Exposure assessments of internally displaced infants to Aflatoxin \mathbf{M}_1 through breast milk feeding, in Damaturu Yobe State.

¹Gide S ¹Warodi F. A ²Alegbe S.D ³Anas G

- Desert Research Monitoring and Control Centre, Yobe State University Damaturu P.M.B 1144.
 - 2) Department of Microbiology, Ahmadu Bello University Zaria
 - 3) Nigerian Institute for Trypanasomiasis Research Kebbi State

gidesuleimans@yahoo.com

Abstract

Aflatoxin M_1 is a biomarker for the detection of breast milk contamination and also a risk factor for early infant's exposure to the toxin. Exposure assessment of 50 internally displaced infants to aflatoxin M_1 through breast-milk feeding was carried out between (June 2016 to October 2016), High performance liquid chromatography (HPLC) was used to evaluate the level of AFM₁ in mother's breast milk samples and the infant's urine samples respectively. Results obtained from the study showed that 96% of the breast milk samples have maximum concentration of $0.0879\mu g/L$ with mean value of $0.0582\mu g/L$ while, the minimum and maximum excretion concentration of AFM₁ in urine sample of infants was $0.0400\mu g/L$ and $0.0651\mu g/L$ respectively with mean value of $0.05005\mu g/L$ at 88%.

The study indicates that the occurrence of AFM_1 in breast milk samples of mothers with the types of food the consumed within 24-48hrs prior to sample collection that predispose the infant's exposure to AFM_1 showed 40% of the women consumed rice and 32% consumed local food

(brabisko/biski) and 24% consumed corn meal with statistically significant with P value less

than 0.05. From the study the 96% of the infants were exposed to the toxin while 18% of the

infants were undetectable this possess a concern that internally displaced infants that were on

admission in the selected facility where exposed to AFM₁.

Keywords: IDP, breast milk, Aflatoxin M1, HPLC

Introduction

Breast feeding is the best and recommended form of infant feeding. Six month is the

recommended period of exclusive feeding for infants. Breastfeeding promotes the mother-child

relationship and ensures better growth and development of the neonates given them required

nutrients, strengthen antibodies and leukocytes (Brasil, 2010; Ishikawa 2016). Aflatoxins are

metabolites of a Fungus of Aspergilus flavus and Aspergilus parasiticus there are various types

of Aflatoxin discovered meanwhile the commonest are Aflatoxin B₁, B₂, G₁,G₂, M₁ and M₂ are

the secondary metabolites of B₁ and B₂ respectively (Bianco et. al.2012). Aflatoxin M₁ (AFM₁)

as a biomarker of Aflatoxin B₁ (AFM₁) can be used to evaluate aflatoxin exposure through diet

for both humans and animals. As a metabolite human exposure to aflatoxin (AFs) occurs through

the intake of contaminated agricultural products or the consumption of products from animals

that were fed with contaminated feed. This contamination may occur by fungal growth during

harvest or improper storage (Ghiasian, 2012). AFB₁ is considering the most toxic among all

other forms of Aflatoxin (Bhet et al., 2010).

Toxicological evaluation of aflatoxin over food intake is essential to any risk evaluation and

important for determining the relationship observed in humans and exposure to aflatoxin

(Leblanc et. al. 2005: Shundo et. al., 2009). AFB₁ is metabolized in animals and the human liver

into AFM₁ by Cytochrome P450–associated enzymes, and then distributed in serum and excreted into milk and urine. In mammals, AFM₁ can be detected in milk within 12 h after the ingestion of AFB₁ (Battacone *et. al.*,2003 and 2012).

The maximum limits for fluid milk and powdered milk should be 0.5 and 5 ng/g, respectively (E U, 2001). A technical regulation on the Maximum Tolerated Limit for Mycotoxins in Food has not been defined regarding whether the fluid milk is raw or pasteurized. The European guidelines established the maximum AFM₁ levels in both raw milk (0.05 ng/l) and infant formulae (0.025 ng/l) (E C, 2010). Based on the potential hazard to infant (zero to 12 months old) health due to carry-over of the aflatoxin biomarker (AFM₁) into milk, the aim of this study was to evaluate the exposure of infants to AFM₁ through consumption of breast milk.

