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3 ***In Vitro* Cercaricidal Activity of Fractions and Isolated Bioactive Compounds from the**
4 **Root Bark of *Erythrophleum ivorense* (Fabaceae) against *Schistosoma haematobium***
5 **Infection.**

6 **Abstract**

7 **Introduction:** *Schistosoma haematobium* is one of the species of *Schistosoma* responsible for
8 schistosomiasis in humans, a major public health problem worldwide. Praziquantel, the most
9 effective drug against all adult stages of human schistosomiasis, faces the threat of resistance
10 and also has sub-optimal efficacy against cercaria, an immature form of schistosomiasis. This
11 underscores the need to search for an alternative antischistosomal drug with pronounced
12 activity particularly against cercaria.

13 **Aim:** This study investigated anti-cercarial activity of total crude (70% ethanolic extract),
14 fractions (methanolic, ethyl acetate and petroleum ether) and isolated bioactive compounds
15 from the root bark of *Erythrophleum ivorense*.

16 **Study design:** *In vitro* anti-cercarial activity was evaluated using 20 freshly shed cercariae
17 from *Schistosoma haematobium* species transferred into 20 well plates. Cercaricidal effect of
18 the various concentrations (15.6, 31.3, 62.5, 125.0, 250.0 and 500.0 µg/mL) of test extracts
19 and compounds were observed for 3 hours using an inverted microscopy. The results showed
20 that extracts and compounds of the plant decreased percentage viability of cercariae in a
21 dose-dependent manner.

22 **Results:** Within two hours of incubation, all cercariae died at the various concentrations of
23 test compounds and extracts with the exception of methanol extract and the bioactive
24 compound erythroivorensin at 15.6 µg/mL. The least potent extract, methanol, had an IC₅₀ of
25 2.11±0.10 µg/mL. Eriodictyol, being the most active compound had an IC₅₀ of 1.23 ± 0.05
26 µg/mL.

27 **Conclusion** It is evident from the results obtained that fractions and isolated bioactive
28 compounds of *Erythrophleum ivorense* can be a potential cercaricidal agent and therefore
29 should be investigated further.

30 **Keywords :** Cercariae, Schistosomiasis, Erythroivorensin, Eriodictyol, Betulinic acid
31 *Erythrophleum ivorense*

33 **Introduction**

34 Schistosomiasis also known as bilharziasis or snail fever is a parasitic disease caused by
35 flukes (trematodes) of the genus *Schistosoma*. It is prevalent in tropical and subtropical areas,
36 especially, in poor communities with no access to safe drinking water and adequate sanitation
37 [1]. People become infected by being in contact with fresh water bodies infested with free-
38 swimming larval forms of the parasite (cercariae) shed from freshwater snail intermediate
39 hosts [2, 3].

40 The disease is better known for its chronicity and debilitating morbidity which results in high
41 costs in public health and economic productivity in developing countries [4]. Globally, more
42 than 207 million people, 85% of whom live in Africa, are infected with schistosomiasis, and
43 an estimated 700 million people are at risk of infection in 76 countries [5]. 200,000 deaths are
44 globally attributed to schistosomiasis annually, and about 10 million women in Africa are
45 infected during pregnancy [6].

46 There is no available vaccine currently and the chemotherapeutic agent of choice which is
47 Praziquantel (PZQ), already faces drawback of drug resistance in some *Schistosoma* isolates
48 [7, 8]. Complementing existing chemotherapy with synthetic molluscicides to eliminate the
49 possibility of re-infestation of water bodies with cercariae faces the challenge of cost as well
50 as environmental pollution [9]. It is based on these reasons that the search for affordable,
51 readily available, less toxic schistosomicidal plant-derived products have become essential.
52 This is because plants have timelessly served as good source for the discovery and
53 development of newer drugs with about 25% of current medicines derived from them [10].
54 Artemisinin, quinine and licochalcone A are examples of plant-derived products in clinical
55 use particularly against parasitic infections [11]. One of such promising plants is
56 *Erythroleum ivolense* which is also known as ‘potrodum’ among the Akans in Ghana, and

