## **Original Research Article**

# Evaluation of a low-cost camera for agricultural applications

## ABSTRACT

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This study aimed to modify a webcam by replacing its near-infrared (NIR) blocking filter to a low-cost red, green and blue (RGB) filter for obtaining NIR images and to evaluate its performance in two agricultural applications. First, the sensitivity of the webcam to differentiate normalized difference vegetation index (NDVI) levels through five nitrogen (N) doses applied to the Batatais grass (Paspalum notatum Flugge) was verified. Second, images from maize crops were processed using different vegetation indices, and thresholding methods with the aim of determining the best method for segmenting crop canopy from the soil. Results showed that the webcam sensor was capable of detecting the effect of N doses through different NDVI values at 7 and 21 days after N application. In the second application, the use of thresholding methods, such as Otsu, Manual, and Bayes when previously processed by vegetation indices showed satisfactory accuracy (up to 73.3%) in separating the crop canopy from the soil.

9 Keywords: NDVI; Paspalum notatum fluegge; Otsu; segmentation.

## 10 1. INTRODUCTION

Recent developments in sensor technologies have made digital cameras more and more efficient and affordable. These systems have been widely used as a versatile remote sensing tool for many applications due to its advantages over film-based aerial photography and satellite imagery [1]. The main advantage of digital photography lies in simplified image processing [2]. Among the advantages of digital photography from these cameras are its relatively low cost, high spatial resolution and near-real-time availability of imagery for visual assessment and image processing.

18 Digital cameras are fitted with either a charge-coupled device (CCD) sensor or a 19 complementary metal oxide semiconductor (CMOS) sensor that are photoconductive 20 devices. These sensors are sensitive to near-infrared (NIR) wavelengths, however, most of these cameras are fitted with a blocking filter to this wavelength. Thus, typically these 21 22 images present only the red, green, and blue (RGB) bands, which are sufficient to represent colors in the visible portion of the spectrum (400 - 700 nm), as recognized by the human 23 24 vision [3]. In most cases, the digital photographs are recorded in joint photographic experts' 25 group (JPEG) or tagged image file format (TIFF), and the RGB channels are obtained 26 through image processing.

The use of images with RGB and NIR bands is very common in agricultural applications, especially for vegetation monitoring. Many vegetation indices, such as the normalized difference vegetation index (NDVI) [4] require spectral information in the NIR and red bands, even though the RGB bands could be sufficient for some applications [5]. Since most consumer-grade cameras only provide RGB bands, NIR filtering techniques can be used to

32 convert an RGB camera into a NIR camera. Moreover, it is possible to replace the blocking

filter by a long-pass infrared filter on standard CCD or CMOS sensors for obtaining NIR

34 images [6].

35 Over the years, numerous systems for collecting images based on cameras or webcams 36 have been developed and modified to obtain NIR information across multiple domains. Most 37 systems included analysis of the nutritional status of agricultural crops [7], disease detection [8], yield estimation [9], and weed identification [10]. In addition, other authors highlight the 38 39 possibilities of using vegetation indices combined with segmentation techniques and texture 40 analysis for obtaining data of interest, such as crop canopy and soil [11, 12]. Furthermore, 41 these cameras can be mounted in a stationary installation [13] or onboard a light aircraft or unmanned aerial vehicle, a deployment which was made possible due to its low weight [14, 42 43 15].

44 Given the many possibilities of using images from RGB or modified cameras to access the 45 NIR band, the use of artificial vision systems through image processing has enabled the 46 extraction of information of interest, which proves to be a great tool for application in the 47 agricultural environment. However, there are still factors, such as different ambient lighting 48 conditions, plant shading and complex background that are challenges to the success of 49 using low-cost images for agricultural applications as described in other studies [16, 17]. 50 Therefore, in view of the challenge to obtain these images with good quality for solving 51 problems, the present study aimed to modify a webcam to obtaining data from the NIR band 52 and to evaluate its performance over different agricultural applications.

