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# **Content of photosynthetic pigments and leaf gas exchanges of young coffee plants under light restriction and treated with paclobutrazol**

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17 **ABSTRACT**  
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The use of shading and paclobutrazol in coffee plants can be an important cultivation strategy to mitigate the negative effects of high solar radiation and atmospheric temperature. Therefore, the levels of photosynthetic pigments and foliar gas exchanges of young coffee plants submitted to doses of paclobutrazol were evaluated, in environments with artificial light restriction. Five experiments were performed: one in full sunlight and four in artificially shaded environments with black polyethylene meshes at 20%, 40%, 60% and 80% levels of light restriction. In each of these environments, an experiment was carried out, consisting of five treatments, defined by the application of paclobutrazol via substrate, at doses of 0, 10, 20, 30 and 40 mg of active ingredient per plant. Joint analysis of experiments and analysis of variance of the regression were made, for the study of levels of shading and doses of paclobutrazol. The light restriction optimized the photosynthetic apparatus of the plants, mainly at levels close to 60%, and considerably favored leaf gas exchanges of arabica coffee. The application of paclobutrazol in the studied dosages resulted in little or no effect on photosynthetic pigment contents and did not influence leaf gas exchanges of coffee plants.

19  
20 *Keywords: Coffea arabica L., shading, triazole, physiological changes.*  
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23 **1. INTRODUCTION**  
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25 In several farming regions of Brazil, cultivated plants are constantly exposed to climatic  
26 adversities that limit their initial establishment in the field, negatively reflecting the yield  
27 potential. Among these, intense solar radiation, high temperatures, and low rainfall are  
28 limiting factors.

29 When plants are exposed to light energy higher than that required by photosynthesis, there  
30 may be energy imbalance that results in photoinhibition. This may promote the biosynthesis  
31 of reactive oxygen species and, consequently, cause oxidative stress. Photoinhibition may  
32 also be a result of photophysical parameters, which include response to light intensity or  
33 wavelength [1-2].

34 Coffee is a native species of understory regions and therefore is considered as a shade  
35 plant [3], with low point of light saturation. Thus, shading of coffee plantations may be an  
36 alternative cultivation method to mitigate negative effects of direct exposure to the sun, in  
37 order to favor the initial establishment of the crop and optimize its development in  
38 subsequent stages.

39 In general, coffee tree presents low rates of assimilation of CO<sub>2</sub> (A) when compared to other  
40 tropical trees. Shading may favor certain environmental factors, such as temperature  
41 attenuation and reduction of water vapor pressure deficit, in order to benefit the gas  
42 exchange of coffee plants [4-5].

43 However, existing information on the effects of shading on gas exchange of coffee plants is  
44 contrasting and depends on the level of light restriction [6-7].

45 In general, comparison to full sunlight exposure, leaves under shading contain greater  
46 amount of chlorophyll per reaction center, more developed antenna complexes, smaller ratio  
47 between chlorophyll a and b, and lower content of carotenoids [8-9].

48 Plant growth regulators, especially inhibitors of biosynthesis of gibberellins, have been  
49 applied to the traditional management of high technological standards of crops. These  
50 products can influence various aspects of plant metabolism, both morphologically and  
51 physiologically, reducing susceptibility to biotic and abiotic stresses [10]. This capacity of  
52 modulation provided to the plants has substantial importance in face of the climatic  
53 adversities verified in cropping environments.

54 Paclobutrazol (PBZ) [(2*RS*, 3*RS*)-1-(4-chlorophenyl)-4,4-dimethyl-2-(1,2,4-triazol-1-yl)-  
55 pentan-3-ol] is triazole capable of inhibiting cytochrome P450 dependent mono-oxygenases  
56 and, consequently, biosynthesis of gibberellins [11]. The changes in plant hormonal balance  
57 caused by triazole, such as elevated levels of cytokinins and abscisic acid, can interfere with  
58 foliar gas exchange and photosynthetic pigment content [12-15].

59 Several studies have demonstrated the ability of paclobutrazol to mitigate the damage  
60 caused by abiotic stresses [16-21], including high temperature stress [11, 21].

61 The effects of this growth regulator have variations according to dosage, phenological stage,  
62 and form of application [22]. For coffee plants, however, the knowledge about such  
63 technology is still incipient, and there is a need for systematic and analytical studies on the  
64 subject.

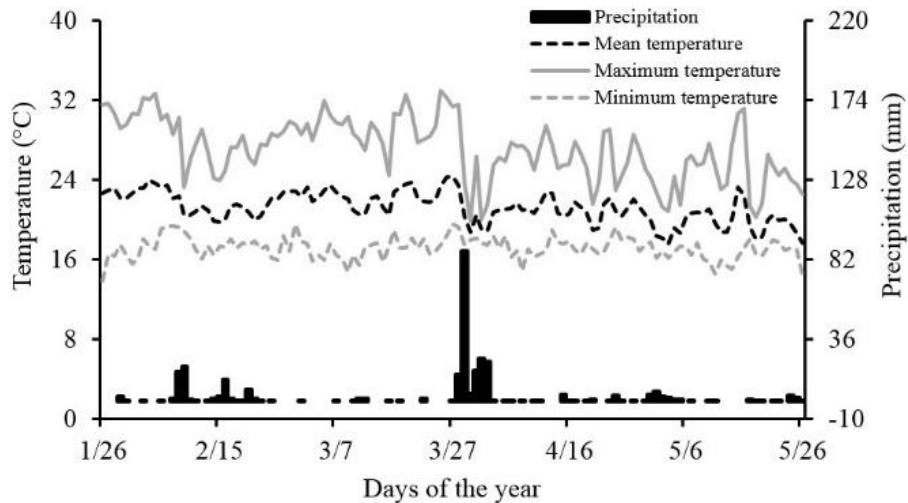
65 Therefore, the use of shading and the application of paclobutrazol in coffee plants is an  
66 important strategy to minimize negative factors related to high solar radiation index and  
67 elevation of atmospheric temperature. The objective of this study was to evaluate the levels  
68 of photosynthetic pigments and foliar gas exchange of young coffee plants submitted to  
69 doses of paclobutrazol in environments with artificial light restriction.