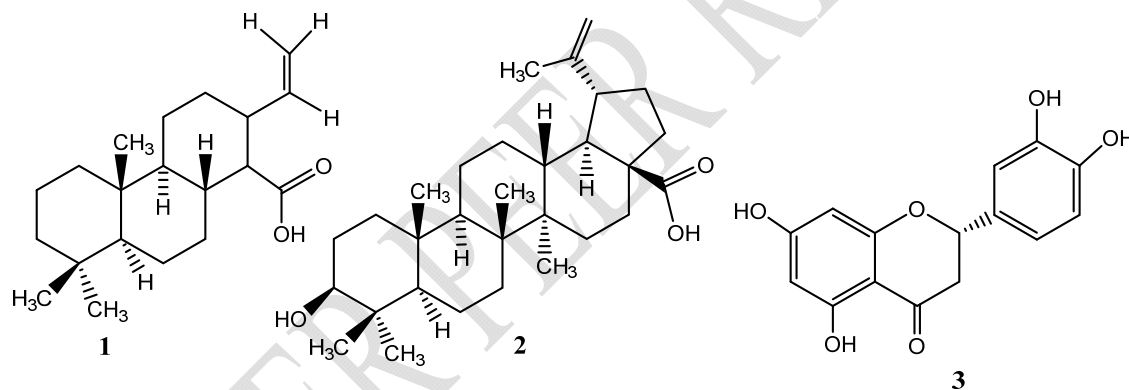
57 “Epoobo” among Yoruba people of South Western Nigeria. The stem-bark and roots of *E.*
58 *ivorensis* are particularly used in the treatment of convulsive pain, disorders, edema, emesis,
59 constipation, smallpox as well as helminthic infestations [12]. A 70% ethanol extract of the
60 stem bark of the plant has been reported to show moderate activity against a wide range of
61 gram positive and gram negative organisms [13]. Wakeel et al., [14] reported on the anti-
62 convulsant and sedative properties of *E. ivorensis* stem bark extract. We have previously
63 reported on the anti-inflammatory activity of the novel phytochemical, erythroivorensin,
64 together with eriodictyol and betulinic acid isolated from the plant [15]. Additionally, we
65 have earlier reported on the leishmanicidal activity of the root bark of the plant and
66 identification of some of its compounds by ultra-performance liquid chromatography
67 quadrupole time of flight mass spectroscopy (UPLC-QTOF-MS/MS)[16]. Despite the fact
68 that the effect of the leaf and stem bark extracts of *Erythrophleum ivorensis* have been
69 screened for antishistosomal activity against *Schistosoma mansoni* [17], this current research,
70 in addition to using the various fractions of the root bark of the plant, focusses also on three
71 isolated bioactive compounds: erythroivorensin, betulinic acid and eriodictyol against
72 immature infective stage of *Schistosoma haematobium* Cercariae.

73 **2.0 Materials and Methods**

74 **2.1 Plant collection and extraction**

75 The root bark of *Erythrophleum ivorensis* was harvested from Adukrom in Nzema-East
76 Metropolis of Ghana, in August 2017 and was authenticated using an earlier collected
77 samples with voucher number BHM/Eryth/017R/2014, which had been deposited at the
78 Herbarium unit of the Department of Herbal Medicine, Kwame Nkrumah University of
79 Science and Technology, Kumasi-Ghana.

80 The root bark of *E. ivorensis* collected was air dried at room temperature (25–27 °C) for two
81 weeks. The dried root bark was pulverized by milling into a coarse powder. 1 kg of the
82 powdered air-dried root bark was cold macerated with 70% ethanol for 72 hours. The
83 resulting extract was filtered and concentrated under reduced pressure (40 °C) using rotary
84 evaporator (Buchi Rotavapor, R 200) to give a crude yield of 9% ^{w/w}. 80 g of the plant extract
85 was successively partitioned with petroleum ether (4 L), ethyl acetate (4 L) and methanol (4
86 L) to obtain three fractions with the yield of 5.8 g, 22.7 g and 38.3 g respectively. Activity-
87 guided isolation and characterization carried out as described previously [15] yielded the
88 following pure compounds: erythroivorensin (1), betulinic acid (2) and eriodictyol (3) as
89 shown in Figure 1.



90
91 Figure 1: Chemical structure of erythroivorensin (1), betulinic acid (2) and eriodictyol (3) compounds isolated.

92

93 2.2 Collection of snails

94 The snails, *Bulinus species*, the intermediate host for *S. haematobium*, were collected from
95 endemic areas in their natural habitats from Tomefa along the Weija River in Ghana. The
96 snails were kept in a plastic aquarium with 50 snails per each aquarium containing clean pond
97 water at room temperature (25 °C) and fed with lettuce at the Biomedical Science Laboratory

98 of University of Cape Coast, Ghana. They were later washed with deionised water and
99 examined for cercariae shedding using inverted microscopy as described previously by
100 Amoani et al. [18].

