

# Hepatitis B core IgM detection in neonates born to HBsAg and HBeAg positive mothers in Maiduguri, Nigeria

## Abstract

Hepatitis B virus is a serious global public health problem and is endemic in Africa, including Nigeria. Infection of pregnant women during the second and third trimester poses a threat of 10 and 90% respectively for vertical transmission. A total of ninety two blood samples from consenting pregnant women were screened for HBsAg and HBeAg using ELISA kit (Cortez Diagnostic Inc, USA). Cord blood samples collected from five neonates of women positive for HBsAg and HBeAg were screened for HBcIgM using the same test kit. Questionnaire was used to collect data on demography, history of blood transfusion and presence of tribal mark from the pregnant women. Overall prevalences of 8.7% (8/92) and 5.4% (5/92) for HBsAg and HBeAg seromarkers, respectively were obtained. Mean age of the pregnant women was 25.75 with 27.2% within 20–24 years and 28.3% within 25–29 years, however this distribution was not statistically significant ( $p=0.6840$ ). Fifty percent (50%) of HBsAg positive women were within the age group of 25–29 years while 80% of HBeAg positive women were within the age group of 20 - 24 years. Blood transfusion ( $p=0.002791$ ) and tribal mark ( $p=0.00265$ ) were found to be associated with acquisition of the virus. Eighty percent (80%: 4/5) of the neonates screened from HBsAg and HBeAg positive women were reactive for HBcIgM. The prevalence of surface antigen in this study suggests the endemicity of hepatitis B virus in the study area while the presence of both surface and envelope antigens in pregnant women portend infectivity. These results suggest the utmost need for establishment of a sustainable intervention measure that would protect not only pregnant women but women of childbearing age in order to mitigate spread of the virus. Screening for other hepatitis B virus seromarkers besides HBsAg before blood transfusion is also advocated.

**Key words:** Hepatitis B surface antigen, Hepatitis B envelop antigen, pregnant women, cord blood, Maiduguri.

## Introduction

Hepatitis B virus (HBV) is a member of the hepadnaviridae family. This virus has a partially circular double stranded DNA and a core antigen surrounded by a shell containing hepatitis B surface antigen (HBsAg), hepatitis B envelop antigen (HBeAg) and hepatitis B core antigen (HBcAg) (Zuckerman *et al.*, 1996; Gasim *et al.*, 2013). Corresponding antibodies to each of these antigens are hepatitis B surface antibody (anti-HBs or HBsAb), hepatitis B envelope antibody(anti-HBe or HBeAb) and hepatitis B core IgM and IgG antibodies ( anti-HBc or HBcAb) (Eke *et al.*, 2011)

Infection with hepatitis B virus (HBV) is a serious public health problem worldwide and the major cause of chronic hepatitis, cirrhosis, and hepatocellular carcinoma (HCC). It has been estimated that there are over 350 million hepatitis B virus (HBV) carriers worldwide. The majority of these carriers resides in the developing countries of South East Asia and sub Saharan Africa, where the lifetime risk of infection is estimated to be greater than 60% with an excess carriage rate of 8% (WHO, 2012). Over 600,000 persons die each year worldwide from complications of HBV infection including liver cirrhosis and hepatocellular carcinoma (Michael *et al.*, 2013).

Transmission of HBV from carrier mothers to their babies can occur during the perinatal period, and appears to be the most important factor in determining the prevalence of infection in high endemicity areas. Nigeria is classified among the countries endemic for HBV infection and currently 18 million Nigerians are infected (Mbaawuaga *et al.*, 2008; Olokoba *et al.*, 2011). When a pregnant woman is infected with HBV, there is a chance of mother-to-child transmission to occur. It has been reported that 10 -20% of women seropositive for HBsAg transmit the virus to their neonates, but in women who are seropositive for both HBsAg and HBeAg; vertical

transmission is approximately 90% (Vranckx *et al*, 1999; Ugwuja, 2010). Vertical transmission from infected mother to infants is thought to be partially responsible for high prevalent of infection in certain high risk group (Ndako *et al.*, 2012). The risk of transmission depends on the degree of maternal infectivity and the genomic type of the virus (Ezegebudo *et al.*, 2004).

