Yield and Nutritional Value Of *Abelmoshus esculentus* L. (Okra) and *Telfairia occidentalis* Hook, F. (Fluted pumpkin) As Influence By Beauty Salon Wastewater.

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Abstract

5 The effect of beauty salon wastewater on yield and nutritional quality of *Abelmoschus esculentus* L. and Telfairia occidentalis F. was investigated. Plants were grown in perforated polythene bags filled 6 7 with 3kg of top soil and irrigated with 0 (control), 25, 50, 75 and 100% of the wastewater. Plants 8 were laid out in a completely randomized design (CRD) consisting of 5 treatments with 6 replicates 9 per treatment. Results showed that the yield parameters of *Abelmoschus esculentus* including number of fruits/plant, fruit fresh weight and dry weight increased at 25-75% concentrations but decreased at 10 11 highest concentration of 100% wastewater in comparison with the control. Similarly, the total biomass of Telfairia occidentalis and Abelmoschus esculentus increased at 25-75% concentrations of 12 13 wastewater. The N, K, Ca and Na composition of the leaf of T. occidentalis and fruits of A. esculentus 14 increased at all wastewater treatment levels. The percentage ash and protein increased in both plants 15 treated with the wastewater. Lipid content increased while the fibre content decreased in T. occidentalis whereas in A. esculentus fibre content increased while the lipid content decreased in 16 17 comparison to the control. Percentage carbohydrate increased in both plants treated with the wastewater. The use of beauty salon wastewater in irrigation of vegetables would not only reduce 18 19 environmental pollution but also serve as an alternative source of fertilizer for vegetable production.

Keywords: Beauty Salon wastewater, *Telfairia occidentalis, Abelmoschus esculentus*, pollution,
 fertilizer.

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24 **1. Introduction**

The use of wastewater for irrigation is widely seen in many cities of developing countries 25 26 where urban wastewater becomes the irrigation source for farmers in urban and semi-urban areas 27 (Raschid-Sally and Jayakody, 2008). Wastewater use for agriculture is an important management 28 strategy in areas with limited freshwater resources, yielding potential economic and environmental 29 benefits. The practice has manifold benefits in the form of water conservation, nutrient recycling and prevention of surface and ground water pollution (Farahat and Linderholm, 2015). The reuse of 30 31 wastewaters for purposes such as agricultural irrigation can reduce the amount of water that needs to 32 be extracted from environmental water sources (Heidarpour et al., 2007). The reuse of wastewater for 33 irrigation purposes gives it a different fate as agricultural crops can make use of the extra water and 34 nutrients.

Wastewater has a potential to supply carbon nutrients (NPK) and micro nutrients to support 35 crop/plant growth (Singh et al., 2011). It serves as a valuable source of plant nutrients and organic 36 37 matter needed for maintaining fertility and productivity levels of the soil (Rusan et al., 2007). 38 Wastewater can have a positive effect on soil and eventually plant growth, due to its being rich in 39 organic matter and nutrients (Ghanbari et al., 2007; Mohammad and Ayadi, 2004). Application of 40 wastewater to cropland and forested lands is an attractive option for disposal because it can improve physical properties and nutrient contents of soils (Kiziloglu et al., 2007). Wastewater irrigation not 41 only provides water, N, and P but also organic matter (OM) to the soils (Siebe, 1998). The results of 42 43 Aghtape et al. (2011) and Tavassoli et al. (2010) experiments showed that irrigation with wastewater significantly increased the fresh and dry forage yield of corn than that of irrigation with well water. 44 Abu Nada (2009) undertook study to assess the long term impacts of wastewater irrigation on 45 46 different parameters of soil and crop. Long term wastewater irrigation increased salt, organic matter and plant nutrients in both soil layers. Khurana and Singh (2012) summarized the available data on 47 chemical composition of different wastewaters and their effects on soil fertility, soil heavy-metal 48 49 content, crop yield and quality. Field application of all types of wastewaters significantly increased 50 soil OC percentage and cation exchange capacity (CEC). Nadav et al. (2013) indicated that the

51 physico-chemical properties of soils were altered by wastewater irrigation, as a result of long-term 52 accumulation of organic matter in the soil profiles. High level of organic matter in wastewater acts as

53 cement for the building up of soil aggregates.

