

44 In many developing countries, chemical coagulants, such as aluminium sulphate and
45 synthetic poly-electrolytes are usually unavailable [7]. Moringa tree seeds, when crushed
46 into powder, can be used as a water-soluble extract resulting in an effective natural
47 clarification agent for highly turbid and untreated pathogenic surface water [8]. Besides
48 improving water drinkability, this technique reduces water turbidity (cloudiness) resulting
49 in water being both aesthetically as well as microbiologically more acceptable for human
50 consumption [9]. The application of this low cost *Moringa oleifera* seeds is
51 recommended for eco-friendly, nontoxic, simplified water treatment for rural and peri-
52 urban people living in extreme poverty.

53 Charcoal filters have been used for several hundred years and are considered one of the
54 oldest means of water purification [10]. Historians have shown evidence that carbon
55 filtration may have been used in ancient Egyptian cultures for medical purposes and as a
56 purifying agent [11]. The first recorded use of a charcoal filter to purify potable water on
57 a large scale occurred in 19th century England [11]. Currently, carbon filters are used in
58 individual homes as point-of-use water filters, groundwater remediation, landfill leachate,
59 industrial wastewater and, occasionally, in municipal water treatment facilities.

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Broad objective

62 The main objective of the study was to evaluate the effectiveness of using *Moringa*
63 *oleifera* seed powder as a coagulant and wattle stem charcoal as filter material in
64 purification of stream water from unprotected sources in Kapseret.

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Specific objective

67 To compare the antimicrobial activity of *Moringa oleifera* seed extract integrated with
68 wattle stem charcoal filtration alongside the independent performance of *Moringa*
69 *oleifera* and charcoal filter system against microbial populations in raw water sample.

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MATERIALS AND METHODS

Study Area

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73 The study was conducted in Kapseret division, Uasin Gishu County, Kenya. The region
74 covers an area of 148.30 sq. Km. It comprises of Simat, Chepkatet and Lemook locations.
75 It receives an average rainfall ranging between 900-1200mm and this occurs between
76 March and September with two distinct peaks in May and August. The dry spells begin in
77 November and end in February while temperatures range between 8.4 and 26°C but these
78 features are changing probably due to climate change [20]. According to the 2009
79 Population and Housing Census, the total population of area stood at 31,030.

80 The area is a peri-urban setup with an increasing population owing to outward expansion
81 of Eldoret town and rural-urban migration. Major water sources in the area include
82 streams, shallow wells and springs. These sources are usually unprotected and therefore
83 exposed to pollution.