Even though studies have been carried out on HBV infection in different parts of Nigeria, and in different cohorts, the prevalence of both HBsAg and HBeAg among pregnant women and especially information regarding the vertical transmission rate is scanty from the north-east region of the country. Therefore, this study was aimed at assaying for HBsAg and HBeAg in pregnant women; HBcIgM in cord blood of babies born to HBsAg and HBeAg positive mothers and to determine the contributory effect of tribal mark and history of blood transfusion to maternal infection in the study area.

## **Materials and Methods**

### **Study area**

This research was carried out at General Mamman Shuwa Memorial Hospital, Maiduguri, Borno State, Nigeria. Maiduguri, also called Yerwa by locals, lies on the geographical coordinates of 11° 50' 42" N, 13° 9' 35" E. It is the capital and the largest city of Borno State in north-eastern Nigeria. The city sits along the seasonal Ngadda River which disappears into the Firki swamps in the areas around Lake Chad. The indigenes are predominantly Kanuri by tribe.

### **Ethical clearance**

Ethical approval for the study was obtained from the Ethical Board of the hospital. Informed oral consent was obtained from all subjects recruited into the study.

### **Study design**

A cross-sectional, hospital –based design was utilized. Ninety two pregnant women whose consent was obtained participated in the research and structured questionnaire was used to obtain the necessary clinical as well as demographic data such as age, tribal mark, history of blood transfusion and trimester.

#### **Exclusion criteria**

Non pregnant women were excluded from the research.

#### **Inclusion criteria**

Only pregnant women were included in the research.

#### **Collection and Processing of Blood Samples**

Five milliliter (5ml) of blood sample was aseptically collected from the pregnant women by venipuncture while the cord blood was collected by the midwives on duty. The blood samples were transferred into clean plain tubes and allowed to clot at room temperature before centrifugation at 300rpm for 5minutes to obtain serum. The serum was aseptically aspirated into a labeled sterile container and stored at -20°C until further analysis.

#### **Determination of HBsAg and HBeAg**

#### **Procedure**

The assay for HBsAg and HBeAg was carried out according to manufacturer's (Cortez Diagnostic Inc, USA) instructions. The reagents and samples were allowed to cool down to room temperature (18-30°C) for at least 15-30 minutes. Wash buffer concentration was checked for the presence of salt crystals and re-solubilized by warming at 37°C until the crystals have been properly dissolved. The strips were set in a strip-holder and sufficient numbers of wells were numbered including three Negative Control (B1, C1, D1) two Positive Control (E1 and F1) and

one Blank (A1). Fifty microlitres (50 µl) of each of the Positive Control, Negative Control and Samples were added into their respective wells using separate disposable pipette tip in order to avoid cross-contamination. Fifty microlitres (50 µl) HRP of conjugate was added to each well except for the Blank well and mixed by tapping the plate gently.

The plate was then covered with a plate cover and incubated for 60 minutes at 37° by using thermostat-controlled water tank in order to assure the temperature stability as well as humidity. At the end of the incubation, the plate cover was discarded and each well was washed 5times with diluted wash buffer and each time, the microwells were allowed to soak for at least 30-60 seconds. After the last wash, the plate was turned down onto blotting paper, and then taped in order to remove the remaining solution.

Fifty microlitres (50µl) of chromogen A and B solution were dispensed into each well including the blank well, mixed by tapping the plate gently and incubated at 37°C for 15 minutes in the dark. At the end of the incubation, the enzymatic reactions between chromogen A and B solutions and the HRP-conjugate produced blue color in the control and HBsAg/HBeAg positive sample wells. Fifty microlitre (50µl) of stop solution was added into each well and properly mixed. The plate reader was calibrated using a blank well and the absorbance was read at 450nm. The reference wavelength of the dual filter instrument was set at 630nm. The cut-off value was calculated. The absorbance was read within 5minutes after stopping the reaction.