101 **2.3 *In vitro* Cercaricidal Activity Test**

102 Cercaricidal effect of the various concentrations (15.6, 31.3, 62.5, 125.0, 250.0 and 500.0
103 µg/mL) of the crude (70% ethanolic) extract, its fractions (methanol, ethyl acetate and pet-
104 ether) pure compounds (erythroivorensin, betulinic acid and eriodictyol) of the root bark of *E.*
105 *ivorensis* as well as control praziquantel were evaluated as described previously [4].

106 An average of 20 freshly shed cercariae were transferred into each of the 20 well plates
107 (Costar) using micropipette. Various concentrations of the extracts and bioactive compounds
108 were freshly prepared and transferred into one well on the plate. The negative control well
109 contained the same number of cercariae and distilled water only. All experiments were
110 carried out in triplicates. Mobility and viability of the *Schistosoma* infectious stage
111 (cercariae) were observed for 3 hours.

112 Unaffected free-swimming larvae, immobile and dead cercariae at the bottom of the wells
113 were observed at 4× magnification using an inverted microscope (Olympus CK 300).
114 Survival and mortality at a successive interval of 15, 30, 60, 120, and 180 min were recorded.
115 Cercariae were presumed dead when they stopped moving and sank down and their tail were
116 detached.

117 The % viability was calculated using the equation below and this was used to plot the
118 survival curves for each of the fractions and compounds.

$$119 \quad \% \text{ Viability} = \left(\frac{\text{Initial count of live cercariae} - \text{number of dead cercaria}}{\text{Initial count of live cercariae}} \right) \times 100$$

