

Original Research Article
THE EFFECT OF INDUCERS AND INHIBITORS OF
MONOOXYGENASE ON THE ACTIVITY NITRERGIC SYSTEM IN
THE MICROSOMES IN THE ISCHEMIC LIVER

Abstract

Experiments were carried out on 62 white male rats and average weighing was 180-220g. We found that inducers and inhibitors of monooxygenase showed opposite effects on the activity of NOS in the ischemic liver microsomes. Benzonal and cimetidine, after 1 night of their introduction, had no significant effect on all studied parameters. After 3 and 10 daily using drug metabolism inducer that benzonal makes slow speed nitrate reductase system, stimulates nitroxyls system (eNOS), and cimetidine, on the contrary – even more nitrate reductase activates the speed system, inhibits eNOS nitroxyls. Now, in connection with the growth of liver disease and aggressive exposure to xenobiotic with induction and inhibitory action, this problem acquires a special urgency and, surely, requires further study.

Keywords: ischemia, liver, monooxygenases, NO-system, benzonal, cimetidine.

INTRODUCTION

An important aspect of the modern drug therapy is a personalized medicine based on research and implementations in practical health care of medicines influencing on the system of biotransformation of xenobiotics in the liver (Sivkov et al., 2010; Archakov et al. 2008). The inducers and inhibitors of drug metabolism is to regulate activity of the monooxygenase system (MOS) of the liver and it plays the key role in this issue (Kukes et al. 2007; Villeneuve et al. 2004). In the last decade, we appreciate basic research in molecular biology and medicine which were found that in vascular endothelium, the synthesis of nitric oxide (NO^o) is the family of cytochrome P-450-like hemoproteins - NO-synthase in 5-electron oxidation of L-arginine with the formation of L-citrulline

30 and NO^o (Minamiyama et al., 2001; Manuhina et al., 2000). A family of
31 isoenzymes of NO-synthase (NOS) synthesize NO from L-arginine by three
32 major isoforms that two constitutive (neuronal (nNOS) and endothelial (eNOS)
33 and one inducible (iNOS) (Ivashkin et al., 2000). In order to producing NO ther
34 are some processes are important like utilizing NOS along with a variety of
35 cofactors, substrates such as arginine, oxygen and oxidized nicotinamide
36 dinuceotid phosphate (NADPH) (Markov, 2005; Vinogradov et al., 2005). In
37 pathological processes accompanied by hypoxia or ischemia, the role of NO-
38 sinus mechanism is reduced and induced activity nitrate reductase systems
39 (Reutov, 2000). It is now established that NOS and inactive nitrate reductase
40 system (LDCs) is found in hepatocytes, endothelium of sinusoids, the Kupffer
41 cells/macrophages (Habib and Ali, 2011), as well as in the endothelium of the
42 portal vein and hepatic artery (Hirst and Robson, 2011; Jaeschke et al., 2001).
43 The presence of NOS in hepatocytes suggests a correlation with the enzymes of
44 MOS. However, in the literature there is practically few data on the effect of
45 inducers and inhibitors of drug metabolism on the activity of NOS in
46 microsomes isolated from hepatocytes in the development of liver pathological
47 process.

48 In connection with the above mentioned case, the purpose of the study
49 was to study the activity of NOS in the liver microsomes after administration to
50 animals in the dynamics of postischemic period of benzonal and cimetidine.

51 MATERIAL AND METHODS

52 The study was carried out on 62 male rats of mixed population weighing
53 180-220g, which were divided into 3 groups. First group animals after 1, 2 and
54 3 days ischemia/hypoxia of the liver caused by occlusion during the 180 min of
55 the vascular pedicle of the left lateral and middle lobes.

56 The study drugs were administered after restoration of blood flow to the
57 liver. An inducer of drug metabolism benzonal was administered
58 intragastrically in the form of 1% solution in 0.5% starch gel single dose of
59 50mg/kg for 1, 3 and 10 days in a row (2ml). Inhibitor of drug metabolism
60 cimetidine also was injected intraperitoneally in 0.1% aqueous solution daily,
61 once daily for 1, 3 and 10 days in a row (2nd group). Control for all research
62 groups served as data of intact animals. Each group consisted of 6-8 animals.

