## <u>Original Research Article</u> Physiological quality in Leucaena leucocephala seeds conditioned with potassium nitrate submitted to saline and water stresses

## ABSTRACT

Leucaena leucocephala is a species that occurs in semiarid regions capable of developing physiological and biochemical strategies in adverse environmental situations, such as saline soils and water deficiency. Even so, there are still losses in germination and initial development of plants mainly due effects the abiotic stresses, with this we have looked for alternatives that diminish the impact on the seeds with the use of potassium nitrate. The objective of this work was to evaluate the physiological quality in L. leucocephala seeds conditioned in potassium nitrate solution and submitted to saline and water stresses. After scarification, the unconditioned leucine seeds were separated and identified, and the conditioners were immersed in the 1.0 Mmol/L solution of potassium nitrate, 24 hours. The treatments were NaCI: 0; 30; 60; 90; 120 and 150 Mmol/L and the water potential: 0,0; -0.2; -0.4; -0.6, and -0.8 MPa. The seeds were distributed over two leaves of Germitest®, moistened with distilled water 2.5 times the dry weight and incubated in B.O.D. at 25 °C for 10 days. Then, normal seedlings, germination speed index, seedling length and seedling mass were measured. Eleven treatments with four replicates of 50 seeds were used. The design was completely randomized, and the statistical analyzes were done in SAS. The use of potassium nitrate (KNO<sub>3</sub>) contributed to the maintenance of the physiological guality of L. leucochephala seeds under conditions of saline stress and water restriction.

### 10

11 Keywords: Leucena; water potencial; salinity

### 12

### 13 1. INTRODUCTION

14

The *Leucaena leucocephala* (Lam.) Belongs to the family Fabaceae (Subfamily Mimosidae), is fast growing, is nitrogen fixer and has been highlighted as a promising alternative for the recovery vegetation cover and rehabilitation of degraded areas, since it has symbiosis with nitrogen-fixing bacteria, improving soil fertility [1].

19

Its use has met a wide variety of demands, including reforestation disturbed areas for erosion control, animal feed, green manuring, fence posts, poles and cellulose. Sowing can be done either by seedlings or directly on the site [2], however, sowing saline soils in regions with a low frequency of rainfall can restrict soil moisture, resulting in slow and irregular emergence, with direct reflexes in the development of seedling [3].

25

The effects of salinity on germination can be noticed both by interference of salts in the cellular metabolism and by reduction the osmotic potential of seed, causing water stress and making water absorption difficult [4]. Salinity may also cause injury, such as chlorophyll degradation and changes in protein metabolism, important amino acid levels linked to thegermination process, and seedling initial development [5].

The presence of water is essential, and its scarcity is one of the adversities encountered by species of plants that develop in saline or dry environments, since availability of water to the seed, as well the speed of absorption, is directly influenced of the water potential difference between seed and soil [6]. Therefore, seeds of species the Fabaceae family even in arid and semiarid regions develop physiological and biochemical strategies capable of developing in adverse environmental situations, such as, in soils with high salt concentration or affected by water deficiency [7,8].

38

39 Due to the severity of losses caused by the effects of saline and water stress on germination 40 and initial plant development, alternatives and treatments have been sought to reduce their 41 impact on seeds. Among the substances with the potential to induce a resistance to these 42 types of stress in the seeds, nitric oxide (NO), inorganic free radical and an extremely 43 versatile biological messenger can be supplied exogenously the seeds through their 44 conditioning in potassium nitrate solutions [9].

Research has demonstrated the involvement of nitric oxide in hormonal signaling [10] and in numerous physiological plant processes, including: protective function against oxidative stress [11]. In seeds, nitric oxide stimulates germination, both under normal conditions and under stress, besides favoring seed dormancy of some species [12] and promoting the elongation and formation of adventitious roots [13].

50 The objective of this work was to evaluate physiological quality in *Leucaena leucocephala* 51 seeds conditioned in potassium nitrate solution and submitted to saline and water stresses.

