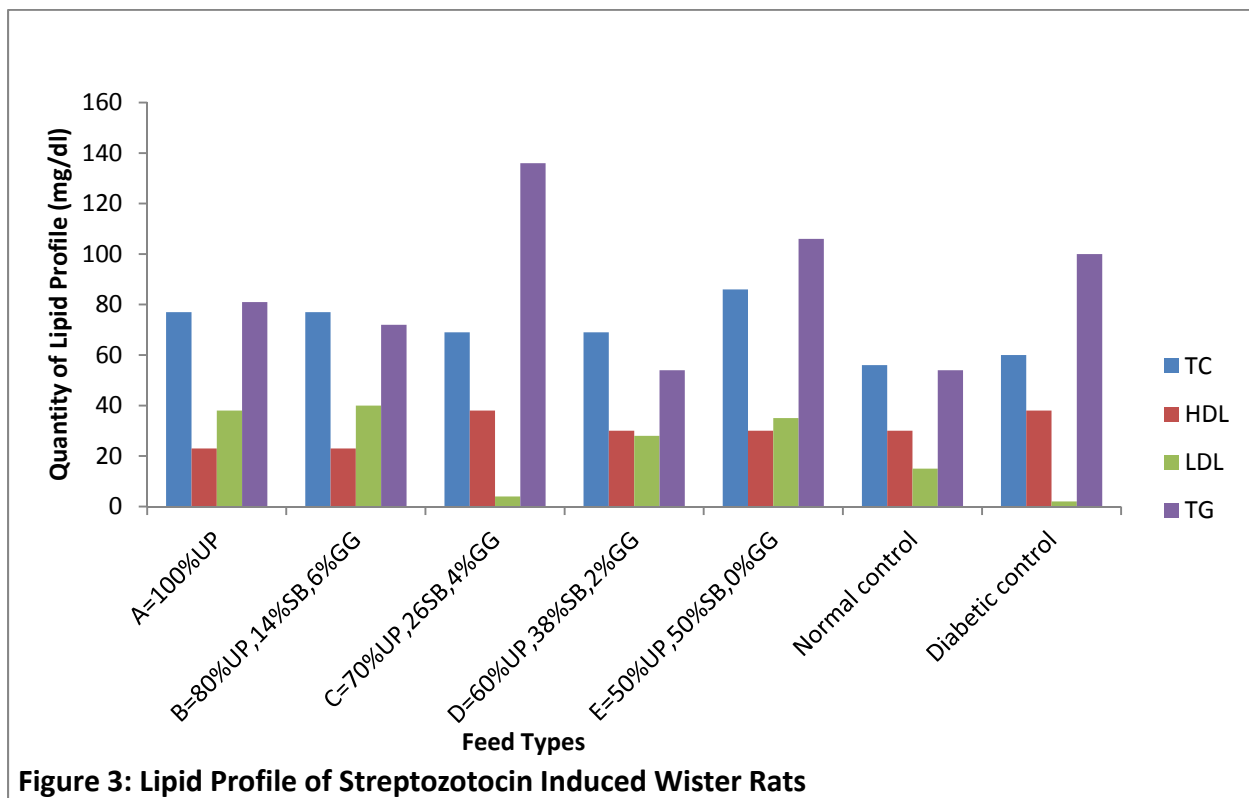
309 There was steady significant increase ($P < 0.05$) in the blood glucose of group A, B, E and
 310 DC throughout the period of this experiment. The results showed that at 0% and 6% inclusion of
 311 ginger in blends A and B respectively, the rats were hyperglycemic. This demonstrated that the
 312 inclusion of ginger at 6% causes rise in BGL. Significant reduction ($P < 0.05$) was observed in the
 313 blood glucose level of rats fed with blends C and D from 286 to 73mg/dl and 307 to 99mg/dl
 314 from day 5 to 10 respectively. This same trend was observed in blood glucose level for 15, 20 and
 315 25 days with rats fed blends C (99, 101 and 85mg/dl) and D (114, 103 and 90mg/dl) respectively,