54 However, apart from plant nutrients contained in wastewater, it may contain various 55 potentially toxic elements and organic matters with highly harmful effects on human and animal health. Municipal wastewater contains relatively high amounts of sodium, which can be accumulated 56 57 in the soil during irrigation with this wastewater and display toxic effects on the plants. If this 58 wastewater is not disinfected or treated in stabilization ponds, it is highly contaminated with microorganisms. Therefore, the utilization of municipal wastewater for the irrigation of crops is 59 associated with a number of risks. Very serious risks are those of crop yields reduction, crops 60 61 contamination with pathogens and intestinal helminthes (Zavadil, 2009).

62 Vegetables play important role in meeting the food requirements of people world-wide, 63 because they are important source of various essential components i.e. minerals, dietary fibers and vitamins (Ogle et al., 2001). They are also potential sources of essential nutrients, constitutes 64 65 functional food components by providing protein, iron and calcium which have noticeable health 66 effects (Arai, 2002). The continuous demand for vegetables has increased the need to cultivate these crops all year round. This in effect leads to the dependence on wastewater during the dry seasons or 67 during periods of drought. Also, due to the light water requirement of some crops, the use of 68 69 wastewater to supplement the freshwater, if any, becomes inevitable.

Okra and fluted pumpkin constitute a major part of the commonly consumed vegetables
which are widely grown on the field and in home gardens where there is high tendency of contact
with beauty salon wastewater. The study is therefore aimed at investigating the impact of beauty salon
wastewater on okra and fluted pumpkin growth.

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75 2. Materials and Methods

76 2.1 Experimental Location

The experiment was conducted in the screen house of the Plant Science and Biotechnology
 Department, Adekunle Ajasin University, Akungba Akoko, Ondo State, Nigeria with latitude 7⁰ 28ⁱN
 and longitude 5⁰44E.

80 **2.2 Planting Materials**

Matured seeds of *Abelmoschus esculentus* were obtained from the Premier Seed Company
 Ibadan, Oyo State, Nigeria, while that of *Telfairia occidentalis* were obtained from a local market at
 Oka-Akoko, Ondo State, Nigeria.

84 2.3 Experimental Set up

85 Top soil used for the experiment was collected from the experimental farm of Plant Science and Biotechnology Department, Adekunle Ajasin University, Akungba Akoko. The soil was air-dried 86 87 and sieved through to remove stones. The beauty salon wastewater was collected from a septic 88 wastewater tank from a beauty salon in Akungba Akoko, Ondo State. Concentrations of 25, 50, 75, 89 and 100% of the wastewater were prepared in a plastic keg just before each treatment by dilution with 90 tap water to make the desired concentrations. Four viable seeds of Abelmoschus esculentus and three 91 viable seeds of *Telfairia occidentalis* were sown in perforated polythene bags containing 3kg of top soil. Seedlings were allowed to establish for three weeks and thinned to one seedling per pot. Plants 92 93 were irrigated with the wastewater at 0 (control), 25%, 50%, 75% and 100% concentrations. Each pot 94 was treated with 250mL (volume enough to keep the soil moist) 2 times in a week; thus each pot 95 received 500mL of wastewater treatment per week. The treatment lasted for 8 weeks. The experiment 96 was carried out from July to October, 2016. Pots were laid in a completely randomized design, with 6 97 replicates per treatments. The experiment ended in October 2016 by harvesting the fruits of 98 Abelmoschus esculentus with the seeds and leaves of Telfairia occidentalis. Their fresh weight was 99 determined after which they were oven-dried at 80°C for the dry weight measurement.

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102 2.4 Measurement of Yield

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Measurement of yield of *Abelmoschus esculentus* and of *Telfairia occidentalis* irrigated with
 different concentrations of beauty salon wastewater was done. The parameters studied include:
 Number of fruits/plant, total fresh weight of fruits/plant, total dry weight of fruits/plant.