52

### 53 2. MATERIAL AND METHODS

54

Seeds of *L. leucocephala* were collected from mature fruits with brown staining from ten matrices, spaced at a distance of 6 meters, located in the vicinity of the municipality of Macaíba - RN (Latitude: -5.86, Longitude: -35.33) in the period from July to August 2017. The region presents Aw tropical rainy climate, with an average temperature of 26 °C, relative air humidity of 74 % and precipitation of 1,134 mm annually, according to Köppen and Geiger [14].

61 After collection, the seeds were manually removed from the pods, selected and mechanically 62 scarified, using N°. 100 sandpaper, on side opposite the thread, only to break the tegument. 63 After scarification, the water content of seeds was determined by greenhouse method at 105 64  $\pm$  3 °C for 24 hours, as recommended by Brazil [15]. And the results were expressed in 65 average percentage of humidity (wet basis).

66 Then, the unconditioned seeds were separated and deposited in identified glass containers. 67 The conditioned seeds were immersed in glass containers containing the 1.0 Mmol/L solution of potassium nitrate for 24 hours. To simulate the stress conditions, following 68 concentrations of NaCl: 0 were used; 30; 60; 90; 120 and 150 Mmol/L [16], and also the 69 70 water potentials: 0; -0.2; -0.4; -0.6 and -0.8 MPa, calculated according to the equation 71 proposed by Michel and Kaufmann [17]. After conditioning and under conditions of saline 72 and water stress, the seeds had their physiological potential evaluated through the tests 73 described below.

74

For germination, four replicates of 50 seeds were used for each treatment, distributed on two sheets of Germitest<sup>®</sup> type paper towels and covered with a third sheet of the same paper, then rolled into a roller. The substrate was moistened with distilled water (AD) in an amount2.5 times the dry substrate weight, according to Brazil [15].

Subsequently, paper rolls with seeds that were not conditioned and conditioned with potassium nitrate were subjected to saline stress and water stress separately and incubated in a type germination chamber (Biochemical Oxygen Demand) at 25 °C for 10 days, when final seedlings were counted. Results were expressed as percentage of germination.

The germination speed was carried out in conjunction with the germination test, distributed on two sheets of Germitest<sup>®</sup> type paper towels and covered with a third sheet of the same paper, then rolled into a roller. The substrate was moistened with distilled water (AD) in an amount 2.5 times the dry substrate weight, according to Brazil [15], with daily counts of seeds that emitted a primary root of at least 2 mm from the first to the tenth day. Determined from the germination velocity index (GVI), calculated according to the formula proposed by Maguire [18].

90

91 The dry mass of seedlings was performed with total of ten normal seedlings per 92 experimental unit, randomly taken at the end of germination test, dried for 24 hours in a 93 greenhouse at  $105 \pm 2$  °C, weighed in precision scale and the results expressed in grams of 94 dry mass of seedlings.

95 The experiment was carried out in a completely randomized design with four replicates of 50 96 seeds, using the factorial arrangement scheme: 2 (unconditioned and conditioned with 97 KNO<sub>3</sub>) x 6 (NaCl concentrations), for salt stress and 2 conditioned and conditioned with 98 KNO<sub>3</sub>) x 5 (osmotic potentials), for water stress. The results were submitted to analysis of 99 variance and, when there was significance, by the F test, submitted to regression analysis. 100 Statistical analyzes were performed using SAS<sup>®</sup> software [19].

101

### 102 3. RESULTS AND DISCUSSION

103

104 According to the analysis of variance, when applying the F test, statistical differences were 105 verified for the variables in which L. leucocephala seeds were submitted, both in relation to 106 the conditioning and concentrations in the saline stress (NaCl) and different potentials 107 osmotic (PEG). Significance was also observed for the interaction between the factors, 108 observing different behaviors for physiological analyzes, except for the germination variable, 109 when subjected to saline stress conditioning and interaction, for which there was no significant effect on the evaluated treatments. When the germination of seeds submitted to 110 water stress was evaluated, no difference was found between the means of the evaluated 111 112 treatments (Table 1).

