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**Original Research Article**  
**Physiological quality in *Leucaena leucocephala***  
**seeds conditioned with potassium nitrate**  
**submitted to saline and water stresses**

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**ABSTRACT**

*Leucaena leucocephala* is a species that occurs in semi-arid regions capable of developing physiological and biochemical strategies under adverse environmental situations, such as saline soils and water deficiency. Even so, there are still losses in germination and initial development of plants, mainly due to abiotic stress effects, making it necessary to implement alternatives which are able to diminish the impact on the seeds with the use of potassium nitrate. The objective of this work was to evaluate the physiological quality in *L. leucocephala* seeds conditioned in potassium nitrate solution and subjected to saline and water stresses. After scarification, the unconditioned leucine seeds were separated and identified, and the conditioners were immersed in the 1.0 Mmol/L solution of potassium nitrate for 24 hours. The treatments were NaCl: 0; 30; 60; 90; 120 and 150 Mmol/L and the water potential: 0,0; -0.2; -0.4; -0.6, and -0.8 MPa. The seeds were distributed over two Germitest<sup>®</sup> sheets, moistened with distilled water 2.5 times the dry weight and incubated in a B.O.D. at 25 °C for 10 days. Then, the germination speed index, seedling length and seedling mass were measured for the normal seedlings. Eleven treatments with four replicates of 50 seeds were used. The design was completely randomized, and the statistical analyzes were performed in SAS. The use of potassium nitrate (KNO<sub>3</sub>) contributed to maintaining the physiological quality of *L. leucocephala* seeds under conditions of saline stress and water restriction.

**Keywords:** *Leucena*; water potential; salinity.

**1. INTRODUCTION**

*Leucaena leucocephala* (Lam.) belongs to the Fabaceae family (Subfamily Mimosidae), is a fast growing specie, nitrogen fixer and has been highlighted as a promising alternative for recovering vegetation and rehabilitating degraded areas, since it has symbiosis with nitrogen-fixing bacteria, improving soil fertility [1].

28 Its use has met a wide variety of demands, including reforestation in disturbed areas for  
29 erosion control, animal feed, green manuring, fence posts, poles and cellulose. Sowing can  
30 be done either by seedlings or directly on the site [2]. However, sowing in saline soils in  
31 regions with a low frequency of rainfall results in slow and irregular emergence, with direct  
32 reflexes on the seedling development [3].

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34 The effects of salinity on germination can be noticed by both interference of salts in the  
35 cellular metabolism and by the reduced osmotic potential of seeds, causing water stress and  
36 making water absorption difficult [4]. Salinity may also cause injury, such as chlorophyll  
37 degradation and changes in protein metabolism, important amino acid levels linked to the  
38 germination process, and initial seedling development [5].

39 The presence of water is essential, and its scarcity is one of the adversities encountered by  
40 species of plants that develop in saline or dry environments, since water availability to the  
41 seed, as well the speed of absorption, is directly influenced by the water potential difference  
42 between seed and soil [6]. Therefore, Fabaceae species seeds, even in arid and semi-arid  
43 regions, may develop physiological and biochemical strategies capable of developing in  
44 adverse environmental situations, such as in soils with high saline concentration or affected  
45 by water deficiency [7,8].

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47 Due to the severity of losses caused by the effects of saline and water stress on germination  
48 and during the initial plant development, alternatives and treatments have been sought to  
49 reduce the impact on seeds. Among the substances able to induce a resistance to these  
50 types of stress in seeds, nitric oxide (NO), inorganic free radical and an extremely versatile  
51 biological messenger can be supplied exogenously to the seeds through their conditioning in  
52 potassium nitrate solutions [9].

53 Research has demonstrated the involvement of nitric oxide in hormonal signaling [10] and in  
54 numerous physiological plant processes, including: protective function against oxidative  
55 stress [11]. In seeds, nitric oxide stimulates germination, both under normal conditions and  
56 under stress, in addition to favoring seed dormancy of some species [12] and promoting the  
57 elongation and formation of adventitious roots [13].

58 The objective of this work was to evaluate physiological quality in *Leucaena leucocephala*  
59 seeds conditioned in potassium nitrate solution and submitted to saline and water stresses.