70

## 71 **2. MATERIAL AND METHODS**

72

73 The experiments were conducted at the Universidade Estadual do Sudoeste da Bahia,  
74 Vitória da Conquista *Campus*, Brazil, between January and May 2017. The experimental  
75 area is located at 14° 53' 05" S and 40° 48' 00" W, at 852 meters of altitude. The climate of  
76 the municipality, according to Köppen-Geiger climatic classification, is of Cwa (tropical of  
77 altitude) type, with mean annual temperature of 20.2°C and a mean annual precipitation of  
78 733.9 mm [23]. The meteorological data obtained during the period of tests can be observed  
79 in Figure 1.



80

81 **Fig. 1. Meteorological data recorded in the automatic meteorological station of the**  
 82 **Universidade Estadual do Sudoeste da Bahia, Brazil, during the experimental period**  
 83 **(INMET).**

84 *Coffea arabica* L. 'Catuaí Red IAC 144' seedlings were obtained in an accredited nursery.  
 85 When they had four pairs of mature leaves (approximately five months old), they were  
 86 individually transplanted to pots with a capacity of 20 dm<sup>3</sup> (32.5 cm high x 34.5 cm higher  
 87 diameter and 22 cm lower diameter).

88 Containers were filled with mix of soil (typical Eutrophic YELLOW LATOSOLO) and humus,  
 89 in the ratio 9: 1, and homogenized through sieve of 5 mm. The chemical analysis of the soil  
 90 used in the mixture showed the following results: pH (H<sub>2</sub>O): 5.4; P: 2.0 mg dm<sup>-3</sup>; K<sup>+</sup>: 0.23  
 91 cmol<sub>c</sub> dm<sup>-3</sup>; Ca<sup>2+</sup>: 2.2 cmol dm<sup>-3</sup>; Mg<sup>2+</sup>: 0.8 cmol<sub>c</sub> dm<sup>-3</sup>; Al<sup>3+</sup>: 0.1 cmol<sub>c</sub> dm<sup>-3</sup>; H<sup>+</sup>: 2.7 cmol<sub>c</sub> dm<sup>-3</sup>.  
 92 Liming and fertilization of the substrate were carried out based on soil chemical analysis,  
 93 and according to the technical recommendation of the Soil Fertility Commission of the State  
 94 of Minas Gerais [24].

95 Immediately after transplanting, the pots were placed in different environments, with 0% (full  
 96 sun), 20%, 40%, 60% and 80% of artificial light restriction. Shaded environments (4 meters  
 97 wide x 8 meters long x 2 meters high) were obtained through black polyethylene meshes. In  
 98 each environment (shaded and in full sun) an experiment was conducted, totaling five  
 99 experiments.

100 Each experiment (0%, 20%, 40%, 60% and 80% of light restriction) consisted of five  
 101 treatments, defined by the application of different doses of paclobutrazol via substrate (0, 10,  
 102 20, 30 and 40 mg of active ingredient per plant). A completely randomized design was used,  
 103 with four replications, totaling 20 plots. Each experimental unit consisted of a pot containing  
 104 a coffee plant. For analysis of leaf gas exchanges, readings were made in blocks, with four  
 105 replications, due to variations occurred during the evaluation period, from 8:00 a.m. to 12:00  
 106 p.m.

107 Applications of paclobutrazol were carried out at 18 days after transplanting of seedlings,  
 108 with the commercial product Cultar 250 SC® (250 g i.a. L<sup>-1</sup> of paclobutrazol), and volume of  
 109 solution of 200 mL per plant, applied directly to the substrate.

110 Management of weeds and pests was performed according to the occurrence along the  
111 experiment conduction. All plants were irrigated every two days, with water volume  
112 determined by the gravimetric method (a control pot for each experiment), in which these  
113 containers were saturated with water, with subsequent gravimetric drainage until constant  
114 weight. First, plant pot of each experiment was weighed to obtain the initial mass (IM). Every  
115 two days, the control pots were weighed again, obtaining the final mass (FM). The volume of  
116 water (V) to be applied at the date of each water replenishment, in liters, was determined by  
117 the difference between the two masses, through the equation:  $V = IM - FM$ , with masses  
118 being expressed in kilograms.

119 At 100 days after application (DAA) of paclobutrazol, SPAD (Soil Plant Analysis  
120 Development) index and photosynthetic pigment content were evaluated. The intensity of  
121 green color of leaf (SPAD index) was determined using a portable chlorophyllometer (SPAD  
122 502, MINOLTA, Japan), with readings at three points of the first fully expanded leaf, from the  
123 apex of the plant, and then the average.

124 The extraction of photosynthetic pigments was performed according to the modified  
125 methodology of [25], by eliminating the stages of maceration and centrifugation of the discs,  
126 described by [26]. The first fully expanded leaf of each plant was collected, from which 10  
127 leaf discs of six millimeters of diameter were removed, with the aid of manual leaf disc  
128 extractor. The material was immediately weighed on analytical balance and filled into  
129 aluminum-coated test tubes containing 20 mL of 80% acetone (v/v). This procedure was  
130 performed in an environment without direct incidence of light. The tubes were then capped,  
131 sealed with plastic film, and kept in the dark for 48 hours to extract the pigments.

132 After this period, absorbance readings of the extracts were performed in spectrophotometer  
133 (700 Plus, Femto, Brazil), at wavelengths of 663 nm, 646 nm and 470 nm. For the calibration  
134 of the spectrophotometer, 80% acetone (v/v) was used as "blank". Concentrations ( $\mu\text{g mL}^{-1}$   
135 of extract) of *a*, *b*, and total chlorophyll, and carotenoids were calculated using specific  
136 equations for each pigment [27]. Depending on the mass of each sample and the volume of  
137 acetone used, the values were converted and the pigment content expressed as  $\text{mg g}^{-1}$  of  
138 fresh leaf matter.