***HBsAg Interpretation of result***

126 The results were calculated by relating each specimen absorbance (A) value to the cut-off value  
127 (C.O) of the plate.

128 Calculation of the cut-off value (C.O) = NC + 0.06 (NC = the mean absorbance value for  
129 three negative controls).

130 **Positive result (A/C.O  $\geq$  1):** Specimens with an absorbance equal to or greater than the cut-off  
131 value were considered reactive, which indicates that hepatitis B virus surface antigen has been  
132 detected.

133 **Negative result (A/C.O  $\leq$  1):** Specimens with an absorbance less than or equal to the cut-off  
134 value were considered reactive, which indicates that hepatitis B virus surface antigen was not  
135 detected using HBsAg ELISA.

136 ***HBeAg Interpretation of results:***

137 **Negative Results (S/C.O. < 1):** Samples with an absorbance less than cut-off value were  
138 considered negative, which indicates that no Hepatitis B virus “e” antigen has been detected with  
139 this HBeAg ELISA kit.

140 **Positive Result (S/C.O.  $\geq$  1):** Samples with an absorbance greater than, or equal to the cut-off  
141 value are initially reactive, which indicates that Hepatitis B virus “e” antigen was detected with  
142 this HBeAg ELISA kit.

143

144

145 **Determination of HBcIgM in cord blood**

HBcIgM was determined using the same method with a slight modification described for HBsAg and HBeAg determination above: One hundred microliter (100µl) of samples, positive and negative controls was added to the respective wells.

#### Interpretation of result

Calculation of cut-off value (C.O) =  $N_c \times 2.1$ ,  $N_c$  = the mean absorbance value for three negative controls.

**Negative result:** ( $S/C.O < 1$ ). Samples with an absorbance less than the cut-off value were considered negative, which indicates that no IgM-class antibodies to hepatitis B core antigen have been detected with this anti HBcIgM ELISA kit.

**Positive Result:** ( $S/C.O > 1$ ) Samples with an absorbance greater than, or equal to the cut-off value were considered reactive, which indicates that IgM-class antibodies to hepatitis B core antigen have probably been detected with this anti HBcIgM ELISA kit.

#### Statistical analysis

Data analysis was performed using an online chi-square calculator with  $p < 0.05$  at 95% confidence interval. Also, any sample reactive for either of the seromarker was considered positive.

#### Results

A total of 92 pregnant women within the age group of 15-39 years have been screened for HBsAg and HBeAg (Table 1). The overall prevalence shows 8.7% and 5.4% for HBsAg and HBeAg respectively. Four (4) women out of 26 pregnant women within the age group of 25-29 years were detected positive for both HBsAg and HBeAg. Similarly, one (1) out of 25 pregnant women within the ages of 20-24 years was found to be positive for both HBsAg and HBeAg.

However, only one (1) pregnant woman was found to be positive for only HBsAg in are of 15-19, 30-34 and 35-39 years. Table 2 shows that blood transfusion (p= 0.002791) and tribal mark (p=0.00265) are associated with transmission of hepatitis B infection. The cord blood analysis for the detection of HBcIgM reveals 4/5 (80%) of babies born to five HBsAg and HBeAg as positive (Table 3).

**Table 1:** Distribution of HBVseromarkers according to age

Age (Years)	n	Positive seromarkers		p-value
		HBsAg(%)	HBeAg(%)	
15 – 19	15	1(6.7)	0(0)	0.684091
20 – 24	25	1(4)	1(4)	
25 – 29	26	4(15.4)	4(15.4)	
30 – 34	14	1(7.1)	0(0)	
35 – 39	12	1(8.3)	0(0)	
>40	00	0(0)	0(0)	
<b>Total</b>	<b>92</b>	<b>8(8.7%)</b>	<b>5(5.4%)</b>	