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## 61 2. MATERIAL AND METHODS

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63 *L. leucocephala* seeds were collected from mature fruits with brown staining from ten  
64 matrices, spaced at a distance of 6 meters, located in the vicinity of the municipality of  
65 Macaíba - RN (Latitude: -5.86, Longitude: -35.33) in the period from July to August 2017.  
66 The region presents an Aw tropical rainy climate, with an average temperature of 26 °C,  
67 relative air humidity of 74 % and precipitation of 1,134 mm annually, according to Köppen  
68 and Geiger [14].

69 After collection, the seeds were manually removed from the pods, selected and mechanically  
70 scarified using No. 100 sandpaper, on the side opposite to the hilum, only to break the  
71 tegument. After scarification, the water content of seeds was determined by the greenhouse  
72 method at  $105 \pm 3$  °C for 24 hours, as recommended by Brazil [15], and the results were  
73 expressed in average percentage of humidity (wet basis).

74 Next, the unconditioned seeds were separated and deposited in identified glass containers.

75 The conditioned seeds were immersed in glass containers containing the 1.0 Mmol/L

76 solution of potassium nitrate for 24 hours. The following concentrations of NaCl were used to  
77 simulate the stress conditions: 0; 30; 60; 90; 120 and 150 Mmol/L [16]; and also the water  
78 potentials: 0; -0.2; -0.4; -0.6 and -0.8 MPa, calculated according to the equation proposed by  
79 Michel and Kaufmann [17]. The seeds had their physiological potential evaluated after  
80 conditioning and under saline and water stress conditions through the tests described below.

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82 For germination, four replicates of 50 seeds were used for each treatment, distributed on two  
83 Germitest® paper towel sheets and covered with a third sheet of the same paper, then rolled  
84 into a roller. The substrate was moistened with distilled water (DW) in an amount 2.5 times  
85 the dry substrate weight, according to Brazil [15].

86 Paper rolls with seeds that were not conditioned and conditioned with potassium nitrate were  
87 subsequently subjected to saline stress and water stress separately and incubated in a  
88 germination chamber (Biochemical Oxygen Demand) at 25 °C for 10 days, when final  
89 seedlings were counted. The results were expressed as a percentage of germination.

90 The germination speed was carried out in conjunction with the germination test, distributed  
91 on two Germitest® paper towel sheets and covered with a third sheet of the same paper,  
92 then rolled into a roller. The substrate was moistened with distilled water (DW) in an amount  
93 2.5 times the dry substrate weight, according to Brazil [15], with daily counts of seeds that  
94 emitted a primary root of at least 2 mm from the first to the tenth day. The germination speed  
95 was determined from the germination velocity index (GVI), calculated according to the  
96 formula proposed by Maguire [18].

97  
98 The dry mass of seedlings was performed with a total of ten normal seedlings per  
99 experimental unit, randomly taken at the end of germination test, dried for 24 hours in a  
100 greenhouse at  $105 \pm 2$  °C, weighed on a precision scale and the results were expressed in  
101 grams of dry mass of seedlings.

102 The experiment was carried out in a completely randomized design with four replicates of 50  
103 seeds using a factorial arrangement scheme: 2 (unconditioned and conditioned with  $\text{KNO}_3$ ) x  
104 6 (NaCl concentrations) for salt stress, and 2 conditioned and conditioned with  $\text{KNO}_3$ ) x 5  
105 (osmotic potentials) for water stress. The results were submitted to analysis of variance, and  
106 when there was significance by the F-test, they were submitted to regression analysis.  
107 Statistical analyzes were performed using SAS® software [19].

### 108 109 **3. RESULTS AND DISCUSSION**

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111 | According to the analysis of variance, when applying the F-test, statistical differences were  
112 verified for the variables in which *L. leucocephala* seeds were submitted, both in relation to  
113 the conditioning and the saline stress (NaCl) concentrations and different osmotic potentials  
114 (PEG). Significance was also observed for the interaction between the factors, observing  
115 | different behaviors for physiological analyzes, except for the germination variable, when  
116 subjected to saline stress conditioning and interaction, for which there was no significant  
117 effect on the evaluated treatments. No difference was found between the means of the  
118 evaluated treatments when the germination of seeds submitted to water stress was  
119 | evaluated (Table 1).