Materials and Method

Study Area: the study area comprises Yobe State Specialist Hospital located in Damaturu in the north eastern of Nigeria.

Ethical Approval and Parents Consent

Ethical clearance was obtained from the ethical and scientific research committee of the ministry of health. A consent form was given to obtain permission from mothers whose babies were on admission. Only those who gave consent were recruited in the study.

Sample size and Study Population

This study was conducted on 50 internally displaced lactating mothers whose infants were on admission in Yobe State Specialist Hospital Damaturu, Yobe State.

Breast Milk Collection:

Breast milk samples were collected by hand expression into glass tubes; the milk samples were stored at 15°C. Samples were thawed gradually to 4°C and then centrifuged at 10°C at 15rpm. Aflatoxins are water-soluble; hence, the upper creamy layer was discarded and the lower phase was used for the quantitative test (Adejumo 2013).

Structured Questionnaire:

Structured questionnaire was design to determine the kinds of food lactating mothers consumed within 24-48hrs that associate them to Aflatoxin exposure.

High performance liquid Chromatography (HPLC), Analysis for Aflatoxin M₁

The concentration of AFM₁ in breast milk was estimated by HPLC configured with LC-10AD pumps, coupled with tungsten detector RF-10Axl. Excitation and emission wavelengths were set at 350 and 440 nm respectively. The stationary phase was a Gemini Column. The mobile phase was isocratic mixture of methanol/acetonitrite/water (25:25:50 v/v/v), with a flow rate of 1ml/min and chromatographic run time of 10 min. The values obtained for recoveries and relative standard deviations of the methods of analysis were in agreement with Commission Regulation (EC) No. 401/2006 for methods of analysis of mycotoxins in foodstuffs (European Union Commission, 2006).

An injector with 50 µl loop was used for the determination of AFM₁. A calibration curve was constructed for AF M₁ using different levels of toxin concentrations with an average of 10 consecutive automated injections of standard solutions of AF M₁ purchased from SUPELCO solutions within USA Involving series of dilutions (Adejumo, 2013).

Preparation of urine samples for high performance liquid chromatography (HPLC)

The extraction and purification of urine samples for AFM₁ determination were performed as follows: A 30 ml volume from the urine sample was filtered through a glass microfiber filter paper. Later, 20 ml of filtered extract was transferred to a 50 ml capacity vial and 20 ml of sodium acetate buffer (pH 5.0) was added. The pH of the mixture was measured and corrected to 5.0 using an appropriate volume of a 0.1 M glacial acetic acid solution whenever necessary. The mixture was directly passed through an immunoaffinity column at a flow rate of approximately 1.0-1.5 ml min⁻¹. After adding the mixture the column was washed with 40 ml of ultrapure water. The column was dried by applying positive pressure with a syringe and bound AFM₁was eluted with 2.0 ml of HPLC-methanol which was recovered in a 4 ml vial previously treated with acid. The eluate was evaporated under nitrogen gas and reconstituted with 500 µl of the mobile phase before liquid chromatography analysis. Detection and quantification of sample extracts were performed by high-performance-liquid chromatography (HPLC) with a liquid chromatography system equipped with a LC-10AT Shimadzu pump, a Shimadzu RF-10AXL fluorescence detector (excitation 250 nm and emission 350 nm), an injection volume of 100µl, and a reverse phase column (250- 4.6mm, particle size of 3 µm) and pre column kept at room temperature. The mobile phase consisted of an isocratic mixture of water and acetonitrile at a volume ratio of 75:25 and a flow rate of 1.0 ml min⁻¹. A calibration curve was prepared using standard AFM₁ solutions in mobile phase at concentrations of 0.005, 0.01, 0.02, and 0.03 ng ml⁻¹ the standard obtained (SUPELCO solutions within USA.) as purified crystalline AFM1 was dissolved in HPLC-grade acetonitrite and its concentration was determined spectrophotometer.