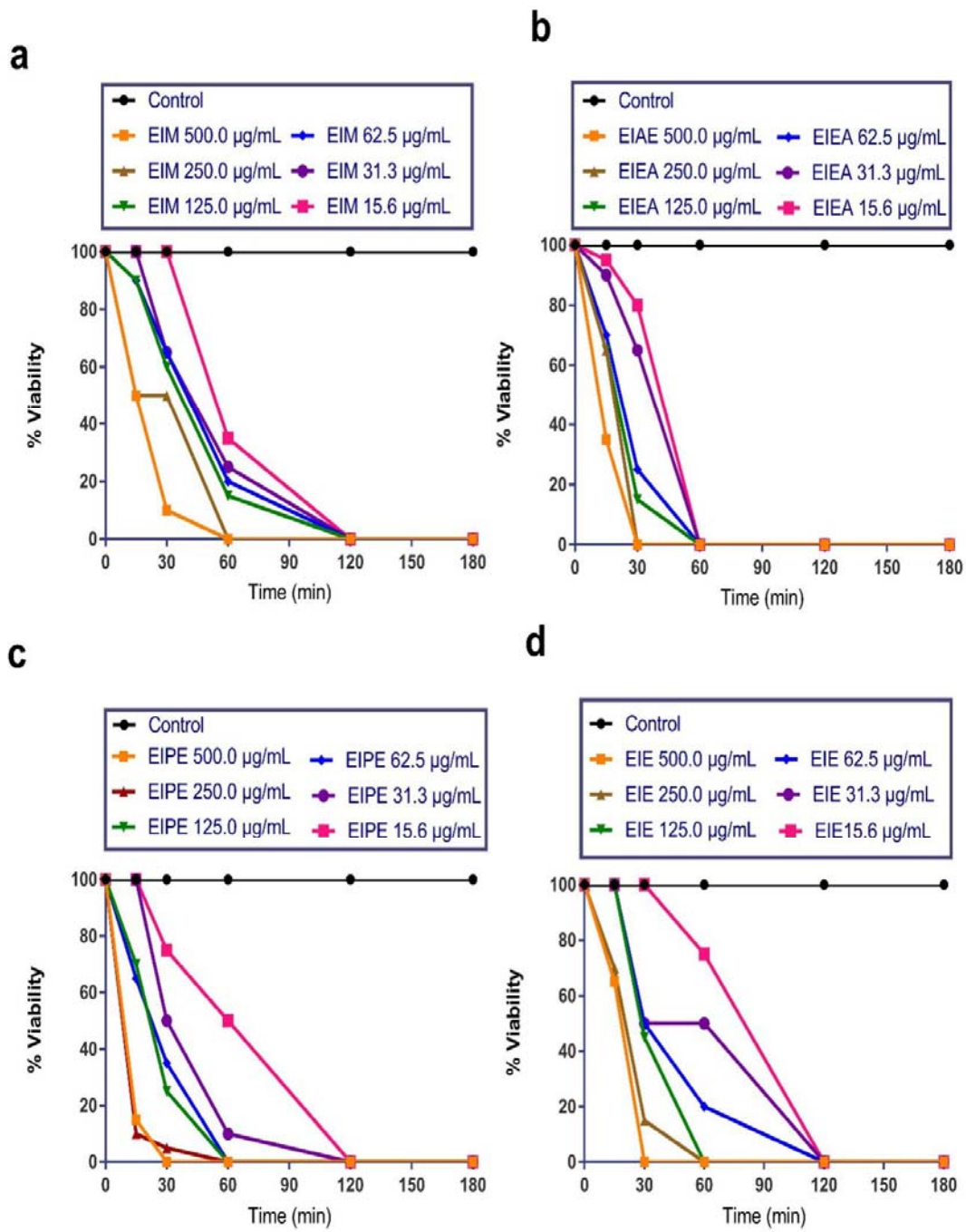
120 **Statistical analysis**

121 Data was presented as mean \pm standard error of mean (SEM). Graphpad® Prism Version 7.0
122 (Graphpad Software, San Diego, CA, USA) for Windows was used to perform all statistical
123 analysis. Time-course curves of percentage viability of the plant extracts against time was
124 plotted. The equation (1) above was used to calculate the percentage viability for each
125 treatment. The concentration at which 50% of the cercariae were inhibited referred to as IC₅₀
126 was determined by plotting a nonlinear regression curve (log concentration of inhibitor verses
127 % viability).

