

1
2 **ANTIMICROBIAL EFFICACY OF SELECTED NATURAL PRODUCTS ON**
3 **MICROORGANISMS ISOLATED FROM THROAT OF PATIENTS WITH**
4 **THROAT INFECTION**

5
6
7 **ABSTRACT**

8 **Introduction:** Natural products have been used in traditional medicines for treatment of infections
9 due to the antimicrobial activity they exhibit. This study therefore evaluates the efficacy of honey,
10 ginger (*Zingiber officinale*) and garlic (*Allium sativum*) extracts on microorganisms isolated from
11 throat of patients with throat infection.

12 **Methods:** The antibacterial and antifungal efficacy of honey, ginger (*Zingiber officinale*) and garlic
13 (*Allium sativum*) extracts was investigated against microorganisms isolated from throats of infected
14 patients at the ENT Department of State Specialist Hospital, Akure. using agar disc diffusion and
15 agar well diffusion technique respectively.

16 **Results:** The bacteria isolated were *Staphylococcus aureus*, *Streptococcus pyogenes*, *Pseudomonas*
17 *aeruginosa*, and *Proteus mirabilis* while the fungi isolated were *Candida albicans* and *Candida*
18 *tropicalis*. The antibacterial and antifungal assay results showed that all the bacterial isolates were
19 inhibited by honey, garlic and ginger extract however, in antibacterial assay, honey, ginger and garlic
20 showed the highest inhibition against *P. mirabilis* (19mm), *P. aeruginosa* (20 mm) and *S. aureus*
21 (23mm) respectively also, antifungal assay results showed that all the extracts had antifungal effect
22 on the fungal isolates. The combination of equal concentrations of honey plus garlic showed the
23 highest inhibitory effect on all the test bacteria followed by honey plus ginger then garlic plus ginger
24 while the combination of honey plus garlic having the highest inhibitory effect on *Candida tropicalis*
25 but garlic plus ginger combination showed the highest inhibitory effect on *Candida albicans*.

26 **Conclusion:** The result of this study therefore showed that the bacteria and fungi isolated from throat
27 of patients with throats infection demonstrated sensitivity towards the tested samples of honey, garlic
28 and ginger and hence, can serve as effective therapeutic agents in the treatment of throat infections.

29 **Keywords:** throat infection, antibacterial, antifungal, natural products
30
31

32 INTRODUCTION

33 In recent years, a lot of attention has been focused on producing medicines and products that are
34 natural. Several plants produce chemicals as primary and secondary metabolites which have
35 beneficial long-term health effects and are used effectively to treat diseases [1]. Specifically, it is the
36 secondary metabolites that exert therapeutic actions in humans. It has been stated that more than 30%
37 of entire plant species, at one time or another, are used for medicinal purposes necessarily due to the
38 amount and type of secondary metabolites they contain. These drugs of plant origin have saved lives
39 of many residents of developing countries because of their good values in treating many infectious
40 and non-infectious diseases [2]. Over the years, plants such as ginger, garlic and honey have been
41 used in traditional medicines for treatment of infections due to the antimicrobial activity they exhibit
42 [3, 4].

43 Ginger (*Zingiber officinale*) mostly used as spice and flavouring, is one of the world's best
44 medicines. Although, native to Asia, ginger is grown throughout the tropics, its therapeutic potentials
45 have been well studied and are reported to be largely due to its volatile oil and oleoresin. It has
46 analgesic, antipyretic and also antibacterial properties [5, 6]. Garlic (*Allium sativum*) is well known
47 for its antifungal, anticancer, antimicrobial activities. The antimicrobial activities of garlic have been
48 related to the presence of growth-inhibiting compounds such as Allicin and related derivatives [3].

49 Honey is the product of flower nectar produced by beehive. It has been proven to have antibacterial
50 activities. It is well-known for its treatment potential of burns and peptic ulcer, infected wounds,
51 bacterial gastroenteritis and eye infection [4]. The high antimicrobial activity of honey has been
52 attributed to its high osmotic effect, pH (3.2 – 4.5), hydrogen peroxide (H₂O₂), bee defensin, and its
53 photochemical nature [5, 7]. High osmolarity has been considered a valuable tool in the treatment of
54 infections, because it prevents the growth of bacteria [5]. Hence, Honey increases the sensitivity of
55 microorganisms to antibiotics and decrease the microbial resistance to antibiotics [4, 8].

