

1 **Original Research Article**

2
3 Comparative Studies on Effectiveness of
4 Branded and Unbranded Disinfectants on *E.*
5 *coli* and *Staphylococcus* species.
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10 **ABSTRACT (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)**
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Aims: To compare the antimicrobial potential of branded and unbranded disinfectants on clinical bacterial isolates.

Study design: the agar-well diffusion and micro broth dilution were adopted for the study. Ten disinfectants of which five were branded (industrial prepared) and five unbranded (indigenous prepared) were used against *E. coli* and *Staphylococcus aureus*.

Place and Duration of Study: Department of Microbiology, Rivers State University. the study was for a period of two months (June-July, 2018).

Methodology: Faecal samples were collected from the University Medical centre and was analyzed in the Microbiology Laboratory for the isolation of *Escherichia coli* and *Staphylococcus aureus* using standard microbiological method. The antimicrobial potential of both branded and unbranded disinfectants on the clinical isolates were evaluated using the micro dilution technique and the well in agar technique

Results: The result in this study showed that both branded and unbranded disinfectants were effective on the *E. coli* and *Staphylococcus* isolates. However, the unbranded were only effective at high concentrations. *E. coli* had zone of inhibition ranging from 0 to 22mm when tested with the unbranded disinfectant, while 0 to 17mm was recorded for *Staphylococcus aureus*. The zones of inhibition of the branded disinfectant on *E. coli* ranged from 0 to 28mm, while zone diameter of *Staphylococcus aureus* ranged from 0 to 21mm. Among the unbranded disinfectants, Lysol produced the highest zone of inhibition. While among the branded disinfectants, Savlon produced the highest zone of inhibition. The positive control was effective against all tested organisms with zones of inhibition ranging from 17-26 mm. On the other hand, as expected, the negative control (sterile distilled water) did not show any zone of inhibition.

Conclusion: The study showed that branded disinfectants were more effective on the clinical isolates than the unbranded disinfectants.

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13 **Keywords:** *Escherichia coli*, *Staphylococcus aureus*, branded disinfectants, unbranded
14 disinfectants, microdilution, well-in-agar diffusion.
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18 **1. INTRODUCTION**

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20 Antimicrobials are substances that have the ability to kill or inhibit the growth or proliferation
21 of microorganisms [17]. This implies that these substances when introduced in objects or
22 other materials or consumed could either be bacteriostatic or bactericidal in action.
23 According to Douglas and Braide [6], antimicrobial substances that when introduced on
24 inanimate objects kills or inhibits the growth of microbes. Thus, a good disinfectant should be
25 able to offer complete and full microbiological sterilization, without harming humans and
26 useful form of life, be inexpensive and noncorrosive. However, most disinfectants are also,
27 by nature, potentially harmful to humans and animals. The choice of disinfectant to be used
28 may depend on the demanding situation. According to Van et al [15], the idea of using
29 disinfectants and antiseptics is to control or reduce the presence of microorganisms. In order
30 to prevent infections as it regards injury, the most vital measure is to kill or inhibit the growth
31 of microorganisms on the skin, wounds and in human body cavity [3]. The antimicrobial
32 potentials of these disinfectants could be influenced by their formulation properties,
33 concentration of organic components, temperature, synergy, rate of dilution and
34 experimental procedures, mode of application, water solubility and pH[5 and 6]. Application
35 factors include the type of surface to be applied, the type of (organic) soil, the temperature
36 and contact time as well as humidity and the method of application (with or without
37 mechanical action) [8]. A disinfectant could be branded or unbranded (Maillard, 2005).
38 These unbranded disinfectants are hawked from place to place and also sold in the local
39 markets [9]. They could be good alternative disinfecting agents if their effectiveness against
40 some clinical isolates is known [10]. Unbranded disinfectants are produced locally by people
41 that are taught how to make different household washing, cleaning and disinfecting agents.
42 When these disinfectants are made by these persons, they are normally packaged in
43 containers (usually liable plastic bottles). There are two different ways by which disinfectants
44 can act on microorganisms: growth inhibition (bacteriostasis and fungistasis) or lethal action
45 (bactericidal, fungicidal or viricidal effects) [2]. Thus, this study is aimed at comparing the
46 antimicrobial potential of branded and unbranded disinfectants on clinical bacterial isolates.

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48 **2. METHODOLOGY**

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50 **2.1 Collection of clinical Samples**

51 Faecal samples were collected from the Rivers State University Medical Center, Port
52 Harcourt in specimen bottle and transported to the Microbiology laboratory of Rivers State
53 University, Port Harcourt.