113

114Table 1. Summary of variance analysis for the variables germination (G), germination115velocity index (GVI), seedling length (SL) and seedling dry mass (SDM) from

116 117 unconditioned seeds and conditioned with potassium nitrate subjected to saline and water stresses.

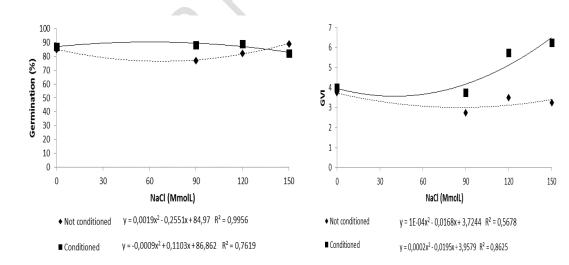
F values – Saline stress							
Source of variation	G (%)	GVI	SL (cm)	SDM (g)			
Conditioning (C)	175,91**	334,67**	387,11**	0,16**			
Saline concentrations (S)	21,10 <sup>ns</sup>	34,08**	4,01**	0,08**			

(C) x (S)	74,83 <sup>ns</sup>	13,73**	25,97**	0,06**			
CV (%)	7,82	14,83	11,83	12,15			
F values – Hydrical stress							
Source of variation	G (%)	GVI	SL (cm)	SDM (g)			
Conditioning (C)	22,94 <sup>ns</sup>	14,88**	14,21**	0,0016**			
Osmotic potentials (O)	26,56 <sup>ns</sup>	51,37**	9,08**	0,0012**			
(C) x (O)	28,02 <sup>ns</sup>	55,43**	120,70**	0,0010**			
CV (%)	7,53	3,90	1,94	7,45			

<sup>ns</sup> not significant, \*\* significant at 1% by the F test.

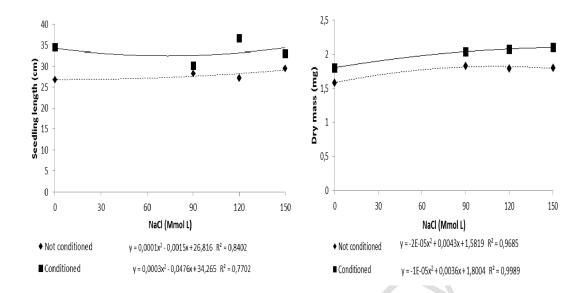
120 The results of the germination test on conditioned or non-saline seeds (Figure 1 A) show that, despite salinity, there was a positive effect of conditioning on this variable. Although the 121 122 seeds did not present significant variation with the progressive increase of the salinity of the 123 substrate, it was observed effect of conditioning on the germination of seeds until the concentration of 90 Mmol/L of NaCl, which presented higher germination than the 124 125 unconditioned seeds. In general, the seeds presented high germination, even when not conditioned. A similar result was verified by Amaro [20], which evaluated Copaifera 126 langsdorffii Desf. (Fabaceae) seeds on a substrate moistened with NaCl solution, with a high 127 percentage of germination, around 95 %. The Leucaena leucocephala (Lam.) De Wit seeds 128 129 Cavalcante and Perez [21] added sodium chloride (NaCl) to substrate and observed high germination percentages of the seeds, which reaffirmed ability of the seeds this species to 130 present a high germinative performance even under saline stress conditions. 131

132



133

<sup>119</sup> 



# Figure 1. (A) Germination, (B) Germination velocity index, (C) Seedling length, and (D) Dry mass of *L. leucocephala* seedlings from not conditioned seeds conditioned with potassium nitrate to saline stress).

138 Thus, L. leucocephala seeds, as well as those of several other species of the Fabaceae 139 family, show resistance or some kind of defense system, capable of synthesizing and 140 accumulating different compounds of small molecular mass such as sugar alcohols, proline 141 and glycine betaine, which are referred to as compatible osmopolymers, osmoprotectants or 142 solutes. And these compounds have an exact function in the plants, being able to be related protection of the plants to abiotic stresses, functioning as a tool for the cellular osmotic 143 144 adjustment, and protection against oxidative damages under adverse conditions, as in 145 drought and salinity tolerance [22].