84 **Sampling and Sample Preparation**

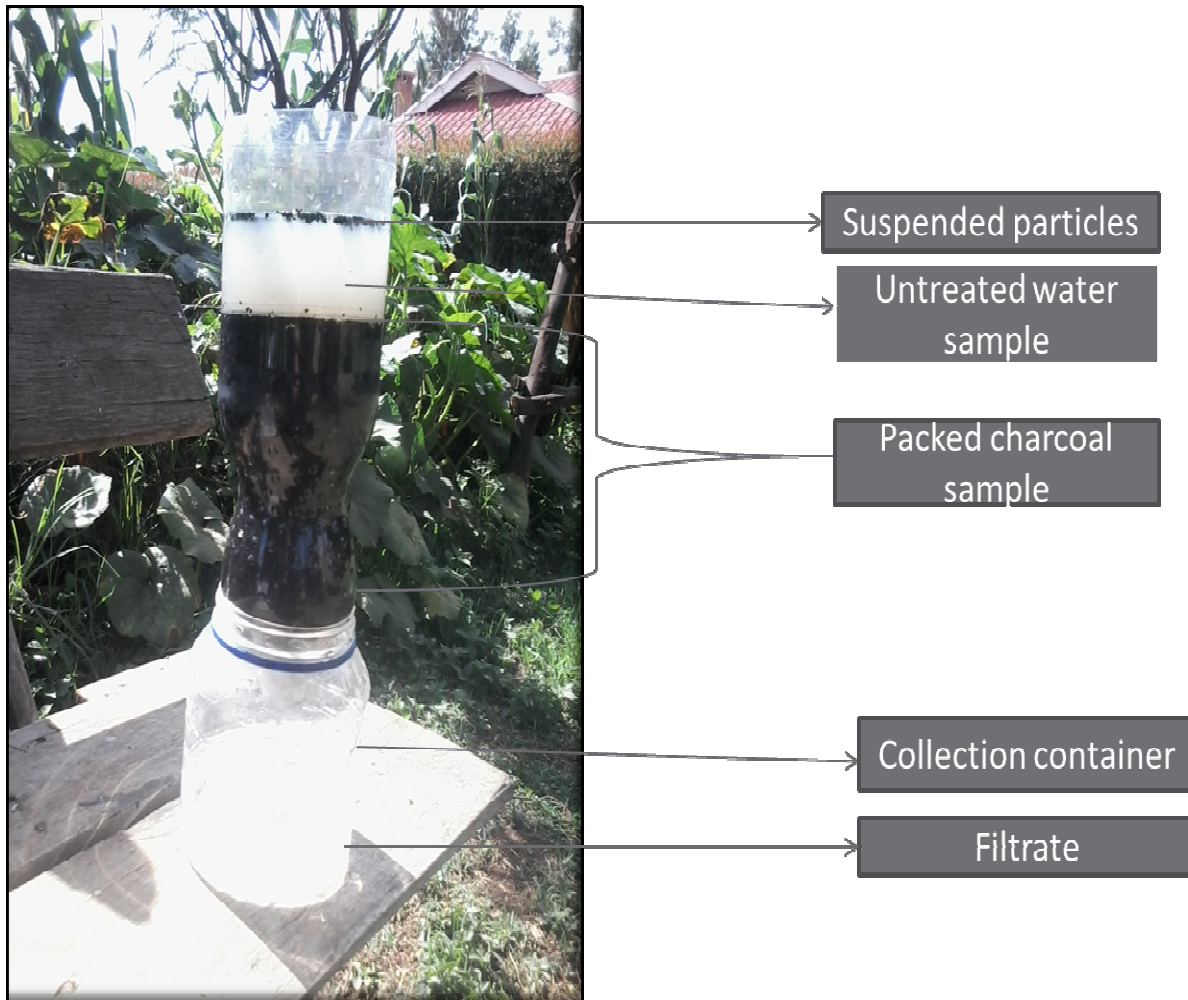
85 Sampling procedures described by American Public Health Association [13] were
86 followed. Glass sample bottles (2000 ml) were sterilized in an autoclave at 121 °C for 15
87 minutes at 121 kPa. A sample of 1.5 litres of water was fetched from each of the five
88 streams i.e Leberio, Malanymaina, Lemook, Nganiat and Kapbodigita in sample bottles
89 and the bottles stoppered. To sterilize the immediate air, flaming was used at the mouth
90 of the sample bottles to avoid possible sample contamination by bacteria in the air around
91 the sampling locations. Samples were collected from these streams in the study area since
92 they were found to be the commonly used water sources by locals. The sampling sites
93 were identified to represent even distribution of unprotected streams across the study
94 area. Random sampling was used in the study. Samples collected were labeled and placed
95 in a cooler box containing ice blocks and then transported within six hours to Eldoret
96 Water and Sanitation (ELDOWAS) laboratories for analysis.

97 **Preparation of *Moringa oleifera* seed extract**

98 Fully matured *Moringa oleifera* seeds were collected from Marigat forest. The seeds
99 were air-dried in direct sun for a week. The shells surrounding the seed kernels were
100 removed using a knife and the kernels were pounded using laboratory mortar and pestle
101 into fine powder. The powder was sieved using a strainer with a pore size of 2.0mm to
102 separate the coarse powder and obtain only the fine powder to achieve solubilization of
103 active ingredients in the seed. This powder was used to prepare *M. oleifera* stock solution
104 for water purification. The stock solution was prepared by mixing 10, 20, 30, 40, 50 and
105 60g of fine seed powder in 1000ml of distilled water and solution later filtered. The
106 suspension was vigorously shaken for 30 min using a stirrer to promote water extraction
107 of the coagulant proteins and this was then passed through filter paper (Whatman No. 1).
108 The filtrate was used within an hour.

109 **Designing an Improved Charcoal Water Filter**

110 Fresh wattle tree charcoal was used as it was readily available and has no known side
111 effects. Crushed charcoal was graded from 0.5 mm to 5mm using standard sieves at the
112 Ministry of Public Works laboratory in Eldoret. The graded charcoal sample was
113 sterilized by boiling in water for 15 minutes before use in the filter. A 2-litre cylindrical
114 plastic container with the lower part cut open was obtained. The smaller opening was
115 covered with a piece of fabric that acted to prevent the charcoal from falling out or
116 running through with the water. Approximately 500g of crushed charcoal of varying sizes
117 was packed into the container tightly. This was meant to create as fine a matrix as
118 possible for the water to drip through slowly, thus trapping more sediment. The crushed
119 charcoal was filled up to about halfway the cylinder. The filter was placed atop a sterile
120 container to collect the filtered water.



