

BIOLOGICAL ATTRIBUTES OF THE SOIL WITH INTERCALED CROPS AND OIL PALM (*Elaeis guineensis* Jacq.): SUSTAINABLE MANAGEMENT IN THE AMAZON

ABSTRACT

In the Brazilian Amazon, the practices and use of agroforestry systems (SAF) are increasingly used, the proper management of them helps to improve soil properties and also to prevent their degradation. In this context, the objective of this work was to evaluate the biological attributes of the soil in the oil palm cultivation systems with intercropping, in an experimental area of São João da Baliza 26, km 12, with geographic coordinates of reference 00°51'13.3"N and 60°00'19.8"W, the altitude of 100 msnm and, distant to 352 km from the capital Boa Vista, state of Roraima realized in 2016. The experimental design used was completely randomized with four repetitions and six treatments: oil palm (*Elaeis guineense* Jacq.) interspersed with pineapple (OPi), bean (OBe), Banana (OBa), yucca (OMa) and *Brachiaria humidicola* (OPa), as well as an adjacent area only with *Brachiaria humidicola* as a witness (Pa). The Tukey test was used at a level of 5% probability in samples analyzed at a depth of 0-0.10 m, to compare the means of the variables evaluated. The COT presented values between 4.70 and 9.45 g kg⁻¹, being the highest values found in the interim systems OMa > Pa > OBa, highlighting the intermediate system OPi that presented the lowest levels. The highest basal respiration values of the soil (RBS) (23.50 mg C-CO₂ kg⁻¹ soil h⁻¹) and carbon from microbial biomass (BMS-C) (116.0 mg C microbiano kg⁻¹ soil) were verified in the pasture system. Likewise, for the urease and acid phosphatase activity, the grass system stands out as a control with values of (148.42 g NH₄⁺ g⁻¹ soil 2 h⁻¹) y (230 µg de p-nitrofenol g⁻¹ soil h⁻¹) followed by palm and grass and cassava systems. However, the β-glucosidase activity (51.22 µg p-nitrofenol g⁻¹ h⁻¹).

It was positively influenced by the oil palm system with cassava. On the other hand, the system interspersed with pineapple showed a higher metabolic coefficient (qCO₂) (0.36 mg C-CO₂ g⁻¹ BMS-C h⁻¹). It can be concluded that the pasture system (Pa) is presented as a more stable environment, followed by interspersed systems of oil palm with grass (OPa) and yucca (OMa).

Keyword: Agroforestry Systems.; Family Farming.; Soil Biology.; Amazon.

1. Introduction

The Brazilian Amazon region is characterized by environments with natural forest and areas in agricultural use (Oliveira et al., 2015), intense deforestation and conversion of forests to pasture areas (Lira et al., 2006; Silva et al., 2006). The degradation of these systems (natural or anthropic) breaks a natural, dynamic and balanced cycle, with a consequence on the chemical, physical and biological attributes of the soils (Peixoto, 2010).

In this context, the agroforestry systems (SAF's) the arboreal component helps to modify the soil environment, since in its constitution there is a greater diversity of root systems which provides greater contributions in the organic matter contents (OM) (Ferreira et al., 2012; Paul et al., 2013). The deposition of the litter generated by the fall of leaves, stems and crop residues, appear as sources and maintenance of the organic

46 matter in the soil (Smiley and Kroschel, 2008; Castro et al., 2009), with subsequent
47 availability and nutrient uptake by plants, especially in tropical soils with low natural
48 fertility (Souza, 2009).

49 Several biological indicators can be recommended to assess changes in
50 ecosystems and to sustain soil biological quality (Burns et al., 2013). Of these, the
51 carbon of the microbial biomass (BMS-C) (Nogueira et al., 2006), basal breathing
52 (Mendes et al., 2012), the metabolic quotient (qCO_2) and, microbial quotient ($qCmic$)
53 derived from BMS (Hungria, 2009; Nunes et al., 2012) and, the enzymatic activity of
54 the soil (Paz-Ferreiro et al., 2010). Palm-oil is also considered as a suitable raw material
55 for biodiesel production (Bhore, 2013). The BMS is one of the key components in soil,
56 with functions in the decomposition of organic residues (Marchiori Júnior and Melo,
57 2000), defined as the living part of the soil organic matter (Nascimento et al., 2009),
58 controlling the flows of C and N (Gosai et al., 2010), this microbial biomass
59 contributes, on average, 2 to 5% of the organic carbon in the soil (Jenkinson and Ladd,
60 1981) and between 1 to 5% of the total N of the soil (Smith and Paul, 1990; Araújo and
61 Monteiro, 2007), serving as a source of nutrients and energy intake (Monokrousos et al.,
62 2006).

63 Other indicators such as basal soil respiration (Mendes et al., 2012), related to
64 the degradation capacity of OM (Pragana et al., 2012), bacteria and fungi are the major
65 responsibility for the release of CO_2 via degradation of OM, in addition to algae,
66 protozoa and, root respiration (Roscoe et al., 2006; Gama-Rodrigues et al., 2008).
67 Related to the degradation capacity of OM (Pragana et al., 2012), bacteria and fungi are
68 the major responsible for CO_2 release via degradation of OM, as well as algae, protozoa
69 and, root respiration (Roscoe et al., 2006; Gama-Rodrigues et al., 2008).

70 The metabolic quotient (qCO_2) (Hungria, 2009; Nunes et al., 2012), refers to the
71 basal respiration of CO_2 incorporated per gram of microbial biomass in a given time,
72 being important in the studies that evaluate the effect of the environmental and
73 anthropogenic conditions on the microbial activity of the soil (Zhang et al., 2011), and
74 may be a good indicator of stress in crop management when BMS-C is affected, as it
75 infers in carbon gains, estimating the efficiency of substrate use by soil microorganisms
76 (Saviozzi et al., 2002; Almeida et al., 2007). In addition, the microbial quotient,
77 providing information on the quality of the organic matter and the amount of carbon
78 immobilized in the microbial biomass (Banning et al., 2008; Nunes et al., 2012).

79 The BMS transformations are mediated and catalyzed by enzymatic processes (Melo
80 et al., 2008), direct mediators in the functioning and catabolism biology of
81 microorganisms (Nielsen and Winding, 2002).

82 In the decomposition of organic residues, the enzyme β -Glucosidase is associated
83 with the breakdown of cellobiosis, involved in the C cycle (Doni et al., 2012), the
84 urease to the urea break in the N cycle, thus acting both in N and C of BMS (García-
85 Ruiz et al., 2008) and phosphatases important in the phosphorus cycle, as they
86 hydrolyze and transform organic P compounds into different inorganic P compounds, in
87 the form of PO_4 of phosphoric esters (Baker et al., 2011; Kedi et al., 2013).