Statistical Analysis of the Data

The Statistical Package for Social Science program (SPSS, Chicago, Illinois, USA) version 21 was used for analysis of data. Data was summarized as mean, standard deviation, for the analysis of the difference in quantitative data between the two groups. The X²-test was used for the analysis of qualitative data. *P value* was considered significant if it was less than 0.05 Simple linear correlations (Pearson's correlation for quantitative data Tumerak, 2011).

Result

Analytical performance

The limit of detection (LOD) for AFM₁ was estimated at $0.01\mu g/mL$ and the limit of quantification (LOQ) was $0.05\mu g/mL$. The linearity of the curve was $0.01-0.05 \mu g/mL$. The calibration curve for AFM₁ had a linear equation of y = 4147.x - 230.3; Fig 1 gives the calibration curve with a correlation coefficient $R^2 = 0.993$ and retention time of 10.0 min.

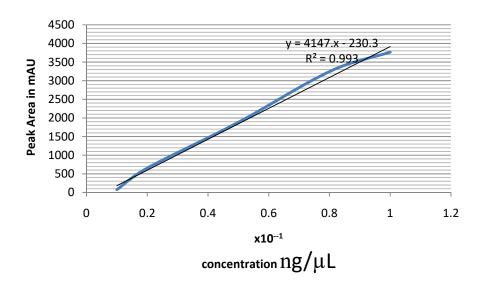


Fig 1 gives calibration curve of AFM₁ with correlation coefficient.

Table 1:Occurrence and concentration of AFM₁ in breast milk consume by infants and excretion in their urine.

Samples	No examined	No. positive (%)	Min conc. (μg/L)	Max conc.(μg /L)	Mean conc(μg/L) (±SD)	Chi square	p- value
Breast Milk	50	48(96)	0.0457	0.0879	0.0582 (±0.012)	2.204	0.02
Infant's Urine (0-6 months)	50	44(88)	0.0400	0.0651	0.05005(±0.0 04781)		

SD.Standard deviation of descriptive quantities

Table 2: Occurrence of AFM₁ in breast milk samples of mothers with the types of food

consumed within 24-48 hours prior to sample collection.

Consumed	No examined	No. positive	Chi square	p-value
foods in 72 hrs	N=50	(%)		
Brabisko/Biski				
(local food)				
Yes	16	(32.0)	8.245	0.045
No	0	(0.0)		
Rice				
Yes	20	(40.0)	9.123	0.034
No	0	(0.0)		
Cornmeal				
Yes	12	(24.0)	10.233	0.013
No	2	(4.0)		

The percentage occurrence and concentration of AFM₁ in breast milk consumed by infants and excretion in their urine as seen in Table 1 shows that 96% of the breast milk consumed by IDP infants have the toxin with minimum concentration of $0.0457\mu g/L$ and maximum concentration of $0.0879\mu g/L$ and the mean concentration of $0.0879\mu g/L$ meanwhile, the minimum and

maximum excretion concentrations of AFM $_1$ in urine samples of infants gave $0.0400\mu g/L$ and $0.0651\mu g/L$ respectively with mean value of $0.05005\mu g/L$ at 88% exposed of the toxin from their mother's breast milk .

Occurrence of AFM₁ in breast milk samples of mothers with the types of food the consumed within 24- 48hrs prior to sample collection that predispose the ID infants exposure to AFM₁ has been illustrated in Table 2; 40% of the women consumed rice and 32% consumed local food (brabisko/biski) and 24% consumed corn meal with statistically significant with P value less than 0.05.