56 Throat infection can be because of various inflammatory and infective causes such as allergies,
57 reflux disease, sinus drainage, and tonsillitis [6]. Throat infections can be of viral or infective
58 etiology, bacteria and fungi has been a challenge for medical practitioners at the ENT department
59 because the infection is difficult to treat with chemotherapy [4]. The difficulty in the treatment is due
60 to the resistant of these microorganisms to antibiotics and the reoccurrence of throat infections after
61 few months or years of treatment with antibiotics has led to increase in the morbidity of the infection
62 [9].

63 Due to the resistance of microorganisms to antibiotics, interest in finding alternative therapeutic
64 measure for the treatment of throat infection has become necessary. In this regard, the present study
65 aims at evaluating the antimicrobial activity of natural products namely honey, ginger and garlic on
66 microorganisms causing throat infections.

67 **MATERIALS AND METHODS**

68 **Study area and period**

69 The study was conducted in the Ear, Nose and Throat (ENT) Department of the State Specialist
70 Hospital, Akure and Federal University of Technology, Akure, Ondo state, Nigeria from March to
71 June, 2017.

72 **Specimen Collection**

73 Swabs from throats and tonsils were collected from patients that attended the ENT clinic for a period
74 of three weeks. Specimens were immediately transported in ice-packed containers to the
75 Microbiology Laboratory of Federal University of Technology Akure, for microbiological analysis.

76 **Ethical Approval**

77 Approval was obtained from the Medical director of the State Specialist Hospital, Akure, Ondo state,
78 Nigeria.

79 **Isolation and Identification of Microorganisms**

80 Swabs from throats were screened and identification of microorganisms was done using standard
81 bacteriological procedures as described by Cheesbrough [10]. Collected swabs were dipped into
82 1.0ml sterile physiological saline and allowed to stand for 10 minutes. It was homogenized and 0.1ml
83 of the suspension was inoculated on MacConkey agar, Mannitol salt agar, Nutrient agar and
84 incubated aerobically at 37°C for 24 hours while Potato Dextrose agar was incubated at 28°C for 48-
85 72 hours. Grown isolates were identified by their colony morphology, Gram staining reaction and
86 biochemical tests including catalase test, citrate utilization test, motility test, indole test, urease test,
87 sugar fermentation test and coagulase test. The fungal isolates were identified based on morphology
88 and microscopic characteristics.

89 **Collection and Authentication of Plant Materials**

90 The ginger, garlic and honey used were purchased at Oja-Oba market, Akure and authenticated at the
91 Museum of the Department of Crop, Soil and Pest Management, FUTA, Ondo state, Nigeria.

92

93

94 **Preparation of Plant Extracts**

95 The crude ginger and garlic extracts were prepared according to the method described by Ogodu and
96 Ekeleme [11]. The ginger and garlic were peeled and washed separately. They were then cut into
97 smaller pieces, weighed and blended in a sterile blender. The blended juice was filtered through a
98 sterile muslin cloth after which the filtrates were purified by passing through Millipore membrane
99 filter paper.

100 **Sterility Check of the extract**

101 Each of the extracts was tested for contaminants by inoculating them on nutrient agar followed by
102 incubation at 37°C for 24 hours after which the plates were observed for growth [12]. No growth in
103 the extracts after incubation indicated that the extracts are sterile after which they were assessed for
104 antimicrobial activity.

105 **Antibacterial Susceptibility Testing**

106 A suspension of 24 hours old pure culture of each bacterial isolate was prepared in nutrient broth
107 (5ml) equivalent to McFarland turbidity standard. The suspensions were spread on to the surface of
108 Mueller-Hinton agar (Oxoid, England) with sterile cotton swabs. The plates were briefly dried and
109 then antibiotic disc of honey, ginger, garlic, honey mixed with garlic (1:1), honey mixed with ginger
110 (1:1), and garlic mixed with ginger (1:1) were added to each plates and incubated over night at 37°C.
111 The diameters of zones of inhibition were measured in millimeters, with a ruler [13].