## 53 2. MATERIAL AND METHODS

The experiment was conducted at the Federal University of Viçosa, Viçosa Campus in Minas Gerais, which is located among the coordinates: 20° 45' 14 "(S) and 42° 52' 54" (W), 649 meters above sea level. The image acquisition system comprised two C3 Tech model HB 2105 webcams that produced images in JPEG format (640x480 pixels).

58 In order to obtain NIR images, a modification was carried out in one of the webcams by 59 removing the NIR blocking filter, and adding an RGB blocking filter, which was made from 60 the magnetic material of a floppy disk (common diskette) as proposed by [18]. Thus, the unmodified webcam, named in this study as RGB webcam and the modified NIR webcam 61 62 were tested on two different applications. First, the performance of the webcam's images to 63 differentiate NDVI values according to different N rates was verified. Second, these images 64 were processed for separating the crop canopy from the soil using different thresholding 65 algorithms.

In the first application, a field experiment was carried out using the Batatais grass (*Paspalum notatum* Flugge), where a randomized block design with five treatments and five replications was adopted. Treatments consisted of five nitrogen (N) doses in the form of ammonium sulfate ( $(NH_4)_2SO_4$ ), which corresponded to 0, 40, 80, 120 and 160 kg ha<sup>-1</sup>. Plot dimensions were 1 m × 1 m.

71 Furthermore, the digital images were captured with both webcams at a height of 3 m from

the ground using a ladder, with the webcam being held by one of the authors, and always

range restring that the image taken cover the entire area of the experimental plot. Data acquisition

74 was performed twice with images being captured at 7 and 21 days after the N application. All 75 images were geometrically corrected through the projective transformation technique using the Matlab<sup>®</sup> software, where reference points were defined at the boundaries of each plot.
 Lastly, the NDVI [4] was calculated by Equation 1 for each experimental plot.

(1)

$$NDVI = \frac{nir - r}{nir + r}$$

79 Where: nir: near-infrared band; and r: red band.

In addition, the portable chlorophyll meter (SPAD-502, Konica Minolta Sensing, Tokyo, Japan) was used to measure the SPAD index (SI) [19]. Thus, at the 7 and 21 days after N application, 30 readings per plot were taken, where the average of all readings was considered as a result. In this study, the SPAD-502 readings were assumed to be the reference of chlorophyll content for the purpose of validating the sensitivity of the webcams in detecting the effect of N doses over the Batatais grass.

In order to verify the significance of the proposed treatments, the results were submitted to analysis of variance (ANOVA) through the F-test. Lastly, regression models were adjusted to assess treatment effects on results of the SPAD index readings and NDVI values. All analyses were carried out using the ASSISTAT, version 7.7 free software [20].

In the second application, the RGB images were used for the ability to differentiate crop canopy from soil under different growing conditions. There were 30 images captured for this study and all of it belonged to maize crops at the V4 vegetative stage (four expanded leaves), which were grown under different soil cover conditions, such as conventional planting system, and no-tillage system with coffee husk and straw residue.

95 The digital images were captured at a height of 1.5 m from the ground and then stored as 96 24-bit colour images with resolutions of 640 × 480 pixels saved in RGB colour space in the 97 JPEG format. Then, to discriminate between the object of interest (plant) and background 98 (soil), algorithms were developed using different thresholding methods, such as Otsu [21], 99 Manual threshold selection, and Bayes [22].

Initially, two methods were used to accentuate the green color of plants in RGB images.
 First, in the absolute green method, the pixel color distance (PCD) value was obtained
 through the euclidean distance (ED) calculation using normalized values from the red and
 green bands of each pixel, as shown in Equation 2 [23].

104 
$$PCD = \sqrt{pixel(r^2) + [pixel(g) - 1]^2}$$
(2)

106 Where: r: pixel value from the red band; and g: pixel value from the green band.

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Second, the excess green normalized index (ExG) was obtained as it is shown in Equation 3
 [24].

$$ExG = \frac{2 \times g - r - b}{r + g + b}$$
(3)

110 Where: g: pixel value from the green band; r: pixel value from the red band; and b: pixel 111 value from the blue band.