54 **2.2 Collection of Disinfectant Samples**

55 The branded disinfectants used were; Purit, Dettol, Ivy's, Savlon and Robert. While the
56 unbranded disinfectants were; Lysol, Pine oil, Morigade, Nigertol, Chlonoxydol. The
57 disinfectants were purchased from different markets within Port Harcourt Metropolis, Rivers
58 State.

59 **2.3 Isolation of Test Organisms**

60 Isolation of the test organisms was carried out as described by Cheesbrough [4]. A thick
61 suspension of the faecal sample was emulsified in 1ml sterile peptone water. Afterwards a
62 loop full of the emulsified sample was inoculated on Mannitol salt agar plates (MSA) and
63 Eosin methylene blue agar plates (EMB). Plates were then incubated at 37°C for 24 hours.

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67 **2.3.1 Confirmation of Test Organisms**

68 Ensuing colonies on the MSA and EMB plates were carefully picked using a sterile wire loop
69 and subcultured on fresh plates of MSA and EMB agar. Pure isolates were then stored in
70 nutrient agar slants and stored in the refrigerator for further use.

71 The respective pure isolates were identified using conventional methods as described by
72 Cheesbrough [4]. Further confirmation of isolates was done by comparing their biochemical
73 results with those presented in Bergey's manual of determinative bacteriology [18]. The
74 conventional methods include; microscopy, motility, coagulase, catalase, oxidase, indole
75 production, methyl red, citrate utilization, **Voges-Proskauer** test and sugar fermentation [4].

76 **2.4 Standardization of Test Inoculum**

77 Test isolates were standardized using the 0.5 McFarland. The test isolates were placed in
78 sterile test tubes containing 4ml distilled water. The turbidity was ascertained using the
79 already prepared McFarland standard [4]. The standardized isolates were carefully spread on
80 prepared sterile Mueller-Hinton agar plates as described by Wemedo and Robinson [16].
81 Plates were allowed to dry before 4 wells using a 6mm well borer were made on the dried
82 seeded plates.

83 **2.4.1 Antimicrobial Assay (Well-in-agar method)**

84 The antimicrobial activity of each disinfectants with different concentration was tested in vitro
85 against *E. coli* and *Staphylococcus aureus*. Aliquots (0.1ml) of 10%, 25%, 50% and 100%
86 concentration of the different disinfectants were transferred using sterile Pasteur pipette in to
87 the four wells. The plates were then incubated at 37°C for 18 to 24hours in an upright
88 position. Autoclaved distilled water was used as negative control while ofloxacin was used
89 as a positive control. After incubation, the plates were observed and the zones of inhibition
90 that developed were read and interpreted [16].

92 **2.4.2 Broth Dilution Method**

93 The minimum inhibitory concentrations of the different disinfectants were **carried out** using
94 the broth dilution method as described by Prescott *et al* (2011). Different concentrations of
95 the disinfectants were prepared (10, 25, 50, 75 and 100) mg/ml [1]. One milliliter (1ml) of the
96 standardized inoculum and the various concentrations of the disinfectants were put into the
97 sterile tubes of nutrient broth respectively. Tubes containing nutrient broth and organisms
98 without the disinfectant served as negative control while the tube containing only the broth
99 and disinfectant without organism served as positive control. These tubes were incubated at
100 37 °C for 18 to 24 hours. Thereafter, the tubes were examined for visible growth or turbidity
101 and recorded. The MIC is the concentration at which no visible growth was observed when
102 compared with the control [9].

105 **3. RESULTS AND DISCUSSION**

107 The result in Table1 showed the characteristics of the two bacterial isolates to some
108 biochemical tests as well as their morphology. The result showed that the isolates were
109 *Escherichia coli* and *Staphylococcus aureus*. In this current study, the antimicrobial activities
110 of both the branded disinfectants and unbranded disinfectants on *Escherichia coli* and
111 *Staphylococcus aureus* using the agar well diffusion showed some level of inhibition. In
112 Table2, the effect of the unbranded disinfectants on *Escherichia coli* showed that the
113 effectiveness of the unbranded disinfectants occurred at the 50% and 100% concentration
114 and at 50% concentration only Lysol, morigade and pine oil were able to produce a clear
115 zone of inhibition while Chlonoxynol and Nigertol showed no antimicrobial effect.
116 Furthermore, all the unbranded disinfectants were able to exert some antimicrobial
117 properties thereby leading to the formation of zones of inhibition at 100% concentration
118 (Table2.). Lyaol and Morigade showed the highest zones of inhibition of 22mm and 20 mm
119 respectively thereby making them the most effective unbraded disinfectants on *E. coli*.