112 For positive control, antibiotic susceptibility pattern of the bacterial isolates was tested with
113 amoxicillin by disc diffusion method on Mueller-Hinton agar (Oxoid, England). The plates were
114 incubated at 37°C for 24 hours and observed for zone of inhibition after which the zones of inhibition
115 were measured and interpreted according to Clinical and Laboratory Standard Institute [14].

116 **Antifungal Susceptibility Testing**

117 A suspension of the pure culture of each yeast isolate was prepared in yeast extract broth. The
118 antifungal susceptibility of the isolates was performed by agar well diffusion method. Six equidistant
119 wells of 5mm in diameter were drilled using a sterile cork borer at different sites on the plates.
120 100µL of each of the extract was aseptically introduced into each holes, and ketoconazole prepared
121 in solution was used as a positive control. The set up was allowed to stabilize for 3 hours before
122 being incubated at 28°C for 48-72 hours after which the zone of inhibition was measured in
123 millimeters [15].

124

125 **Statistical analysis**

126 Results were expressed by means of \pm SD. Statistical significance was established using one-way
 127 analysis of variance (ANOVA). Means were separated according to Duncan’s New Multiple Range
 128 Test ($p < 0.05$) using software SPSS 20.0.

129 **RESULTS**

130 **Isolation and Identification of Microorganisms**

131 A total of 126 isolates were collected from throat swab of patients with throat infections
 132 over a 3 weeks’ period. The bacterial isolates identified from the specimen collected include
 133 *Streptococcus pyogenes*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Proteus mirabilis*
 134 while the fungal isolates include *Candida albicans* and *Candida tropicalis*.

135 The results revealed that the highest numbers of patients with throat infections were the male patients
 136 between the ages 10-20 and the highest microbial count was recorded among the male patients.
 137 Details of the demographic distribution of patients with throat infection and the total viable count of
 138 bacteria and fungi are presented in **Table 1** and **2** respectively.

139 **Table 1: Demographic Distribution of Patients with Throat Infection**

A g e (y e a r s)		M a l e (%)		F e m a l e (%)		T o t a l (%)	
1	- 1	0	10 (20.41)	7	(14.29)	17	(34.69)
1	0 - 2	0	12 (24.49)	9	(18.37)	21	(42.86)
2	0 - 3	0	7 (14.29)	4	(8.16)	11	(22.45)
T o t a l		2	9 (59.18)	2	0 (40.82)	4	9 (100)

140

141 **Table 2: Total Viable Bacterial and Fungal Count of Patients with Throat Infection.**

G e n d e r	Bacterial counts (CFU/ml)	Yeast counts (CFU/ml)	Mould counts (SFU/ml)
M a l e	552.00 \pm 1.15 ^b	300.00 \pm 0.50 ^b	0.00 \pm 0.00 ^a
F e m a l e	450.00 \pm 0.54 ^a	230.00 \pm 1.54 ^a	0.00 \pm 0.00 ^a

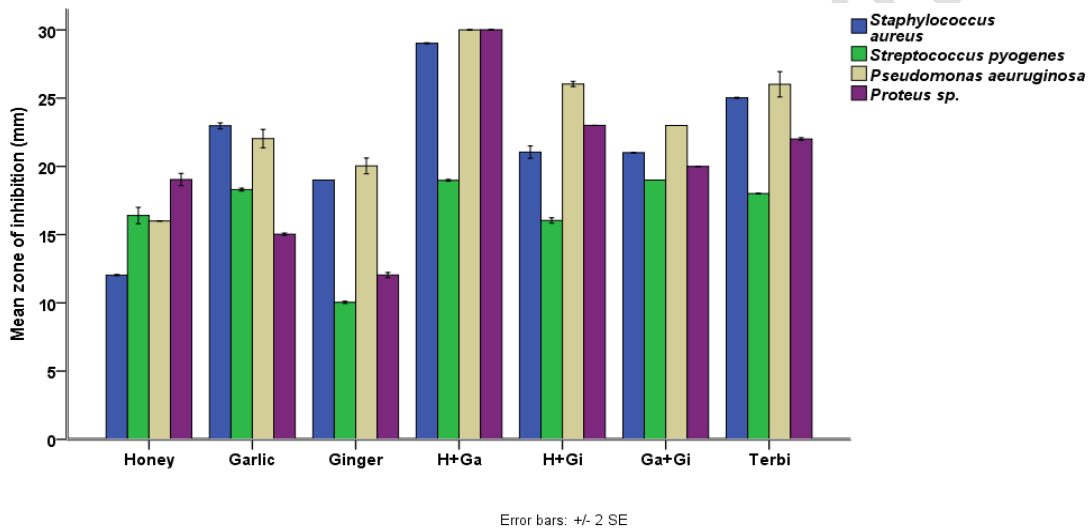
142 Values are presented as mean \pm SE. Values in the same column carrying different superscript are
 143 significantly different at ($p \leq 0.05$) using Duncan’s New Multiple Range test.

