

1
2 **ANTIMICROBIAL EFFICACY OF SELECTED NATURAL PRODUCTS ON**
3 **MICROORGANISMS ISOLATED FROM THROAT OF PATIENTS WITH**
4 **THROAT INFECTION**

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7 **ABSTRACT**

8 **Introduction:** Natural products have been used in traditional medicines for treatment of infections
9 due to the antimicrobial activity they exhibit. This study therefore evaluates the efficacy of honey,
10 ginger (*Zingiber officinale*) and garlic (*Allium sativum*) extracts on microorganisms isolated from
11 throat of patients with throat infection.

12 **Methods:** The antibacterial and antifungal efficacy of honey, ginger (*Zingiber officinale*) and garlic
13 (*Allium sativum*) extracts was investigated against microorganisms isolated from throats of infected
14 patients at the ENT Department of State Specialist Hospital, Akure. using agar disc diffusion and
15 agar well diffusion technique respectively.

16 **Results:** The bacteria isolated were *Staphylococcus aureus*, *Streptococcus pyogenes*, *Pseudomonas*
17 *aeruginosa*, and *Proteus mirabilis* while the fungi isolated were *Candida albicans* and *Candida*
18 *tropicalis*. The antibacterial and antifungal assay results showed that all the bacterial isolates were
19 inhibited by honey, garlic and ginger extract however, in antibacterial assay, honey, ginger and garlic
20 showed the highest inhibition against *P. mirabilis* (19mm), *P. aeruginosa* (20 mm) and *S. aureus*
21 (23mm) respectively also, antifungal assay results showed that all the extracts had antifungal effect
22 on the fungal isolates. The combination of equal concentrations of honey plus garlic showed the
23 highest inhibitory effect on all the test bacteria followed by honey plus ginger then garlic plus ginger
24 while the combination of honey plus garlic having the highest inhibitory effect on *Candida tropicalis*
25 but garlic plus ginger combination showed the highest inhibitory effect on *Candida albicans*.

26 **Conclusion:** The result of this study therefore showed that the bacteria and fungi isolated from throat
27 of patients with throats infection demonstrated sensitivity towards the tested samples of honey, garlic
28 and ginger and hence, can serve as effective therapeutic agents in the treatment of throat infections.

29 **Keywords:** throat infection, antibacterial, antifungal, natural products
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31

32 INTRODUCTION

33 In recent years, a lot of attention has been focused on producing medicines and products that are
34 natural. Several plants produce chemicals as primary and secondary metabolites which have
35 beneficial long-term health effects and are used effectively to treat diseases [1]. Specifically, it is the
36 secondary metabolites that exert therapeutic actions in humans. It has been stated that more than 30%
37 of entire plant species, at one time or another, are used for medicinal purposes necessarily due to the
38 amount and type of secondary metabolites they contain. These drugs of plant origin have saved lives
39 of many residents of developing countries because of their good values in treating many infectious
40 and non-infectious diseases [2]. Over the years, plants such as ginger, garlic and honey have been
41 used in traditional medicines for treatment of infections due to the antimicrobial activity they exhibit
42 [3, 4].

43 Ginger (*Zingiber officinale*) mostly used as spice and flavouring, is one of the world's best
44 medicines. Although, native to Asia, ginger is grown throughout the tropics, its therapeutic potentials
45 have been well studied and are reported to be largely due to its volatile oil and oleoresin. It has
46 analgesic, antipyretic and also antibacterial properties [5, 6]. Garlic (*Allium sativum*) is well known
47 for its antifungal, anticancer, antimicrobial activities. The antimicrobial activities of garlic have been
48 related to the presence of growth-inhibiting compounds such as Allicin and related derivatives [3].