112 Subsequently, the Otsu, Manual, and Bayes methods were applied to each image. As a 113 result, all images showed some noise, which was removed by using a median filter with a 3 m × 3 m window size. Moreover, the ground truth segmentation model for comparison of
 the three algorithms was developed from the K-means method.

Generally, this method can be employed in different areas including image processing, where it can be used as a thresholding method based on data clustering. This method partitions n pixels into k clusters, where k is an integer value that holds k < n. The k-means algorithm classifies pixels in an image into k number of clusters according to some similarity feature, such as the grey level intensity of pixels, and distance of pixel intensities from centroid pixel intensity [25].

122 The algorithm is based on six steps:

- 123 1. Selection of k clusters (k is a user defined parameter);
- 124 2. Calculation of the number of image pixels N;
- 125 3. Selection of k initial pixel intensity centroids µj;
- 126 4. Calculation of distances  $D_{ij}$  between pixel  $x_i$  and each centroid  $\mu j$  as given in Equation 4. 127
- 128

 $D_{ij} = (x_i - \mu_j)^2$ 

129 Where:  $i = 1 \div N$ ; and  $j = 1 \div k$ .

131 Particular pixel  $\mathbf{x}_i$  is then classified to cluster  $c_i$  to which centroid it has the smallest distance.

- 132 5. Recalculation of centroid positions µ<sub>j</sub> as a mean value of all pixel intensities, which
   133 belong to cluster cj as shown in Equation 5.
- 134

130

(5)

(4)

135 Where: I<sub>i</sub> is the number of pixels that belong to cluster c<sub>i</sub>.

136 6. Steps (4) and (5) are repeated until classification of the image pixels does not change.

137 In this study, the value of k (number of clusters) was defined as two, where the first
represented the crop canopy and second the soil. Then, in order to validate the performance
of each thresholding method, the accuracy index, proposed by [26] was computed using
Equation 6.

141  $Accuracy = 100 \times \frac{A \cap B}{A \cup B}$ (6)

Where: A: represents the set of pixels in the ground truth image that is marked as crop
canopy; and; B: represents the set of pixels in the segmentation that is marked as crop
canopy.

145 This measure of accuracy determines how closely the segmentation matches the ground 146 truth, with 100% indicating an exact match and perfect segmentation. Thus, to verify the 147 significance of the proposed methods, the accuracy means were compared by the Students t 148 test at a 5-% significance level ( $\alpha < 0.05$ ).

- 149 3. RESULTS AND DISCUSSION
- 150 **3.1 Application 1**

151 Average values of the SI and NDVI as a function of the nitrogen doses, as well as its respective coefficient of variation (CV), are shown in Table 1. It can be observed that CV 152 values for NDVI index tended to be higher than to SI values at 7 and 21 days, which may be 153 154 justified by the low uniformity of the Batatais grass on the study area. Furthermore, the fact that SPAD readings are done by direct contact with the leaf surface might have decreased 155 156 its CV. In addition, its higher number of readings per plot also contributes to decrease CV values, which is not done in the NDVI calculation, since only one RGB, and NIR images are 157 158 used per plot to obtain the index.

159	Table 1. Descriptive statistics of the SI (SPAD index) and NDVI (normalized difference
160	vegetation index) at 7 and 21 days after N application.