However, it was verified that potassium nitrate (KNO<sub>3</sub>) did not promote seed germination improvement, since these already present high viability even under conditions of salt stress.

148 It was observed that the rate of germination, length and dry mass of seedlings increased as the substrate concentrations increased, evidencing that the potassium nitrate (KNO<sub>3</sub>) and 149 salt stress L. leucocephala seeds (Figures 1 B, C and D), evidencing that potassium nitrate 150 (KNO<sub>3</sub>) concentrations helped to reverse and/or minimize the negative effects of salinity in 151 152 seed germination velocity index and seedling initial development. The plausible explanation 153 for this phenomenon is the possibility of nitric oxide liberators promoting seedling tolerance 154 to salinity by reducing Na<sup>+</sup> and Cl<sup>-</sup> transport to leaves and by the ability to compartmentalize 155 these ions in vacuoles to avoid their accumulation in the cytoplasm or cell walls and thus 156 avoid salt toxicity [35].

157

134

This is because the mechanism of action of potassium nitrate acts on the reception of electrons, reducing the nitrite form inside the seeds, reoxidizing the NADPH and increasing the availability of NADP for the reduction of the dehydrogenases of the cycle of pentose phosphate, helping the overcoming of seed dormancy [3]. In addition to being an excellent growth promoter of seedlings, potassium nitrate can also act to maintain balance in plant cells, promoting respiration and metabolism of carbohydrates [23].

164

165 Regarding the unconditioned seeds, lower performance was observed in all salinity 166 concentrations when the germination rate, length and dry mass of seedlings were evaluated. 167 As for the variable germination, the vigor of the unconditioned seeds was more affected by 168 the salinity. Evaluating the physiological quality of seeds of Crambe abyssinica Hochst. ex R. 169 E. Fr., Nunes et al. [24] found that the increase in saline concentration of the substrate also 170 reduced the rate of germination. This decrease in germination speed was also observed by 171 Sousa Filho [25] in Erythrina mulungu Mart ex Benth. (Fabaceae) seeds under saline stress. 172 These results confirm the negative effect of soil salinity on the vigor of seeds of different 173 species, reducing the emergence speed and the growth of the seedlings, generating 174 damages to agricultural production.

Salinity probably interferes with the physiological quality of seeds as well, in terms of reservoir energy expenditure to absorb water and subsequently not reserving the reservoir for other processes, inducing changes in the activities of catalase, polyphenoloxidase and peroxidase enzymes [26]. In studies carried out by Marques et. [27], the effect of saline stress on emergence and establishment of seedlings seems to have been responsible for the inhibition of cotyledonary reserve depletion and embryonic axis growth, since in the analyzes the seedlings presented excessive accumulation of Na<sup>+</sup> and Cl<sup>-</sup> ions.

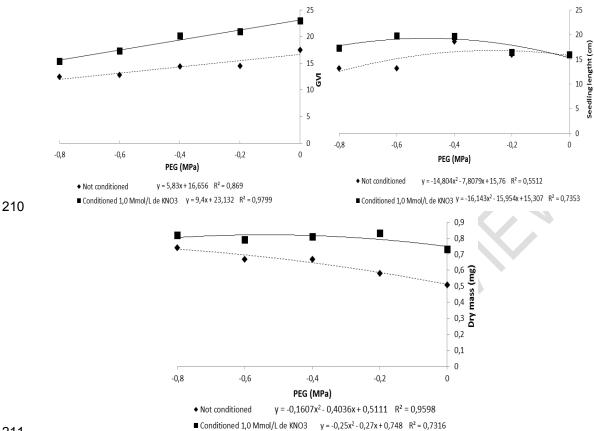
182 The excess Na<sup>+</sup> and Cl<sup>-</sup> ions seem to play a role in reducing the physiological quality of 183 seeds, since they tend to cause a decrease in protoplasmic intumescence (the ions in 184 solution initially cause a decrease in intumescence, and only after absorption and 185 accumulation in the vacuoles and apoplast is that the rate of water absorption is normalized), 186 affecting the enzymatic activity, resulting, mainly, in the inadequate production of energy by 187 disturbances in the respiratory chain [28]. In addition, it is necessary to emphasize its toxic 188 effect, resulting from the concentration of ions in the protoplasm. As the saline concentration 189 increases in the soil solution, there is an increase in the osmotic pressure and, therefore, the 190 plant does not absorb the soil water, causing physiological and morphological disturbances 191 that hinder the survival of the plant in this stress condition [29].