Discussion

Exposure assessment of internally displaced infants to aflatoxin M₁ through consumption of breast milk in Damaturu was 96% with maximum concentration of 0.0879μg/L of the toxin. The mean value concentration of AFM₁ in the breast milk samples was 0.0582μg/L this may be explained that the mothers were exposed to the toxin in their diet. Considerably, another independent screening by Adejumo *et al* (2013) report thatoccurrence of AFM₁ was 82% inbreast milk samples in Ogun state, Nigeria which is lower. While Abdulrazzaq *et al*. (2003) and Sadeghi *et al*. (2009) in United Arab Emirate and Iran respectively reported 99.5% and 98.1% frequencies which is higher. The percentage occurrence in this study is higher than that reported in Cameroun and Egypt, 48%, and 56% respectively (Tchana *et al.*, 2010; Polychronaki *et al.*, 2007). This could be as a result of differences in the type of food consumed, storage condition of foods and lifestyle of the people as well as level of contamination with the secreting *Aspergillus species*.

Infant examined are between the age brackets 0-6 month with minimum concentration of 0.04μg/L and maximum concentration of 0.0651μg/L with mean excretion concentration of 0.05005μg/L. This high concentration implies that the infant were exposed to high level of aflatoxin which is indicative from the levels seen in their mothers' breast milk samples a major food source for the infant which are expected to be breast feed for six months. 88% of the examined children shown exposure to AFM₁ which implies that 18% were undetected in the urine samples of the infant which is in line with the study reported in Brazil by Giolo *et al.*2012. This result can be compared with other researchers that there is a relationship between food intake and urinary excretion of AFM₁ (Mason *et al.*, 2015). Redzwan *et al* (2012) reported that there was significant relationship between consumption of AFM₁ in milk and excretion in the urine.

Conclusion

The mean concentration of examined breast milk samples was $0.0582\mu g/L$. It should be noted that, the concentration of AFM₁ in all the breast milk samples used were higher than the acceptable tolerance level of $0.025\mu g/L$ and $0.05\mu g/L$ for infants milk by the European Communities and Codex Alimentarius respectively. This possess a concern on internally displaced infants that were on admission in the selected facility where exposed to AFM₁. This implies that AFM₁ has the potential to be a public health problem in Damaturu.

References

- Abdulrazzaq, Y.M.,Osman, N.,Yousif, Z.M., Al-Falahi, S.(2003). Aflatoxin M₁ in Breast Milk of UAE women. *Ann.Trop. paediat.*,23:173-179.
- Adejumo, O., Atanda. O., Raiola, A., Somorin, Y., Bandyopadhyay, R. and Ritieni, A., (2013).

 Correlation between Aflatoxin M₁ content of breast milk, dietary exposure to aflatoxin B₁ and socioeconomic status of lactating mothers in Ogun State, Nigeria: Food and Chemical Toxicology, 56:171-177
- Angele N Tchana, Paul F Moundipa, Felicite M Tchouanguep Aflatoxin Contamination in foods and body fluids in relation to malnutrition and cancer status in Cameroun International Journal of Environmental Research and Public Health 7(1), 178-188, 2010
- Barbieri, G., Bergamini, C., Ori, E. ans Pasca, P. (1994). Afaltoxin M₁ in parmesan cheese: HPLC determination *J Food Sci.*, **56** (6):1313-31.
- Battacone G., Nudda A., Cannas A., Borlino A.C., Bomboi G., Pulina G.(2003) Excretion of aflatoxin M₁ in milk of dairy ewes treated with different doses of aflatoxin B₁. *J. Dairy Sci.*;86:2667–2675.
- Battacone G., Nudda A., Rassu S.P.G., Decandia M., Pulina G. (2012) Excretion pattern of aflatoxin M₁ in milk of goats fed a single dose of aflatoxin B₁.J. Dairy Sci.;95:2656–2661.