**Table 2:** The Impact of Tribal mark and Blood transfusion on HBV maternal infection

Predisposing factors	n	Positive seromarkers		p-value
		HBsAg	HBeAg	
<u>Tribal Mark</u>				
Yes	67	5	0	0.00265
No	25	3	5	
<u>Blood Transfusion</u>				
Yes	2	2	2	0.002791
No	90	6	3	



**Table 3:** Hepatitis B seromarkers in mothers and detection of HBcIgM in cord blood

Sample No.	Trimester	HBsAg	HBeAg	HBcIgM in cord blood
3	3 <sup>rd</sup>	+ve	+ve	Reactive
29	3 <sup>rd</sup>	+ve	+ve	Reactive
40	3 <sup>rd</sup>	+ve	+ve	Reactive
72	3 <sup>rd</sup>	+ve	+ve	Reactive
81	3 <sup>rd</sup>	+ve	+ve	Non-Reactive

## Discussion

The high prevalence (8.7%) of hepatitis B infection observed among the pregnant women in the study indicates endemic infection in the study area according to the World Health Organization classification for hepatitis B infection (Chen and Chang, 2010). The prevalence is higher than some previous studies in Nigeria (Ndam *et al.*, 2008; Pennap *et al.*, 2011 and Alegbeleye *et al.*, 2013). The implication of this result is the likelihood of transmission of the virus to the newborn especially by mothers who were both HBsAg and HBeAg positive. In endemic countries, mother-to-child transmission accounts for most cases of infections and is, therefore, the main mechanism that perpetuates the infection in the population (Borgia *et al.*, 2012; Coppola *et al.*, 2010). HBV causes an acute hepatitis that becomes chronic in a percentage that is highly dependent on age of acquisition of the infection (Coppola *et al.*, 2010; Coppola *et al.*, 2013a; Coppola *et al.*, 2013b). The rate of chronicity is about 90% in infants infected at birth or during the first year of life, 30–50% in children aged 1–6 years, and 5–10% in children above 6 years of age and in adults (WHO, 2013; Borgia *et al.*, 2012). Once chronic hepatitis is established, a

percentage ranging from 15–40% evolve to liver cirrhosis and hepatocellular carcinoma (Sagnelli *et al.*, 2012; Coppola *et al.*, 2014).

Age-based distribution of sampled pregnant women shows that they enter marriage/pregnancy as early as 15 years of age, however only those within 20 – 24 years (27.2%) and 25 – 29 (28.3%) years were positive for HBsAg and HBeAg. This is in line with a report from in Port Harcourt, Nigeria (Alegbeleye *et al.*, 2013). The result of this study suggests the necessity of targeting not only pregnant women but all women of childbearing age including those younger than 15years old during vaccination against hepatitis B virus with a view to protecting them against the infection. The sum of the pregnant women who were positive for both seromarkers in the two age groups is more than half (55.4%) of the total sample (Table 1), therefore their babies are at high risk of getting infected with the virus at birth. Transmission of hepatitis B infection occurs either by intrauterine transmission or transmission during delivery. While intrauterine transmission accounts for only the minority of cases of HBV transmission, transmission during delivery is the most frequent method of vertical transmission. It is mostly due to newborn contact with the mother's infected secretions or blood at the time of delivery. (Piratvisuth,2013). Moreover, high viral load and positivity of HBeAg have been associated with an increased risk of transmission through this route (Xu *et al.*, 2002; Xu *et al.* 2013). Therefore, it implies that such infection, where it is not managed properly, will progress to hepatocellular carcinoma later in life.

With respect to tribal mark, statistical analysis ( $p=0.00265$ ) shows that it was a significant factor for hepatitis B transmission. This may be connected to the methods employed in the incision. Low economic status of patients encourages the use and reuse of incision instrument quite often. Therefore, since tribal mark incision is a common cultural practice among the populace, the need

to enlighten the population on the expediency of using sterile instruments is very essential with the view to preventing possible spread of the virus through contaminated instruments.