192

193 When seeds conditioned or not subjected to water stress, no change in seed germination 194 was observed along the reduction of water availability and no difference between the two 195 treatments in any of the potentials tested.

196 However, as the water potential became more negative, there was a decrease in the rate of 197 germination of L. leucocephala seeds conditioned and not conditioned with potassium nitrate 198 (KNO<sub>3</sub>), and conditioned seeds presented higher results in relation to unconditioned seeds 199 (Figure 2 A). KNO<sub>3</sub> under water stress showed a significant interaction and showed a 200 positive influence on the evaluations described in L. leucocephala seeds (Figure 2A), since 201 conditioning has the function of repairing damaged macromolecules and cellular structures 202 with the activation of metabolic events in stages I and II of imbibition without, however, 203 occurring root protrusion [36]. The potassium nitrate (KNO<sub>3</sub>) also helped to reverse and/or 204 minimize the negative effects of seed germination and initial seedling development, in 205 addition to being a very tolerant to negative water potentials. Thus, the conditioning 206 technique establishes favorable conditions to increase seed germinability and vigor of 207 seedlings [37], thus contributing to high-quality seedlings even under stress conditions [38]. 208

209



211 212

# Figure 2. Germination velocity index (A), seedling length (B) and dry mass (C) of *L. leucocephala* from seeds not conditioned and conditioned with potassium nitrate (and submitted to water stress).

216

When evaluating the viability of *Apuleia leiocarpa* (Vogel) J.F.Macbr. (Fabaceae) in different water potentials, Spadeto et al. [31] also observed that the more negative the potentials, the lower germination speed averages were -0.4 and -0.8 MPa obtained 3.80 and 3.74, respectively. When Felix et al. [34] evaluated the effects of water stress, verified that the germination speed in *L. leucocephala* seeds were reduced when submitted to the most negative water potentials, -0.3 and -0.6 MPa, obtaining a mean of 8.0 and 4.0, respectively.

Therefore, the results of these researchers corroborate with those obtained in this research, indicating the negative effect of reducing the availability of water in substrate on the germination of seeds this species. This fact is probably due to decrease in water availability, which limits imbibition and reduction of seed water intake, reducing the metabolic activities expected during germination phases, slowing down the process of mobilization and utilization reserves, leading to reduction of the rate germination [5].

It is observed that *L. leucocephala* seeds presented high rate of germination in the conditions in which the lowest water restrictions were observed. According to Bewley et al. [5] Seeds such as *L. leucocephala* are tolerant to water stress due to their natural occurrence in semiarid regions, as they acquire biochemical and physiological strategies that can be attributed to membrane repair even in situations of low water potential.

Conditioning with potassium nitrate (KNO<sub>3</sub>) has influenced the reversal of harmful changes in cell membranes, caused by inactivation of enzymes and inhibition of protein synthesis [29]. This shows that even under conditions of stress of abiotic origin, *L. leucocephala* seeds had positive effects on the physiological responses involved in the whole germination process, regulating percentage of germination and rate germination, as well as the growth, development and establishment of seedlings.