63 The animals were sacrificed by instant decapitation method under light
64 Rausch-anesthesia. The extracted liver was perfused through the inferior vena
65 cava by chilled ($0\pm4^{\circ}\text{C}$) 50 mM Tris HCl buffer, pH 7.4, containing 0.05 M *KCl*
66 and 0.25 M saccharose. After washing the liver from the blood it was ground
67 and homogenized in the same solution (1:3). From that fraction, which was
68 obtained by centrifugation at VAC-602 (Germany) after 20 minutes of
69 unscrewing, with 12 thousand g, had been beset microsomes thousand at 105 g
70 for 60 min. All procedures were performed in the refrigerating chamber KHS-
71 12(Russia) at $0\pm4^{\circ}\text{C}$. In microsomes, resuspended in 100 mM Tris - HCl buffer;
72 pH 7.4 was evaluated activity of monooxygenase system that content of
73 cytochromes P-450, P-420, and b5 by classic method of T. Omura, R. Sato
74 (1964), the activity of NADPH-reductase (NADPH-op.-ed.) by C. H. Williams,
75 H. Kamin (1961), benzo(α)pyrene hydroxylase (B(a)PG) by C. H. Yang,
76 L.P.Kicha (1978). Aniline hydroxylase (AG) by A. I. Archakov et al. (1975), N-
77 demethylase amidopyrine (N-AP) by A. Bast, J. Nordhosck (1981), glucose-6-
78 phosphatase (G-6-Phase) by N. S. Gnosh, N. C. Kar (1983) were assessed
79 (Table 1).

80 Nitroxygenase activity was determined by the content of stable
81 metabolites nitrite and nitrate NO^- , NO_2^- and NO_3^- - by the method of P. P.
82 Golikov et al.(2000), activity of endothelial NOS (eNOS) by Sumbaev V.V.,
83 Yasinska, I.M. (2000), inducible NOS (iNOS) and the concentration of

84 peroxynitrite (ONO_2^-) in Ravaeva M. Yu, E. N. Chuyan (2011). Content,
85 activity of monooxygenase and oxidoreductase of nitrooxygenase systems were
86 recorded on computerized dual beam spectrophotometer UV-2100 (Ltd, China).
87 The content and activity of oxidoreductase was calculated in microsomes per
88 milligram of protein in 1 ml (mg/ml), which was determined by the method of
89 O. N. Lowry et al. (1951).

90 **STATISTICAL ANALYSIS**

91 The obtained results were subjected to statistical analysis using the
92 software package Excel, Statistic for Windows V.6.0. Normality of distribution
93 of quantitative parameters was checked using the criteria Kolmogorov-Smirnov
94 and Shapiro-Wilk test. Calculated arithmetic mean (M), standard deviation (σ),
95 error arithmetic average (m), sample standard deviation (S). The distribution of
96 the samples was carried out on the basis of student's criterion (t) with the
97 computation of error probability (P). The correlations for the indicators was
98 carried out using correlation analysis Pearson (r). For comparison, samples
99 were used Student's t -test. Data were considered significant at $p < 0.05$.

100 **RESULTS AND DISCUSSION**

101 Benzonal and cimetidine had no significant effect on all studied
102 parameters characterizing the activity of NOS in the liver microsomes after 1
103 night of their introduction and after chemotherapy, compared to other groups,
104 which drugs are not injected, the corresponding term monitoring (1 day) (Table
105 2). In subsequent periods after 3 and 10 days benzonal significantly reduced the
106 expression of NO , iNOS and ONO_2^- on the background of the dynamic of the
107 studied follow-up period of increasing eNOS activity and content of microsomal
108 protein. At the same time, after 3 and 10 of the daily administration of
109 cimetidine in selected microsomal fractions of the liver shows a dynamic period
110 of observation, a decrease in the activity of eNOS and increased expression of

111 NO, iNOS and ONO_2^- , marked inhibition of microsomal protein concentration.
112 Therefore, the introduction of animals with ischemic liver benzonal optimizes
113 the processes of NOS in microsomal system in the body, and cimetidine on the
114 contrary an even greater extent, potentiates the effects of damage to this system.
115 When analyzing the performance of NOS is therefore with the activity of eNOS
116 associated changes in the level of iNOS reaction rate, the content of microsomal
117 NO and ONO_2^- in all studied groups of animals. In this regard, it is quite
118 possible to believe that the increased NO and ONO_2^- is due to inhibition of
119 eNOS and overexpression of iNOS. Benzonal positively influenced changes in
120 the level of NO in microsomes, reduced an activity of iNOS and content of
121 cytotoxic ONO_2^- . You can put that with the decreased activity of iNOS and the
122 level of ONO_2^- was associated, although not significantly increasing the activity
123 of eNOS and restore to control values the concentrations in the ischemic liver
124 microsomes NO administered to animals of benzonal.