316 having normal blood glucose <200mg/dl. This shows the potency of the blends C (70% unripe
317 plantain, 26% Soybean and 4% ginger) and D (60% unripe plantain, 38% soybean and 2%
318 ginger) in the management of the ailment (diabetes). This could be attributed to the combination
319 levels of the plant materials particularly the inclusion of ginger at 4% and 2% in blend C and D
320 respectively. Ginger provides an amount of potassium that could help stroke and diabetes and
321 adult requires 2000mg of potassium each day. Potassium is important for diabetic patients and
322 those at the risk of it. The findings of recent study published by researchers from university of
323 Sydney in 2012 revealed that ginger extract helps to increase cell absorption of glucose even
324 independent of insulin (www.naturalnews.com). The predominant pungent compound in ginger is
325 responsible for its benefit to humans [33]. According to Andalluet *al.* [34] ginger has a
326 therapeutic benefit of lowering fasting serum blood glucose level in Type 2 diabetes. According
327 to Singh *et al.* [35] many of the putative activities of ginger (antioxidant, anti-inflammatory,
328 hepatoprotective, antiobesity) are often associated with the etiology and pathophysiology of
329 Type 2 diabetes, which suggest the possibility that ginger may not have a direct effect on
330 diabetes but acts indirectly by suppressing factors that lead to impaired glucose control. Thus,
331 was supported by a study showing that ginger root powder (200mg/kg body weight) in type 2
332 diabetic rat model reversed symptoms of metabolic syndrome, blood glucose, blood lipid and
333 decreased oxidative stress [36]. Although blend B had ginger inclusion at 6%, the glucose level
334 was >200mg/dl throughout the period of this experiment. This shows that ginger inclusion at 6%
335 could result in hyperglycemic condition. However at day 10, a rise was observed in the glucose
336 level of rats fed blend A (272-334mg/dl), B (245-301mg/dl) and E (247-370mg/dl) with
337 corresponding decrease in body weight (Figure 2). Thus this indicates that the formulation for A,
338 B and E could not control the diabetic condition. The DC rats fed with conventional rat feed
339 increased in blood glucose level and body weight steadily throughout the period of experiment,
340 while the body weight decreased from 228 to 220g and increased from 235 to 241g in rats fed
341 blends C and D respectively.

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Effect of Food Blends (Diet) on Serum Lipid Profile



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Figure 3: Lipid Profile of Streptozotocin Induced Wister Rats

359 Serum lipid concentration of streptozotocin induced rats fed with food blends and
360 conventional feed in this study is shown in figure 3. From the results, serum high density
361 lipoprotein cholesterol (HDLc) concentration in rats fed blends C (38mg/dl) and NC
362 (38mg/dl) were same but higher and significantly different ($P < 0.05$) from HDLc of rats fed with
363 other blends.

364 Total cholesterol (TC) increased in all the blends. However, the lowest concentration in
365 TC was observed in blends C and D (Figure 3). Thus this depicts that blends C (69mg/dl) and D
366 (69mg/dl) are better having lower cholesterol concentration.

367 For low density lipoprotein cholesterol (LDLc) significant increase ($P < 0.05$) was
368 observed in all the blends. However, blend C (4.0mg/dl) was next to NC (2.0mg/dl) while the
369 highest was observed in blend B (40.0mg/dl).

370 The increased in LDLc, TC, and decreased in HDLc agrees with the findings of
371 Adaramoye *et al.* [37] for diabetic rats. Besides, the formulated diets are plant materials
372 containing phytochemicals. [38] reported that action of plant extract in reducing plasma

373 cholesterol concentration could be due to the ability of one or more of the phytochemicals in the
374 plant to activate the functioning enzymes of the rats responsible for cholesterol absorption.

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CONCLUSION

379 In this research work, it was observed that the blends of unripe plantain, soya beans and
380 ginger in adequate proportion (C=70% unripe plantain, 26% soybean, 4% ginger or D= 60%
381 unripe plantain, 38% soybean, 2% ginger) could help to reduce blood glucose, improve
382 haematological parameters and lipid profile. The mineral content (potassium, calcium and iron
383 except sodium) of the blends increased with increasing soybean inclusion level, depicting that
384 soybean is rich in these minerals. Significant reduction ($P < 0.05$) was observed in the blood
385 glucose level of rats fed blends C and D from 286 to 85mg/dl and 307 to 90mg/dl respectively.
386 The lowest concentration of TC was observed in blends C and D. This depicts that blends C
387 (69mg/dl) and D (69mg/dl) are better and preferred to the other blends. In addition, blend C also
388 had the least value (4.0mg/dl) of low density lipoprotein cholesterol (LDLc). Hence, blend C
389 is most preferred to prevent and control diabetes as well as improve the health status of diabetic
390 patients.

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Ethical Approval:

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394 As per international standard or university standard ethical approval has been collected
395 and preserved by the authors.