Time	•	N rates (kg ha <sup>-1</sup> )				
Days	s <u> </u>	40	80	120	160	(%)
		<u> </u>	SI (SPAD-502)			
7	<mark>40.22</mark>	<mark>43.17</mark>	<mark>43.20</mark>	<mark>44.95</mark>	<mark>47.00</mark>	3.67
<mark>21</mark>	<mark>37.95</mark>	<mark>44.92</mark>	<mark>48.12</mark>	<mark>45.82</mark>	<mark>46.95</mark>	6.55
		N	IDVI (webcam)	)		
7	<mark>0.19</mark>	<mark>0.23</mark>	<mark>0.27</mark>	0.31	0.33	<mark>26.4</mark>
<mark>21</mark>	<mark>0.23</mark>	<mark>0.25</mark>	<mark>0.26</mark>	<mark>0.22</mark>	<mark>0.39</mark>	<mark>17.9</mark>
		CV:	Coefficient of va	riation		

162 Even showing sensitivity to the applied N rates, NDVI results from both dates (7 and 21 163 days) were relatively low, which might be associated with low uniformity of the vegetation, 164 and absence of radiometric calibration. [27] highlights that using a reference panel for standardization or the inclusion of a gray Spectralon (or other diffuse reflectors) panel within 165 166 the field of view of the webcam would potentially be of value for calibration under changing 167 illumination conditions (e.g. cloudy vs. sunny days). Thus, a radiometric calibration could increase the sensitivity of the webcam, which would result in higher NDVI values and lower 168 169 weather interference. However, the results obtained here suggest that even without this 170 calibration, the webcam was still capable of detecting differences among treatments.

The regression analyses carried out to access the effect of nitrogen doses on SI and NDVI values at 7 and 21 day after N application showed a linear (7 days) and quadratic (21 days) response for both indices. Moreover, both indices were significant at 1% probability with a coefficient of determination (R<sup>2</sup>) of 0.93 (SI, p-value: 0.0001), and 0.98 (NDVI, p-value: 0.008), respectively. In Figure 1 it is possible to observe the linear increase of the SI and NDVI values as the N doses increases at 7 days after the fertilization.





180 Fig 1. SPAD Index (SI) and NDVI index as a function of topdressing nitrogen doses.

When observing the SI values at 21 days (Figure 1), a linear increase in its values is also observed up to the dose of 80 kg ha<sup>-1</sup> of N. However, from the 120 kg ha<sup>-1</sup> of N, SI values 181 182 showed a decrease, which demonstrates a quadratic response to different N doses with a R<sup>4</sup> 183 of 0.8931 (p-value: 0.0068). Similarly, NDVI values showed a linear increase up to 80 kg ha<sup>-1</sup> 184 of N. Although, when looking at 120 and 160 kg. ha<sup>-1</sup> N doses, NDVI response showed a 185 high variation for both treatments, which resulted in low correlation ( $R^2 = 0.67$ ) (p-value: 186 187 0.0169). This high variation in the NDVI response is possibly associated with the low uniformity of the grass, as well as to changes in weather and illumination conditions, which 188 189 might have influenced the visual guality of the images. Even though there was a high 190 variation in response to these treatments, SI and NDVI values at 21 days were also 191 significant at 1%, and 5% probability, respectively.

192 In general, this quadratic response for both indices at 21 days indicates that, in this range, 193 increasing the nutrient concentration (nitrogen) would not reflect on grass growth, and it 194 represents the plant luxury consumption. According to [28], the luxury consumption is defined as the N storage in the vacuole instead of its participation in the chlorophyll 195 196 molecule. The same authors also point out that, excessive consumption is not always 197 undesirable since it allows plants to accumulate nutrients when its availability is high. In this 198 case, a gradual release is performed by the plant, when the absorption is insufficient to 199 support its growth.

Results obtained in this study showed that the webcam sensor evaluated was capable of detecting the effect of N doses over the Batatais grass for both dates, at 7 and 21 days after N application. The SPAD-502 used here as a reference method presented better results, which was expected due to its higher sensitivity and correlation with the leaf chlorophyll content.

205 Compared to other low-cost, sensor-based methods for monitoring crops phenology, such as 206 radiometric instruments based on LED sensors [29], or light emitting diodes [30], a clear 207 advantage of using webcams is that it can yield images with good spatial resolution. This 208 enables tracking the phenology of different crops by breaking the image into different regions of interest (e.g., crops and weeds) [27]. On the other hand, there is no doubt that higher-209 210 quality spectral imaging could, potentially, be obtained from existing, commercially available 211 multispectral cameras. However, for budget-limited observational and experimental studies, 212 the system proposed here may represent an acceptable compromise, given its low cost and 213 promising performance.