- Bhat, R., Rai, R.V.andKarim, A. A. (2010). Mycotoxins in food and feed: present status and future concerns. *Comprehens Rev. Food Sci. Food Safety*, 9:57–81.
- Biancardi A.(1997). Determination of aflatoxin M₁ residues in milk: A comparative assessment of ELISA and IAC-HPLC methods. In *Alimnet*., 36:870-876.
- Bianco, G., Russo, R., Marzocco, S., Velotto, S., Autore, G. and Severino, L. (2012). Modulation of macrophage activity by aflatoxins B₁ and B₂ and their metabolites aflatoxins M₁ and M₂. *Toxicon*, 59(6):644-650
- Brasil .Amamentaçãoe uso de Medicamentose Outras Substâncias. 2nd ed. EditoradoMinistério da Saúde; Brasília, Brasil: 2010.
- EU Regulation 466/2001 of 8 March 2001 (2001). Setting maximum levels for certain contaminants in food stuff. Official J. of European Commission, L077, 11-12
- European Commission (EC) EUR-Lex; [(accessed on 17 August 2016)]. Commission Regulation (EC) No. 165/2010 of 26 February of 2010 Amending Regulation (EC) No. 1881/2006 Setting Maximum Levels for Certain Contaminants in Foodstuffs as Regards Aflatoxins. Available online: http://eurlex.europa.eu/legalcontent/EN/TXT/PDF/?uri=CELEX:32010R0165&from=EN.
- Ghiasian, S.A. and Maghsood, A.H. (2012). Infants' Exposure to Aflatoxin M₁ from mother's Breast milk in Iran: *Iranian J. Pub. Health*, 41(3):119-126

- Giola M.P; Oliveira C.M., Bertolin., D.A., Lonardi M.C.V., Gouveia M.S., Netto, D.P; Nixdorf, S.L, Machinski M. Aflatoxin M₁ in urine of non carriers and chronic carriers of hepatitis B virus in moringa, Brazil. Braz. J.Pharm. Sci 2012, 48, 447-452
- Ishikawa A., T., Cássia R. Takabayashi-Yamashita, Elisabete Y. S. Ono, Artur K. Bagatin, Fabiana F. Rigobello , Osamu Kawamura, Elisa Y. Hirooka and Eiko N. Itano (2016). Exposure Assessment of Infants to Aflatoxin M₁ through Consumption of Breast Milk and Infant Powdered Milk in Brazil. *Toxins*, 8(9): 246
- Leblanc J.C., Tard A., Volatier J.L., Verger P.(2005) Estimated dietary exposure to principal food mycotoxins from the first French Total diet study. *Food Addit. Contam.*;22:652–672.
- Mason, S., Hajimohammadi, B., Ehrampoush, M.H., Khabiri, F., Soltani, M. (2015). A Survey on relationship between diet and urinary excretion of aflatoxin M1: a screening pilot study on Iranian population. *Journal of Food Quality and Hazards Control*, 2:66-70
- Redzwan, S. M., Rosita, J., Sokhini, A. M. M., & Aqilah, A. R. N. (2012). Association between aflatoxin M₁ excreted in human urine samples with the consumption of milk and dairy products. *Bulletin of Environmental Contamination and Toxicology*,89(6), 1115-1119.
- Shundo L., Navas S.A., Lamardo L.C.A., Ruvieri V., Sabino M. Estimate of aflatoxin M₁ exposure in milk and occurrence in Brazil. Food Control.2009;20:655–657.\

- Sadeghi N, Oyeisi MR, Jannat B, Hajimahmoodi M, Bonyani H. Jannat F. Incidence of aflatoxin M₁ in human breast milk in Tehran, Iran Food Control. 2009 20:75-8
- Tomerak R.H., Shaban, H.H., Khalafallah, O.A, and EL Shazly. M.N. (2011). Assessment of exposure of Egyptian infants to aflatoxin M₁ through breast milk; *J Egypt Public Health Assoc.*, **86**:51; 0013-2446
- Polycrhonaki N., Wild C.P; Mykkanen H.; Amra, H.; Abdelwahhab M.; Sylla, A; Diallo, M; Elnezemi H; Turner, P.C Urinary biomarkers of aflatoxin exposure in young children from Egypt and Guinea. Food chem. Toxicol 2008. 46, 519-526