120 The antimicrobial activities of the unbranded disinfectant on *Staphylococcus aureus* showed
121 that the unbranded disinfectants were not effective at 10 and 25 % concentrations. Also, at
122 50% concentration only pine oil was able to inhibit the staphylococcal isolates at a zone of

123 10mm. whereas at 100% concentration, all unbranded disinfectants except Pine oil exerted
 124 some level of antimicrobial activities showing visible zones (Table 3).
 125 The result of the antimicrobial activities of the branded disinfectant on *Escherichia coli*
 126 is presented in Table 4. The result showed that only Robert and Savlon were able to inhibit the
 127 isolates of *E. coli* at 10% with zones of inhibition observed to be 12mm and 7mm
 128 respectively. While at 25, 50 and 100%, all branded disinfectants produced visible zones of
 129 inhibition on the isolates. At 25% concentration, Robert was the most effective having zone
 130 of 15mm while Ivy's and Savlon were the most effective disinfectants at the 50%
 131 concentration with zones observed around 18mm and 20mm respectively. At 100% only
 132 Purit produced the least zone of inhibition of 18mm on the isolates while other disinfectants
 133 had greater zones of inhibitions.

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 135 **Table 1: Colonial morphology and biochemical characteristic of the bacterial isolates**

Isolate	Morphology	microscopy	G	L	S	M	F	Cat	Coa.	Ind.	MR.	Cit.	Mot	Identity
A	Metallic-silver small round flat	-ve bacilli	+	+	+	-	-	+	-	+	+	-	+	<i>Escherichia coli</i>
B	Golden yellow round smooth	+ve clustered cocci	+	+	+	+	+	+	+	-	+	+	-	<i>Staphylococcus aureus</i>

136 **Key:** G; glucose, L; Lactose, S; Sucrose, M; Maltose, Cat. ;Catalase, Coa.; Coagulase, Ind.;Indole;
 137 MR; Methyl red, Mot.; Motility, Oxi.; Oxidase,
 138 SSA; Salmonella and Shigella Agar, MSA; Manitol salt Agar

139 **Table2:Effect of unbranded disinfectants on *Escherichia coli***

Con.	Control				Disinfectants										
	Positive (Ofloxacin)		Negative (Sterile water)		Chlonoxyinol		Lysol		Morigade		Nigertol		Pine oil		
	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	
10%	8	10	00	00	00	00	00	00	00	00	00	00	00	00	00
25%	12	11	00	00	00	00	00	00	00	00	00	00	00	00	00
50%	16	18	00	00	00	00	12	12	10	10	00	00	6	6	
100%	10	22	00	00	00	00	22	22	20	20	12	10	10	14	

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155 **Table 3: Effect of unbranded disinfectants on *Staphylococcus aureus***

Con.	Control				Disinfectants										
	Positive (Ofloxacin)		Negative (Sterile water)		Chlonoxydol		Lysol		Morigade		Nigertol		Pine oil		
	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	
10%	12	10	00	00	00	00	00	00	00	00	00	00	00	00	00
25%	17	16	00	00	00	00	00	00	00	00	00	00	00	00	00
50%	21	20	00	00	00	00	00	00	00	00	00	00	00	10	10
100%	28	28	00	00	00	00	17	19	12	10	14	14	00	00	

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158 **Table 4: Effect of branded disinfectants on *Escherichia coli***

Con.	Control				Disinfectants										
	Positive (Ofloxacin)		Negative (Sterile water)		Dettol		Ivy's		Purit		Robert		Savlon		
	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	
10%	8	10	00	00	00	00	00	00	00	00	00	12	10	5	7
25%	12	11	00	00	12	10	12	14	11	9	13	15	12	14	
50%	16	18	00	00	14	14	18	16	12	14	15	17	20	18	
100%	10	22	00	00	22	20	26	25	16	18	24	26	26	28	

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163 **Table 5: Effect of branded disinfectants on *Staphylococcus aureus***

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Con.	Control				Disinfectants									
	Positive (Ofloxacin)		Negative (Sterile water)		Dettol		Ivy's		Purit		Robert		Savlon	
	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2	P1	P2
10%	12	10	00	00	12	10	00	00	9	11	00	00	8	8
25%	17	16	00	00	16	18	00	00	21	17	16	14	16	14
50%	21	20	00	00	20	22	10	8	23	21	17	18	22	20
100%	28	28	00	00	22	24	14	16	24	26	21	23	22	24