In most pre-transfusion blood screening exercises in Nigeria, only HBsAg is routinely tested for without assaying for other seromarker(s) such as the rare HBcAg or the readily assessable anti-HBc (where present) which indicate window phase of infection. Therefore, the reason for the statistical significance of blood transfusion as an important factor in the spread of hepatitis B virus in the study population can premise on the possible transfusion of infected blood. Hence, it is advised that any blood meant for transfusion be subjected to comprehensive screening in order to prevent transmission of the virus.

## **Conclusion**

The prevalence of HBsAg (8.7%) and HBeAg (5.4%) in this study corroborates with the results of previous studies in Nigeria that classify hepatitis B infection as endemic. The age groups (20-24 years and 25-29 years) that recorded highest rate of infection bring to the fore the at-risk-population that should be targeted during vaccination exercise even though younger women may be included. Also, 80% (4/5) detection rate of HBcIgM in cord blood of babies born to infected mothers is an indication of either transplacental immunoglobulin transfer or infection of the newborn during delivery. This exposes such babies to risk of complications if infection is not remedied promptly.

## **Acknowledgement**

The authors thank all the pregnant women who participated in the study and wish to extend profound gratitude to the midwives who helped in the collection of cord blood.

## References

- Alegbeleye, O.J., Nyengidiki, K.T., and Ikemdo, I.J., (2013) Maternal and neonatal seroprevalence of hepatitis B surface antigen in a hospital based population in south-south Nigeria . *International Journal of Medicine and Medical Sciences*. 5(5): 241 – 246,
- Borgia, G., Carleo, M.A., Gaeta, G.B., Gentile, I. (2012). Hepatitis B in pregnancy. *World Journal of Gastroenterology*, 18(34):4677–4683
- Chen, C. and Chang, M. (2010). Hepatitis B and pregnancy: The scientific basis for perinatal prevention. *Cambridge Journal Online*, 21: 89 – 113.
- Coppola, N., Masiello, A., Tonziello, G. (2010 ). Factors affecting the changes in molecular epidemiology of acute hepatitis B in a Southern Italian area. *Journal of Viral Hepatitis*, 17(7):493–500.
- Coppola, N., Loquercio, G., Tonziello, G. (2013a). HBV transmission from an occult carrier with five mutations in the major hydrophilic region of HBsAg to an immunosuppressed plasma recipient. *Journal of Clinical Virology*, 58(1):315–317.
- Coppola, N., Tonziello, G., Colombatto, P. (2013b). Lamivudine-resistant HBV strain rtM204V/I in acute hepatitis. *British Journal of Infection*, 67(4):322–328.
- Coppola, N., Potenza, N., Pisaturo, M. (2013c). Liver microRNA hsa-miR125a-5p in HBV chronic infection: correlation with HBV replication and disease progression. *PLoS One*, 8(7):e65336.
- Coppola, N., Marrone, A., Pisaturo, M. (2014). Role of interleukin 28-B in the spontaneous and treatment-related clearance of HCV infection in patients with chronic HBV/HCV dual infection. *European Journal of Clinical Microbiology and Infectious Disease*, 33(4):559–567.
- Eke, A.C, Eke, U.A, Okafor, C.I. Ezebialu, I.U. Ogbuagu, C. (2011). Prevalence, correlates and pattern of Hepatitis B surface antigen in a low resource setting. *Virology Journal*. 8:12.

288 Ezeqbudo, C.N., Agbonlahor, D.E, Nwobu, G.O., Igwe, C.U., Agba, M.I., Okpala, H.O. (2004).  
 289 The seroprevalence of Hepatitis B surface antigen and Human immunodeficiency virus  
 290 (HIV) among pregnant women in Anambra state. *Shiraz E-medical Journal*, 5(2):1-8.