139 At 99 DAA of paclobutrazol, leaf gas exchanges were evaluated. These evaluations were  
140 performed on the same leaf used for the other physiological analyzes, using an infrared gas  
141 analyzer (IRGA), LCPro, ADC, UK coupled to an actinic light source of  $1000 \mu\text{mol photons}$   
142  $\text{m}^{-2} \text{s}^{-1}$  of photosynthetically active radiation.

143 Rate of  $\text{CO}_2$  assimilation ( $A$ ,  $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ), transpiration rate ( $E$ ,  $\text{mmol water vapor m}^{-2} \text{ s}^{-1}$ ),  
144 stomatal conductance ( $g_s$ ,  $\text{mol m}^{-2} \text{ s}^{-1}$ ), and the internal  $\text{CO}_2$  concentration in the leaf ( $C_i$ ,  
145  $\mu\text{mol CO}_2 \text{ mol}^{-1} \text{ air}$ ). Carboxylation efficiency ( $A/C_i$ ) was calculated by the ratio of  $\text{CO}_2$   
146 assimilation rate to internal  $\text{CO}_2$  concentration in the leaf.

147 Data were submitted to normality tests (Lilliefors) and homogeneity of variances (Cochran).  
148 After the analysis of variance of each experiment (each level of light restriction) was carried  
149 out, joint analysis of experiments was performed, respecting for each variable relation  
150 between mean squares of residue less than or equal to 1:7, according to [28]. When joint  
151 analysis presented significance ( $p < 0.05$ ), regression analysis was performed for the study  
152 of paclobutrazol doses and levels of shading. The regression models were defined based on  
153 the significance ( $p < 0.05$ ), the highest coefficient of determination ( $R^2$ ) and the biological  
154 response for each characteristic studied. For statistical analysis, the program Statistical and  
155 Genetic Analysis System (SAEG), version 9.1 was used.  
156

157 **3. RESULTS AND DISCUSSION**

158

159 Light restriction was the factor with the greatest impact on the variables related to  
 160 photosynthetic pigments (content of chlorophyll *a*, *b*, and total, carotenoid content, and  
 161 chlorophyll *a:b* ratio), and intensity of green color in the leaf. For chlorophyll *a* and total  
 162 content, there was interaction between the studied factors (levels of light restriction and  
 163 doses of paclobutrazol). The environment with 60% of light restriction was not grouped for  
 164 the analysis of the parameters chlorophyll *b* content and carotenoid content (Table 1).

165 For the unfolding of interaction between the factors, a cubic model for the relationship  
 166 between chlorophyll *a* content and light restriction levels (LR) in coffee plants treated with 0  
 167 and 30 mg of paclobutrazol (PBZ) was delineated. For the coffee plants submitted to 10, 20  
 168 and 40 mg of the regulator, a linear model was established increasing as a function of levels  
 169 of shading (Figure 2A).

170 Chlorophyll *a* content of plants not treated with PBZ (0 mg) was lower than treatment in full  
 171 sun at levels below 33.6% of LR. From this level, the values were higher than the control,  
 172 with an estimated maximum content of 2.34 mg g<sup>-1</sup> of chlorophyll *a* (65.7% LR). The  
 173 maximum levels of chlorophyll *a* estimated for the treatments with 10, 20 and 40 mg of PBZ  
 174 (2.45, 2.43 and 2.34 mg g<sup>-1</sup>, respectively), remained close to the estimated maximum value  
 175 for coffee plants without regulator application.

176 **Table 1. Analysis of variance summary and coefficients of variation (CV) of leaf**  
 177 **greening (SPAD), chlorophyll *a* content (Chl *a*), total chlorophyll content (Chl *a+b*),**  
 178 **ratio of chlorophyll *a* to *b* (Chl *a:b*), chlorophyll *b* content (Chl *b*) and carotenoid**  
 179 **content (Car) of *Coffea arabica* L. ‘Catuaí Vermelho IAC 144’ plants submitted to**  
 180 **different light restriction levels (LR) and paclobutrazol doses (D), evaluated at 100**  
 181 **days after the application of the regulator. Vitória da Conquista - BA, 2017.**

SV	df	MEAN SQUARES				df	Chl <i>b</i>	Car
		SPAD	Chl <i>a</i>	Chl <i>a+b</i>	Chl <i>a:b</i>			
LR	4	391.1**	3.2**	5.9**	4.0*	3	0.2915**	0.042*
D	4	97.5 <sup>ns</sup>	0.2 <sup>ns</sup>	0.5 <sup>ns</sup>	0.3 <sup>ns</sup>	4	0.0008 <sup>ns</sup>	0.003 <sup>ns</sup>
LR*D	16	63.7 <sup>ns</sup>	0.3*	0.6**	1.0 <sup>ns</sup>	12	0.0137 <sup>ns</sup>	0.011 <sup>ns</sup>
Wn	75	66.1	0.1	0.3	0.7	60	0.0085	0.011
CV (%)		12.3	18.5	20.5	17.6		23.5	25.5