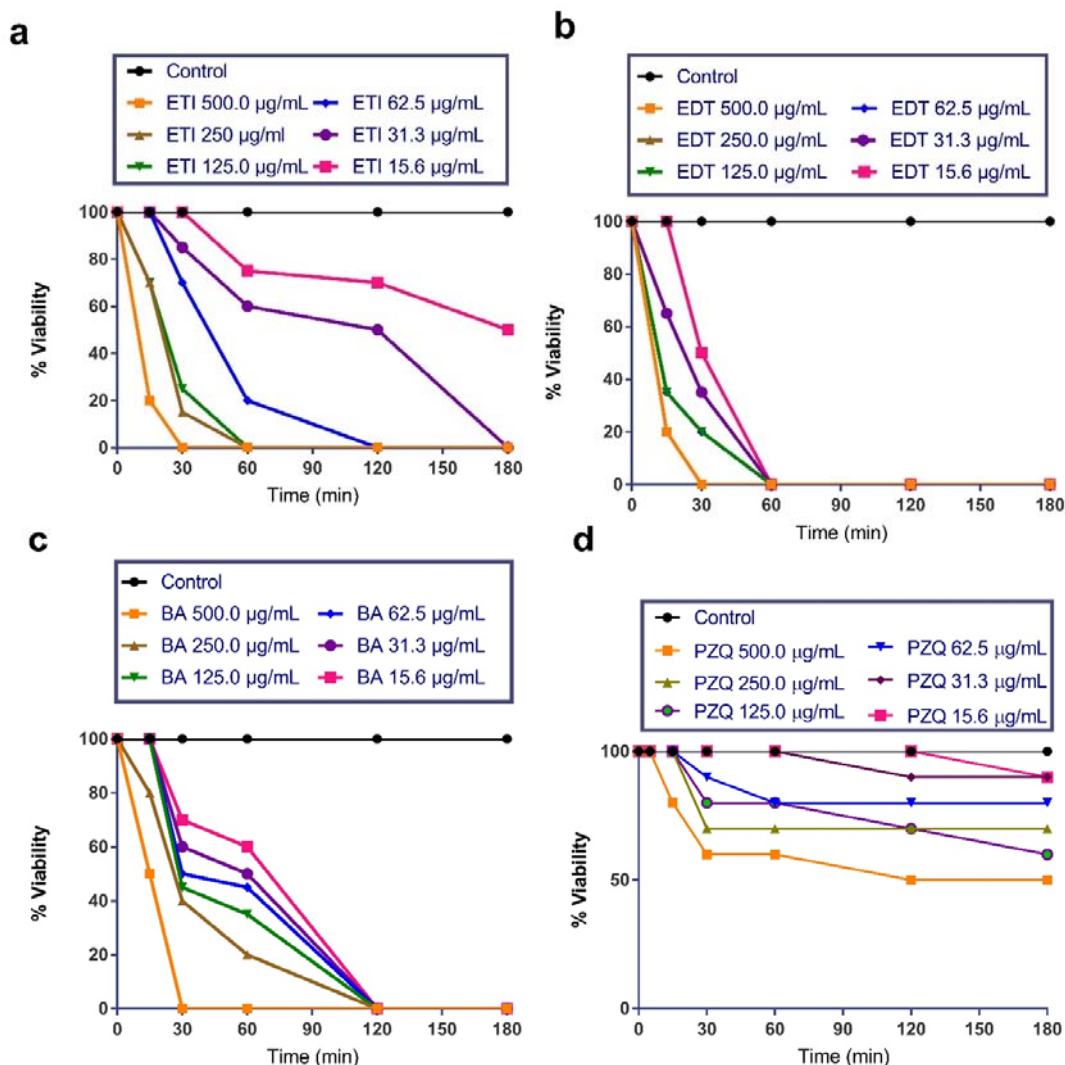
128 **3.0 Results and Discussion**

129 Exposure of *S. haematobium* cercariae to the crude hydro-ethanolic extract of *E. ivorense*, its
130 fractions and compounds, showed concentration dependent increase in mortality (Figures 2-
131 4). The ethyl acetate fraction and one of its isolates, eriodictyol, showed higher mortality rate
132 than the other fractions and compounds tested against the cercariae of *S. haematobium*. With
133 the exception of erythroivorensin and betulinic acid (at 15.6 $\mu\text{g/mL}$), all the various fractions
134 and eriodictyol at all concentrations achieved 100% mortality of cercaria within 180 min of
135 incubation (Figures 2 and 3). In the absence of the plant extract, cercariae showed normal
136 viability without any morphological changes (tail loss) throughout the entire duration of the
137 experiment as was observed in the control sample. Though 40% mortality of cercariae was
138 achieved at the maximum concentration of praziquantel (PZQ 500 $\mu\text{g/mL}$), none of the
139 various concentrations of the standard antischistosomal drug could eliminate all the cercariae.

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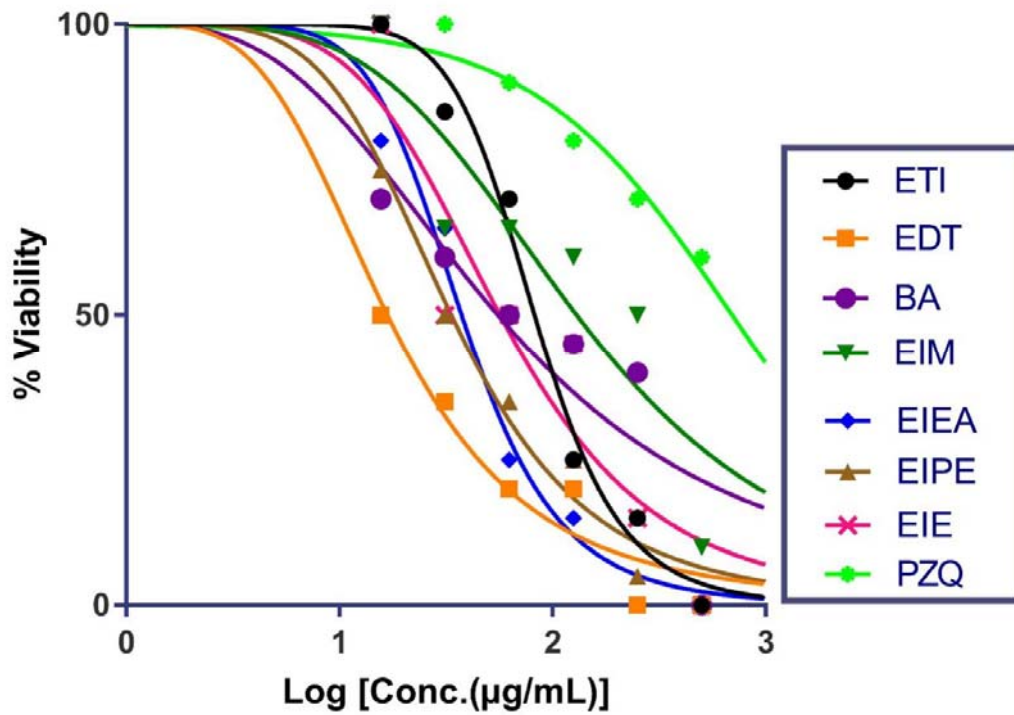
141
 142 Figure 2: Effect of different concentrations of (a) methanol (EIM) (b) ethyl acetate (EIEA) (c) petroleum ether
 143 (EIPE) fractions and (d) 70% crude ethanol (EIE) extract of *E. ivorense* root bark on the viability of *S.*
 144 *haematobium* cercariae



145
 146 Figure 3: Effect of different concentrations of (a) erythroivorenin (ETI) (b) eriodictyol (EDT) (c) betulinic acid
 147 (BA) isolated from the root bark of *E. ivorense* and (d) praziquantel (PZQ) on the viability of *S. haematobium*
 148 cercariae.