144

145 **Susceptibility Pattern of the Isolates to Honey, Ginger and Garlic**

146 The antimicrobial activities of honey, garlic, ginger and their synergistic effects are presented for
147 bacteria and fungi in **Figure 1 and 2** respectively. The highest inhibitory effect of honey was
148 observed with *Proteus mirabilis*, garlic with *Staphylococcus aureus* while ginger showed the highest
149 inhibitory activity against *Pseudomonas aeruginosa*. The synergistic effect of honey and garlic
150 produced the highest inhibitory effect on the bacterial isolates compared to honey/ginger mixture and
151 garlic/ginger mixture.

152 *Candida albicans* showed the highest sensitivity to garlic and ginger while the most sensitivity to
153 honey was observed with *Candia tropicalis*. The synergistic effects of the natural products inhibited
154 all the yeast isolates.

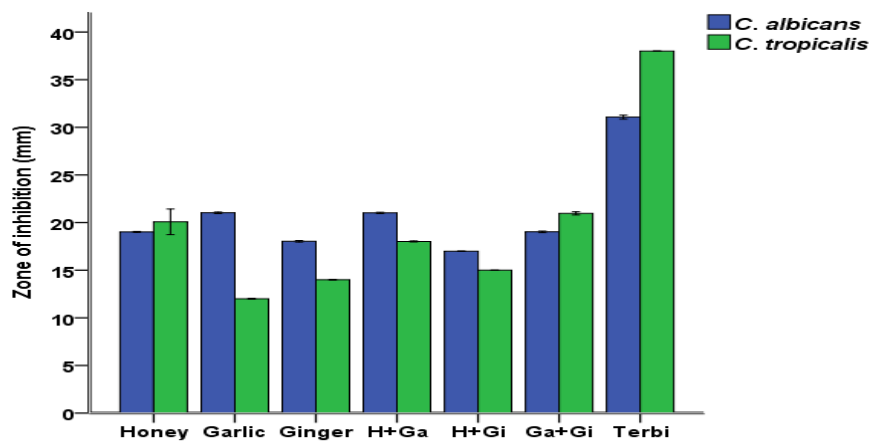


155

156 **Fig. 1: Antibacterial susceptibility pattern of ginger, honey and garlic on bacterial isolated**
157 **from throats of infected patients.**

158 *Key: H+Ga = Honey plus garlic, H+Gi = honey plus ginger, Ga+Gi = garlic plus ginger, Terbi =*
159 *Antibiotic*

160



161

UNDER PEER REVIEW

162 **Fig. 2: Antifungal susceptibility of fungal isolates from throat infection to honey, garlic and**
163 **ginger**

164 *Key: H+Ga = Honey plus garlic, H+Gi = honey plus ginger, Ga+Gi = garlic plus ginger, Terbi =*
165 *Antibiotic*

166 **DISCUSSION**

167 This study has shown that throat infections are caused by bacteria and fungi. However, there were
168 differences in the microbial load of male patients to that of the female patient at State Specialist
169 Hospital Akure. The total viable bacterial and fungal counts observed in male patients was higher
170 than what was observed in female patients. Variations in microbial load may be attributed to the
171 differences in anatomy, lifestyle and socioeconomic differences [16]. The result of this work also
172 revealed that different bacteria such as *Streptococcus pyogenes*, *Staphylococcus aureus*,
173 *Pseudomonas aeruginosa*, and *Proteus mirabilis* and yeast such as *Candida tropicalis* and *Candida*
174 *albicans* may be responsible for causing throat infections. This data collaborates with the previous
175 work [17]. The presence of these bacteria in the throat could be as a result of contamination of the
176 food and water that individuals eat or drink, environmental factors, or by the microflora of the throat
177 [18].