291 Gasim, G.I., Murad, I.A. and Adam, I. (2013). Hepatitis B and C virus among pregnant woman  
 292 in Arab and African Countries. *Journal of Infection Developing Countries* 7(8);566-578

293 Mbaawuaja, M.E., Enenebeaku, O.N.M., Okopi, A.J. and Damen, G.J.A. (2008). Hepatitis B  
 294 virus (HBV) infection among pregnant womam in Makurdi, Nigeria. *African Journal of*  
 295 *Biomedical Research*, (11): 155-159

296 Michael, A. (2013). Prevalence of hepatitis B e antigen among human immunodeficiency virus  
 297 and hepatitis B virus co-infected patients in Jos, Nigeria. *Journal of Infection in*  
 298 *Developing Countries*, 7(12):951-959.doi:10.3855/jidc.2747

299 Ndako, A.J.,Echeonwu, N.O.G., Nwankiti, O.O., Onovoh, M.E., Ujah, A., Ikani, A.P. and Paul,  
 300 A.G.(2012). Hepatitis B virus seroprevalence among pregnant females in Northern  
 301 Nigeria. *Research Journal of Medical Sciences*, 6(3); 129-133.

302 Ndams, I.S., Joshua, I.A., Luka, S.A. and Sadiq, H.O. (2008). Epidemiology of hepatitis B  
 303 infection among pregnant women in Minna, Nigeria. *Journal of World Science*. 3(3): 5–8

304 Olokoba, A.B., Salawu, F.K., Danburam, A., Olokoba, L.B., Midala, J.K., Badung, L.H. and  
 305 Olatinwo, A.W.O. (2011). Hepatitis B virus infection amongst pregnant women in North  
 306 Eastern Nigeria. A call for action. *Nigeria Journal of Clinical Practice*, 14(1).  
 307 [www.njcponline.com](http://www.njcponline.com)

308 Pennap, G.R, Osanga, E.T, Ubam, A, (2011). Seroprevalence of hepatitis B surface antigen  
 309 among pregnant women attending antenatal clinic in Federal Medical Center, Keffi,  
 310 Nigeria. *Research Journal of Medical Science* 51(2):80-82.

311 Piratvisuth, T. (2013). Optimal management of HBV infection during pregnancy. *Liver Int.*, 33  
 312 (1):188–194.

313 Sagnelli E, Stroffolini T, Mele A. (2012). Impact of co-morbidities on the severity of chronic  
 314 hepatitis B at presentation. *World Journal of Gastroenterology*, 18(14):1616–1621

315 Ugwuja, E.I. (2010). Seroprevalence of Hepatitis B surface antigen and liver function tests  
 316 among adolescents in Abakaliki, Southern Nigeria. *International Journal of Tropical.*  
 317 *Medicine*, 6(2):1-6.

318 Vranckx, R., Alisjahbana, A, Meheus, A. (1999). Hepatitis B. Virus Vaccination and ante natal  
 319 transmission of HBV markers to neonates. *Journal of Viral Hepatitis*, 6 (2): 135 – 39.

320 World Health Organization (2012) Hepatitis B. Fact Sheet No. 204. Available:  
 321 <http://www.who.int/mediacentre/factsheets/fs204/en>

322 World Health Organization [webpage on the Internet]. Hepatitis B. Geneva, Switzerland: World  
 323 Health Organization; 2013 [updated July 2013]. Available from:  
 324 <http://www.who.int/mediacentre/factsheets/fs204/en/index.html>.

325 Xu DZ, Yan YP, Choi BC. (2002). Risk factors and mechanism of transplacental transmission  
 326 of hepatitis B virus: a case-control study. *Journal of Medical Virology*, 67(1):20–26

327 Xu, G., Wei Giuo, Y., Zhang, C., Zhang, C., Zhang N. Wang, G. (2013) “An Analysis of the  
 328 molecular evolution of hepatitis B viral Genotypes A/B/D using a Bayesian Evolutionary  
 329 Method. *Virology Journal*, 10 (1). 256,

330 Zuckerman, A.J. (1996). “Hepatitis virus” In Baron S. *et al. Baron's Medical Microbiology* (4th  
 331 ed.). University of Texas Medical Branch. ISBN 0-9631172-1-1