240 There was a tendency to increase the length and dry mass of L. leucocephala seedlings as 241 the water restriction was increased, regardless of the potassium nitrate (KNO<sub>3</sub>) or not. 242 However, the seeds presented different results for length and dry mass of seedlings when 243 compared to conditioned and unconditioned seeds. With the reduction of the water potential, 244 the length of the seedlings resulting from conditioned seeds presented results with a 245 tendency to increase the difference in relation to the unconditioned seeds, indicating an 246 increase in the size conditioned seedlings in relation the untreated seedlings under 247 conditions of lack of water (Figure 2 B). On the other hand, the dry matter of seedlings 248 showed a decrease in the water potential (Figure 2 C), in relation to the uncon solved seeds. 249

The ability to develop roots and aerial parts, even under conditions of stress and water scarcity, varies widely between species and may be related to the hypothesis of adaptation, due to the proliferation and the increase of the roots increase the capacity of water absorption in a limit situation, or may be related to the growth rate of a given plant [32].

Many proteins that are induced in the early stages of water stress are also involved in root morphogenesis and carbon/nitrogen metabolism, which may contribute to the nullification of stress by increasing root growth [33]. In the case of *L. leucocephala*, the literature reports that it is a plant adapted to inhospitable conditions and such rusticity can favor the tolerance of the seeds under different conditions.

259

### 260 **4. CONCLUSION**

261

The use of potassium nitrate (KNO<sub>3</sub>) showed a positive action on the reversal of salt stress
 and the conditioning of the seeds under low osmotic potential, influencing the maintenance
 of the physiological quality of *L. leucocephala.*

### 266 COMPETING INTERESTS

267268 Authors have declared that no competing interests exist.

### 269 270 AUTHORS' CONTRIBUTIONS

271 272

This work was carried out in collaboration with all authors.

# 273274 REFERENCES

- 275
- Nogueira NO, Oliveira OM de, Bernardes CA da SM, Bernardes C de O. Utilização de leguminosas para recuperação de áreas degradadas. Enciclopédia Biosfera. 2012;8(14):2121.
- Cardoso EA, Alves AU, Cavalcante ÍHL, Farias SGG, Santiago FEM. Métodos para superação de dormência em sementes de leucena. Revista Ciências Agrárias. 2012;55(3):220–4. PortugueseAvailable: http://dx.doi.org/10.4322/rca.2012.062
- 3. Mariano LG, Somavilla A, Silveira AG da, Salamoni AT. Análise de superação de dormência de sementes de *Leucaena leucocephala* e desenvolvimento inicial de