214 3.2 Application 2

215 Initially, performance analyses of segmentation algorithms were based on visual analysis by 216 comparing the proposed methods to the reference binary image. Then, the accuracy index 217 (equation 6) was used for comparing each result with that obtained through the K-means. In 218 general, segmentation methods when combined with the ExG index showed higher accuracy results than those methods preceded by the euclidean distance (ED). Moreover, the highest 219 220 overall mean accuracy (80.3%) was obtained using the Otsu method preceded by ExG index. On the other hand, the lowest accuracy mean was observed using the Manual 221 222 method with the ED index (73.3%).

These results corroborate with [31], which observed that images segmented by the Otsu with the ExG index showed 88% accuracy when compared to other indices using RGB bands. In another study [32], these authors when using the Otsu method preceded by different indices, such as ExG, ExR (excess of red), and another index based on the CIE I\*a\*b color space obtained accuracies of 74%, 77.2%, and 62%, respectively. This demonstrates that the contrast provided by vegetation indices is of great use to highlight the crop canopy from the soil, and could yield in high accuracy segmentation.

When analyzing the accuracy of each image, the highest values were observed for the Manual and Otsu method when preceded by the ED index, which resulted in 95.9% of accuracy for both methods. According to [23], the ED method is based on the search for homology among plants, where after obtaining the spectral energy of plant content; its similarity is verified through the Euclidean distance measurement. Figure 2 shows examples of resulting images from the proposed segmentation algorithms.



- Fig 2. Images processed by the proposed segmentation algorithms. (a) RGB image,
  (b) Euclidean distance, (c) ExG index, (d) K-means, (e) Bayes with ED, (f) Bayes with
  ExG, (g) Manual with ED, (h) Manual with ExG, (i) Otsu with ED, and (j) Otsu with ED.
- 240 In order to determine the most accurate method, the data set was submitted to the Student t-
- test at 5% significance level. Results from the ANOVA showed that statistically, there was no
- 242 difference in performance among the proposed methods when compared to each other.
- Although, the highest CV values were obtained through Bayes (34.72%), and Otsu methods (33.28%), when preceded by the ED index as it is shown in Table 2.

### Table 2. Accuracy results from the proposed segmentation algorithms.

Methods		Accuracy (%)			
_	Max	Min	SD	CV	Mean
Otsu + ED	95.9	32.0	25.65	33.28	77.1
Otsu + ExG	90.9	61.6	9.09	11.33	80.3
Manual + ED	95.9	32.0	23.43	31.99	73.3
Manual ExG	93.5	55.9	13.05	17.12	76.2
Bayes + ED	93.7	22.5	26.15	34.72	75.3
Bayes + ExG	90.9	61.6	16.11	21.19	76.0

246 Max: maximum; Min: minimal; SD: Standard deviation; CV: coefficient of variation. ED: Euclidean 247 distance; ExG: Excess of green

248 These results can be justified by the adverse illumination conditions during the image 249 acquisition period, which resulted in erroneous segmentation due to shaded areas in 250 images. Thus, the Otsu, manual, and Bayes segmentation methods presented satisfactory 251 accuracy (up to 73.3%) for separating crop canopy from the soil when preceded by the ExG 252 and ED indices. Even though a satisfying performance has been achieved, there are still 253 factors, such as the lighting conditions, plant shading and complex background that are 254 challenges to the success of segmentation.

255 Thus, the application of low-cost consumer cameras for process control as an element of 256 precision farming could save fertilizer, pesticides, machine time, and labor force. Although 257 research activities on this topic have increased over the years, high camera prices still reflect 258 on low adaptation to applications in all fields of agriculture. Smart cameras adapted to 259

agricultural applications can overcome this drawback.