125 According to the literature data, iNOS, ONO_2^- and NO are components of
126 the expression system of nitrate reductase. During ischemia and/or hypoxia
127 involves an increase in the cytotoxic compounds, including NO and ONO_2^-
128 which block the active centers of cytochrome P450 in microsomes ischemic
129 liver [11]. Cimetidine as followed data, reinforces these processes in
130 microsomes of animals with ischemic liver and suppresses NOS way. However,
131 as shown by some researchers during ischemia/hypoxia blockade the active site
132 of the isoforms of cytochrome P-450 activated oxygen metabolites, including
133 NO and ONO_2^- have a fragile relationship [12]. In this regard, we can assume
134 that the inducer of drug metabolism benzonal, promotes the release of the
135 connection of active center of cytochrome P-450 with NO and ONO_2^- ischemic
136 liver. As a result of increased accessibility to the substrates of oxidation in
137 particular L-arginine, which plays a major role in the regulation of functional
138 metabolic and regenerative functions of liver [13, 14]. This is evidenced by the

increase of eNOS activity in microsomes when administered to animals with ischemic liver of benzonal. Therefore, benzonal as an inducer of drug metabolism when administered to animals with ischemic liver microsomes increases in NOS activity, through mechanisms of oppression nitrate reductase components, thus reducing the level in hepatocytes toxic compounds, the overexpression of NO and ONO_2^- .

DISCUSSION

We considered that a number of basic factors like covariates in these analyzes, which can lead to an association between NO and ONO_2^- , including age, race/ethnicity, diabetes, hypertension, hyperlipidemia, cardiac vascular diseases (cardiovascular diseases), body mass index (BMI), smoking history, alcohol consumption, physical activity, prior use of hormone therapy (HT), and in the longitudinal analysis of the active HT hand for participants in clinical trials of HT. Diabetes mellitus was defined as self-administration of pills or insulin and / or serum glucose on an empty stomach $> 126 \text{ mg/dl}$ [3, 8]. Hypertension was defined as a systolic blood pressure less than 140 mmHg or diastolic blood pressure $> 90 \text{ mmHg}$ or took pills for hypertension [4]. Hyperlipidemia was defined as total cholesterol level $> 240 \text{ mg/dl}$ or LDL $> 160 \text{ mg/dl}$ or taking cholesterol-lowering drugs [11]. The survey history, alcohol use, previous cardiovascular diseases and previous use and duration of hormones were set in the questionnaire. Physical activity was determined using personal habits data and classified into a total metabolic equivalent (MET) per week [9, 13]. This lack of effect was confirmed by Western blot, which demonstrated that the expression of these enzymes was not altered by L-NAME. Discourse. Our results show that chronic treatment of rats with L-NAME is effective in hypertrophy of the walls of the arterial vessel wall (envelopes), together with perivascular fibrosis associated with the deposition of collagen fibers [14]. In addition, a connection between sinusoidal lumen and interstitial

expansion (increase in cellularity, mainly of fibroblasts, and connective ability in the portal space) was achieved [12, 16]. A decrease in sinusoidal calibration is associated with an imbalance in vasoactive mediator production in sinusoids when exposed to L-NAME. Acute or chronic treatment with L-NAME results that are invasive due to the lack of NO to counteract the suppression of peptides, such as angiotensin-sin and endothelin that ultimately leads to hypertension. Cells that are important regulators of the sinusoidal capillary layer are very sensitive to their predecessors (Rockey 2001), and their contraction with vasoconstrictors can reduce the sinusoidal capillary space. These cells may be an important factor in the increase in intrahepatic resistance observed in portal hypertension. By agreement with studies Dupuis et al. (2004) reported enhanced gene expression associated with the regulation of cell proliferation, extracellular matrix remodeling, and NO/cGMP signaling in aortic tissue of rats treated with L-NAME for 15 or 30 days [15, 17].

CONCLUSION

Therefore, inducers and inhibitors have opposite effects on the activity of NOS in the ischemic liver microsomes. Benzonal makes slow speed of nitrate reductase system, stimulates nitroxygenase system (eNOS), and cimetidine, on the contrary that even more nitrate reductase system activates the speed system, inhibits eNOS nitroxyls. The difference in activity of benzonal and cimetidine explain through what mechanisms can regulate the enzyme monooxygenase, thereby positively impact on pathological processes in the liver that is critical to its hypoxic conditions. At the present time in connection with the growth of liver disease and aggressive exposure to xenobiotic with induction and inhibitory action on the person, this problem acquires a special urgency and, of course, requires further study.