107 2.5 Chlorophyll content analysis

The total chlorophyll content of leaves of *Abelmoschus esculentus* and *Telfairia occidentalis* was determined using the (Arnon, 1949) method. One gram of fresh leaves was ground with acid leached sand, (sand washed with concentrated sulphuric acid (H_2SO_4) and thoroughly rinsed with distilled water to remove all nutrients). The chlorophyll content was extracted using 10ml of 80% acetone and centrifuged at 2000rpm for 15minutes. The clear supernatant liquid was decanted and the absorbance read with photo spectrophotometer at 663nm and 645nm respectively. The 80% acetone served as reference blank. The total chlorophyll content was calculated using the formula:

115 Total chlorophyll content= $\{20.2 \text{ x } D_{645} + 8.02 \text{ x } D_{663}\} \text{ x } \{(50/100 \text{ x } (100/5) \text{ x} 1/2)\}$

117 2.6 Fresh and dry weight determination

Plants were carefully uprooted at the end of experiment by soaking the soil with water for easy uprooting without any damage to the root. The roots were washed and each plant was separated into stems, roots and leaves. The parts were taken to the laboratory for weighing to determine the fresh weight. Dry weight was also determined after drying in an oven at 80°C to constant weight. The weight was measured using sensitive weighing balance. Also, fruits of plant were harvested and weighed in the laboratory to determine the fresh weight. The fruits dry weight was determined after drying in an oven at 80°C.

126 2.7 Plant Analysis

Dried seeds of *Abelmoschus esculentues* and dried leaves and stems of *Telfairia occidentalis* were ashed and dissolved with 10ml of 20% sulphuric acid. The Solution was placed on a hot plate preset at 30^oC to falcilitate its dissolution. It was removed from hot plate, filtered into 10ml capacity volumentric flask and marked to volume with distilled water. This filtrate was used for Ca2+, Mg2+, Na+, K+. Na+ and K+ analysis by flame photometer, and Mg2 and Ca2+ analysis by EDTA. Seeds and leaves were also assayed for proximate compositions: crude protein, fat and carbohydrate, crude fiber and total ash following the method of AOAC.

135 2.7.1 Sodium and potassium determination

A 2.54g and 1.9067g of pure oven dried NaCl and KCl respectively were dissolved in water to make 11itre. 10ml each of the reagent was then pipette to make 100ml with ammonia acetate to have 100ppm solution from the 100ppm, 0,2,4,6,8 and 10ml were pipette into 100ml flask and made each to mark, to have 0,2,4,6,8,10ppm working standard for two stocks. The ammonium acetate extract was employed for the determination. The flame photometer was adjusted according its instruction manual. The standards were aspirated to obtain reliable curves before aspirating the samples. The blank (ppm) is the ammonium acetate.

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145 2.7.2 Calcium and Magnesium determination

A 10ml aliquot of the plant samples was carefully transferred in duplicate into 150ml conical flask capacity. A drop of potassium cyanide and hydrochloride were introduced to both beakers. To one set of the beaker, 5ml of ammonium buffer, three drops of erichrome black, and indicator were added and titrated against 0.05M EDTA solution. The titre value obtained gave a combine result for calcium and magnesium while to the second set of beaker; 5ml of 8M of NaOH and a pinch of cal-red indicator were added and titrated against 0.05mDTA solution. The titre value obtained gave directly 152 the result of calcium concentration. The difference between the first and the second value obtained 153 gave the amount of magnesium.

155 2.7.3 Determination of crude protein

The estimation of crude protein involves the determination of total nitrogen usually by kjedahl procedure. The amount of crude protein is obtained by multiplying the nitrogen content by 6.25. This factor is based on the assumption that all the nitrogen in the tissue is present as protein.

160 2.7.4 Crude fibre determination

161 About 0.50-2.0g of the milled sample was measured into 1 litre conical flask (W_0). 200ml of 162 boiling 1.25% H₂SO₄ was added and boiled gently for 30 minutes using cooling fingers to maintain a 163 constant volume, This was filtered through muslin cloth or poplin material stretched over 9cm 164 Buchner funnel. It was rinse well with hot distilled water added and the material was scraped back into flask with spatula. About 20ml of boiling 1.25% NaOH was added and boiled gently for 30 165 166 minutes using cooling fingers to maintain a constant volume. This was filtered through poplin cloth 167 and residues washes with10% HCl and twice with industrial methylated spirit, acetone or ethanol. 168 This was allowed to drain, dry and the residue scraped into a crucible or silica dish. This was dried overnight at 105° C in the oven and then cooled in a desiccator. The sample was weighed (W), ashed at 169 170 550° C for 90 minutes in a muffle furnace, cooled in a desiccators and weighed again (W₂).

172 Calculation

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173 % crude fibre = $W_1 - W_2 \times 100$

175 2.7.5 Determination of soluble carbohydrate (nitrogen free extract)

The nitrogen- free extractive (NFE) referred to as soluble carbohydrate is not determined directly but obtained as a difference between crude protein and the sum of ash, protein, crude fat and crude fibre.