49 Honey is the product of flower nectar produced by beehive. It has been proven to have antibacterial
50 activities. It is well-known for its treatment potential of burns and peptic ulcer, infected wounds,
51 bacterial gastroenteritis and eye infection [4]. The high antimicrobial activity of honey has been
52 attributed to its high osmotic effect, pH (3.2 – 4.5), hydrogen peroxide (H₂O₂), bee defensin, and its
53 photochemical nature [5, 7]. High osmolarity has been considered a valuable tool in the treatment of
54 infections, because it prevents the growth of bacteria [5]. Hence, Honey increases the sensitivity of
55 microorganisms to antibiotics and decrease the microbial resistance to antibiotics [4, 8].

56 Throat infection can be because of various inflammatory and infective causes such as allergies,
57 reflux disease, sinus drainage, and tonsillitis [6]. Throat infections can be of viral or infective
58 etiology, bacteria and fungi has been a challenge for medical practitioners at the ENT department
59 because the infection is difficult to treat with chemotherapy [4]. The difficulty in the treatment is due
60 to the resistant of these microorganisms to antibiotics and the reoccurrence of throat infections after
61 few months or years of treatment with antibiotics has led to increase in the morbidity of the infection
62 [9].

63 Due to the resistance of microorganisms to antibiotics, interest in finding alternative therapeutic
64 measure for the treatment of throat infection has become necessary. In this regard, the present study
65 aims at evaluating the antimicrobial activity of natural products namely honey, ginger and garlic on
66 microorganisms causing throat infections.

67 **MATERIALS AND METHODS**

68 **Study area and period**

69 The study was conducted in the Ear, Nose and Throat (ENT) Department of the State Specialist
70 Hospital, Akure and Federal University of Technology, Akure, Ondo state, Nigeria from March to
71 June, 2017.

72 **Specimen Collection**

73 Swabs from throats and tonsils were collected from patients that attended the ENT clinic for a period
74 of three weeks. Specimens were immediately transported in ice-packed containers to the
75 Microbiology Laboratory of Federal University of Technology Akure, for microbiological analysis.

76 **Ethical Approval**

77 Approval was obtained from the Medical director of the State Specialist Hospital, Akure, Ondo state,
78 Nigeria.

79 **Isolation and Identification of Microorganisms**

80 Swabs from throats were screened and identification of microorganisms was done using standard
81 bacteriological procedures as described by Cheesbrough [10]. Collected swabs were dipped into
82 1.0ml sterile physiological saline and allowed to stand for 10 minutes. It was homogenized and 0.1ml
83 of the suspension was inoculated on MacConkey agar, Mannitol salt agar, Nutrient agar and
84 incubated aerobically at 37°C for 24 hours while Potato Dextrose agar was incubated at 28°C for 48-
85 72 hours. Grown isolates were identified by their colony morphology, Gram staining reaction and
86 biochemical tests including catalase test, citrate utilization test, motility test, indole test, urease test,
87 sugar fermentation test and coagulase test. The fungal isolates were identified based on morphology
88 and microscopic characteristics.

89 **Collection and Authentication of Plant Materials**

90 The ginger, garlic and honey used were purchased at Oja-Oba market, Akure and authenticated at the
91 Museum of the Department of Crop, Soil and Pest Management, FUTA, Ondo state, Nigeria.

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94 **Preparation of Plant Extracts**

95 The crude ginger and garlic extracts were prepared according to the method described by Ogoto and
96 Ekeleme [11]. The 500g of ginger and garlic were peeled and washed separately. They were then cut
97 into smaller pieces, weighed and blended in a sterile blender. The blended ginger and garlic yielded
98 126ml and 173ml of juice respectively, the juice was filtered through a sterile muslin cloth after
99 which the filtrates were purified by passing through Millipore membrane filter paper.

100 **Sterility Check of the extract**

101 Each of the extracts was tested for contaminants by inoculating them on nutrient agar followed by
102 incubation at 37°C for 24 hours after which the plates were observed for growth [12]. No growth in
103 the extracts after incubation indicated that the extracts are sterile after which they were assessed for
104 antimicrobial activity.