88 The use of soil management studies using associated systems is a technique that is
89 currently used with a boom to prevent soil degradation and at the same time prevent soil
90 deterioration, being the biological indicators of soil used as a result of management.
91 However, for tropical soils there are still many studies. In this sense, the objective of
92 this work was to evaluate in an oil palm plantation, the effect of different associated
93 sowing systems (pineapple, cowpea, banana, manioc and pasture) on the biological
94 attributes (COT, RBS, BMS-C, qCO_2 , $qCmic$ and enzymatic activity) to improve soil

95 properties under agroecological production in a Red-Yellow Argisol of the Amazon
96 (Brazil).

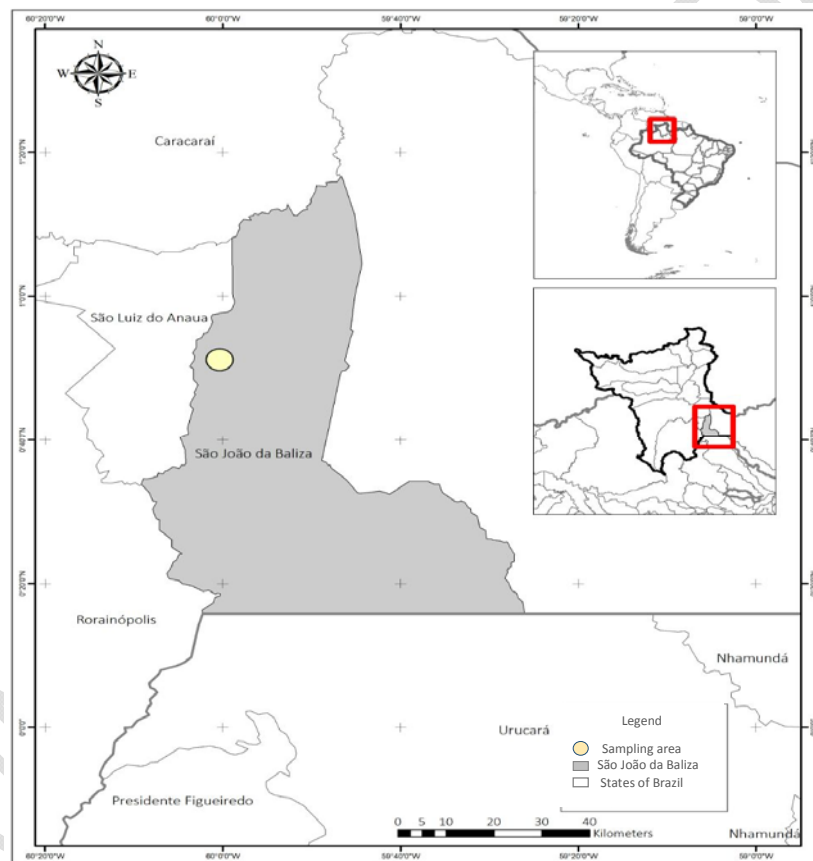
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99 2. Materials and methods

100 2.1. Study of area

101 The experiment was carried out in an experimental area of Embrapa (Brazilian
102 Agricultural Research Company), located in the municipality of São João da Baliza,
103 Roraima, Brazil. In the vicinal 26, Km 12, with geographic coordinates of reference 00°
104 51' 13,3"N and 60° 00' 19,8"W and altitude of 100 m, distant to 352 km of the capital
105 Boa Vista, state of Roraima (Figure 1).



106

107 Figure 1 – Experimental area where the research was developed.

108

109 2.2. Soil, vegetation and climate of the study area

110 The experimental area is located in a type of dense Ombrophilous forest, with
111 the dominant soil type of that area being argissolo According to the Brazilian soil
112 classification system (Morais et al., 2012; Santos et al., 2018), the relief according to the
113 Köppen classification is undulated with an inclination between 2-10% and Aw, humid
114 tropical climate, according to that type of region of the south of the State of Roraima
115 (Barbosa, 1997; Cruz et al., 2014), a precipitation varying from 1,800 to 1,900 mm,

116 relative humidity with an annual average between 85 to 90%, luminosity of 1,500 to
117 3,000 hours per year and the average annual temperature is 27°C (Lopes, 2014).

118 2.3. Management history of the study area

119 Before the implementation of the experiment, the system was established as a
120 12-year-old pasture of *Brachiaria humidicola*, used to graze cattle, and then intervened
121 to install the experimental plots in the period of 2012, and an area of approximately 1
122 ha⁻¹ was demarcated and used, and, the practice of clearing and weeding.

123 After the Brazilian Agricultural Research Company (EMBRAPA), responsible
124 for the experimental area, performed chemical analyzes for the 0-0.20 layer and 0.20-
125 0,40 m with the following results: pH (H₂O) = 5.4 - 5.0; organic matter content (%) =
126 2.6 - 1.3; exchangeable bases (cmol / dm³): Ca = 0.34 - 0.18; Mg = 0.10-0.05; K =
127 0.07-0.04; Al = 0.36 - 0.59; the sorptive complex: SB (cmol / dm³) = 0.52-0.28; CTCt
128 (cmol / dm³) = 4.73-4.04; V (%) = 11-7; m (%) = 41-68; P-available (mg/dm³) = 1.54 -
129 0.94. Subsequently liming was performed throughout the area, with dolomitic limestone
130 at a dose of 1.5 t ha⁻¹ (100% PRNT).

131 The planting of the oil palm was carried out with a density of 143 plants ha⁻¹, in
132 a system with the plants arranged in equilateral triangle with a spacing of 9 m of the
133 side between the plants in the line and of 7.8 m between the lines, in pits of 40 x 0.40 x
134 0.40 m. During the same period, intercropping was established between the lines of the
135 palm tree, measuring 36 x 39 m (width x length), with 27 plants, making up an
136 experimental area per plot of 1404 m². The pit was initially fertilized with 500 g of
137 triple superphosphate (45% P₂O₅).

138

139 2.4. Systems evaluated

140 The systems evaluated were six; T1: OPa (Oil palm interspersed with pasture).
141 In this system, the palm was intercalated with *Brachiaria humidicola*, with no
142 fertilization in the pasture, only in the oil palm.