121
122 **Figure 3: An improvised Charcoal Filter**

123 **Sample Filtration**

124 A 500 ml sample of raw water was slowly poured into the filter and allowed to slowly
 125 percolate through. The filtrate was collected in a sterilized beaker. The raw and the
 126 filtered samples were later analysed for total coliforms, fecal coliforms and biological
 127 oxygen demand. To determine the effectiveness of combined activity of *M. oleifera* and
 128 charcoal filter, stream water was initially treated with optimum stock solution i.e. 40g of
 129 *M. oleifera* seed powder in 1000ml of distilled water. The treated sample was then passed
 130 through the charcoal filter in a similar procedure of filtration undertaken above.

131 **Determination of the antimicrobial activity of Wattle tree charcoal-*Moringa oleifera***
 132 **seed filter in water purification.**

133 **Estimation of Total and fecal coliforms**

134 Analysis of collected raw water samples and treated water samples to estimate the
 135 populations of total coliforms and fecal coliforms was done using the Colilert-18 test
 136 procedure. This analysis represented one aspect of water quality whose findings were

137 used to draw inferences about the suitability of the water for use based on average
138 microbial populations as per WHO recommendations.

139 One pack of Colilert reagent was added to a 100 ml room temperature water sample in a
140 sterile water container. The container was capped and shaken until its contents dissolved.
141 The sample/ reagent mixture was poured into a quanti tray and sealed in a quanti tray
142 sealer. The quanti-tray 2000 of 97 wells was used. The sealed tray was incubated at 37°C
143 for 18 hours. The results were read according to an interpretation table as described by
144 [14].

145 Fluorescence to detect the presence of *Escherichia coli* was checked using a 6-Watt, 365-
146 nm Ultra violet light lamp within 5 inches of the sample in a dark environment. This
147 procedure ensured that the UV light was directed away from the experimenter's eyes and
148 towards the sample. Colilert results were read after 18 hours, however if the results were
149 ambiguous based on the initial reading, incubating up to additional four hours to allow
150 the color and/or fluorescence to intensify was done. Only sterile, none buffered, oxidant
151 free water for dilutions was used. Aseptic techniques were followed during analysis and
152 good laboratory practice GLP for disposal. Sample tests were stored at 25°C away from
153 light.

154 **Measurement of Dissolved Oxygen**

155 Dissolved Oxygen (DO) was measured using a SX716 Dissolved Oxygen (DO) meter.
156 The machine calibrations were adjusted to read or display 100% active air concentration
157 and the tip of the probe was immersed into the sample in a container and the machine
158 allowed to stabilize before obtaining the actual level of oxygen in parts per million (ppm)
159 which is equivalent to mg/l (APHA, 2017). To reduce errors that might affect oxygen
160 levels during transportation from the field, the initial measurement of dissolved oxygen
161 was done in the area where samples were collected. The Dissolved Oxygen of the treated
162 samples was also taken in the laboratory for analysis to determine the effectiveness of
163 each treatment process.

164 **Measurements of Biochemical Oxygen Demand (BOD)**

165 Biochemical Oxygen Demand (BOD) is a measure of the oxygen in the water that is
166 required by the aerobic organisms. The biodegradation of organic materials exerts oxygen
167 tension in the water and increases the biochemical oxygen demand (14).

168 In the current study initial DO values were recorded in the field and the same samples
169 incubated at 20 °C for 5 days in dark bottles. This was done in order to avoid some
170 processes like photosynthesis and respiration that could have released or consumed

171 oxygen hence affecting its concentration. Final DO was recorded at the end of 5 days.
 172 Biological Oxygen Demand after the 5th day was determined in the formula given below:

173 $BOD_5 = \text{Final DO} - \text{Initial (12)}$. Similar procedure was done for *Moringa oleifera* treated
 174 samples and charcoal filtered samples in the laboratory.

175 **Data processing and analysis**

176 Analysis of variance (ANOVA) was conducted to assess whether significant ($p < 0.05$)
 177 variations existed among the treatments given to assess their effectiveness as water
 178 coagulants. Analysis of data was computed using GenStat Discovery Edition III, 2008.

179
 180 **RESULTS**

181 Based on the objectives of the study, the findings of the study were as follows;
 182 **Effects of *Moringa oleifera* and charcoal filter on assessment of microbiological**
 183 **parameters**

184 In this study, the microbiological parameters under investigation were total coliforms,
 185 fecal coliforms and biological oxygen demand. Summaries of the findings for these
 186 parameters are shown in tables below.

187 The interactions between *Moringa oleifera* and charcoal filter had significant ($p \leq 0.05$)
 188 effects on all microbiological parameters.