Ethical Approval:

As per international standard or university standard written ethical approval has been collected and preserved by the author(s).

Consent: NA

Table 1. Dynamics of activity indicators of monooxygenase inhibitors of liver after acute ischemia/hypoxia and establishment in different dates (days) benzonal and cimetidine, M±m

Groups	P-450, nm/mg	P-420, nm/mg	b ₅ , nm/mg	NADFH- cyt.c-red, nm/min/mg	B(a)PG, nm/min/mg	AG, nm/min/mg	N-AP, nm/min/mg	G-6-Phase, nm/min/mg
Control group	0,97±0,031	0,036±0,001	0,63±0,026	106,9±3,95	1,69±0,078	0,88±0,023	4,85±0,151	79,8±3,06
Ischemia: 1 day	0,30±0,018*	0,262±0,009*	0,15±0,006*	8,4±0,29*	0,65±0,022*	0,30±0,017*	1,57±0,062*	24,7±1,03
3 day	0,37±0,015*	0,191±0,008*	0,20±0,007*	13,7±0,44*	0,89±0,035*	0,37±0,015*	1,85±0,061*	39,5±1,65
10 day	0,45±0,018*	0,170±0,006*	0,25±0,008*	21,1±0,87*	0,93±0,042*	0,41±0,018*	1,91±0,067*	43,3±1,17
Ischemia+ B: 1 day	0,35±0,017*	0,231±0,010*	0,17±0,007*	8,9±0,36*	0,73±0,039	0,35±0,019*	1,62±0,048*	25,3±1,23*
3 day	0,68±0,021* ^Δ	0,107±0,013* ^Δ	0,31±0,009* ^Δ	68,1±2,48*	1,05±0,044* ^Δ	0,51±0,022*	2,24±0,079* ^Δ	45,9±1,34* ^Δ
10 day	1,55±0,059* ^Δ	0,015±0,002* ^Δ	0,77±0,032* ^Δ	120,4±6,35*	2,01±0,095* ^Δ	1,46±0,061* ^Δ	6,35±0,330* ^Δ	82,7±3,56* ^Δ
Ischemia+ C: 1 day	0,29±0,019*	0,266±0,008*	0,16±0,005*	8,5±0,33*	0,68±0,027*	0,32±0,018*	1,59±0,055*	23,9±1,16*
3 day	0,31±0,014* ^Δ	0,243±0,009* ^Δ	0,17±0,006*	13,1±0,59*	0,80±0,031*	0,34±0,015*	1,73±0,059*	37,2±1,48*
10 day	0,28±0,011* ^Δ	0,285±0,005* ^Δ	0,16±0,005* ^Δ	17,5±0,58* ^Δ	0,71±0,026* ^Δ	0,29±0,016* ^Δ	1,54±0,048* ^Δ	28,6±0,89* ^Δ

* - P<0.05 compared with control, Δ- P<0.05 compared to hypoxia of the corresponding period

Table 2. Dynamics of indicators of activity of NO – system in the liver microsomes after playing it acute ischemia/ hypoxia and different periods (day) of benzonal and cimetidine, M±m.

Group	NO, mkM/mg	eNOS, mkM/min/mg	iNOS, mkM/min/mg	ONO2-, mkM/mg	Protein mc, mg/ml
Control	5,5±0,16	17,4±0,62	0,10±0,002	0,080±0,016	36,8±1,22
Ischemia: 1 day	8,6±0,33*	7,9±0,29*	0,35±0,017*	0,23±0,010*	29,5±1,13*
3 day	8,1±0,27*	8,5±0,35*	0,23±0,009*	0,19±0,009*	30,8±1,09*
10 day	7,6±0,28*	9,7±0,42*	0,17±0,006*	0,14±0,007*	31,2±1,18
Ischemia+B 1 day	8,7±0,29*	8,3±0,21*	0,32±0,019*	0,22±0,011*	29,1±1,26*
3 day	6,3±0,26* ^Δ	12,5±0,43* ^Δ	0,17±0,005* ^Δ	0,16±0,006* ^Δ	31,7±1,31
10 day	5,8±0,22 ^Δ	18,4±0,59 ^Δ	0,11±0,004* ^Δ	0,07±0,005* ^Δ	37,5±1,42
Ischemia+C: 1 day	8,9±0,39*	8,1±0,28*	0,36±0,019*	0,25±0,013*	28,7±1,26*
3 day	10,6±0,37* ^Δ	8,4±0,15*	0,33±0,012* ^Δ	0,21±0,011*	28,3±1,33*
10 day	13,5±0,52* ^Δ	7,2±0,18* ^Δ	0,46±0,021* ^Δ	0,35±0,014* ^Δ	32,6±1,40