143 T2: OPi (Oil palm interspersed with pineapple). Before planting the pineapple, a
144 planting of peanuts was carried out in 2012, with 11 lines (0.50 x 0,30 m) interspersed
145 by a palm oil strip, with fertilization in the pit using 10 g of simple superphosphate and
146 later harvested. The pineapple (Vitória) was then permanently implanted until the
147 moment of the evaluation, being 4 double lines in the spacing of 0.40 between plants
148 and 1 m between double lines, totalling 3,600 seedlings, with fertilization of 1 kg of the
149 formulated 8-28-16 (NPK) per line and 10 g per plant.

150 T3: OBe (Oil palm intercalated with beans). It was initially composed by the
151 consortium beans (Guariba) and maize (BR 106), established between each palm line, 3
152 lines of cowpea, 5 maize lines and 3 lines of cowpea. The line spacing was 0.5 m, and 8
153 plants of linear bean cowpea and 5 plants of corn per linear meter. The planting
154 fertilization was 10 g of the formulated 8-28-16 (NPK) and later planted only the bean
155 culture.

156 T4: OBa (Oil palm intercalated with banana), 120 seedlings of the cultivar Japira
157 were planted at the centre of the palm interline, with a spacing of 1.50 m between
158 plants, consorted with 25 rows of cowpea (Guariba), both sides of the banana line at a
159 spacing of 0.50 m between rows and a density of 8 plants per linear meter. The
160 fertilization consisted of 200 g of limestone and 100 g of simple superphosphate per
161 well for the banana. beans were used 10 g of NPK (8-28-16) per pit.

162 T5: OMa (Oil palm intercalated with manioc). Initially the system was
163 confirmed with rows of maize in double rows spaced 0.50 m and density of 5 plants per

164 linear meter (Cv.BR 106), spaced 1.00 m from single rows of manioc (3 rows) in the
165 spacing of 1.00 x 1.00 m, following the arrangement of 2 rows of corn and 3 rows of
166 manioc successively. The total area of the parcel had an arrangement of 20 maize lines
167 combined with 15 lines of cassava, involving 1,440 corn pits and 135 manioc pits.
168 Fertilization was performed in the pit, using 50 g of the formulation 8-28-16 (NPK).

169 T6: Pa (single pasture as a control). The *Brachiaria humidicola* pasture 12 years
170 after deforestation, without intervention or record of use of mechanization, corrective
171 and fertilizers, with continuous pasture use.

172 The fertilization for the oil palm was carried out during the vegetative
173 development from the first to the third year with the beginning of the production,
174 according to the results of the soil analysis obtained and according to recommendations
175 according to Embrapa-Roraima, being carried out per year two maintenance fertilizers,
176 with cover application per plant of: 200, 300 and 500 g of urea; 500, 600, 750 g of triple
177 superphosphate; 200, 300 and 400 g of potassium chloride; 100, 100 and 200 g of
178 magnesium sulfate; 30, 50 and 60 g of borax and 15, 15 and 50 g of zincop 101.
179 Fertilizations were carried out in two periods at the beginning and end of the rainy
180 season. During the conduction of the experiment to ensure the good development of the
181 palm and the crops, cultural practices such as weeding, thinning, defoliation of
182 intercropping, crowning of the oil palm and scrubbing of the spontaneous vegetation
183 between the lines were carried out.

184

185 2.5. Soil sampling and biological analyses

186 The experimental design was the completely randomized (DIC) with four
187 replicates being the readings made in triplicate. Each area consisted of the oil palm with
188 the different intercrop cultures, distributed in plots with 27 plants. Twelve plants were
189 randomly identified in each plot, in which sampling and evaluations were performed.

190 For the collection of the soil samples It was made in March 2016 in 3 mini-
191 trenches were opened per experimental unit, with dimensions of 0.50 x 0.50 x 0.50 m
192 and, allocated between the lines 0.5 m from the centre of the palm frond, collected the
193 samples only in the 0-0.10 m layer; after being conditioned, identified and kept in a
194 thermal box, later transported to the soil laboratory of the Agricultural Sciences Center
195 of the Federal University of Roraima, capital of Boa Vista, Brazil. Subsequently, a 2
196 mm mesh analysis was carried out, the roots were removed and then the breathing
197 experiment simulating the field conditions was mounted in the laboratory.

198 In the laboratory, the following biological analyzes were determined: soil
199 microbial biomass carbon (BMS-C), according to the fumigation-extraction
200 methodology proposed by Vance et al. (1987), with second adaptations Silva et al.
201 (2007). Samples were then weighed and fumigated directly with 1 mL of chloroform
202 and samples without the presence of chloroform (non fumigated samples). The
203 calculation of the carbon released in the titration by the excess of dichromate with
204 ammoniacal ferrous sulfate.

205 Soil basal respiration (RBS), determined according to the methodology proposed
206 by Jenkinson and Powlson, (1976), modified by Silva et al. (2007), adaptations. The
207 samples were weighed and incubated with 1 mol L⁻¹ NaOH for 24 days, with
208 subsequent quantification of C-CO₂ released every 3 days, totaling 8 titrations (3, 6, 9,
209 12, 15, 18, 21 and 24 days), the values being summed, to obtain a general reference of
210 accumulated C-CO₂ during the incubation period.

211 The metabolic quotient (q_{CO_2}), determined by the ratio of soil microbial activity
212 (C-CO₂ released) in relation to microbial carbon biomass (BMS-C), following the

213 methodology proposed by [Anderson and Domsch, \(1993\)](#), modified by [Silva et al.](#)
214 [\(2007\)](#). The microbial quotient ($qCmic$) was calculated using the expression proposed
215 by [Sparling \(1992\)](#), being necessary the determination of the COT present in the soil,
216 and later calculated.

217 In the determination of β -glucosidase activity, 0.05 M p-nitrophenol- β -D-
218 Glucopyranoside (0.05 M PNG) was used as the substrate in the reaction described
219 second [Tabatabai, \(1994\) and Dick \(1996\)](#), with adaptations. The determination of the
220 amount of p-nitrophenol in each sample was performed on the basis of a standard curve
221 prepared with concentrations of 0, 10, 20, 30, 40, 50 μg of p-nitrophenol mL^{-1} . The
222 readings were determined by molecular spectrophotometry, UV-Visible was used using
223 a SHIMADZU UV-1800 model at an absorbance of 400 nm and expressed in
224 micrograms of p-nitrophenol released per hour per gram of dry soil ($\mu\text{g p-nitrophenol h}^{-1}$
225 g^{-1} dry soil).