120

121 **Table 1. Summary of variance analysis for the variables of germination (G),**  
122 **germination velocity index (GVI), seedling length (SL) and seedling dry mass (SDM)**  
123 **from unconditioned seeds and conditioned with potassium nitrate subjected to saline**  
124 **and water stresses.**

F values – Saline stress				
Source of variation	G (%)	GVI	SL (cm)	SDM (g)
Conditioning (C)	175.91**	334.67**	387.11**	0.16**
Saline concentrations (S)	21.10 <sup>ns</sup>	34.08**	4.01**	0.08**
(C) x (S)	74.83 <sup>ns</sup>	13.73**	25.97**	0.06**
CV (%)	7.82	14.83	11.83	12.15
F values – Hydrical stress				
Source of variation	G (%)	GVI	SL (cm)	SDM (g)
Conditioning (C)	22.94 <sup>ns</sup>	14.88**	14.21**	0.0016**
Osmotic potentials (O)	26.56 <sup>ns</sup>	51.37**	9.08**	0.0012**
(C) x (O)	28.02 <sup>ns</sup>	55.43**	120.70**	0.0010**
CV (%)	7.53	3.90	1.94	7.45

125 <sup>ns</sup> not significant, \*\* significant at 1% by the F test.

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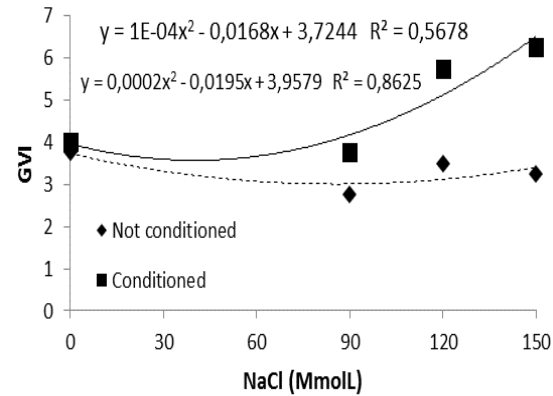
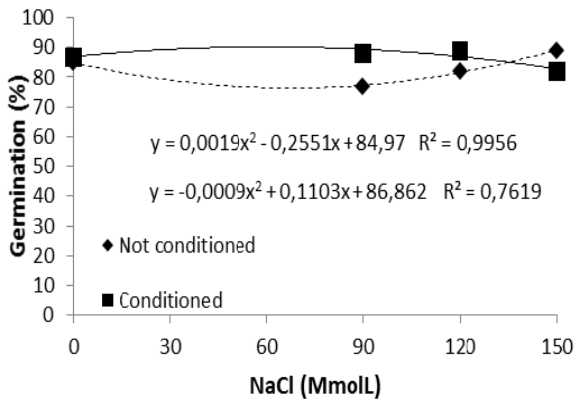
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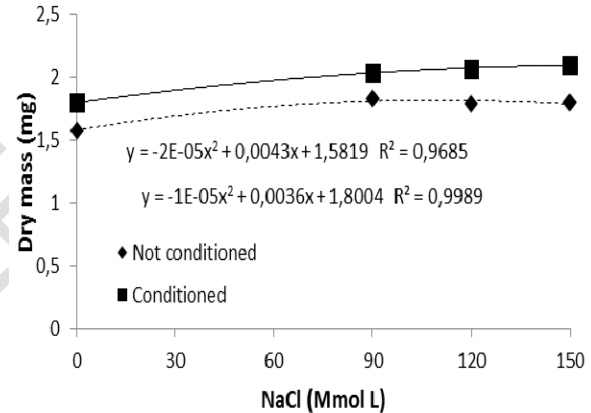
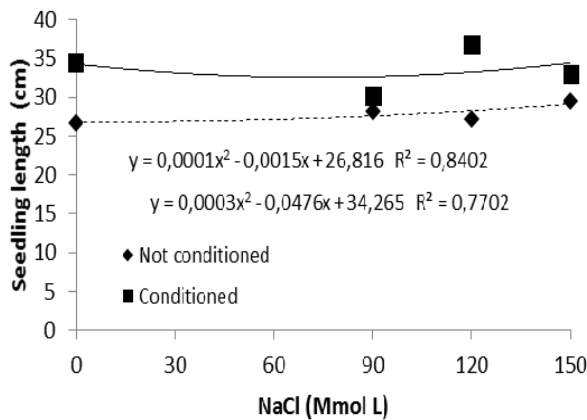
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The results of the germination test on conditioned or non-saline seeds (Figure 1 A) show that, there was a positive effect of conditioning on this variable despite salinity. Although the seeds did not present significant variation with the progressive increase of the salinity of the substrate, a conditioning effect on the germination of seeds was observed until the concentration of 90 Mmol/L of NaCl, which presented higher germination than the unconditioned seeds. In general, the seeds presented high germination, even when not conditioned. A similar result was verified by Amaro [20], who evaluated *Copaifera langsdorffii* Desf. (Fabaceae) seeds on a substrate moistened with NaCl solution, with a high germination percentage of around 95 %. For *Leucaena leucocephala* (Lam.) De Wit seeds, Cavalcante and Perez [21] added sodium chloride (NaCl) to a substrate and observed high germination percentages of the seeds, which reaffirms the ability of the seeds of this species to present high germinative performance even under saline stress conditions.