189 **Table 1.0 Analysis of variance (ANOVA) summary on the effect of treatment on**
 190 **percentage reduction of biological parameters (TC, FC and BOD) in sampled water**
 191

Source of variation	Total Coliforms		Fecal Coliforms		BOD	
	F-Value	P-Value	F-Value	P-Value	F-Value	P-Value
Treatment	23.38	0.000*	60.996	0.000*	29.402	0.000*
		*		*		*

192
 193 **Total coliforms**

194 The results of total coliforms in the sample water were as shown in Table 2.0. There
 195 were significant differences in total coliforms count ($p < 0.05$) among the different
 196 treatments of the sample water in the area of study. *Moringa oleifera* reduced the total
 197 coliforms by approximately 33%. Filtration over charcoal reduced the population

198 significantly by a further 33%. A combination of *M. oleifera* and charcoal filtration
 199 reduced the total coliform population by 92%. This reduction was significantly different
 200 from either using charcoal or *M. oleifera* singly (Table 2.0).

201 **Fecal coliforms**

202 The results of fecal coliforms (Table 2.0), shows that there were significant differences in
 203 fecal coliforms count ($p < 0.05$) among the different treatments. *Moringa oleifera* reduced
 204 the population by 21%. Charcoal filtration further reduced the population by a significant
 205 82%. A combination of the two treatments reduced the population by approximately
 206 99%. This reduction was significantly different from either using charcoal or *M. oleifera*
 207 singly (Table 2.0).

208 **Biological Oxygen Demand (BOD)**

209 The BOD levels were found to be significantly different ($p < 0.05$) among the different
 210 treatments (Table 2.0). BOD reduction by *Moringa oleifera* was 20%. Filtration over
 211 charcoal reduced the BOD concentration by a further 12%. A combination of the two
 212 treatments reduced the BOD concentration by 51%.

213
 214 **Table 2.0 Mean (\pm) percent reductions of microbiological parameters using *Moringa oleifera*, charcoal
 215 filter and *Moringa oleifera* and charcoal filter combined in water treatment.**

Treatment	Total Coliforms	Fecal Coliforms	BOD
A(<i>M. oleifera</i>)	32.56 \pm 15.88a	21.37 \pm 16.94a	19.95 \pm 9.36a
B(Charcoal)	66.05 \pm 10.68b	82.44 \pm 11.19b	31.51 \pm 4.76b
C(Combined)	92.36 \pm 14.48c	99.23 \pm 0.84c	50.66 \pm 3.52c

216 Means followed by different letters within a column are significantly different at $p < 0.05$

217

218 **DISCUSSION**

219 **Effects of *Moringa oleifera* and charcoal filter on microbiological parameters**

220 *Moringa oleifera* reduced the total coliforms by approximately 33%. Filtration over
 221 charcoal reduced the population significantly by a further 33%. A combination of *M.*
 222 *oleifera* and charcoal filtration reduced the total coliform population by 92%. This
 223 reduction was significantly different from either using charcoal or *M. oleifera* singly.
 224 Processing the water by coagulation using *M. oleifera* as natural coagulant showed that
 225 the treatment with *M. oleifera* provided additional advantage of reduced total coliforms

226 *Moringa oleifera* seeds can be applied to treat water on two levels, acting both as a
 227 coagulant and an antimicrobial agent [16]. It is generally accepted that *Moringa* plant
 228 works as a coagulant that leads to the formation of “flocs” that settle at the bottom of
 229 water [16]. The antimicrobial aspects of *Moringa* plant continue to be investigated [15].
 230 While there are on-going research work being conducted on the nature and characteristics
 231 of these components, it is accepted that treatments with *Moringa* solutions remove 90-
 232 99.9% of the impurities in water [17]. A viable alternative to the chemical coagulants is

233 natural coagulant [18]. *Moringa* seed pods are allowed to dry naturally on the tree prior to
234 harvesting. The mature seeds are readily removed from the pods, easily shelled and then
235 may be crushed and sieved using traditional techniques such as those employed for the
236 production of maize flour [19]. The crushed seeds' powder, when mixed with water,
237 yields a solution [20]. To treat surface water, the equivalent weight of seed powder
238 required to make up a crude extract solution is dependent upon the turbidity [9].

239 *Moringa oleifera* derived coagulants offers several advantages over conventional
240 coagulants such as aluminium sulphate [20]. This includes its activity being maintained
241 over a wide range of influent pH values i.e. no pH correction is required. Natural
242 alkalinity of the raw water also remains unchanged following coagulation i.e. no addition
243 of alkalinity is required. Sludge production is also greatly reduced and is essentially
244 organic in nature with no aluminium residuals sludge volumes are reduced by a factor of
245 up to 5 [21].