226 The urease activity was determined by the quantification of the ammonium
227 released by the urea hydrolysis (0.08 M solution), using the colorimetric method
228 recommended by [Kandeler and Gerber \(1988\)](#) adaptations. Determination of the amount
229 of N-NH_4^+ in each sample was performed based on a standard curve with known
230 concentrations of 0; 0.25; 0.5; 1.0; 2.0; and 4.0 $\mu\text{g mL}^{-1}$ of N-NH_4^+ , then the readings
231 performed by UV-Visible spectrophotometry at an absorbance of 660 nm and,
232 expressed in micrograms of N-NH_4^+ , released for two hours per gram of dry soil ($\mu\text{g N-}$
233 $\text{NH}_4^+ 2\text{h}^{-1} \text{g}^{-1}$ dry soil).

234 Likewise, acid phosphatase activity, determined using as a substrate in the
235 reaction 0.05 M p-nitrophenyl phosphate (0.05 M PNP), according to [Tabatabai, \(1994\)](#)
236 [and Dick \(1996\)](#), adaptations. The determination of the amount of p-nitrophenyl in each
237 sample was performed based on a standard curve of p-nitrophenyl (0, 10, 20, 30, 40, 50
238 μg of p-nitrophenyl mL^{-1}), then the readings determined by spectrophotometry
239 molecular UV-Visible at an absorbance of 400 nm, and values expressed in micrograms
240 of p-nitrophenyl released per hour per gram of dry soil ($\mu\text{g p-nitrophenyl h}^{-1} \text{g}^{-1}$ dry
241 soil).

242 For the statistical analysis, the data were first tabulated in Excel spreadsheets
243 and submitted to the verification of homogeneity of variances according to Levene test
244 and normality of the errors by the Shapiro-Wilk test, after which variance and Tukey
245 test were performed at level 5 % of probability, also performed Pearson and multivariate
246 correlation analysis (PCA and HCA), with the aid of statistical software SAS and
247 INFOSTAT.

248

249

250 3. Results and discussion

251 The results of the analyzes of variance obtained showed significant differences for the
252 evaluated treatments ([Table 1](#)) The COT presented values between 4.70 and 9.45 g kg^{-1} ,
253 being the highest values found in the interim systems OMa > Pa > OBa, highlighting the
254 intermediate system OPi that presented the lowest levels. The intercalary systems
255 contribute to the maintenance of the agrosystems, maintaining a balance of soil fertility
256 ([DELARMELINDA et al., 2010](#)) by promoting coverage, biomass production, carbon
257 accumulation and nutrient removal from the deeper layers, in a cycling process that
258 becomes more efficient reducing leach losses and erosion. The accumulation of biomass
259 in these systems came mainly from the prunings made to the oil palm plants, as well as
260 the fall of leaves and branches of the other species of the systems, contributing to the
261 maintenance of the levels of organic matter and biological activity in these production

262 models (SMILEY; KRUSCHEL, 2008; CASTRO et al., 2009). Another relevant factor
 263 to understand the best results presented in these areas of agricultural use is the response
 264 of limestone application, which may have favored, accelerating the decomposition
 265 process, mineralization and availability of organic matter with subsequent increase of
 266 biological activity in the soil generally.

268 **Table 1** - The values of total organic carbon (TOC), basal respiration of soil
 269 accumulated in 24 days (RBS), soil microbial biomass carbon (BMS-C), metabolic
 270 quotient (qCO_2) and the quotient of microbial carbon ($qCmic$), in a Red-Yellow Argisol
 271 with different systems of use in the 0-0.10 m layer, São João da Baliza, Boa Vista -
 272 Roraima, 2017.

Systems of use	Total organic carbon and microbial attributes				
	COT (g ^{kg⁻¹})	RBS (mg of C-CO ₂ Kg ⁻¹ soil h ⁻¹)	BMS-C (mg C microbial Kg ⁻¹ soil)	qCO ₂ (mg C-CO ₂ g ⁻¹ BMS-C h ⁻¹)	qCmic (%)
OPa	8.02 bc	20,28 c	105,75 b	0,19 c	1,34 a
OPi	4.70 d	19,31 d	53,36 e	0,36 a	1,15 ab
OBe	7.19 c	19,65 d	81,35 d	0,24 b	1,16 ab
OBa	8.41 abc	21,44 b	85,22 d	0,25 b	1,02 b
OMa	9.45 a	21,49 b	95,93 c	0,22 bc	1,02 b
Pa	8.64 ab	23,50 a	116,00 a	0,20 c	1,36 a

273 The average value for the readings made in triplicate is presented. Means followed by the same lowercase
 274 letter in the columns and upper case in the rows do not differ statistically from each other by the Tukey
 275 test at a 5% probability level.

277 For soil basal respiration (RBS), which relates the degradation capacity of the
 278 OM, being defined as the sum total of all the metabolic functions in which the CO₂ is
 279 produced (Pragana et al., 2012). Variable results were observed after 24 days of
 280 incubation, with the highest values found in the Pa area (23.50 mg C-CO₂ kg⁻¹ soil⁻¹)
 281 and the lowest values in the OBe and OPi intercalary systems (23,50 ; 19.65 and 19.31
 282 mg of C-CO₂ kg⁻¹). Although, without differences between the OBa and OMa systems
 283 (21.44 and 21.49 mg C-CO₂ kg⁻¹ dry soil⁻¹) presenting lower values when compared to
 284 the Pa area and higher with respect to the OPa system (20.28 mg of C-CO₂ kg⁻¹ soil⁻¹).
 285 Therefore, the microbial activity in the soil with intercalated cropping systems was
 286 lower when compared to the Pa system (Table 1).

287 Many works have been developed in the southwest of the Amazon, among them
 288 Mazzetto et al. (2016), evaluating biological attributes in an Argisol, found higher
 289 values of RBS in the pasture area compared to areas of agricultural use and native
 290 forest, observing a great similarity between values of respiration rates between native
 291 areas and pasture areas, with significant differences for areas used as agricultural
 292 systems. These results corroborate Silva et al. (2014), observing higher values of RBS
 293 in the pasture area when they evaluated the biological activity of soils in organic,
 294 agroforestry and pasture systems in the southwest of the Amazon. Still, Silva et al.
 295 (2012), when investigating RBS, verified the highest values in pasture areas and three
 296 forest fragments in comparison with areas of annual and perennial crops. According to
 297 these authors, the factors responsible for the renewal of plant and microbial biomass and
 298 nutrient cycling may have promoted lower respiration rates in soils under annual and
 299 perennial crops, and in systems in which land use changes the dynamics of the organic
 300 matter, there were significant differences in the biological attributes.