#### 260 4. CONCLUSION

The webcam sensor was capable of detecting the effect of nitrogen doses over the Batatais 261 grass through different NDVI values at 7 and 21 days after N application. Regarding the use 262 263 of webcam images in agricultural applications through thresholding methods, it was possible 264 to observe that the segmentation process over RGB images becomes challenging due to 265 non-uniform illumination conditions, and complex image background. Thus, the use of 266 thresholding methods, such as Otsu, Manual, and Bayes when previously processed by the ExG and ED indices can satisfactorily separate the crop canopy from the soil. As a 267 268 recommendation for future studies, both images (NIR and RGB) can be used to calculate 269 vegetation indexes to perform studies on phenology or plant's nutritional status. Also, the RGB images can be processed using segmentation algorithms to quantify plant diseases or 270 271 leaves damaged by pests in crops.

#### 272 **COMPETING INTERESTS**

273 Authors have declared that no competing interests exist.

#### AUTHORS' CONTRIBUTIONS 274

275 This work was carried out in collaboration between all authors. All authors read and 276 approved the final manuscript.

#### REFERENCES 277

278 1. Yang C, Westbrook JK., Suh CPC, Martin DE, Hoffmann WC, Lan Y, & Goolsby JA. 279 2014. An airborne multispectral imaging system based on two consumer-grade

280 281 282		cameras for agricultural remote sensing. Remote Sensing, 6(6), 5257-5278. DOI: https://doi.org/10.3390/rs6065257.
283 284 285 286	2.	Lebourgeois V, Bégué A, Labbé S, Mallavan B, Prévot L, & Roux B. 2008. Can commercial digital cameras be used as multispectral sensors? A crop monitoring test. Sensors, 8(11), 7300-7322. DOI: https://doi.org/10.3390/s8117300.
287 288 289 290 291	3.	Sonnentag O, Hufkens, K, Teshera-Sterne C, Young AM, Friedl M, Braswell BH, Milliman T, O'keefe J, & Richardson AD. 2012. Digital repeat photography for phenological research in forest ecosystems. Agricultural and Forest Meteorology, 152(1), 159–177. DOI: https://doi.org/10.1016/j.agrformet.2011.09.009.
292 293 294 295	4.	Rouse JW, Haas Jr. RH, Schell JA, & Deering DW. 1974. Monitoring vegetation systems in the Great Plains with ERTS, NASA SP-351. Third ERTS-1 Symposium, Vol. 1, pp. 309 – 317, NASA, Washington, DC.
296 297 298 299 300	5.	Nijland W, De Jong R, De Jong SM, Wulder MA, Bater CW, & Coops NC. 2014. Monitoring plant condition and phenology using infrared sensitive consumer grade digital cameras. Agricultural and Forest Meteorology, 184(1), 98-106. DOI: https://doi.org/10.1016/j.agrformet.2013.09.007.
301 302 303 304 305	6.	Rabatel G, Gorretta N, & Labbé N. 2014. Getting simultaneous red and near- infrared band data from a single digital camera for plant monitoring applications: Theoretical and practical study. Biosystems Engineering, 117(1), 2–14. DOI: https://doi.org/10.1016/j.biosystemseng.2013.06.008.
306 307 308 309	7.	Jia B, He H, Ma F, Diao M, Jiang G, Zheng Z, Cui J, & Fan H. 2014. Use of a digital camera to monitor the growth and nitrogen status of cotton. The Scientific World Journal, 2014(1), 1-12. DOI: http://sci-hub.tw/10.1155/2014/602647.
310 311 312 313	8.	Castro A I, Ehsani R, Ploetz RC, Crane JH, & Buchanon S. 2015. Detection of laurel wilt disease in avocado using low altitude aerial imaging. PloS one, 10(4), 1-13. DOI: https://doi.org/10.1371/journal.pone.0124642.
314 315 316 317 318 319	9.	Stroppiana D, Migliazzi M, Chiarabini V, Crema A, Musanti M, Franchino C, & Villa P. 2015. Rice yield estimation using multispectral data from UAV: A preliminary experiment in northern Italy. In <i>Geoscience and Remote Sensing Symposium (IGARSS), IEEE International</i> (pp. 4664-4667). DOI: https://doi.org/10.1109/IGARSS.2015.7326869.
320 321 322 323 324	10.	Romeo J, Guerrero JM, Montalvo M, Emmi L, Guijarro M, Gonzalez-De-Santos P, & Pajares G. 2013. Camera sensor arrangement for crop/weed detection accuracy in agronomic images. Sensors, 13(4), 4348-4366. DOI: http://sci-hub.tw/10.3390%2Fs130404348.
325 326 327 328 329	11.	Montalvo M, Guerrero JM, Romeo J, Emmi L, Guijarro M, & Pajares G. 2013. Automatic expert system for weeds/crops identification in images from maize fields. Expert Systems with Applications, 40(1), 75-82. DOI: https://doi.org/10.1016/j.eswa.2012.07.034.
330 331	12.	Torres-Sánchez J, López-Granados F, & Peña M. 2015. An automatic object- based method for optimal thresholding in UAV images: Application for vegetation