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140 **Figure 1. (A) Germination, (B) Germination velocity index, (C) Seedling length, and (D)**  
 141 **Dry mass of *L. leucocephala* seedlings from unconditioned seeds and those**  
 142 **conditioned with potassium nitrate (submitted to saline stress).**

143 Thus, *L. leucocephala* seeds, as well as those of several other species of the Fabaceae  
 144 family, show resistance or some kind of defense system which makes them capable of  
 145 synthesizing and accumulating different compounds of small molecular mass such as sugar  
 146 alcohols, proline and glycine betaine, which are referred to as compatible osmopolymers,  
 147 osmoprotectants or solutes. These compounds have an exact function in the plants, being  
 148 able to protect the plants from abiotic stresses, functioning as a tool for the cellular osmotic  
 149 adjustment, and protection against oxidative damage under adverse conditions, such as in  
 150 drought and salinity tolerance [22].

151 However, it was verified that potassium nitrate did not promote seed germination  
 152 improvement, since these seeds already present high viability even under salt stress  
 153 conditions.

154 It was observed that the germination rate, length and dry mass of seedlings increased as the  
155 substrate concentrations increased, evidencing that the potassium nitrate and salt stress in  
156 *L. leucocephala* seeds (Figures 1 B, C and D) and the potassium nitrate concentrations  
157 helped to reverse and/or minimize the negative effects of salinity in seed germination  
158 velocity index and initial seedling development. The plausible explanation for this  
159 phenomenon is the possibility of nitric oxide liberators promoting seedling tolerance to  
160 salinity by reducing Na<sup>+</sup> and Cl<sup>-</sup> transport to leaves and by the ability to compartmentalize  
161 these ions in vacuoles to avoid their accumulation in the cytoplasm or cell walls and thus  
162 avoid salt toxicity [35].

163  
164 This is because the mechanism of action of potassium nitrate acts on the reception of  
165 electrons, reducing the nitrite form inside the seeds, reoxidizing the NADPH and increasing  
166 the availability of NADP which reduces the dehydrogenases in the pentose phosphate cycle,  
167 helping to overcome seed dormancy [3]. In addition to being an excellent growth promoter of  
168 seedlings, potassium nitrate can also act to maintain balance in plant cells, promoting  
169 respiration and metabolism of carbohydrates [23].

170  
171 Regarding the unconditioned seeds, lower performance was observed in all salinity  
172 concentrations when the germination rate, length and dry mass of seedlings were evaluated.  
173 Regarding the variable germination, the vigor of the unconditioned seeds was more affected  
174 by the salinity. In evaluating the physiological quality of *Crambe abyssinica* Hochst. ex R. E.  
175 Fr. seeds, Nunes et al. [24] found that the increase in saline concentration of the substrate  
176 also reduced the germination rate. This decrease in germination speed was also observed  
177 by Sousa Filho [25] in *Erythrina mulungu* Mart ex Benth. (Fabaceae) seeds under saline  
178 stress. These results confirm the negative effect of soil salinity on the vigor of seeds of  
179 different species, reducing the emergence speed and the growth of the seedlings,  
180 generating damage to agricultural production.