301 In agreement with this work, in all the evaluated systems the highest respiration
 302 rates were observed in the Pa area compared to the areas of agricultural use (Table 1).

303 Soils under anthropic interference as in cultivated areas, undergo changes in their
304 composition and metabolic activity, and the microbial population is under stress
305 (Moreira and Siqueira, 2006). Thus, the highest rate of respiration may or may not be
306 desirable and may indicate both disturbance and high level of ecosystem productivity
307 and should be analyzed in each context (Islam and Weil, 2000). It can thus be
308 interpreted as a desirable characteristic when it is considered that the decomposition of
309 organic residues will provide nutrients to the plant, in addition, it promotes processes
310 such as: aggregation, cation exchange capacity and water retention (Roscoe et al.,
311 2006). These increases in the rate of respiration in the pasture area can also be explained
312 by the preference of the microorganisms for certain types of organic materials, besides
313 the presence of feces and urine of the cattle promoting a high metabolic activity, being
314 also that the grasses release C in the form of CO₂ because they present a larger root
315 system, more aggressive and exploratory, with a high respiratory activity.

316 In relation to the soil microbial biomass carbon (BMS-C), which represents the
317 amount of carbon that the microbial biomass of the soil immobilizes in its cells, it is
318 verified that the different systems evaluated promoted variations in these contents, area
319 Pa (116.00 mg C microbial kg⁻¹ alone). OPa was the system that presented the highest
320 values of 105.75 mg C microbial kg⁻¹ soil, followed by OMa with 95.93 mg C microbial
321 kg⁻¹ soil, OBa and OBe presented close values and no significant statistical differences
322 (85.22 and 81.35 mg C microbial kg⁻¹ alone). Taking into account the interim system of
323 OPi that presented the lowest values of 53.36 mg C microbial kg⁻¹ soil, differing
324 statistically from all other treatments, and the Pa and OPa systems were two times
325 higher (Table 1).

326 Factors such as clay content, moisture and water dynamics, types of cover with
327 the continuous addition of residues, modifications of the microclimate of the area, root
328 distribution, incorporated organic carbon contents directly influencing the activity and
329 biological diversity and management practices such as incorporation of inorganic
330 fertilizers explain the results of the present study. In this context, research developed by
331 several authors in the southwest of the Amazon also reported the highest values of
332 BMS-C in the pasture area compared to areas of agricultural use and native forest.
333 These results were attributed to fine root biomass as a factor that can influence the
334 response of microbiological attributes in the pasture system, in addition, the age of
335 establishment as a source of carbon accumulation (Silva et al., 2014; Mazzetto et al.,
336 2016).

337 In pasture areas, this effect occurs because the root system is abundant and
338 extensive, presenting a continuous renewal and a strong rhizospheric effect, promoting
339 greater biological activity (Alves et al., 2011). Similarities between pasture and native
340 areas may be related to equivalent carbon stock, while the differentiated behavior
341 presented by the areas in agriculture may be related to the soil management and the
342 different types of crops that are found in the region (pineapple, banana, beans, corn,
343 rice, cocoa, coffee, among others). It should be noted that the evaluation of BMS-C or
344 RBS alone provides only limited information on certain responses in the ground
345 interference to stress or disturbances. Therefore, it is necessary to follow other
346 evaluations and can be conducted together with the determination of these
347 characteristics, such as the metabolic quotient ($q\text{CO}_2$), microbial carbon quotient
348 ($q\text{Cmic}$) and the enzymatic activity of the soil.

349 Thus, the metabolic quotient ($q\text{CO}_2$) was estimated from the values of RBS.
350 accumulated in 24 days and BMS-C, being verified the highest value in the agricultural
351 use system OPi (0.36 mg C-CO₂.g⁻¹ BMS-C. h⁻¹) differing from the other systems,
352 followed by OBa, OMa and OBe, which did not differ statistically from each other

353 (0.25, 0.22 and 0.24 mg C-CO₂.g⁻¹ BMS-C. h⁻¹), and the lowest values were found in
354 the Pa and OPa areas (0.20 and 0.19 mg C-CO₂.g⁻¹ BMS-C) (Table 1). According of
355 Chaer (2011) e Souza et al. (2013), low values of *q*CO₂ indicate a more stable
356 environment with better quality in the physical, chemical and biological attributes,
357 which demonstrates a more balanced ecosystem in the pasture areas, however, in the
358 cultures intercalated with the replacement of the cover occurs more rapid decomposition
359 of the vegetal residues, increasing the metabolic quotient (Balota et al., 1998).

360 On the other hand, Anderson e Domsch (1993), report that this attribute serves to
361 estimate the efficiency of substrate use by soil organisms, so the high value found in
362 agricultural areas and more specifically in the pineapple (OPi) palm system indicates the
363 occurrence of disturbances where the microbial population is oxidizing of their own
364 cells, and these high values correspond to the need for a high energy demand for their
365 maintenance, survival and adaptation to the soil, therefore, the microbial population is
366 in adverse or stressful conditions.

367 Still, Martins et al. (2010), explain that the higher values of *q*CO₂ found in the
368 systems indicate more carbon losses in the system per unit of microbial biomass and are
369 related to the microbial biomass mineralization response. Where lower values of *q*CO₂
370 and higher value of BMS-C suggest that microbial biomass was more efficient in the
371 use of organic compounds, releasing less carbon, like CO₂ and incorporating more to
372 the microbial tissues (Cunha et al., 2011). In this way, in relation to the mineralization
373 of the microbial biomass, it can be inferred that, in comparison to the Pa system, the
374 OPa, OMa and OBe systems were very similar in the use of the organic compounds,
375 incorporating C to their tissues and releasing less as CO₂. It can also be concluded that
376 the soil microbial population in these areas of agricultural use OPa, OMa and OBe,
377 demanded similar amounts of energy to maintain, indicating that the intercalated
378 systems studied can reduce the emission of CO₂ over time, since they are more stable
379 environments for the soil microbial community.