12. Torres-Sánchez J, López-Granados F, & Peña M. 2015. An automatic object-based method for optimal thresholding in UAV images: Application for vegetation

332 333		detection in herbaceous crops. Computers and Electronics in Agriculture, 114(6), 43-52. DOI: https://doi.org/10.1016/j.compag.2015.03.019.	
334 335 336 337 338 339	13.	Sakamoto T, Shibayama M, Kimura A, & Takada E. 2011. Assessment of digital camera-derived vegetation indices in quantitative monitoring of seasonal rice growth. ISPRS Journal of Photogrammetry and Remote Sensing, 66(6), 872-882. DOI: https://doi.org/10.1016/j.isprsjprs.2011.08.005.	
340 341 342 343 344 345	14.	Caturegli L, Corniglia M, Gaetani M, Grossi N, Magni S, Migliazzi M, Angelini L, Mazzoncini, M, Silvestri N, Fontanelli M, Raffaelli M, Peruzzi A, & Volterrani M. 2016. Unmanned aerial vehicle to estimate nitrogen status of turfgrasses. PloS one, 11(6), 1-13. DOI: https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0158268.	
346 347 348 349 350	15.	Levin N, Ben-Dor E, & Singer A. 2005. A digital camera as a tool to measure colour indices and related properties of sandy soils in semiarid environments. International Journal of Remote Sensing, 26(24), 5475–5492. DOI: https://doi.org/10.1080/01431160500099444.	
351 352 353 354	<mark>16.</mark>	Vesali F, Omid M, Kaleita A, & Mobli H. 2015. Development of an android app toestimate chlorophyll content of corn leaves based on contact imaging. ComputersandElectronicsinAgriculture, 116,211-220.DOI:https://doi.org/10.1016/j.compag.2015.06.012.	
355 356 357 358 359 360	17.	Aureliano Netto AF, Martins RN, Aquino de Souza GS, Araujo GDM, Hatum de Almeida SL, & Capelini VA. 2018. Segmentation of RGB images using different vegetation indices and thresholding methods. NATIVA, 6 (4), 389-394. DOI: DOI: http://dx.doi.org/10.31413/nativa.v6i4.5405	
361 362 363 364 365	18.	Micha DN, Penello G, Kawabata RMS, & Camarott T. 2011. Vendo o invisível. Experimentos de visualização do infravermelho feitos com materiais simples e de baixo custo. Revista Brasileira de Ensino de Física, 33(1), 1501,2011. DOI: http://sci-hub.tw/10.1590/S1806-11172011000100015.	Field Code Changed
366 367 368	<mark>19.</mark>	Minolta Camera Co. Ltd. Chlorophyll meter SPAD-502 Instructional Manual. Minolta, Osaka, Japan, 1989. 22p.	Termatted: Portuguese (Brazil)
369 370 371 372	20.	Silva FAS, & Azevedo CAV. 2016. The Assistat Software Version 7.7 and its use in the analysis of experimental data. African Journal of Agricultural Research, 11(39), 3733-3740. DOI: https://doi.org/10.5897/AJAR2016.11522,	<b>Formatted:</b> English (United States)
373 374 375 376	21	Otsu N. 1979. A threshold selection method from gray-level histogram. IEEE Transactions on System Man Cybernetics, 9(1), 62-66. DOI: https://doi.org/10.1109/TSMC.1979.4310076.	
377 378 379 380	22	Gonzales RC, & Woods RE. 1992. Digital image processing (vol.2). Addison- Wesley Publishing Company.	