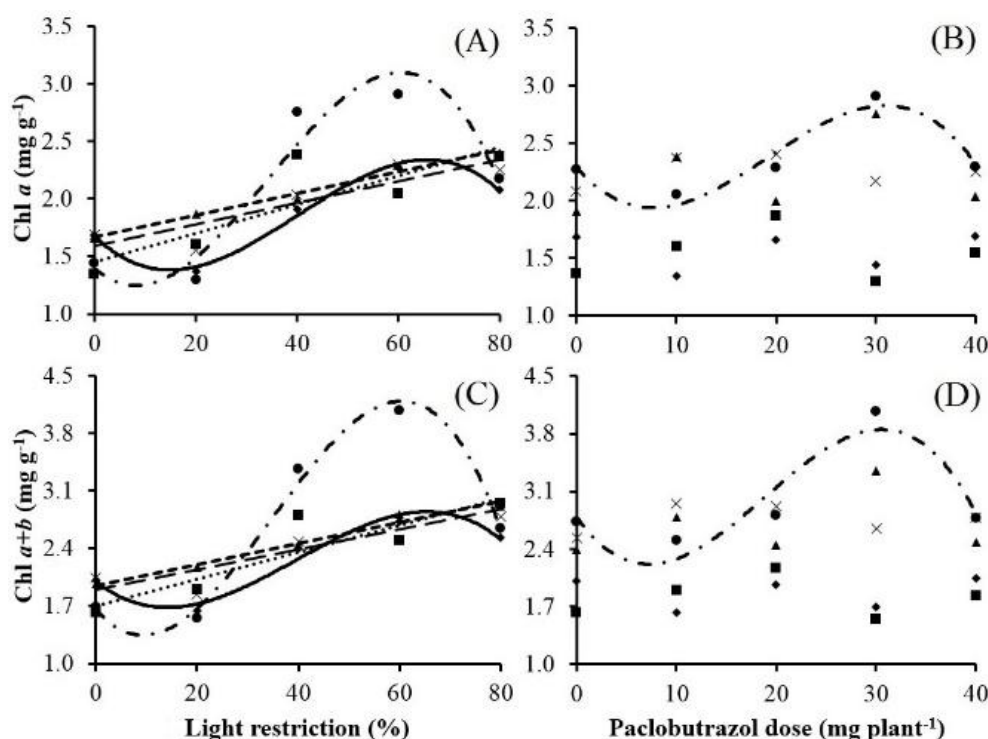
182 <sup>ns</sup>, \* e \*\*: non-significant, significative by “F” test at 5% and 1% of probability, respectively.

183 On the other hand, in coffee plants treated with 30 mg of PBZ, the effect of shading on  
 184 increasing chlorophyll *a* content was potentiated. There was an expressive increase in the

185 content of this pigment promoted by shading at levels above 17.2%, with an estimated  
 186 maximum value of 3.09 mg g<sup>-1</sup> of chlorophyll a (121.75% higher than the full sun treatment),  
 187 at the level of 60.4% of LR.

188 It was not possible to delineate a mathematical model to express the relationship between  
 189 the chlorophyll a content and the PBZ doses of coffee plants conducted under levels of 0,  
 190 20, 40 and 80% of LR. A cubic model was designed to express the effect of PBZ doses on  
 191 the chlorophyll a content of coffee plants kept under 60% of light restriction. The values were  
 192 higher than the control (without PBZ application) at doses higher than 17.7 mg of the  
 193 regulator per plant, with an estimated maximum value for the dose of 31.0 mg PBZ (2.82 mg  
 194 g<sup>-1</sup>) (Figure 2B).

195 For the unfolding of interaction between LR levels and PBZ doses, in the evaluation of the  
 196 total chlorophyll content, a similar trend was observed for chlorophyll a (Figure 2C and 2D).  
 197 Maximum levels of total chlorophyll as a function of LR levels were estimated at 2.85, 3.00,  
 198 2.97, 4.19 and 2.87 mg g<sup>-1</sup> of fresh matter for plants treated with 0, 10, 20, 30 and 40 mg of  
 199 PBZ, respectively. As with chlorophyll a, coffee plants treated with 30 mg of PBZ via soil  
 200 showed a more significant increase in total chlorophyll content as a function of shade levels  
 201 compared to other doses (Figure 2C). For this treatment, maximum total chlorophyll content  
 202 (4.19 mg g<sup>-1</sup>), estimated at 60.4% of LR level, was approximately 2.5 times higher than the  
 203 treatment in full sun.



204

205 **Fig. 2. Chlorophyll a and total chlorophyll content in leaves of coffee plants (*Coffea***  
 206 ***arabica* L. ‘Catuaí Vermelho IAC 144’) in response to different light restriction levels**  
 207 **(LR) and paclobutrazol doses (D), at 100 days after the application of the regulator. (A,**  
 208 **B) chlorophyll a content (Chl a): (A) ♦0 mg –  $\hat{Y}^* = 1.67811 - 0.0428795X + 0.00176607X^2$**   
 209 **– 0.0000146094X<sup>3</sup> (R<sup>2</sup> = 0.9874); ■10 mg –  $\hat{Y}^{**} = 1.451 + 0.0124625X$  (R<sup>2</sup> = 0.7246); ▲20**  
 210 **mg –  $\hat{Y}^{**} = 1.664 + 0.0095125X$  (R<sup>2</sup> = 0.9839); ●30 mg –  $\hat{Y}^{**} = 1.39532 - 0.0378676X +$**

211  $0.00264821X^2 - 0.0000257552X^3$  ( $R^2 = 0.9276$ ); x40 mg -  $\hat{Y}^{**} = 1.592 + 0.0093125X$  ( $R^2 =$   
212  $0.7836$ ). (B) ♦0%; ■20%; ▲40%; ●60% -  $\hat{Y}^{**} = 2.29157 - 0.0999643X + 0.00811786X^2 -$   
213  $0.00014X^3$  ( $R^2 = 0.9199$ ); x80%. (C, D) total chlorophyll content (Chl a+b): (C) ♦0 mg -  $\hat{Y}^*$   
214  $= 1.99146 - 0.0472693X + 0.00204464X^2 - 0.0000171615X^3$  ( $R^2 = 0.9584$ ); ■10 mg -  $\hat{Y}^{**} =$   
215  $1.706 + 0.016225X$  ( $R^2 = 0.8051$ ); ▲20 mg -  $\hat{Y}^{**} = 1.952 + 0.0127625X$  ( $R^2 = 0.9797$ ); ●30  
216 mg -  $\hat{Y}^{**} = 1.66129 - 0.0719911X + 0.00446161X^2 - 0.0000426562X^3$  ( $R^2 = 0.9875$ ); x40  
217 mg -  $\hat{Y}^{**} = 1.8975 + 0.0122X$  ( $R^2 = 0.7984$ ). (D) ♦0%; ■20%; ▲40%; ●60% -  $\hat{Y}^{**} = 2.78125$   
218  $- 0.171187X + 0.0146375X^2 - 0.000258125X^3$  ( $R^2 = 0.8583$ ); x80%. \* e \*\*: significant by  
219 regression analysis at 5% e 1% of probability, respectively.