380 In addition, it is also observed that the OPi system is undergoing some
381 environmental stress due to its relationship with *q*CO₂ and low microbial activity, unlike
382 the Pa, OPa and OMa systems, which besides stimulating microbial development
383 (BMS-C), present a good quality of the organic matter (*q*Cmic), increasing the
384 efficiency of use of the substrates (Table 1). In this aspect, the microbial quotient
385 (*q*Cmic) is an important indicator of impacts (Klumpp et al., 2003). Providing
386 information on the quality of organic matter and the amount of C immobilized in
387 microbial biomass (Banning et al., 2008; Nunes et al., 2012). The highest values of
388 *q*Cmic observed in the Pa and OPa environments (1.36 and 1.34%) were higher than the
389 values found in the areas destined to the conventional use OPi and OBe (1.15 and
390 1.16%) and, with the lower values observed in OBa and OMa systems (1.02 and 1.02%)
391 (Table 1). High levels of *q*Cmic indicate that there is an increase in BMS-C against the
392 amount of organic C available, that is, a greater efficiency in its use by microorganisms,
393 being reported as an indicator of the quality of OM, allowing to accompany
394 disturbances promoted by ecological imbalance and variations in OM levels caused by
395 (Alvarez et al. 1995).

396 In an experiment carried out by Mazzetto et al. (2016), when assessing soil
397 biological attributes in southwestern Amazonia, observed higher *q*Cmic values in
398 pasture areas compared to areas of agricultural use and native forest. In this study, the
399 lowest values were found in the systems of agricultural use in relation to the systems
400 submitted under pasture, indicating a greater disturbance in these environments, either
401 by type of management or by anthropic intervention. Jekinson e Ladd (1981) e

402 [Jakelaitis et al. \(2008\)](#), establish normal values of $qCmic$ between 1 and 4%, being the
 403 values observed in this study in this range, for the different areas studied.

404 On the other hand, it was also evaluated the enzymatic activity β -glucosidase,
 405 urease and phosphatase for the different systems studied ([Table 2](#)).

406

407 **Table 2** - Activity of β -glucosidase, urease and acid phosphatase, in a Red-Yellow
 408 Argisol with different systems of use in the 0-0.10 m layer, São João da Baliza, Boa
 409 Vista - Roraima, 2017.

Systems of use	Microbial attributes		
	β -Glucosidase ($\mu\text{g p-nitrophenol.}$ $\text{g}^{-1} \text{ soil h}^{-1}$)	Urease ($\mu\text{g NH}_4^+$ $\text{g}^{-1} \text{ soil}^{-1}$)	Acid phosphatase ($\mu\text{g p-nitrofenol.}$ $\text{g}^{-1} \text{ soil h}^{-1}$)
OPa	39.59 abc	121.11 b	145.83 bc
OPI	22.97 d	104.49 c	47.42 d
OBe	37.90 bc	117.15 b	119.42 c
OBa	32.16 cd	118.34 b	132.04 c
OMa	51.22 a	126.45 b	168.83 b
Pa	47.83 ab	148.42 a	230.33 a

410 The average value for the readings made in triplicate is presented. Means followed by the same lowercase
 411 letter in the columns and upper case in the rows do not differ statistically from each other by the Tukey
 412 test at the 5% probability level.

413

414 In the OMa system with the highest activity of β -glucosidase 51.22 $\mu\text{g p-}$
 415 $\text{nitrophenol g}^{-1} \text{ soil h}^{-1}$, followed by Pa and OPa systems with values of 47.83 and 39.59
 416 $\mu\text{g p-nitrophenol g}^{-1} \text{ soil h}^{-1}$, behaving statistically the same. On the other hand, lower
 417 values were observed in the OBe and OBa systems (37.90 and 32.16 $\mu\text{g p-nitrophenol g}^{-1}$
 418 soil h^{-1}) and the OPI system with values much lower than 22.97 $\mu\text{g p-nitrophenol g}^{-1}$
 419 soil h^{-1} ([Table 2](#)). The incorporation of OM in these systems provides the necessary
 420 substrate for the action of β -glucosidase, maintaining and protecting the enzymes
 421 in their active forms, due to the formation of complex enzymes-humic compounds ([Deng](#)
 422 [and Tabatabai, 1997](#)).

423 In addition, areas formed by grasses have a dense root area exploring greater
 424 depths when compared to agricultural areas, favoring the microbial biomass of the
 425 rhizosphere and consequently stimulating a greater activity of the microorganisms.
 426 Regarding the type of cover, in the PS area besides forming these complexes, the
 427 incorporation and decomposition of the organic residues become slower due to chemical
 428 conditions, presenting lower pH values when compared to the intercalated areas,
 429 allowing to maintain material for periods long time with more action and biological
 430 breathing activity. In addition, the quality of the incorporated residue influences the
 431 enzymatic activity, the intercalary systems with the use of cover plants, present material
 432 with recalcitrant constituents to the microbial decomposition, such as lignins, waxes and
 433 phenolic compounds of high molecular weight, with greater difficulty to be broken , and
 434 less amount of easily decomposed and incorporated material such as carbohydrates
 435 ([Cunha et al., 2011](#)).

436 According [Moscatelli et al. \(2012\)](#), the land use systems that provide greater
 437 diversity and quantity of organic waste favor the development of microorganisms and
 438 promote an increase in enzymatic activity. This explains the greater activity found in the
 439 DM, PS and DP systems, being that these systems presented the highest TOC levels in
 440 the soil ([Tablet 1](#)), and, the greater activity of the enzyme β -glucosidase. According with
 441 [Dick et al. \(1996\)](#), in experiments developed establish values of variation for the

442 enzyme β -Glucosidase of 38 to 720 $\mu\text{g p-nitrophenol g}^{-1} \text{ soil h}^{-1}$, observing in this range
443 the values found in the different systems evaluated in this work [Almeida et al. \(2016\)](#),
444 when evaluating the fauna and microbiological attributes of an Argisol under cover
445 systems in southern Brazil, reported maximum values of 72.4 $\mu\text{g p-nitrophenol g}^{-1} \text{ h}^{-1}$
446 soil for β -Glucosidase activity and values of 23.6 $\mu\text{g NH}_4^+ \text{ g}^{-1} \text{ soil 2h}^{-1}$ for the urease
447 activity, in a consortium of lablab (*Dolichos lablab*) with corn. In this sense [Bandick e](#)
448 [Dick \(1999\)](#), when studying the effect of fertilizers and crop rotation on the enzymatic
449 activity in the soil, establish activity values of 40 to 270 $\mu\text{g NH}_4^+ \text{ g}^{-1} \text{ solo 2 h}^{-1}$. Results
450 found in this research are within this range of variations in urease activity ([Table 2](#)).