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REFERENCES

- 399 1. Akah JA, Lemji JA, Salawa OA, Okoye TC, Offiah NV. (2009). Effects of
400 *Vernonia amygdalina* on Biochemical and Haematological Parameters in Diabetic
401 Rats. Asian Journal of Medicinal Science. 1(3): 108-113.
- 402 2. Amos A, McCarty D, and Zimmet P. (1997). The rising global burden of diabetes and
403 its complications: estimates and projections to the year 2010. Diabetic Med. ;14: S1-S85
- 404 3. Zimmet P, Shaw J, and Albert KG. (2003). Preventing Type 2 diabetes and the
405 dysmetabolic syndrome in the real world: a realistic review. Diabet. Med.; 20: 693-702
- 406 4. Wild S, Roglic G, Green A, Sicree R, and King H. (2004). Global prevalence of diabetes:
407 Estimates for the year 2000 and projections for 2030. Diabetes Care; 27 (5): 1047-1053.
- 408 5. Guariguata L, Whiting DR, Humbleton I, Beagley J, Linnenkamp U, and Shaw JE.
409 (2014). Global estimates of diabetes prevalence for 2013 and projections for 2035.

- 410 Diabetes Research and Clinical Practice; 103 (2): 137-149.
- 411 6. Nweze EI. (2009). Justification for the use of *Ocimum gratissimum* in herbal medicine and
412 its alteration with disc antibiotics. www.biomedicine.com.
- 413 7. Sofowora A. (1993). Medicinal Plants and Traditional Medicine in Africa. John Wiley
414 and Sons Ltd. Pp. 33-34.
- 415 8. Akinmoladun AC. Ibukun EO. Afor E. Akirinlola B.L. Onibon T.R. Akinboboye
416 A.O.Obuotor EM. Farombi EO. (2007). Chemical constituents and antioxidant activity
417 of *Alstonia boonei*. *Afr. J. Biotechnol.* 6(10): 1197-1201.
- 418 9. Ahenkora K. Kyei MA. Marfo EK. Banful B. (1998). Nutritional composition of false
419 horn plantain during ripening and processing. *J. Food Chem.* pp. 455- 458.
- 420 10. Kumar RS. Sivakuma T. Sunderem RS. Gupta M. Murujesh K. Rajeshwa Y. Kumar MS.
421 Kumar KA. (2005). Antioxidant and antimicrobial activities of *Bauhinia Recemosa* L Stem
422 Bark. *J. Med. Biol. Res.* 38: 1015 -1024.
- 423 11. Marthur NK. and Marthur V. (2001). Antioxidants: Natural ingredients and additives for
424 food. *Beverage Food World*, 5: 13-15.
- 425 12. Ng SP. Fong CS. (2000). Banana enhances your anti-cancer power. In: Health
426 discovery. Petaling Jaya Malalaysia: Life Publisher Berhad. of cocoyam
427 (*Colocassia esculenta*) leaf with some green leafy vegetables. *Nig J. Nutr Sci.* 27 (1), 22 –
428 26.
- 429 13. Kadam ML., Salve RV. Mehrajfatema ZM. and More SG. (2012). Development
430 and evaluation of composite flour for messi roti/ chapatti. *Journal of Food Process*
431 *Technology.* 3(1): 01-07.
- 432 14. Srinivasan K. (2005). Plant foods in the management of diabetes mellitus: Spices as
433 beneficial anti diabetic food adjunct. *International Journal of Food Science and Nutrition*;
434 56 (6): 399-414.
- 435 15. Grant KI. (2000). Ginger. *Am. J. Health Sys. Pharm.*; 57: 945-957.
- 436 16. Ursell A. (2000). The complete guide to healing foods: pp 112-114, Dorling Kindersley
437 Ltd, London.
- 438 17. Suekawa M. Ishige A. Yuansa K. Sudo K. Aburada M. and Hosoya E. (1984).
439 Pharmacological studies on ginger- Pharmacological actions of pungent constituents of
440 6-gingerol and 6-shogaol. *J. Pharmacobiodyn*; 7: 836-848.
- 441 18. Arablou T. Aryaeian N. Valizadeh M. Shariffi F. Hosseini A and Djalali M. (2014). The
442 effect of ginger consumption on glycemic status, lipid profile and some
443 inflammatory markers in patients with Type 2 diabetes mellitus. *International Journal of*
444 *Food Sciences and Nutrition*; 65 (4): 515-520.
- 445 19. Mozaffari-Khosravi H. Taleai B. Jalali B-A. Najarzadeh A. and Mozayan MR. (2014).
446 The effect of ginger powder supplementation on insulin resistance and glycemic indices
447 in patients with Type 2 diabetes: A randomized double-blind placebo-controlled
448 trial. *Complementary Therapies in Medicine*; 22 (1): 9-16.
- 449 20. Mahluji S. Attari VE. Mabassori M. Payahoo L. Ostadrahimi A. and Golzari SEJ
450 (2013). Effect of ginger (*Zingiber officinale*) on plasma glucose level, HbA1c and
451 insulin sensitivity in type 2 diabetic patients. *International Journal of Food Sciences and*
452 *Nutrition*; 64 (6): 682-686.
- 453 21. Son MJ. Miura Y. and Kazum Y. (2014). Mechanism of anti diabetic effect of gingerol in
454 cultured cells and obese diabetic model mice. *Cytotechnology*.