220 As a strategy to increase the efficiency of light absorption processes, plants grown under  
221 lower radiation levels tend to have higher density of light-picking complexes when compared  
222 to plants kept in full sunlight [29]. In addition, the increase in chlorophyll content under low  
223 light conditions may be associated, in part, with higher nitrogen allocation to photosystems  
224 [30].

225 The total chlorophyll content of coffee plants conducted under 60% of LR, as a function of  
226 doses of PBZ applied, was higher than the control at dosages above 16.5 mg of the inhibitor,  
227 with a maximum point estimated for 30.6 mg of PBZ. Any mathematical model among those  
228 studied expressed the effect of PBZ treatment on the total chlorophyll content of coffee  
229 plants kept under 0, 20, 40 and 80% shading (Figure 2D).

230 Treatment with PBZ may result in increases cytokinin levels [14]. It is known that elevation in  
231 cytokinin levels can accelerate chloroplast differentiation and chlorophyll biosynthesis, and  
232 maintain the integrity of this molecule [31]. Working with *Solenostemon rotundifolius*, [32]  
233 observed that PBZ treatment resulted in higher number of chloroplasts per cell unit in the  
234 leaves when compared to the control treatment.

235 It was observed increase in SPAD index of arabica coffee plants, due to the increase in  
236 shading levels, with maximum value (68.99) estimated for the level of 62.7% of LR (Figure  
237 3A). There is positive correlation between SPAD index and chlorophyll content in leaves of  
238 different plant species [33-36]. Therefore, the increase observed in the SPAD index in this  
239 study was associated with higher chlorophyll content in leaves of the shaded plants (Figure  
240 2C).

241 Generally, treatment with PBZ provides higher SPAD index in plants. This fact is commonly  
242 associated with the increase of chlorophyll content, or the higher number of chloroplasts per  
243 unit of leaf area, in response to increases of leaf thickness and decrease of leaf area [32, 13].  
244 However, for the present study, there was no effect of the PBZ doses applied via soil, on the  
245 coffee plants SPAD index (Figure 3B).

246 It should be noted that SPAD index is based on a unit of green light reflectance area, while  
247 the chlorophyll content determined in the present work was defined based on the mass of  
248 the leaf blade. Due to the existence of an impact intensity differential of PBZ on leaf  
249 morphology (area reduction and thickness increase) and chloroplast metabolism (chlorophyll  
250 synthesis, movement, distribution, and anatomy of these plastids), different associations  
251 between these effects may interfere with the intensity of the green color of the leaf.

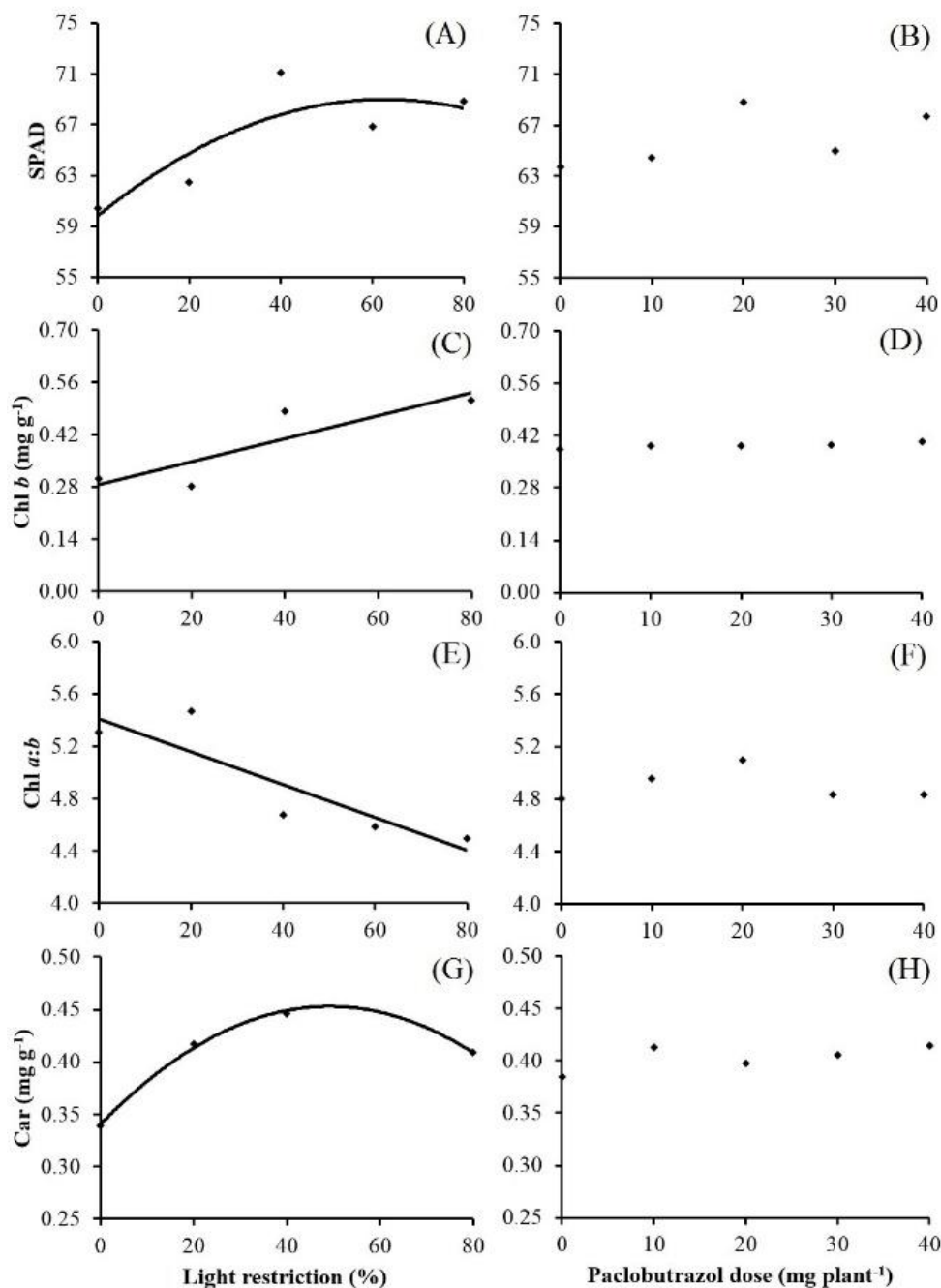
252 There was a tendency for linear increase of chlorophyll *b* content as a function of the  
253 increase in LR levels (Figure 3C). [29] associated the decrease of the chlorophyll *b* content  
254 in *Illicium floridanum* cultivated in full sun to the degradation of this pigment by the excess of  
255 irradiation. In addition, the higher development of LHCI and LHCII antenna complexes in  
256 shaded plants [8] may be associated with this response.