178 All the tested bacterial and fungal isolates were completely susceptible to the tested samples of
179 honey, ginger and garlic and their mixtures. This study further revealed that honey-garlic mixture
180 produced the highest inhibitory effect on the test bacterial and fungal isolates compared to the single
181 effects and the other combinations i.e. honey-ginger and ginger-garlic mixtures. This can be
182 explained to be due to the synergistic effects of honey and garlic on the isolates as many compounds
183 present in both the honey and garlic combined to inhibit the organisms. This result is in close
184 proximity to the other results [3, 11].

185 In previous study, local residents have been found to use honey for pharyngitis and respiratory
186 ailments [4]. The antimicrobial activity of honey is highly complex due to the involvement of multiple
187 compounds and due to the large variation in the concentrations of these compounds among honeys.
188 The use of honey where antibiotic treatments had failed to clear infection have been demonstrated in
189 many studies [3, 4]. The control of infection by honey is said to be attributed to its high osmolarity
190 while its hydrogen peroxide content, low pH, content of phenol (inhibin) and other unidentified
191 properties are responsible for its antibacterial properties [19, 20, 21]. Acidity is also one of the
192 factors that contributes to the antibacterial property of honey [20]. The medicinal properties of ginger
193 are due to variety of bioactive compounds such as tannins, flavonoid, glycosides, essential oils,
194 saponins, phytosterols, amides and alkaloids [3, 11]. The antimicrobial properties of garlic may be

195 due to its potentially active chemical constituents as it contains at least 33 sulphur compounds and
196 several enzymes. One of the most biologically active compounds in garlic is allicin (diallyl
197 thiosulfinate or diallyl disulfide) has been largely attributed to be responsible for the medicinal
198 effects of garlic [3].

199 CONCLUSION

200 The single and combined samples of honey, ginger and garlic showed a high degree of antimicrobial
201 activity on the tested bacterial and fungal isolates from throat infections, therefore, these natural
202 products can serve as effective therapeutic agents and a natural alternative to conventional antibiotics
203 in the treatment of throat infections. The combination of honey and garlic however show much
204 promise in the development of phytomedicines in the treatment of throat infections.

205

206 REFERENCES

- 207 1 Adesina, S. K., Illoh, H. C., Jonny, I. I. and Imo, E. J. (2013). African Milstletoes
208 (Loranthaceae); Ethnopharmacology, chemistry and medicinal values: An update. *African*
209 *Journal of Traditional Complement Alternative Medicine*. **10**(3):161-170
- 210 2 Tala, S. D., Gatsing, D., Fodouop, S. P. C., Fokunang, C., Kengni, F. and Djimeli, M. N.
211 (2015). *In vivo* anti-salmonella activity of aqueous extract of *Euphorbia prostrata* Aiton
212 (Euphorbiaceae) and its toxicological evaluation. *Asian Pacific Journal of Tropical*
213 *Biomedicine*. **5**(4): 310-318
- 214 3 Zakia, M., Nihal, A. and Reem El B. (2014). A Natural Alternative to Conventional
215 Antibiotics. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. **5**(4):
216 588- 599
- 217 4 Nanda, M. S., Mittal, S. P. and Gupta, V. (2016). Role of honey as adjuvant therapy in
218 patients with sore throat. *National Journal of Physiology, Pharmacy and Pharmacology*, **7**(4):
219 412-415
- 220 5 Yahaya, O., Yabefa, J. A., Umar, I. O., Datshen, M. M., Egbunu, Z. K. and Ameh, J. (2012).
221 Combine antimicrobial effect of ginger and honey on some human pathogens. *British Journal*
222 *of Pharmacology and Toxicology*, **3**(5): 237-239.
- 223 6 Metwali, Z., Abdalla, N. and El Barrani, R. (2014). A natural alternative to conventional
224 antibiotics. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*,
225 **5**(4):588-599.
- 226 7 Mshelia, B. M., Adeshina, G. O. and Onaolapo, J. A. (2017). The antibacterial activity of
227 honey and lemon juice against *Streptococcus pneumoniae* and *Streptococcus pyogenes*