149 The dose response curves of the effects of the various fractions and isolated compounds from
 150 *E. ivorense* on *S. haematobium* cercariae demonstrates that the activity of these isolates and
 151 compounds are dose-dependent. The cercaricidal activities of the various fractions and
 152 extracts were quantified using IC_{50} . From the results presented on Table 1 and Figure 4,
 153 eriodictyol was found to be most potent with an IC_{50} of 1.23 $\mu\text{g/mL}$ whereas the methanol
 154 fraction was found to be the least potent with IC_{50} of 2.11 $\mu\text{g/mL}$. The activity of the ethyl

155 acetate fraction was higher than the total crude ethanol extract but lower than its isolate
156 eriodictyol. Thus purification of the ethyl acetate fraction afforded higher anti-cercarial
157 activity.



158

159 Figure 4: Dose-response curves of the effects of the crude extract (EIE), various fractions (EIM, EIEA, EIPE)
160 isolated compounds (ETI, EDT and BA) from *E. ivorensis* and praziquantel (PZQ) on *S. haematobium* cercariae.

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167 Table 1. IC₅₀ values of fractions and isolated compounds from *E. ivorense*.

Compound/Fraction	IC ₅₀ (μg/mL)
EIE	1.75 ± 0.08
EIM	2.11 ± 0.10
EIEA	1.53 ± 0.02
EIPE	1.59 ± 0.03
ETI	1.92 ± 0.02
EDT	1.23 ± 0.05
BA	1.74 ± 0.10
PZQ	695.50 ± 0.05

168 Crude ethanol (EIE), Methanol (EIM), ethyl acetate (EIEA), petroleum ether (EIPE) extracts, erythroivorenin
169 (ETI), eriodictyol (EDT), betulinic acid (BA) and Praziquantel (PZQ).

170

171 This current study investigated cercaricidal activities of methanol, alcoholic, petroleum ether,
172 ethyl acetate fractions and isolated compounds (erythroivorenin, betulinic acid and
173 eriodictyol) obtained from *Erythrophleum ivorense* on *Schistosoma haematobium* cercariae *in*
174 *vitro*. We have earlier reported the anti-inflammatory and anti-leishmanial activity of these
175 compounds and fractions from the plant [15, 16]. It is an indication that the plant will have an
176 activity against cercaria from *Schistosoma haematobium*, another parasitic disease. Also, its

177 anti-inflammatory property is essential since inflammation is an important component of
178 infectious diseases [19]. The current study has demonstrated that the various fractions and
179 compounds isolated from the plant have potent cercaricidal activity and that ethyl acetate
180 fraction and the compound eriodictyol are the most potent. It is not surprising that the
181 compounds cassane diterpene erythroivorensin, triterpene betulinic acid and flavanone
182 eriodictyol which showed marked activity were all isolated from the ethyl acetate fraction of
183 the plant.

184 The results obtained indicate a potent cercaricidal activity of the various fractions and
185 compounds with ethyl acetate fraction and the compound eriodictyol being the most potent.
186 The cassane diterpene erythroivorensin, triterpene betulinic acid and flavanone eriodictyol
187 which showed marked activities were all isolated from the ethyl acetate fraction of the plant.
188 The cercaricidal activity of the flavanone eriodictyol was relatively higher than that of the
189 ethyl acetate fraction implying that the erythroivorensin and betulinic acid had a relatively
190 little effect on the cercaricidal ability of the extract. The crude ethanolic extract
191 comparatively recorded lower activity than its ethyl acetate fraction, probably because some
192 compounds, present in the root bark, may have antagonistically functioned to reduce the
193 cercaricidal potency of the extract. That notwithstanding, the crude alcoholic extract, various
194 fractions and isolated compounds produced 100% mortality of *Schistosoma haematobium*
195 cercariae at higher concentrations within the 3 h study period.