257 A linear decreasing effect was observed for the relationship between chlorophyll *a* and *b*  
258 ratio (Chl *a:b*) and LR levels (Figure 3E). In general, the size of the antenna complexes  
259 (LHCI and LHCII) of plants increases under low irradiation, while under high irradiation, it is  
260 reduced to avoid overexcitation of the photosystems [8]. It is well-known that photosystems  
261 only contain chlorophyll *a*, while antenna complexes present both chlorophyll *a* and *b* [9].  
262 Thus, increase in LHCI and II complexes in shaded plants may result in lower chlorophyll *a:b*  
263 ratio when compared to sun leaves.

264 Under shading conditions, the environment under the canopy of shading plants is enriched  
265 with green light, as this is the predominant wavelength in the light transmitted and reflected  
266 by leaves. Chlorophyll *b* shows the maximum absorption peak closest to green wavelength,  
267 compared to chlorophyll *a*. Therefore, the reduction of chlorophyll *a:b* ratio is an important  
268 strategy to increase the use of the predominant green light, which affects the leaves of  
269 shaded plants [37].

270 A quadratic model for the relationship between carotenoid content of coffee plants and the  
271 levels of shading was delineated. The conduction of coffee plants in shaded environments  
272 resulted in higher levels of carotenoids, with maximum value estimated at 49.3% of LR  
273 (Figure 3G). Generally, very intense solar radiation induces elevation of carotenoid levels,  
274 since this pigment is involved in protecting the damage caused by excessive light [9], which  
275 was not observed in the present study.



276

277 **Fig. 3.** SPAD index, chlorophyll *b* content, ratio between chlorophyll *a* e *b*, and  
 278 carotenoid content in leaves of coffee plants (*Coffea arabica* L. 'Catuai Vermelho IAC  
 279 144') in response to different light restriction levels (LR) and paclobutrazol doses  
 280 (D), at 100 days after the application of the regulator. (A, B) SPAD index (SPAD): (A)  
 281  $\hat{Y}^{**} = 59.8123 + 0.292836X - 0.00233598X^2$  ( $R^2 = 0.7305$ ); (C, D) chlorophyll *b* content  
 282 (Chl *b*): (C)  $\hat{Y}^{**} = 0.2856 + 0.00307571X$  ( $R^2 = 0.7583$ ); (E, F) ratio of chlorophyll *a* to *b*  
 283 (Chl *a:b*): (E)  $\hat{Y}^{**} = 5.4074 - 0.01257X$  ( $R^2 = 0.7853$ ); (G, H) carotenoid content (Car):  
 284 (G)  $\hat{Y}^* = 0.339864 + 0.00457841X - 0.0000464205X^2$  ( $R^2 = 0.9959$ ). \* e \*\*: significant by  
 285 regression analysis at 5% e 1% of probability, respectively.

286 However, the effect verified in the present study corroborates with [3], who observed higher  
287 carotenoid content in arabica coffee leaves conducted under 85% of light restriction, when  
288 compared to those grown in full sun.

289 The higher carotenoid content in shaded coffee plants observed in this work may have  
290 occurred due to the greater amount of light absorption complexes per unit of leaf area in  
291 these plants [29], which has carotenoids as components of the complex antenna.

292 The content of chlorophyll *b*, chlorophyll *a:b* ratio and carotenoid content of coffee plants  
293 were not altered by the application of paclobutrazol (Figure 3D, 3F, and 3H). According to  
294 [38], the treatment with triazoles can increase abscisic acid and cytokinins, resulting in  
295 increase in chlorophyll and carotenoid contents in leaves. However, for the present work, the  
296 dosages of PBZ used were not effective in inducing such changes.

297 It should be emphasized again that the anatomical effect induced by PBZ in increasing  
298 thickness and reducing leaf area may interfere with pigment contents when considering the  
299 quantification based on the mass of the leaf blade.

300 Light restriction influenced all the characteristics related to leaf gas exchange, with the  
301 exception of internal CO<sub>2</sub> concentration in the substomatic chamber. However, no  
302 characteristics were affected by PBZ doses or the interaction between LR and PBZ (Table  
303 2).

304 Often, limitations of leaf gas exchange in coffee plants are strictly associated with the  
305 sensitivity of stomata to the increase in the vapor pressure deficit between leaf and  
306 atmosphere [39, 5]. Air temperature attenuation is an important environmental change  
307 promoted by shade cultivation [4], and may reduce the above limitations [40].