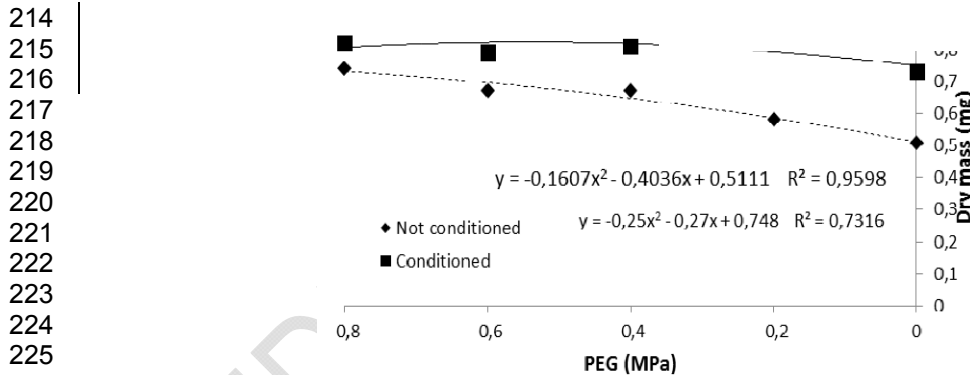
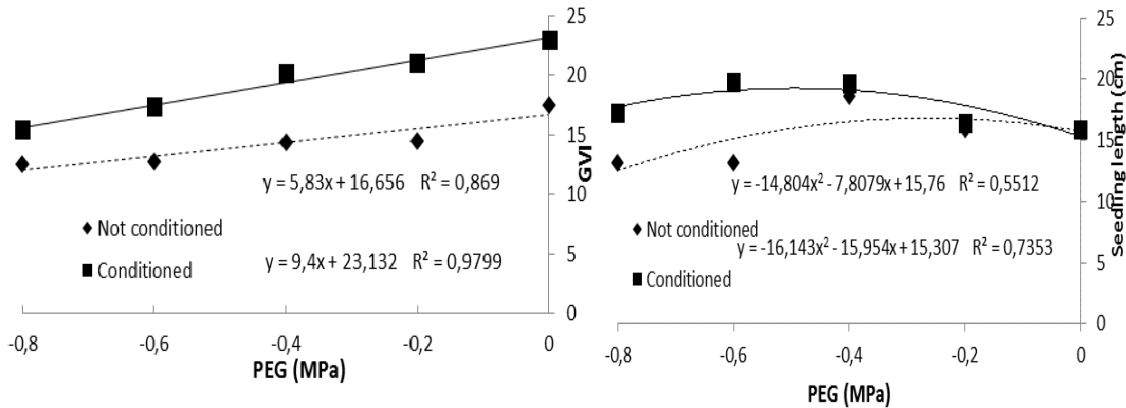
181 Salinity probably interferes with the physiological quality of seeds, and also in terms of  
182 reservoir energy expenditure to absorb water and subsequently does not reserve the  
183 reservoir for other processes, inducing changes in the activities of catalase,  
184 polyphenoloxidase and peroxidase enzymes [26]. In studies carried out by Marques et al.  
185 [27], the effect of saline stress on emergence and establishment of seedlings seems to have  
186 been responsible for inhibiting cotyledonary reserve depletion and embryonic axis growth,  
187 since the seedlings in these analyzes presented excessive accumulation of Na<sup>+</sup> and Cl<sup>-</sup> ions.

188 The excess Na<sup>+</sup> and Cl<sup>-</sup> ions seem to play a role in reducing the physiological quality of  
189 seeds, since they tend to cause a decrease in protoplasmic intumescence (the ions in  
190 solution initially cause a decrease in intumescence, and only after absorption and  
191 accumulation in the vacuoles and apoplast is the rate of water absorption normalized),  
192 affecting the enzymatic activity, mainly resulting in the inadequate production of energy by  
193 disturbances in the respiratory chain [28]. In addition, it is necessary to emphasize its toxic  
194 effect, resulting from the concentration of ions in the protoplasm. As the saline concentration  
195 increases in the soil solution, there is an increase in the osmotic pressure, and therefore the  
196 plant does not absorb the soil water, causing physiological and morphological disturbances  
197 that hinder the plant's survival in this stress condition [29].

198  
199 No changes in seed germination were observed along the reduction of water availability  
200 when seeds were conditioned or not subjected to water stress, and no differences between  
201 the two treatments for any of the potentials were tested.