196 Thus the present study has highlighted the ethyl acetate fraction and its flavanone constituent
197 eriodictyol as clear drug candidates in the development of agents to obstruct the life cycle of
198 the parasite through its asexual aquatic stage (cercaria) and thus could be considered in
199 biological control programs. In Ghana and other African countries, due to the large
200 dependence of the populace on herbal medicine use, consideration could be made in
201 formulating the ethyl acetate fraction or eriodictyol as an ointment to be used prior to decent

202 into these water bodies. Research into the safety of these products on other aquatic life is thus
203 welcome.

204 Praziquantel, the most commonly used antischistosomal drug, increases the permeability of
205 the membranes of Schistosome cells towards calcium ions. It induces contraction of the
206 parasites which results in paralysis in the contracted state and also causes focal
207 disintegrations [20]. However, this effect is not well expressed in cercariae hence its
208 ineffectiveness against cercariae as was observed in the results presented in Figure 3. The
209 extracts and isolated compounds of *E. ivorensis*, caused focal disintegration (loss of tail) and
210 paralysis of the cercariae and subsequently, death. Further research on possible mechanism of
211 action of the fractions and compounds isolated from the plant is recommended. Since
212 standard anti-cercarial agents are not widespread, the present study brings to the fore,
213 extracts, fractions and compounds of *E. ivorensis* as potential biological drug leads for the
214 development of eco-friendly cercaricides for the mitigation of schistosomiasis. This will help
215 reduce the incidence and prevalence of the second most important human parasitic disease after
216 malaria, on the wane.

217 **4.0 Conclusions**

218 The various fractions and compounds of *Erythrophleum ivorensis* exhibited a marked
219 cercaricidal activity. Thus, the study may provide some scientific justification for the
220 ethnomedicinal uses of the root bark of *Erythrophleum ivorensis* in Ghana. Therefore, it is
221 recommended that the isolated bioactive compounds of this plant should be further evaluated
222 and developed into a cercaricidal formulation for prophylactic use especially before one
223 descends into infested water body.

224 **Ethical consideration**

225 All authors hereby declare that "Principles of Laboratory Animal Care" (NIH
226 Publication No. 85-23, Revised 1985) were followed. All protocols used in the study
227 were approved by the Department of Biomedical Sciences' ethics committee.

228 **Conflicts of Interest**

229 The authors declare that there is no conflict of interest regarding the publication of this paper.

230 **Data Availability Statement**

231 The authors declare that all data have been included in the manuscript.

232

233 **References**

- 234 [1] WHO. (2016). Schistosomiasis facts sheet. Retrieved from
235 http://www.who.int/neglected_diseases/resources/schistosomiasis/en/&ved
- 236 [2] J. M. Naples, C. Shiff, and R. U. Halden. "Reduction of infectivity of schistosome
237 cercariae by application of cercaricidal oil to water." *The American Journal of*
238 *Tropical Medicine and Hygiene*, vol. 73, no. 5, pp. 956-961, 2005
- 239 [3] M. Mengistu, T. Shimelis, W. Torben, A. Terefe, T. Kassa, and A. Hailu. "Human
240 intestinal schistosomiasis in communities living near three rivers of Jimma town,
241 south Western Ethiopia." *Ethiopian Journal of Health Sciences*, vol. 21, no. 2, pp.
242 111-118, 2011.
- 243 [4] E.M. Tekwu, K. M. Bosompem, W. K. Anyan, R. Appiah-Opong, K. B.-A. Owusu,
244 M. D. Tettey, F. A. Kissi, A. A. Appiah, V. P. Beng, and A. K. Nyarko. "In vitro
245 assessment of anthelmintic activities of *Rauwolfia vomitoria* (Apocynaceae) stem
246 bark and roots against parasitic stages of *Schistosoma mansoni* and cytotoxic
247 study." *Journal of Parasitology Research*, vol. 2017, pp. 1-11, 2017, Article ID
248 2583969.
- 249 [5] B. A. Obare. "Evaluation of Cercaricidal and Miracidal Activity of Selected Plant
250 Extracts Against Larval Stages of *Schistosoma Mansoni*." *Journal of Natural*
251 *Sciences Research*, vol. 6, no. 22, 24–31, 2016.