* - P<0.05 compared with control, Δ- P<0.05 compared to hypoxia of the corresponding period

REFERENCES

1. Sivkov A.S., Paukov S.V., Ruvinov Yu.V., Kukes I.V. Individual safety of pharmacotherapy in assessing the activity of cytochrome P-450 3A4 isoenzyme (CYP3A4). Klin.med. - 2010. - №2. - P. 61-67.
2. Archakov A.I., Lisitsa A.V., Petushkova N.A., Karuzina I.I. Cytochrome P-450, drug disease and personified medicine. Part I. Clin.Med.-2008.-No. 2.- S. 4-8.
3. Kukes V.G., Sychev D.A., Shikh E.V. The study of biotransformation of drugs is a way to increase the effectiveness and safety of pharmacotherapy. Doctor. - 2007. - №1. - P. 2-5.
4. Minamiyama Y., Jmaoka S., Takemura S. Escape from tolerance of organic Nitrite by induction of cytochrome P450.Free Radical Biology et Medicine. – 2001. – Vol. 31, №11. – P. 1498-1508.
5. Manukhina E.B., Malyshev I.Yu., Archipenko Yu.V. Nitric oxide in the cardiovascular system: a role in adaptive protection. Vestn. RAMS. -2000. -№4. -C.16-21.

- 226 6. Ivashkin V.T., Drapkina O.M. Nitric oxide in the regulation of the
227 functional activity of physiological systems. *Rus. Jour. Gastroenter., hepat.,*
228 *coloproctol.* -2000. -№4. -P. 16-21.
- 229 7. Markov H.M. Molecular mechanisms of vascular endothelial dysfunction.
230 *Cardiology.* -2005. -12. -P. 62-72.
- 231 8. Pokrovsky V.I., Vinogradov N.A. Nitric oxide, its physiological and
232 pathophysiological properties. *Ter.arch.* -2005. -№1. -P. 82-87.
- 233 9. Reutov V.P. Medico-biological aspects of cycles of nitric oxide and
234 superoxide anion radical. *Herald of RAMS.* -2000. -№4. -P.35-41.
- 235 10. Yakovlev S.V., Rudakova A.V. Model pharmacoeconomic studies in
236 antimicrobial chemotherapy: the methodology for conducting and the need to
237 take into account additional factors. *Wedge. Pharmacol. and ter.* -2004. -T.13,
238 No.2. -P.27-31.
- 239 11. Minamiyama Y., Takemura S., Imaoka S., Funae Y., Tanimoto Y., Inoue
240 M. Irreversible Inhibition of Cytochrome P450 by Nitric Oxide. *J. Pharmacol.*
241 *Exp. Ther.*-1997. -Vol.283(3). -P.1479-1485.
- 242 12. Lyakhovich VV, Vavilin VA, Zenkov NK, Men'shchikova EB Activated
243 oxygen metabolites in monooxygenase reactions. *Bulletin SO RAMN.* - 2005. -
244 №4. - P. 7-12.
- 245 13. Villeneuve J.P., Pichette V. Cytochrome P-450 and liver diseases. *Curr.*
246 *Drug Metab.* -2004. -Vol.5. -P.273-282.
- 247 14. Yasui, H., Hayashi S., Sakurai H. Possible involvement of singlet oxygen
248 species as multiple oxidants in P450 catalytic reactions. *Drug Metab.*
249 *Pharmacokinet.* -2005. -Vol.20. -P.1-13.
- 250 15. Habib S., Ali A. Biochemistry of Nitric Oxide. *Ind. J. Clin. Biochem.* -
251 2011. - Vol. 26, №1. - P. 3-17.
- 252 16. Hirst D.G., Robson T. Nitric oxide physiology and pathology. *Methods.*
253 *Mol. Biol.* -2011. -Vol.704. -P.1-13.

254 17. Jaeschke H., Gores G.J., Cederbaum A.J., Hinson J.A., Pessayre D.,
255 Lemasters J.J. Mechanisms of Hepatotoxicity. Hepatology. -2001. –Vol.15. –P.
256 718-724.