105 **Antibacterial Susceptibility Testing**

106 A suspension of 24 hours old pure culture of each bacterial isolate was prepared in nutrient broth
107 (5ml) equivalent to McFarland turbidity standard. The suspensions were spread on to the surface of
108 Mueller-Hinton agar (Oxoid, England) with sterile cotton swabs. The plates were briefly dried and
109 then a circular paper disc which has been soaked overnight in concentrated honey, ginger, garlic,
110 antimicrobial susceptibility assay for the combinations of the selected natural products were carried
111 out by mixing 100ml of concentrated honey with 100ml of concentrated garlic and mixed thoroughly
112 to give a mixture of honey mixed with garlic (1:1), this was repeated for; honey mixed with ginger
113 (1:1), and garlic mixed with ginger (1:1) were added to each plates and incubated over night at 37°C.
114 The diameters of zones of inhibition were measured in millimeters, with a ruler [13].

115 For positive control, antibiotic susceptibility pattern of the bacterial isolates was tested with
116 amoxicillin by disc diffusion method on Mueller-Hinton agar (Oxoid, England). The plates were
117 incubated at 37°C for 24 hours and observed for zone of inhibition after which the zones of inhibition
118 were measured and interpreted according to Clinical and Laboratory Standard Institute [14].

119 **Antifungal Susceptibility Testing**

120 A suspension of the pure culture of each yeast isolate was prepared in yeast extract broth. The
121 antifungal susceptibility of the isolates was performed by agar well diffusion method. Six equidistant
122 wells of 5mm in diameter were drilled using a sterile cork borer at different sites on the plates.
123 100µL of each of the extract was aseptically introduced into each holes, and ketoconazole prepared
124 in solution was used as a positive control. The set up was allowed to stabilize for 3 hours before

125 being incubated at 28°C for 48-72 hours after which the zone of inhibition was measured in
 126 millimeters [15].

127

128 **Statistical analysis**

129 Results were expressed by means of ±SD. Statistical significance was established using one-way
 130 analysis of variance (ANOVA). Means were separated according to Duncan’s New Multiple Range
 131 Test (p< 0.05) using software SPSS 20.0.

132 **RESULTS**

133 **Isolation and Identification of Microorganisms**

134 A total of 126 isolates were collected from throat swab of patients with throat infections
 135 over a 3 weeks’ period. The bacterial isolates identified from the specimen collected include
 136 *Streptococcus pyogenes*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Proteus mirabilis*
 137 while the fungal isolates include *Candida albicans* and *Candida tropicalis*.

138 The results revealed that the highest numbers of patients with throat infections were the male patients
 139 between the ages 10-20 and the highest microbial count was recorded among the male patients.
 140 Details of the demographic distribution of patients with throat infection and the total viable count of
 141 bacteria and fungi are presented in **Table 1** and **2** respectively.

142 **Table 1: Demographic Distribution of Patients with Throat Infection**

A g e (y e a r s)		M a l e (%)		F e m a l e (%)		T o t a l (%)	
1	- 1	0	1 0 (2 0 . 4 1)	7	(1 4 . 2 9)	1 7	(3 4 . 6 9)
1	0 - 2	0	1 2 (2 4 . 4 9)	9	(1 8 . 3 7)	2 1	(4 2 . 8 6)
2	0 - 3	0	7 (1 4 . 2 9)	4	(8 . 1 6)	1 1	(2 2 . 4 5)
T o t a l		1	2 9 (5 9 . 1 8)	2 0	(4 0 . 8 2)	4 9	(1 0 0)

143

144 **Table 2: Total Viable Bacterial and Fungal Count of Patients with Throat Infection.**

G e n d e r	Bacterial counts (CFU/ml)	Yeast counts (CFU/ml)	Mould counts (SFU/ml)
M a l e	5 5 2 . 0 0 ± 1 . 1 5 ^b	3 0 0 . 0 0 ± 0 . 5 0 ^b	0 . 0 0 ± 0 . 0 0 ^a

F e m a l e 4 5 0 . 0 0 ± 0 . 5 4 ^a 2 3 0 . 0 0 ± 1 . 5 4 ^a 0 . 0 0 ± 0 . 0 0 ^a