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167 **Table 6: Concentration of Activity of Branded Disinfectants (MIC)**

Organisms	Branded Disinfectants	Concentration (%)
<i>Staphylococcus aureus</i>	Purit	75
	Dettol	50
	Ivy's	75
	Salvon	75
	Robert	75
<i>Escherichia coli</i>	Purit	50
	Dettol	75
	Ivy's	75
	Salvon	50
	Robert	75

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MIC: minimal inhibitory concentration

170 **Table 7: Concentration of Activity of Unbranded Disinfectants (MIC)**

Organisms	Unbranded Disinfectants	Concentration (%)
<i>Staphylococcus aureus</i>	Lysol	75
	Pine oil	75
	Morigade	75
	Nigertol	75
	Chlonoxynol	75
<i>Escherichia coli</i>	Lysol	50
	Pine oil	75
	Morigade	50
	Nigertol	75
	Chlonoxynol	75

171 *MIC: minimal inhibitory concentration*

172 The bacterial isolates in this current study have shown some level of resistance and
173 susceptibility to the various form of disinfectants.

174 *Staphylococcus aureus* is a known cause of various form of infections ranging minor skin
175 infections, such as pimples, impetigo, boils, cellulitis, folliculitis, carbuncles, scalded skin
176 syndrome, and abscesses, to life-threatening diseases such as pneumonia, meningitis,
177 osteomyelitis, endocarditis, toxic shock syndrome, gastrointestinal diseases, bacteremia and
178 sepsis [13 and 20]. While some strains of *Escherichiacoli* are virulent and are responsible for
179 diarrheal infections worldwide as well as neonatal meningitis, septicemia, and urinary tract
180 infections (UTIs) [14].

181 The result in this study showed that the branded disinfectants are very much effective than
182 the unbranded disinfectants. There is also a dearth of information on the effectiveness as
183 well as the composition of unbranded disinfectants. However, Douglas and Braide (2015) in
184 a study of the effectiveness of Locally Formulated Unbranded disinfectants on clinical
185 bacterial isolates reported that unbranded (locally formulated) disinfectants are more potent
186 when not diluted and that the differences in the activities of the unbranded and branded
187 disinfectants may be due to the different substances used in formulations, as well as the
188 structure and nature of the cell wall of the microbes. The disinfectants in this current study
189 showed some level of activity on both Gram negative and positive bacterial isolates
190 indicating that they have broad spectrum of activity. This is in agreement with Douglas and
191 Braide [6] who had earlier reported that disinfectants show a broad spectrum of activity
192 against different bacterial isolates.

193 Effectiveness of Dettol and Savlon has been reported by [9] who carried out a study on the
194 efficacy of some disinfectants on clinical isolates including *Escherichia coli* and
195 *Staphylococcus aureus*.in their study, Dettol was more active against the isolates compared
196 with Savlon and other tested disinfectants. Other studies carried out by Olowe [12] and
197 Olasehinde *al* [11] also reported Dettol to be a strong disinfectant. Furthermore, El-
198 Mahmood and Doughari[7] in a study of Bacteriological examination of some diluted
199 disinfectants routinely used in the specialist hospital Yola, Nigeria reported that Purit has a
200 higher activity on *E. coli* than *S. aureus* whereas in this study Purit was more effective
201 against *S. aureus* than *E. coli*.

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204 **4. CONCLUSION**

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206 The antimicrobial effectiveness of five unbranded disinfectants and five branded
207 disinfectants *Staphylococcus aureus*, and *Escherichia coli* was evaluated. Despite some level

208 of antimicrobial actions observed in the unbranded disinfectants, the findings in this study
209 **have** shown that the branded disinfectants are more effective than the unbranded
210 disinfectants. Also, since the unbranded disinfectants have shown some level of
211 antimicrobial actions, increasing the formulation or the quantity for disinfection would be
212 necessary.

213 **COMPETING INTERESTS DISCLAIMER:**

214 **Authors have declared that no competing interests exist. The products used for this**
215 **research are commonly and predominantly use products in our area of research and**
216 **country. There is absolutely no conflict of interest between the authors and producers of**
217 **the products because we do not intend to use these products as an avenue for any**
218 **litigation but for the advancement of knowledge. Also, the research was not funded by**
219 **the producing company rather it was funded by personal efforts of the authors.**

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221 **Competing interests**

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223 Authors have declared that no competing interests exist.

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226 **ETHICAL APPROVAL (WHERE EVER APPLICABLE)**

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228 The permission to undertake this study was obtained from the Rivers State Health Research
229 Ethical Committee.

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