451 These results indicate that OM, besides serving as a substrate for the microbiota,
452 may be protecting this enzyme against the action of proteolytic enzymes naturally
453 present in the soil, maintaining the potential of urease activity for longer periods of
454 time. In the Pa area there is consequently a greater root system when compared to the
455 areas with intermediate cultures OMa, OPa, OBa, OBe and OPi, which increases the
456 rhizosphere stimulating the activity of the organisms in these places, in addition, there is
457 a continuous contribution of organic residues, even in areas under agricultural use, low
458 activity may also be related to changes in the composition of microbial communities
459 present. Although, organic nitrogen can be found as urea occurring in the natural form
460 through animal excretions and as a product of nucleic acid mineralization ([Metting](#)
461 [Junior, 1992](#)), indicating that the ammonification is normally occurring in the PS area,
462 and the chemical conditions are favorable with low values of pH, clay contents that may
463 be influencing the retention of the ammonium cation, by adsorption processes to the soil
464 colloids making it relatively stationary. It should be pointed out that the interim systems
465 OMa, OPa, OBa, OBe did not present significant differences between them in the
466 urease activity, with values very close to the Pa system, but it is also important to
467 highlight that the coverage has a great influence in this process, stable conditions of pH,
468 as well as a more diversified coverage that provides a better source of energy for the
469 microorganisms, thus favoring the activity of this enzyme.

470 In relation to acid phosphatase activity, the Pa system with the highest values of
471 230.33 $\mu\text{g p-nitrophenol g}^{-1} \text{ h}^{-1}$ is also highlighted, followed by the OMa and OPa
472 systems (168.83 and 145.85 $\mu\text{g p-nitrophenol g}^{-1} \text{ soil h}^{-1}$), but without significant
473 differences. Already, decreasing values were observed in the OBa and OBe intercalated
474 systems (132.04 and 119.42 $\mu\text{g p-nitrophenol g}^{-1} \text{ h}^{-1}$ soil) and also lower values in the
475 OPi system with 47.42 $\mu\text{g p-nitrophenol g}^{-1} \text{ soil h}^{-1}$ ([Table 2](#)). The activity of acid
476 phosphatase is influenced mainly by the chemical characteristics of the soil, OM has
477 beneficial effects since it increases the microbial community. In addition, the highest
478 activity of this enzyme is found with pH of the soil around 5.0 with tendency to
479 decrease with increases of pH to values of 7.0 ([Dick et al., 1988](#)). Being strictly
480 controlled by biological demand, so that the lower the amount of inorganic phosphorus
481 the greater activity of the enzyme ([Speir and Ross, 1978](#); [Allison et al., 2007](#)), this was
482 verified in the systems with OMa > OPa > OBa > OBe > OPi, where the acid phosphatase
483 activity was lower, in this sense, observing ideal conditions in native areas and pasture
484 areas when compared to cultivated areas ([Table 2](#)).

485 It is also known that the higher activity of acid phosphatase is related to the
486 decrease of soil P contents ([Gatiboni et al, 2008](#)), corroborating with the results found in
487 the Pa area, as control with the highest activity without the presence of inorganic
488 fertilizers, followed by the OMa and OPa intercalated systems. Consequently, in the
489 cultivated areas, localized applications of phosphate inorganic fertilizers cause a
490 decrease in the enzymatic action, occurring in zones of low activity and concentration in
491 this enzyme, contrary in the native and pasture areas, maintaining this dynamic balance.

502 These observed differences between the different evaluated systems can also be
 503 explained to the qualitative changes in the diversity of the microbial communities and
 504 the stress caused by the incorporated inorganic fertilizers. Like this, [Peixoto \(2010\)](#),
 505 when evaluating microbial biomass and enzymatic activity in soils of the state of São
 506 Paulo under native and cultivated vegetation, found values for acid phosphatase activity
 507 of 8.61 $\mu\text{g p-nitrophenol kg}^{-1}\text{soil h}^{-1}$ in culture condition at 191, 79 $\mu\text{g p-nitrophenol kg}^{-1}$
 508 soil h^{-1} in grazing condition. For forest soils found values of 19.63 mg p-nitrophenol
 509 $\text{kg}^{-1}\text{soil h}^{-1}$ at 158.22 $\mu\text{g p-nitrophenol}$.

510 In this sense, ([Table 3](#)), it presents the correlation between the organic matter
 511 contents and the soil biological attributes, for the different evaluated treatments.

512 **Table 3** – Pearson's correlation between total organic carbon (TOC) and biological
 513 attributes, studied in a Red-Yellow Argisol with different systems of use in the 0-0.10 m
 514 layer, São João da Baliza, Boa Vista - Roraima, 2017.

	TOC	RBS	C-BMS	$q\text{CO}_2$	$q\text{Cmic}$	$\beta\text{-Glu}$	Ure	Fos Ac
TOC	~							
RBS	0.60**	~						
C-BMS	0.74**	0.74**	~					
$q\text{CO}_2$	-0.76**	-0.55**	-0.94**	~				
$q\text{Cmic}$	-0.32ns	0.19ns	0.38ns	-0.30ns	~			
$\beta\text{-Glu}$	0.70**	0.58**	0.74**	-0.71**	-0.06ns	~		
Ure	0.59**	0.88**	0.84**	-0.68**	0.33ns	0.69**	~	
Fos Ac	0.71**	0.87**	0.90**	-0.79**	0.27ns	0.75**	0.91**	~

515 Where: n.s. - not significant ($p > 0.05$); * - significant at the 5% level ($p \leq 0.05$); ** - significant at
 516 the 1% level ($p \leq 0.01$).

517 Thus, a positive and highly significant correlation between RBS and OM was
 518 observed (r) of 0.60 ($p \leq 0.01$). Thus, the availability and continuous addition of organic
 519 residues to the soil increases the OM content, favoring and promoting greater biological
 520 activity with a subsequent increase in the metabolism of microorganisms and respiratory
 521 rate. Positive and significant correlation of BMS-C with OM and RBS, (r) 0.74 and 0.74
 522 ($p \leq 0.01$). [Carneiro \(2010\)](#), reported a significant and positive correlation between BMS-
 523 C and OM, indicating that the microbial attribute is influenced by the OM contents,
 524 being an expected fact since, the greater incorporation of organic residues increases the
 525 biological activity, releasing C as CO_2 by microbial metabolism ([Table 3](#)).

526 Highly significant but negative correlation was observed between $q\text{CO}_2$ and
 527 OM, (r) of -0.76 ($p \leq 0.01$), and a highly significant and negative correlation between
 528 $q\text{CO}_2$ and RBS and BMS-C with (r) -0.55 and -0.94 ($p \leq 0.01$) ([Table 3](#)), data
 529 corroborated by [Pimentel et al., \(2006\)](#); [Souza et al., \(2013\)](#) and [Colodel, \(2014\)](#).
 530 Positive and significant correlation between the OM, BMS-C with the enzyme β -
 531 glucosidase, (r) of 0.70 and 0.74 ($p \leq 0.01$). The higher the soil carbon content in the
 532 studied areas, the higher the β -Glucosidase activity found. [Stott et al. \(2010\)](#) e [Silva et al. \(2012\)](#),
 533 also report the same sequence in their studies.