- 455 22. Sukalingam K.Ganesan K. and Gani S.B. (2013). Hypoglycemic effect of 6-gingerol,
456 anactiveprinciple of ginger in streptozotocin-induced diabetic rats. *Journal of*
457 *Pharmacology andToxicological Studies*; 1 (2): 23-30.
- 458 23. Abdulrazaq NB. Cho MM. Win NN.Zaman R. and Rahman MT. (2012). Beneficial
459 Effectsof ginger (*Zingiberofficinale*) on carbohydrate metabolism in streptozotocin-
460 induceddiabetic rats. *British Journal of Nutrition*; 108 (7): 1194-1201.
- 461 24. Jafri SA.Abass S. and Qasim M. (2011). Hypoglycemic effect of Ginger (*Zingiber*
462 *officinale*)inalloxan-induced diabetic rats (*Rattusnorvagicus*). *Pakistan Veterinary*
463 *Journal*; 31 (2):160-162.
- 464 25. Dacie JV. and Lewis SM .(1994).*Practical Haematology*. 8th ed., p. 49, Longman Group
465 Ltd., Hong Kong
- 466 26. Price KR Johnson IT, Fenwic CR. (1987). The chemical and biological significance
467 ofsaponins in food and feeding stuff. Unpublished Crit. Rev. Food Sci. Nutr. 26: 27-135.
- 468 27. Robbins R J. (2003). Phenolic acids in foods. An overview of analytical methodology. *J.*
469 *Agric. Food Chem.* 51: 2886-2887.
- 470 28. Eleazu C O. Okafor PN.Amajor J. Awa E. Ikpeama AIandEleazu KC.(2011).Chemical
471 Composition, antioxidant activity, functional properties and inhibitory actionof unripe
472 plantain (*M. Paradisiacae*) flour.*African Journal of Biotechnology* Vol.10(74), pp.
473 16948-16952,
- 474 29. Thomas MC. Maclsaac R.J. Tsalamandris C. (2003). Unrecognized anemia in patients
475 with diabetes: a cross sectional survey. *Diabetes care.* 26:1164-1169
- 476 30. Onat A. Can G. Kaya H. Hergene G. (2010) Atherogenic index of plasma (\log_{10}
477 triglycerides/highdensity lipoprotein cholesterol) predict high blood pressure, diabetes
478 and vascular event. *Journal clin. Lipid* 4:89-98
- 479 31. Palacios I. Lozano M. Moro C. (2011). Antioxidant properties of phenolic compounds
480 occurring in edible mushrooms. *Food chem.*128:674-678
- 481 32. Egunyomi A. Moody JO. Eletu OM. (2009). Antisickling activities of two
482 ethnomedicinal plant recipes used for the management of sickle cell anaemia in Ibadan,
483 Nigeria. *Afri. J.Biotechnol.* 8:20-25
- 484 33. Choudhari SS.Kareppa BM.(2013) Identification of bioactive compounds of zingiber
485 *officinale* roscoe rhizomes through gas chromatography and mass spectrometry.
486 *Int J Pharm Res Dev*5:16-20.
- 487 34. Andallu B. Radhika B.Suryakantham V. (2013). Effect of aswagandha, ginger
488 andmulberry on hyperglycemia and hyperlipidemia. *Plant Foods Hum Nutr.* 58:1-7
- 489 35. Singh AB. Akanksha SN. Maurya R. Srivastava AK. (2009). Anti-hyperglycemic,
490 lipidlowering, and anti-oxidant properties of [6]-gingerol in db/db mice. *Int J Med*
491 *MedSci*1:536-44.
- 492 36. Madkor HR. Mansour SW.Ramdan G. (2010). Modulatory effect of garlic, ginger,
493 turmeric and their mixture on hyperglycemia, dyslipidemia and oxidative stress in
494 streptozotocin-nicotinamide diabetic rats. *Br. J.Nutr* 1:105-107
- 495 37. Adaramoye OA. Nanneri VO. Anyaanwu KC. (2005). Possible anti atherogenetic effect
496 ofKolaviron (a *Garcinia kola* seed extract) in hypercholesterolemia rats. *Clin.*
497 *Exp.Pharmacol. Physiol.* 32 (1-2) ;40-46
- 498 38. Ezekwe CI.Obidoa O. (2001). Biochemical effect of *Vernoniaamygdalina*on rat liver
499 microsomes. *Niger. J. Biochem. Mol. Biol.*16:1745-1798.
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