- 228 isolates from respiratory tract infections. *Advances in Biotechnology and Microbiology*, **4**(5):
229 001-008.
- 230 8 Vallianou NG, Gounari P, Skourtis A, Vallianou NG, Gounari P, Skourtis A. (2014). Honey
231 and its anti-inflammatory, anti-bacterial and anti-oxidant properties. *General Med.* **2**:132.
- 232 9 Del Mar CB, Glasziou PP, Spinks AB. (2006). Antibiotics for sore throat. Cochrane Database
233 Syst Rev. 4:CD000023.
- 234 10 Cheesbrough, M. (2006). *District Laboratory Practice in Tropical Countries*. Part 2, UK;
235 Cambridge University Press.
- 236 11 Ogodo, C. and Ekeleme, U. G. (2013). *In-vitro* antibacterial activity of garlic cloves and
237 ginger rhizomes on food-borne pathogens. *International Journal of Basic and Applied*
238 *Sciences*, **2**(4): 387-392.
- 239 12 Arekemase, M. O., Kayode, R. M. O. and Ajiboye, A. E. (2011). Antimicrobial activity and
240 phytochemical analysis of *Jatropha curcas* plant against some selected microorganisms.
241 *International Journal of Biology*, **3**(3): 52-59.
- 242 13 Amel, A. S., Fadwa, M. E., Smah, A. S. and Nazar, A. O. (2015). Antimicrobial activity of
243 *Zingiber officinale* (Ginger) oil against bacteria isolated from children throat. *Journal of*
244 *Microbiology and Biomedical Research*,
- 245 14 Clinical and Laboratory Standard Institute, CLSI. (2014). Performance Standards for
246 Antimicrobial Susceptibility Testing. *Twenty-fourth Informational Supplement*. CLSI
247 document M 100-S24. Wayne, P. A: 550-84.
- 248 15 Owoseni, A. A. and Ajayi, A. (2010). Antimicrobial properties of ethanolic and aqueous
249 extracts of *Cymbopogon citratus* on selected bacteria and fungi. *J. Med. Appl. Biosci.* **2**(4),
250 64–73.
- 251 16 Belstrom D., Holmstrum P., Nielsen C. H., Kirbkby N. and Twetman S. (2014). Bacterial
252 profiles of saliva in relation to diet, lifestyle factors and socioeconomic status. *Journal of oral*
253 *microbiology.* **6**(1): 23609
- 254 17 Kenealy T. (2015). Acute Infective Sore Throat. *Clinical Evidence Handbook.* **91**(10):526-
255 527.
- 256 18 Ahmad M. M., Kurawa Z. M., Shuaibu I. and Yahaya G. (2016). Microbiological assessment
257 of bacterial isolates from ear, nose and throat (ENT) among patients attending Aminu Kano
258 Teaching hospital. *Nigerian Journal of basic and applied science.* **24**(1): 15-18
- 259 19 Bansal V, Medhi B, Pandhi P. (2005). Honey – A remedy rediscovered and its therapeutic
260 utility. *Kathmandu Univ Med J (KUMJ).* **3**(3):305-9.

- 261 20 Olaitan PB, Adeleke OE, Ola IO. (2007). Honey: A reservoir for microorganisms and an
262 inhibitory agent for microbes. *Afr Health Sci.* 7(2):159-65.
- 263 21 Kamaruddin MY, Zainabe SA, Anwar S, Razif MA, Yassim MY. (2012). The efficacy of
264 honey dressing on chronic wounds and ulcers. In: Juraj M, editor. *Honey: Current Research
265 and Clinical Uses.* New York: Nova Science Publishers; 185-96.
- 266
- 267

UNDER PEER REVIEW