202 However, as the water potential became more negative, there was a decrease in the  
203 germination rate of *L. leucocephala* seeds conditioned and not conditioned with potassium

204 nitrate (KNO<sub>3</sub>), and conditioned seeds presented higher results in relation to unconditioned  
 205 seeds (Figure 2 A). KNO<sub>3</sub> under water stress showed a significant interaction and showed a  
 206 positive influence on the evaluations described in *L. leucocephala* seeds (Figure 2A).  
 207 Potassium nitrate also helped to reverse and/or minimize the negative effects of seed  
 208 germination and initial seedling development, in addition to being very tolerant to negative  
 209 water potentials. Thus, the conditioning technique establishes favorable conditions to  
 210 increase seed germinability and vigor of seedlings [36], contributing to high-quality seedlings  
 211 even under stress conditions [37].  
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 229 **Figure 2. Germination velocity index (A), seedling length (B) and dry mass (C) of *L.***  
 230 ***leucocephala* from unconditioned seeds and seeds conditioned with potassium nitrate**  
 231 **(and submitted to water stress).**

232  
 233 When evaluating the viability of *Apuleia leiocarpa* (Vogel) J.F.Macbr. (Fabaceae) in different  
 234 water potentials, Spadeto et al. [31] also observed that more negative potentials of -0.4 and -  
 235 0.8 MPa obtained lower germination speed averages of 3.80 and 3.74, respectively. When  
 236 Felix et al. [34] evaluated the effects of water stress, they verified that the germination speed  
 237 in *L. leucocephala* seeds were reduced when submitted to the most negative water  
 238 potentials of -0.3 and -0.6 MPa, obtaining a mean of 8.0 and 4.0, respectively.

239 Therefore, the results of these researchers corroborate with those obtained in this research,  
240 indicating the negative effect of reducing the water availability in substrate on the  
241 germination of seeds for this species. This fact is probably due to a decrease in water  
242 availability, which limits imbibition and reduces seed water intake, reducing the metabolic  
243 activities expected during germination phases, slowing down the process of mobilization and  
244 utilization reserves, leading to a reduction in the germination rate [5].

245 It is observed that *L. leucocephala* seeds presented a high germination rate in the conditions  
246 in which the lowest water restrictions were observed. According to Bewley et al. [5], seeds  
247 such as *L. leucocephala* are tolerant to water stress due to their natural occurrence in semi-  
248 arid regions, as they acquire biochemical and physiological strategies that can be attributed  
249 to membrane repair even in situations of low water potential.

250 Conditioning with potassium nitrate ( $\text{KNO}_3$ ) has influenced the reversal of harmful changes  
251 in cell membranes caused by inactivation of enzymes and inhibition of protein synthesis [29].  
252 This shows that even under stress conditions of abiotic origin, *L. leucocephala* seeds had  
253 positive effects on the physiological responses involved in the whole germination process,  
254 regulating germination percentage and germination rate, as well as the growth, development  
255 and establishment of seedlings.

256 There was a tendency to increase the length and dry mass of *L. leucocephala* seedlings as  
257 the water restriction was increased, regardless of the potassium nitrate ( $\text{KNO}_3$ ) or not.  
258 However, the seeds presented different results for length and dry mass of seedlings when  
259 compared to conditioned and unconditioned seeds. With the reduction of the water potential,  
260 the length of the seedlings resulting from conditioned seeds presented results with a  
261 tendency to increase the difference in relation to the unconditioned seeds, indicating an  
262 increase in the size of conditioned seedlings in relation to the untreated seedlings under  
263 water stress conditions (Figure 2 B). On the other hand, the dry matter of seedlings showed  
264 a decrease in the water potential (Figure 2 C) in relation to the unconditioned seeds.  
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266 | The ability to develop roots and shoots even under conditions of stress and water scarcity,  
267 varies widely between species and may be related to the adaptation hypothesis, due to the  
268 proliferation and increase of the roots which increase the capacity of water absorption in a  
269 limiting situation, or may be related to the growth rate of a given plant [32].

270 Many proteins that are induced in the early stages of water stress are also involved in root  
271 morphogenesis and carbon/nitrogen metabolism, which may contribute to the nullification of  
272 stress by increasing root growth [33]. In the case of *L. leucocephala*, the literature reports  
273 that it is a plant which can adapt to inhospitable conditions, and such rusticity can favor the  
274 tolerance of its seeds under different conditions.



275 **4. CONCLUSION**

276

277 The use of potassium nitrate (KNO<sub>3</sub>) showed a positive action on the reversal of salt stress  
278 and the conditioning of the seeds under low osmotic potential, influencing the maintenance  
279 of the physiological quality of *L. leucocephala*.

280

281 **COMPETING INTERESTS**

282

283 The authors declare they have no competing interests.

284

285 **AUTHOR CONTRIBUTIONS**

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287 This work was carried out in collaboration with all of the authors.

288

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