246 With proper mixing, the moving particles enlarged and formed flocs that fall to the
247 bottom of the vessel due to gravity. This confirms the effectiveness of *Moringa oleifera*
248 as coagulant for the purification of dirty water. Furthermore, the decrease in total
249 coliform number was also affected by alkaline condition generated by *Moringa oleifera*.
250 Most microorganisms grow well at pH 6.0-8.0, but some of them can grow well at pH 3
251 (acidophiles) and at pH 10.5 (alkaliphiles). Coliform bacteria are facultative anaerobic
252 microorganisms that can grow in aerobic environments and in fermentation condition that
253 produces lactic acid. Therefore these bacteria can still grow at low pH environment,
254 coliform bacteria can still grow, but they cannot survive alkaline pH 14. Additions of
255 *Moringa* as coagulant affect the increase in pH which in turn stops bacteria from
256 growing.

257 Bacterial species *S. faecalis* and *P. aeruginosa* which were cultured in water, stop
258 growing back after *M. oleifera* seeds were added [22]. When the seeds of *M. oleifera* are
259 crushed and dissolved into the water, protein produces a positive charge that acts like a
260 magnet and attracts dominant negatively charged particles such as clay, silk, and other
261 toxic particles. This is in accordance with the invention that the flocculation process
262 removes about 90-99% of bacteria that are usually attached to solid particles, so the
263 bacteria will be aggregated together to form flocs and can be removed from the water
264 [22]. The control treatment had the highest counts of coliform. This affirms earlier stated
265 recommendation above that raw water without treatment is not safe for drinking.

266 It was observed that the BOD of raw water was very high. This was due to the presence
267 of high amount of decomposable organic matter in the water samples. Generally, use of
268 *M. oleifera* jointly with charcoal filters had significant effects on nutrients and BOD.
269 BOD reduction by *Moringa oleifera* was 20%. Filtration over charcoal reduced the BOD
270 concentration by a further 12%. A combination of the two treatments reduced the BOD
271 concentration by 51%. This was probably due to the fact that the phosphates and nitrates
272 were filtered out mechanically by adsorption and retention in the charcoal filter.

273 The porosity and large surface area of charcoal provides a multitude of reactive sites for
274 the attachment of dissolved compounds. These reactive sites can bind non-problematic
275 dissolved organic compounds as well as targeted hazardous contaminants. Background
276 dissolved organic matter, present in all natural waters, can occupy sites on charcoal
277 surfaces and thereby exclude contaminants of concern. This is called "fouling." Fouling

278 in charcoal filters is mitigated by upstream unit processes – in our case, the *Moringa* seed
279 treatment – that act to remove a substantial portion of background dissolved organic
280 matter from the source water before it encounters the charcoal. The principle is to achieve
281 a high level of treatment prior to the charcoal filter, in order to “save the carbon” for
282 removal of targeted problematic dissolved compounds that make it through the previous
283 treatment steps.

284 The charcoal filter in this case functions as a post-coagulation adsorber. The charcoal
285 filter is placed after the *Moringa* seed treatment in order to target specific components of
286 background organic matter (for example, compounds that cause undesirable tastes, odors,
287 or appearance) or synthetic organic compounds (SOCs) such as pesticides,
288 pharmaceuticals, fuel compounds, etc., that are not well removed by the preceding unit
289 processes.

290 The two most important factors affecting the efficiency of charcoal filtration are the
291 amount of charcoal in the unit and the amount of time the contaminant spends in contact
292 with it. The more the charcoal used the better. Similarly, the lower the flow rate of the
293 water, the more time that the contaminants will be in contact with the charcoal, and the
294 more absorption that will take place. Particle size also affects removal rates. The effective
295 lifetime of the charcoal filter media depends upon the quality of the charcoal, as well as
296 the characteristics of the source water and efficacy of upstream treatment steps.

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CONCLUSIONS AND RECOMMENDATION

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Based on this study, the following conclusions were made;

300 i) *Moringa oleifera* seed powder demonstrated the presence of coagulating
301 properties in water treatment.

302 ii) There was enhanced improvement in water quality when *Moringa oleifera* seed
303 extracts were used in combination with charcoal filter against the test
304 microorganism.

6.2 Recommendations

306 i) The *Moringa oleifera* seed extracts can be used in the formulation of a chemical
307 coagulant in water treatment only after scientific validation of their safety.

308 ii) There is need to further elucidate phytochemical components present in the
309 *Moringa oleifera* seed extracts which might be responsible for the antimicrobial
310 activity.

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