534 Positive and highly significant correlation between OM, BMS-C and, urease
 535 activity, (r) of 0.59 and 0.84 ($p \leq 0.01$). Research carried out by different authors as
 536 [Zornoza et al. \(2006\)](#), in soils of the Mediterranean region in Spain; [Silveira \(2007\)](#),
 537 evaluating the quality of agricultural soils of Rio Grande do Sul, also observed a
 538 positive and highly significant correlation between organic matter and urease activity.
 539 Positive and significant correlation between OM, BMS-C and, the enzyme acid
 540 phosphatase, (r) of 0.71 and 0.90 ($p \leq 0.01$) respectively. Data that corroborate with those
 541 obtained by [Nahas et al., \(1994\)](#); [Fernandez et al., \(1998\)](#) and [Peixoto \(2010\)](#), when they

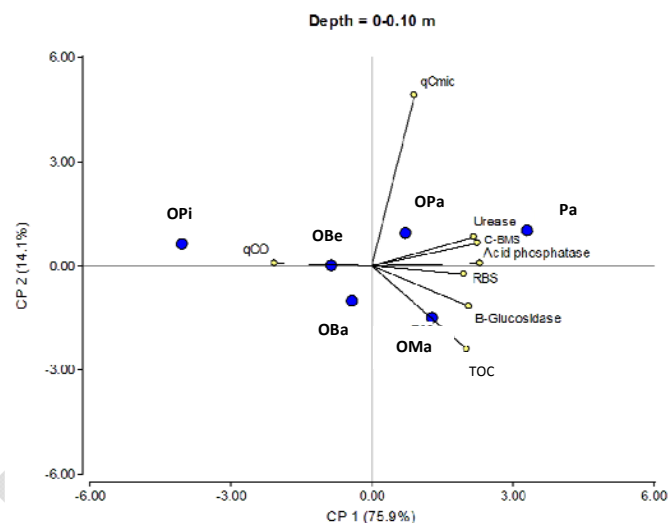
534 found significant correlations between OM and acid phosphatase activity. These types
535 of correlations are important from the point of stability of the soil fertility of the
536 different systems.

537 Furthermore, analyzes of the main components were carried out jointly for the
538 different OPa, OPi, OBe, OBa, OMa and Pa systems in the 0-0.10 m layer in order to
539 evidence and analyze the interdependence of the variables between carbon (TOC), and
540 the biological attributes studied, trying to find with minimal loss of information, a new
541 set of variables (main components) that explain the structure of the variation, being
542 represented the weight of each variable analyzed in each component (axes).

543 Being represented in the (Figure 2), the results of the analysis of the main
544 components (PCA) of the evaluated variables: TOC and microbiological and
545 biochemical attributes, the sum of the variability retained in these components
546 explained 90% of the original variability of the data.

547

548 **Figure 2** - Distribution of the original variables between the total organic carbon
549 (TOC) and the biological attributes in the 0-0.10 m layer on the first and second main
550 component (CP 1 and CP 2).



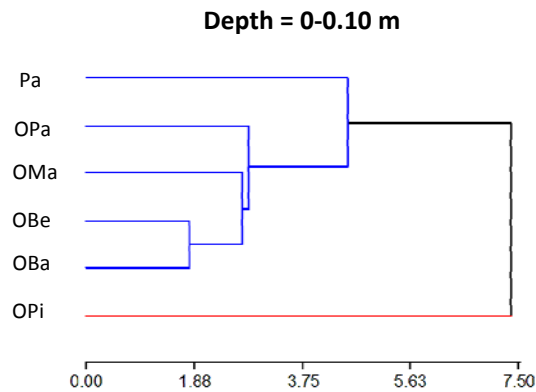
551

552 The first major component (CP 1) contributed with 75.9% of the total variance
553 explained, however, most of the variables that were strongly affected, among them:
554 urease, BMS-C and acid phosphatase, contributing positively to the CP 1, and inverse
555 with the variable qCO_2 . In addition, $qCmic$ did not influence all the different systems
556 evaluated in this component. These results indicate that CP 1 allowed to distinguish the
557 cultures that are associated to these variables, being the Pa, OPa and OMa systems that
558 contributed the most to improve the microbiological and biochemical conditions of the
559 soil. However, the second main component (CP 2) explained 14.1% of the total and was
560 related to the variable $qCmic$, in negative projection with the variables RBS, β -
561 glucosidase and, TOC. The analysis showed that the OPi system was more related to
562 this component and, with lower effects for OBe and OBa (Figure 2).

563

564 **Figure 3**. Analysis of hierarchical components (HCA) for the variables studied
565 in the different systems evaluated.

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In the HCA hierarchical dendrogram (Figure 3), the variables that were grouped are OBe and OBa with a Euclidean distance of 1.88; a second grouping between OMa and the group formed by OBe and OBa, a third group formed by OPa, a larger group composed of Pa together with the other groups (OBa, OBe, OMa and OPa) can still be visualized. We can say that the variable OPi has no correlation with the other elements of the group, because there is a separation distance very high.

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In this case, the analyzes of PCA and HCA showed a great sensitivity to the categorical selection of the variables in the different evaluated systems, suggesting that the continuous incorporation and decomposition of residues and the exudates released by the roots, promote an increase in microbial activity as a final result in greater biochemical cycles, as observed in the Pa, OPa and OMa areas. In addition, the $qCmic$ attribute was negatively correlated with the qCO_2 attribute, with the OPi system being more strongly associated with this component and, with smaller effects for the OBe and OBa systems, indicating that the systems were strongly affected, implying a high metabolic activity (qCO_2) with higher energy supply ($qCmic$) (Figure 2 and 3).

585 4. Conclusions

586 In agreement with the results obtained in this work, the interleaved systems improve
587 soil conditions in relation to the Pa control system. The balance and dynamics in the
588 OBe and OBa systems, modified by the anthropic action, present intermediate values
589 related to the microbiological and biochemical attributes. The OPi system presents as
590 the less stable environment, with a high stress and lower quality of microbiological and
591 biochemical attributes. All the microbiological and biochemical attributes analyzed in
592 the different agricultural systems are influenced by the OM contents.
593

594 **Conflicts of Interest:** The authors declare no conflict of interest.
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