- 252 [6] D.U. Olveda, Y. Li, R.M. Olveda, A.K. Lam, D.P. McManus, T.N. Chau, D.A. Harn,
253 G.M. Williams, D.J. Gray and A.G. Ross. “Bilharzia in the Philippines: past, present,
254 and future.” *International Journal of Infectious Diseases*, vol. 18, pp.52-56, 2014.
- 255 [7] D. P. McManus, and A. Loukas. “Current status of vaccines for schistosomiasis.”
256 *Clinical Microbiology*, vol. 21, pp. 225–242, 2008.
- 257 [8] D. Cioli, L. Pica-Mattocchia, A. Basso, and A. Guidi. “Schistosomiasis control:
258 praziquantel forever.” *Molecular Biochemistry Parasitology*, vol. 195 pp. 23–29,
259 2014
- 260 [9] A. A. Hassan, A. E. Mahmoud, R. A. Hassan and E. M Huseein, “Evaluation of
261 Euphorbia aphylla, Ziziphus spina. Christi and Enterolobium contortisliquum as
262 molluscicidae agents.” *Journal of American Science*, vol. 7, pp. 511–520, 2011.
- 263 [10] S. Wachtel-Galor and I.F.F. Benzie. Herbal medicine: an introduction to its history,
264 usage, regulation, current trends, and research needs. In: Benzie IFF, Wachtel-Galor
265 S, editors. Herbal medicine: biomolecular and clinical aspects. 2nd ed. Boca Raton
266 (FL): CRC Press, Taylor & Francis; p. 1–10, 2011.
- 267 [11] O. Kayser, A. F. Kiderlen, and S. L. Croft. ‘Natural products as antiparasitic drugs.’
268 *Parasitology Research*, vol. 90, no. 2, pp. S55–S62, 2003.
- 269 [12] Oliver-Bever, B. Medicinal plants in tropical West Africa: Cambridge university
270 press, 1986.
- 271 [13] L. Adu-Amoah, E. Kesseih, C. Agyare, and A. Hensel. Antimicrobial and cytotoxicity
272 studies of the methanolic extracts of Erythrophleum ivorense leaf and stem bark.
273 *Planta Medica*, 79(13), pp. 1153, 2013.
- 274 [14] O.K. Wakeel, S. Umukoro, O.T. Kolawole, E.O. Awe, O.G. Ademowo, “Anti-
275 convulsant and sedative activities of extracts of Erythrophleum ivorense stem bark in
276 mice.” *Asian Journal of Biomedicine and Pharmaceutical Sciences*, vol. 4, pp. 43–47,
277 2014
- 278 [15] F. A. Armah, K. Annan, A. Y. Mensah, I. K. Amponsah, D. A. Tocher, and S.
279 Habtemariam. Erythroivorenin: A novel anti-inflammatory diterpene from the root-
280 bark of Erythrophleum ivorense (A Chev.). *Fitoterapia*, vol. 105, pp. 37-42, 2015.
- 281 [16] F. A. Armah, I. K. Amponsah, A. Y. Mensah, R. A. Dickson, P. A. Steenkamp, N. E.
282 Madala, and C. K. Adokoh. “Leishmanicidal activity of the root bark of
283 Erythrophleum Ivorense (Fabaceae) and identification of some of its compounds by
284 ultra-performance liquid chromatography quadrupole time of flight mass

- 285 spectrometry (UPLC-QTOF-MS/MS)". *Journal of Ethnopharmacology*, vol. 211, pp.
286 207-216, 2018.
- 287 [17] G. Kyere-Davies, C. Agyare, Y. D. Boakye, B. M. Suzuki, and C. R. Caffrey. "Effect
288 of Phenotypic Screening of Extracts and Fractions of *Erythrophleum ivorense* Leaf
289 and Stem Bark on Immature and Adult Stages of *Schistosoma mansoni*." *Journal of*
290 *Parasitology Research*, vol. 2018, 2018.
- 291 [18] B. Amoani, E. O. Ameyaw, D-B. Asante, F. A. Armah, J. Prah, C.P.K. Botchey and
292 J.N. Boampong. Effect of pre-existing *Schistosoma haematobium* infection on
293 *Plasmodium berghei* multiplication in Imprinting Control Region (ICR) mice. *Asian*
294 *Pacific Journal of Tropical Biomedicine*, vol. 5 no. 1, pp. 930-934, 2015
- 295 [19] E. Ricciotti and G. A. FitzGerald. Prostaglandins and inflammation. *Arteriosclerosis,*
296 *Thrombosis and Vascular Biology*, vol. 31, no. 5, pp. 986-1000, 2011.
- 297 [20] K. Wolters. "Praziquantel." The American Society of Health-System Pharmacists.
298 2016 Retrieved from <http://www.drugs.com/monograph/praziquantel.html>