284 plântulas. Electronic Journal of Management, Education and Environmental 285 Technology. 2016;20(1):398-404. Portuguese. Available: 286 http://dx.doi.org/10.5902/2236117019719 287 4. Begum MAJ, Selvaraju P, Venudevan B. Saline stress on seed germination. 288 Scientific Research and Essavs. 2013;8(30):1420-3. Available: 289 http://dx.doi.org/10.5897/SRE2013.5600 290 5. Bewley JD, Bradford K, Hilhorst H, Nonogaki H. Seeds: physiology of development, 291 germination and dormancy. 2013. 292 6. Cavallaro V, Barbera AC, Maucieri C, Gimma G, Scalisi C, Patanè C. Evaluation of 293 variability to drought and saline stress through the germination of different ecotypes 294 of carob (Ceratonia siliqua L.) using a hydrotime model. Ecological Engineering. 295 2016;95:557-66. Available: http://dx.doi.org/10.1016/j.ecoleng.2016.06.040 296 7. Elias SG, Copeland LO, McDonald MB, Baalbaki RZ. Seed testing: principles and 297 practices. East Lansing: Michigan State University Press.; 2012. 368 p. Available: 298 http://dx.doi.org/1611860393,9781611860399 299 8. Guedes RS, Alves EU, Viana JS, Goncalves EP, Lima CR de, Santos SDRN dos. 300 Germinação e vigor de sementes de Apeiba tibourbou submetidas ao estresse 301 hídrico e diferentes temperaturas. Ciência Florestal. 2013;23(1). Portuguese. 302 Available: http://dx.doi.org/10.5902/198050988438 303 9. Martins CC, Pereira MRR, Lopes MTG. Germinação de sementes de eucalipto sob 304 estresse hídrico e salino. Bioscience Journal. 2014;30(3):318-29. 305 10. Matakiadis T, Alboresi A, Jikumaru Y, Tatematsu K, Pichon O, Renou J-P, et al. The 306 Arabidopsis Abscisic Acid Catabolic Gene CYP707A2 Plays a Key Role in Nitrate 307 Control of Seed Dormancy. Plant Physiology. 2008;149(2):949-60. Available: 308 http://dx.doi.org/10.1104/pp.108.126938 309 11. Zheng C, Jiang D, Liu F, Dai T, Liu W, Jing Q, et al. Exogenous nitric oxide improves 310 seed germination in wheat against mitochondrial oxidative damage induced by high 311 salinity. Environmental and Experimental Botany. 2009;67(1):222-7. Available: 312 http://dx.doi.org/10.1016/j.envexpbot.2009.05.002 313 13. Pereira BLC, De Lima E Borges EE, Oliveira AC, Leite HG, De Carvalho Gonçalves JF. Influência do óxido nítrico na germinação de sementes de Plathymenia reticulata 314 315 Benth com baixo vigor. Scientia Forestalis. 2010;(88):629-36. Portuguese. 316 14. Köppen W, Geiger R. Klimate der Erde. Gotha: Verlag Justus Perthes. 1928. 317 15. Brasil. Ministério da Agricultura, Pecuária e Abastecimento. Regras para análise de 318 sementes. Brasília, DF: MAPA/ACS; 2009. 399 p. Portuguese. 16. 319 Betoni R. Scalon S de PQ. Mussury RM. Salinidade e temperatura na germinação e 320 vigor de sementes de mutambo (Guazuma Ulmifolia Lam.) (Sterculiaceae). Revista 321 Árvore. 2011;35:605–16. Portuguese. Available: http://dx.doi.org/10.1590/S0100-322 67622011000400004 323 17. Michel BE, Kaufmann MR. The osmotic potential of polyethylene glycol 6000. Plant 324 Physiology. 1973;51(5):914-6. Available: http://dx.doi.org/10.1104/PP.51.5.914 325 Maguire JD. Speed of germination-aid selection and evaluation for seedling 18. 326 Science. 1962;2:176-7. emergence and vigor. Crop Available: 327 http://dx.doi.org/http://dx.doi.org/10.2135/cropsci1962.0011183X000200020033x 328 SAS Institute (2011) SAS/STAT Versão 9.3: User's Guide. SAS Institute Inc., 8621. 19. 329 Amaro MS. Germinação de sementes e mobilização de reservas em plantas de 20. 330 copaibasobr estresse hídrico e salino. Tese (Doutorado em Fitotecnia). Universidade 331 Federal do Ceará: 2012. Portuguese. 332 21. Cavalcante A de MB, Perez SCJG de A. Efeitos dos estresses hídrico e salino sobre 333 a germinação de sementes de Leucaena leucocephala (Lam.) de Wit. Pesquisa 334 Agropecuária Brasileira. 1995;30(2):281–9. Portuguese. 22. Hurtado Salazar A. Tolerância ao estresse abiótico (salinidade e seca) e influência 335 336 de porta-enxertos na qualidade de frutos de Passiflora spp. Tese (Doutorado em