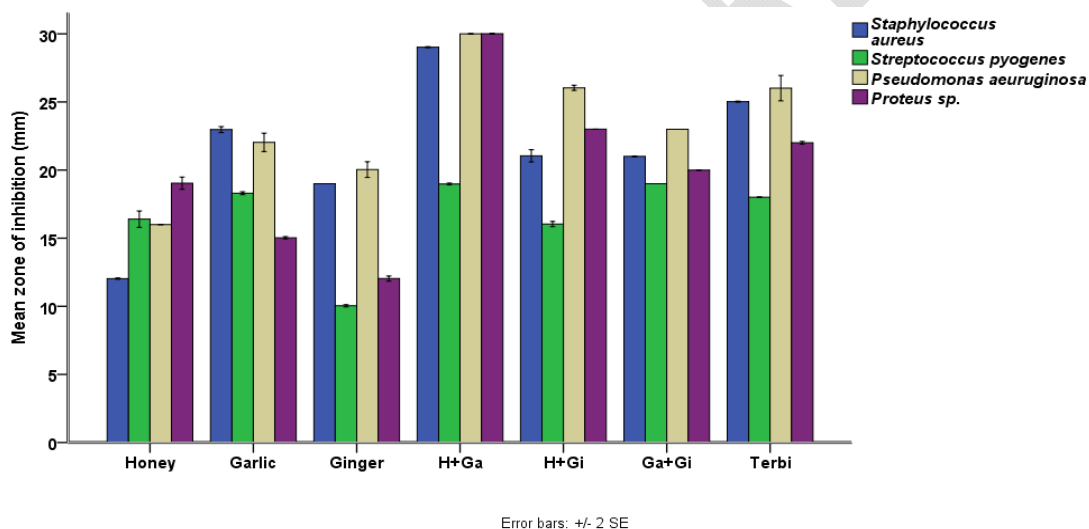
145 Values are presented as mean ±SE. Values in the same column carrying different superscript are
 146 significantly different at (p≤ 0.05) using Duncan’s New Multiple Range test.

147

148 **Susceptibility Pattern of the Isolates to Honey, Ginger and Garlic**

149 The antimicrobial activities of honey, garlic, ginger and their synergistic effects are presented for
 150 bacteria and fungi in **Figure 1 and 2** respectively. The highest inhibitory effect of honey was
 151 observed with *Proteus mirabilis*, garlic with *Staphylococcus aureus* while ginger showed the highest
 152 inhibitory activity against *Pseudomonas aeruginosa*. The synergistic effect of honey and garlic
 153 produced the highest inhibitory effect on the bacterial isolates compared to honey/ginger mixture and
 154 garlic/ginger mixture.

155 *Candida albicans* showed the highest sensitivity to garlic and ginger while the most sensitivity to
 156 honey was observed with *Candia tropicalis*. The synergistic effects of the natural products inhibited
 157 all the yeast isolates.

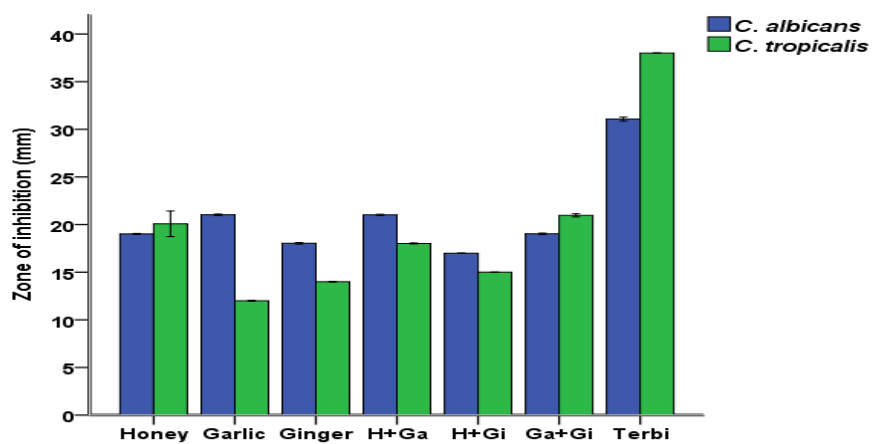


158

159 **Fig. 1: Antibacterial susceptibility pattern of ginger, honey and garlic on bacterial isolated**
 160 **from throats of infected patients.**