308 A quadratic model was designed to express stomatal conductance ( $g_s$ ) response of coffee  
309 plants as a function of the levels of light restriction. Initially, it is observed a slight decrease  
310 of the values up to the level of 15.3% of shading (3.5% lower than the control). However, the  
311 increase was more expressive from the 30.5% of LR level, with maximum  $g_s$  at 80% of  
312 shading, 58.93% higher than the full sun treatment (Figure 4A).

313 Shading provides a modification in the microclimate of the growing environment, in order to  
314 decrease wind speed and leaf temperature, and increase relative humidity of the air. This  
315 results in a reduction in vapor pressure deficit and, therefore, reduces stomatal limitations of  
316 coffee trees [40]. The highest values of  $g_s$  in coffee plants conducted under higher levels of  
317 light restriction in this study were associated with this fact.

318 Direct relationship between the increases in light restriction levels and the potential net  
319 assimilation rate of CO<sub>2</sub> (*A*) and transpiration rate (*E*) of the coffee plants were verified. The  
320 elevation of *A* and *E* values in 73.04 and 43.27%, respectively, was observed for the highest  
321 levels of shading (Figures 4C and 4E).

322 Stomatal conductance is the main limiting factor of the photosynthetic rate in plants grown in  
323 full sun [40], a fact that may be associated to the increase in *A* values of shaded coffee  
324 plants, since  $g_s$  was also elevated under these conditions.

325 It is worth mentioning that the increase of *A* under light restriction was similar to the increase  
326 in the content of photosynthetic pigments under these conditions. Thus, the higher content of  
327 chlorophylls and carotenoids (Figures 2A, 2C, 3C and 3G) may also have contributed to the  
328 increase in photosynthetic rates.

329 **Table 2. Analysis of variance summary and coefficients of variation (CV) of stomatal**  
 330 **conductance ( $g_s$ ), net CO<sub>2</sub> assimilation rate ( $A$ ), transpiration rate ( $E$ ), internal CO<sub>2</sub>**  
 331 **concentration ( $C_i$ ), and carboxylation efficiency ( $A/C_i$ ) of *Coffea arabica* L. ‘Catuai**  
 332 **Vermelho IAC 144’ plants submitted to different light restriction levels (LR) and**  
 333 **paclobutrazol doses (D), evaluated at 99 days after the application of the regulator.**  
 334 **Vitória da Conquista – BA, 2017.**

SV	df	MEAN SQUARES				
		$g_s$	$A$	$E$	$C_i$	$A/C_i$
LR	4	0,1488**	37,93**	2,38**	622,66 <sup>ns</sup>	0,000428**
D	4	0,0431 <sup>ns</sup>	5,07 <sup>ns</sup>	0,11 <sup>ns</sup>	254,59 <sup>ns</sup>	0,000061 <sup>ns</sup>
LR*D	16	0,0229 <sup>ns</sup>	3,89 <sup>ns</sup>	0,30 <sup>ns</sup>	399,76 <sup>ns</sup>	0,000049 <sup>ns</sup>
BL	3	0,5144**	11,86*	0,99*	3100,54**	0,000076 <sup>ns</sup>
Wn	72	0,0357	3,25	0,32	266,98	0,000036
CV (%)		60,76	27,84	24,49	5,31	28,34

335 <sup>ns</sup>, \* e \*\*: non-significant, significant by “F” test at 5% and 1% of probability, respectively.

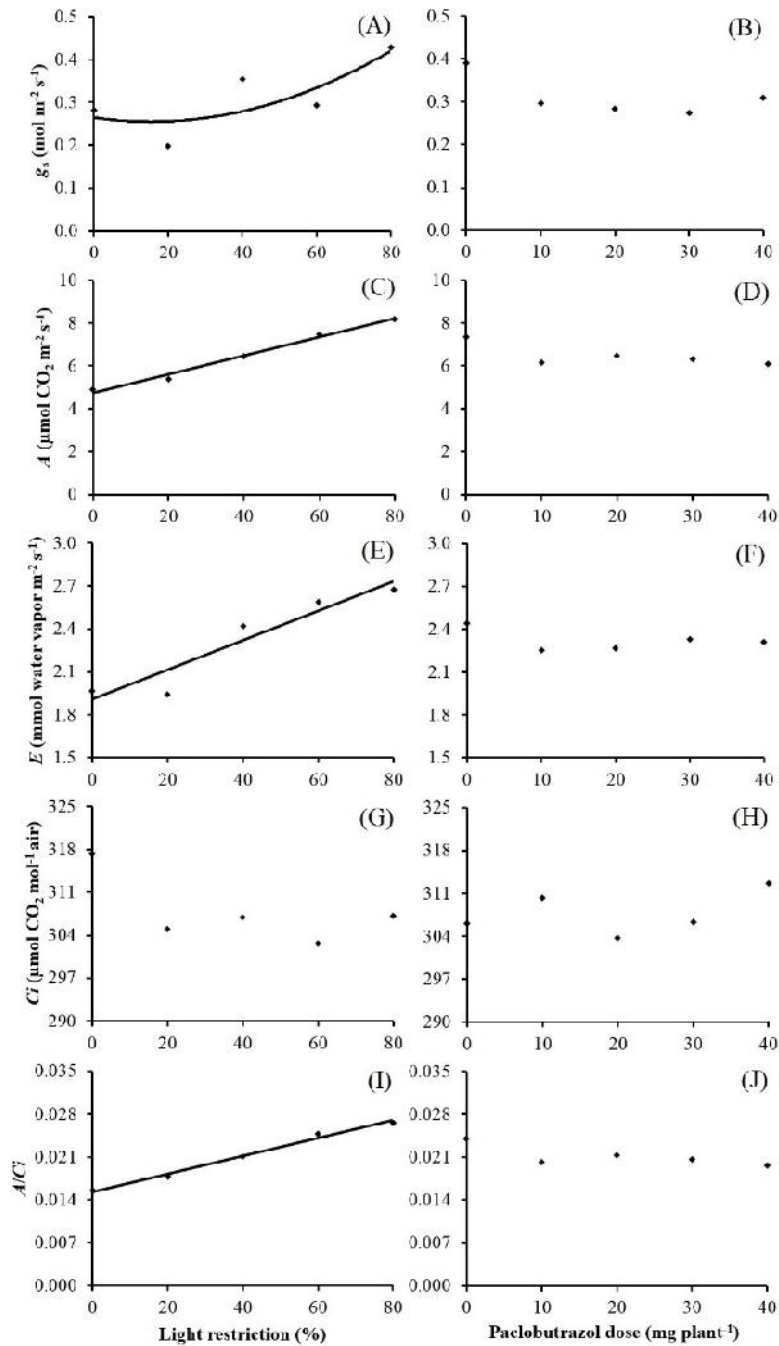
336 The increase of  $E$  observed in the shaded coffee plants was related to the higher values of  
 337  $g_s$  (less leaf stomatal resistance), since this process results mainly from the diffusion of  
 338 water vapor through stomatal opening.