- 337 Fitotecnia). Universidade Federal de Viçosa; 2016. Portuguese.
- Brito SF, Bezerra AME, Pereira DS. Efeito da Temperatura e do KNO<sub>3</sub> na germinação de *Acnistus arborescens* (Solanaceae). Floresta e Ambiente.
  2016;22(3):406–12. Portuguese.
- Nunes A da S, Souza LCF de, Scalon S de PQ, Pagnoncelli J. Nitrato de potássio e retirada do pericarpo na germinação e na avaliação do vigor de sementes de crambe. Semina: Ciências Agrárias. 2015;36(3):1775–82. Portuguese. Available: http://dx.doi.org/10.5433/1679-0359.2015v36n3Supl1p1775
- Sousa Filho PH de, Pereira DP, Marques JB, Moreira ÉFA, Carvalho M, NAVES MF.
  Efeito do estresse salino na germinação de sementes de *Erythrina mulungu* Mart. ex
  Benth. Anais do I Seminário de Pesquisa e Inovação Tecnológica. 2017;1(1).
  Portuguese.
- Maia JM, Ferreira-Silva SL, Voigt EL, Macêdo CEC de, Ponte LFA, Silveira JAG.
  Atividade de enzimas antioxidantes e inibição do crescimento radicular de feijão caupi sob diferentes níveis de salinidade. Acta Botânica Brasilica. Portuguese.
  2012;26(2):342–9. Available: http://dx.doi.org/10.1590/S0102-33062012000200010
- 353 27. Marques EC, Freitas VS, Bezerra MA, Prisco JT, Gomes-Filho E. Efeitos do estresse
  354 salino na germinação, emergência e estabelecimento da plântula de cajueiro anão
  355 precoce. Revista Ciência Agronômica. 2011;42(4):993–9. Portuguese. Available:
  356 http://dx.doi.org/10.1590/S1806-66902011000400023
- 357 28. Larcher W. Ecofisiologia vegetal. Editora Rima. 2000. Portuguese.
- 358 29. Taiz L, Zeiger E. Fisiologia Vegetal. 3rd ed. Editora Artmed. 2013. 918 p.
- 30. Lopes JC, Freitas AR de, Beltrame RA, Venancio LP, Manhone PR, Silva FRN e.
  Germinação e vigor de sementes de pau d'alho sob estresse salino. Pesquisa
  Florestal Brasileira. 2015;35(82):169. Portuguese. Available: http://dx.doi.org/10.4336/2015.pfb.35.82.631
- 363 31. Spadeto C, Mengarda LHG, Paulucio MC, Lopes JC, Matheus MT. Embebição, osmocondicionamento e viabilidade de sementes de *Apuleia leiocarpa* (VOGEL.) J.
  365 F. MACBR. Ciência Florestal. 2018;28(1):80. Portuguese. Available: http://dx.doi.org/10.5902/1980509831582
- 367 32. Forti, VA, Cicero, SM, Pinto, TLF Efeitos de potenciais hídricos do substrato e teores
  368 de água das sementes na germinação de feijão. Revista Brasileira de Sementes.
  369 2009;31(2):63-70. Portuguese. Available: http://dx.doi.org/10.1590/S0101370 31222009000200007
- 371 33. Masetto TE, Quadros J de B, Ribeiro DM, Rezende RKS, Scalon S de PQ. Potencial hídrico do substrato e teor de água das sementes na germinação do crambe. Revista Brasileira de Sementes. Portuguese. 2011;33(3):511–9. Available: http://dx.doi.org/10.1590/S0101-31222011000300014
- 375 34. Felix FC, Araújo FS, Silva MD, Ferrari CS, Pacheco MV. Estresse hídrico e térmico na germinação de sementes de *Leucaena leucocephala* (Lam.) de Wit. Revista Brasileira de Ciências Agrárias. 2018;2(13):e5515. Portuguese. Available: http://dx.doi.org/10.5039/agraria.v13i2a5515
- 37935.Munns R. Comparative physiology of salt and water stress. Plant, Cell and380Environment, 2002;25(1):239-250. Available: 10.1590/0034-737X201663010006
- 38136.Bradford, KJ. Manipulation of seed water relations via osmotic priming to improve382germination under stress conditions. HortScience. 1986;21(5):1105-1112.
- 383 37. Cardoso NSN, Oliveira LM, Fernandez LG, Pelacani CR, Souza CLM, Oliveira ARMF.
   384 Osmocondicionamento na germinação de sementes, crescimento inicial e conteúdo
   385 de pigmentos de *Myracrodruon urundeuva* fr. Allemão. Revista Brasileira de
   386 Biociência. Portuguese. 2012;10(4):457-461.
- 387 38. Santos MCA, Aroucha EMM, Maracajá PB, Souza MS, Souza PA. Condicionamento
   388 osmótico de sementes: revisão de literatura. Revista Caatinga. Portuguese.
   389 2008;21(2):1-6.