161 Key: H+Ga = Honey plus garlic, H+Gi = honey plus ginger, Ga+Gi = garlic plus ginger, Terbi =
 162 Antibiotic

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165 **Fig. 2: Antifungal susceptibility of fungal isolates from throat infection to honey, garlic and**
166 **ginger**

167 *Key: H+Ga = Honey plus garlic, H+Gi = honey plus ginger, Ga+Gi = garlic plus ginger, Terbi =*
168 *Antibiotic*

169 **DISCUSSION**

170 This study has shown that throat infections are caused by bacteria and fungi. However, there were
171 differences in the microbial load of male patients to that of the female patient at State Specialist
172 Hospital Akure. The total viable bacterial and fungal counts observed in male patients was higher
173 than what was observed in female patients. Variations in microbial load may be attributed to the
174 differences in anatomy, lifestyle and socioeconomic differences [16]. The result of this work also
175 revealed that different bacteria such as *Streptococcus pyogenes*, *Staphylococcus aureus*,
176 *Pseudomonas aeruginosa*, and *Proteus mirabilis* and yeast such as *Candida tropicalis* and *Candida*
177 *albicans* may be responsible for causing throat infections. This data collaborates with the previous
178 work [17]. The presence of these bacteria in the throat could be as a result of contamination of the
179 food and water that individuals eat or drink, environmental factors, or by the microflora of the throat
180 [18].

181 All the tested bacterial and fungal isolates were completely susceptible to the tested samples of
182 honey, ginger and garlic and their mixtures. This study further revealed that honey-garlic mixture
183 produced the highest inhibitory effect on the test bacterial and fungal isolates compared to the single
184 effects and the other combinations i.e. honey-ginger and ginger-garlic mixtures. This can be
185 explained to be due to the synergistic effects of honey and garlic on the isolates as many compounds
186 present in both the honey and garlic combined to inhibit the organisms. This result is in close
187 proximity to the other results [3, 11].

188 In previous study, local residents have been found to use honey for pharyngitis and respiratory
189 ailments [4]. The antimicrobial activity of honey is highly complex due to the involvement of multiple
190 compounds and due to the large variation in the concentrations of these compounds among honeys.
191 The use of honey where antibiotic treatments had failed to clear infection have been demonstrated in
192 many studies [3, 4]. The control of infection by honey is said to be attributed to its high osmolarity
193 while its hydrogen peroxide content, low pH, content of phenol (inhibin) and other unidentified
194 properties are responsible for its antibacterial properties [19, 20, 21]. Acidity is also one of the
195 factors that contributes to the antibacterial property of honey [20]. The medicinal properties of ginger
196 are due to variety of bioactive compounds such as tannins, flavonoid, glycosides, essential oils,
197 saponins, phytosterols, amides and alkaloids [3, 11]. The antimicrobial properties of garlic may be

198 due to its potentially active chemical constituents as it contains at least 33 sulphur compounds and
199 several enzymes. One of the most biologically active compounds in garlic is allicin (diallyl
200 thiosulfinate or diallyl disulfide) has been largely attributed to be responsible for the medicinal
201 effects of garlic [3].

202 CONCLUSION

203 The single and combined samples of honey, ginger and garlic showed a high degree of antimicrobial
204 activity on the tested bacterial and fungal isolates from throat infections, therefore, these natural
205 products can serve as effective therapeutic agents and a natural alternative to conventional antibiotics
206 in the treatment of throat infections. The combination of honey and garlic however show much
207 promise in the development of phytomedicines in the treatment of throat infections.

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