- 381 23. Nejati H, Azimifar Z, & Zamani M. 2008. Using fast fourier transform for weed detection in corn fields. In Systems, Man and Cybernetics. IEEE 382 383 International Conference on (pp. 1215-1219). DOI: 384 https://doi.org/10.1109/ICSMC.2008.4811448.
- 386 24. Woebbecke DM, Meyer G.E., Von Garden K, Mortensen DA. 1995. Color indices for weed identification under various soil, residue and lighting 388 conditions. Transactions of the ASAE 38, 259-269. DOI: http://sci-389 hub.tw/10.13031/2013.27838.
  - 25. Dass R, Priyanka, Devi S. 2012. Image Segmentation Techniques. International Journal of Electronics & Communication Technology, 3(1), 1-5.
  - 26. Coy A, Rankine D, Taylor M, Nielsen DC, & Cohen J. 2016. Increasing the accuracy and automation of fractional vegetation cover estimation from diaital photographs. Remote Sensing, 8(7), 474-488. DOI: https://doi.org/10.3390/rs8070474.
  - 27. Petach AR, Toomey M, Aubrecht DM, & Richardson AD. 2014. Monitoring vegetation phenology using an infrared-enabled security camera. Agricultural 143-151. and Forest Meteorology, 195(9), DOI: https://doi.org/10.1016/j.agrformet.2014.05.008.
- 28. Baesso MM, de Carvalho Pinto FDA, de Queiroz D, Santos NT, & de Souza 404 Carneiro JE. 2013. Avaliação da deficiência de nitrogênio no feijoeiro 405 usando um medidor portátil de clorofila. Engenharia na Agricultura, 21(2), 406 407 122-128. DOI: https://doi.org/10.13083/reveng.v21i2.318.
- 29. Ryu Y, Lee G, Jeon S, Song Y, & Kimm H. 2014. Monitoring multi-layer 409 canopy spring phenology of temperate deciduous and evergreen forests 410 using low-cost spectral sensors. Remote Sensing of Environment, 149(6), 411 227-238. DOI: http://sci-hub.tw/10.1016%2Fj.rse.2014.04.015. 412
- 413 414 30. Ryu Y, Baldocchi DD, Verfaillie J, Ma S, Falk M, Ruiz-Mercado I, Hehn T, & 415 Sonnentag O, 2010. Testing the performance of a novel spectral reflectance sensor, built with light emitting diodes (LEDs), to monitor ecosystem 416 metabolism, structure and function. Agricultural and Forest Meteorology, 417 150(12), 1597–1606. DOI: https://doi.org/10.1016/j.agrformet.2010.08.009. 418
- 31. Hamuda E, Glavin M, & Jones E. 2016. A survey of image processing 420 techniques for plant extraction and segmentation in the field. Computers and 421 422 Electronics Agriculture, 125(7), 184-199. DOI: in 423 https://doi.org/10.1016/j.compag.2016.04.024.
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402 403

425 32. Bai X, Cao Z, Wang Y, Yu Z, Hu Z, Zhang X, & Li C. 2014. Vegetation segmentation robust to illumination variations based on clustering and 426 morphology modelling. Biosystems engineering, 125(9), 80-97. DOI: 427 https://doi.org/10.1016/j.biosystemseng.2014.06.015. 428