339 Higher values of  $g_s$  and  $E$  were verified in coffee plants conducted under artificial light  
 340 restriction, compared to coffee plants grown in full sun [6]. Higher photosynthetic rates in  
 341 shaded coffee plants were verified by [41], compared to plants conducted without light  
 342 restriction.

343 In the present work, although the shaded coffee plants presented less resistance to gas  
 344 diffusion, internal CO<sub>2</sub> concentration ( $C_i$ ) was not altered by light restriction levels (Figure  
 345 4G). [7] also observed no difference between the  $C_i$  of shaded coffee trees and full sun.

346 On the other hand, carboxylation efficiency ( $A/C_i$ ) showed a tendency of linear increase as a  
 347 function of the increase of shading levels (Figure 4I). This parameter was elevated up to  
 348 75.9% at the level of 80% of shading, compared to the control treatment (full sun). This  
 349 result was related, in part, to the temperature attenuation in shaded environments.

350 Ribulose-1,5-bisphosphate-carboxylase/oxygenase (rubisco) enzyme present in chloroplasts  
 351 can catalyze both photosynthesis and photorespiration. The rates of each of these  
 352 processes depend on the activity of rubisco as carboxylase or oxygenase, and they are  
 353 modified by the environmental conditions [9].



354

355 Fig. 4. Leaf gas exchanges of coffee plants (*Coffea arabica* L. 'Catuaí Vermelho IAC  
 356 144') in response to different light restriction levels (LR) and paclobutrazol doses  
 357 (D), at 99 days after the application of the regulator. (A, B) stomatal conductance ( $g_s$ ):  
 358 (A)  $\hat{Y}^* = 0,2644 - 0,0012025X + 0,000039375X^2$  ( $R^2 = 0,6289$ ); (C, D) net CO<sub>2</sub>  
 359 assimilation rate ( $A$ ): (C)  $\hat{Y}^{**} = 4,7421 + 0,043295X$  ( $R^2 = 0,9884$ ); (E, F) transpiration  
 360 rate ( $E$ ): (E)  $\hat{Y}^{**} = 1,9066 + 0,0103125X$  ( $R^2 = 0,8940$ ); (G, H) internal CO<sub>2</sub> concentration  
 361 ( $C_i$ ); (I, J) carboxylation efficiency ( $A/C_i$ ): (I)  $\hat{Y}^{**} = 0,015343 + 0,000145525X$  ( $R^2 =$   
 362  $0,9892$ ). \* e \*\*: significative by regression analysis at 5% e 1% of probability,  
 363 respectively.

364 Although the activity of the enzyme as carboxylase increases with temperature, the affinity of  
365 rubisco by CO<sub>2</sub>, as well as the solubility of CO<sub>2</sub>, decrease. This results in increases in  
366 photorespiratory activity at higher temperatures and, consequently, lower carboxylation  
367 efficiency [42].

368 On this way, increases in the carbon assimilation rate due to the light restriction, associated  
369 to a constant *C<sub>i</sub>* between the treatments, resulted in higher carboxylation efficiency in  
370 shaded coffee plants.

371 PBZ treatment can alter several aspects of leaf gas exchange in many species [12, 43-44].  
372 In the present study, however, PBZ application via soil did not influence any of the  
373 parameters related to gas exchange of coffee plants (Figures 4B, 4D, 4F, 4H and 4J).

374 According to [12], the increase in abscisic acid contents resulting from triazole application  
375 may result in partial stomatal closure and reduction in the transpiration rate of treated plants.  
376 On the other hand, PBZ application did not alter stomatal conductance in coffee plants [45].

377 The effect of PBZ on increasing [44] or reducing [46] the rate of CO<sub>2</sub> assimilation is  
378 modulated by dosage and form of application. In coffee plants, [45] found that application of  
379 lower concentrations of PBZ via leaf yielded higher photosynthetic rates and carboxylation  
380 efficiency, while higher concentrations restricted both processes.

381 The absence of the effect of PBZ on leaf gas exchanges of coffee plants, in this work, was  
382 possibly due to the fact that the dosages studied were too low to alter these parameters.

383

#### 384 **4. CONCLUSION**

385

386 Light restriction optimized the photosynthetic apparatus of the plants, mainly at levels close  
387 to 60%, and favored the leaf gas exchanges of arabica coffee in initial growth. The  
388 application of paclobutrazol in the dosages studied resulted in little or no effect on the levels  
389 of photosynthetic pigments, and did not influence the leaf gas changes of young arabica  
390 coffee plants.

391

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398

#### 399 **AUTHORS' CONTRIBUTIONS**

400

401 André F. F. Ribeiro developed the study, participated in the collection of data and the  
402 accomplishment of the statistical analysis, of the writing of the manuscript and carried out  
403 the bibliographic research. Sylvana N. Matsumoto guided the first author during the  
404 development of the study, and participated in performing the statistical analysis and writing  
405 of the manuscript. The other authors assisted in the conduction of the experiment, data  
406 collection and analysis. All authors read and approved the final manuscript.

407

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