179 NFE= 100-(%ash + % crude fibre +% crude fat + % crude protein)

 W_0

181 2.8 Statistical analysis

The data obtained were subjected to one-way analysis of variance (ANOVA) and means were
 separated with Tukey HSD Multiple Range tests at 5% level of probability using SPSS 21.0.

185 **3. Results**

Table 1. shows the effect of beauty salon wastewater on the yield of *Abelmoschus esculentus*. Beauty salon wastewater at 25-75% concentrations increased the yield of *Abelmoschus esculentus*. But the yield reduced at 100% concentration in comparison with the control. Plants irrigated with 75% concentration of the wastewater yielded an average of 4.00 fruits per plant compared to average of 3.85 fruits per plant in control and average of 3.67 fruits per plant in higher concentration of 100% of the wastewater. Similarly, the fruits fresh and dry weight increased at 75% concentration and reduced at 100% concentration of the wastewater in comparison with the control.

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200 Table 1. Yield of Abelmoschus esculentus (Okra) grown in soil irrigated with water

201 containing different proportions of beauty salon wastewater under screen house condition

Yield	Quantity of beauty salon wastewater applied (%)					
parameters	0	25	50	75	100	
Number of fruits/plant	3.85 ^a	4.00 ^a	3.87 ^a	4.00 ^a	3.67 ^a	
Fruit fresh weight/plant (g)	31.64 ^a	36.41 ^a	34.04 ^a	41.48 ^a	31.56 ⁸	
Fruit dry weight/plant (g)	4.31 ^a	4.76 ^a	4.66ª	6.36 ^a	3.81 ^a	

202 Each value is a mean± S.E of 6 replicates. For each value, means with the same letter(s) in superscript

203 on the same row are not significantly different at P≥0.05 (Tukey HSD test)

204 This present experiment shows significant effects of beauty salon wastewater on fresh and dry 205 weight of Plants parts. Table 2 and 3. illustrate the impact of beauty salon wastewater on the fresh and dry weights of roots, stems and leaves of Abelmoschus esculentus and Telfairia occidentalis irrigated 206 207 with beauty salon wastewater. Fresh and dry weights parts of Abelmoschus esculentus increased with 208 a significant different (p>0.05) at 25-75% concentrations of the beauty salon wastewater but 209 decreased at highest concentrations of 100% when compared with the control(Table 2). Similarly, the 210 fresh weights of the different parts of Telfairia occidentalis increased at 25-75% concentrations of 211 beauty salon wastewater compared with the control (Table 3).

Table 2. Effect of beauty salon wastewater on the fresh and dry weight (g) of plant parts in
 Abelmoschus esculentus.

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Parameter	Plant part	Concentration of beauty salon wastewater applied(%)							
		0	25	50	75	100			
Fresh	Root fresh weight	12.53 ^a	15.10 ^b	13.63 ^b	13.85 ^b	11.41 ^a			
weight (g)	Leaf fresh weight	5.62 ^{ab}	5.56 ^{ab}	9.08 ^b	5.82 ^{ab}	3.56 ^a			
	Stem fresh weight	11.67 ^a	14.90 ^a	13.23 ^a	15.85 ^a	8.72 ^a			
	Root dry weight	2.49 ^{ab}	3.29°	2.70 ^{abc}	2.88 ^{bc}	2.03 ^a			
Dry weight (g)	Leaf dry weight	1.38 ^a	1.41 ^a	2.49 ^b	1.44 ^a	0.98 ^a			
	Stem dry weight	2.26 ^a	2.60 ^a	2.58 ^a	2.99 ^a	1.59 ^a			

Comment [ASP1]: Table format is differs.....

215 Each value is a mean± S.E of 6 replicates. For each value, means with the same letter(s) in superscript

216 on the same row are not significantly different at $P \ge 0.05$ (Tukey HSD test)

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Table 3. Effect of beauty salon wastewater on the fresh and dry weight (g) of plant parts in *Telfairia occidentalis*

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Parameter	Plant part	Concentration of beauty salon wastewater applied (%)						
		0	<mark>25</mark>	<mark>50</mark>	<mark>75</mark>	<mark>100</mark>		
	Root fresh weight	18.80 ^{ab}	18.96 ^{ab}	19.91 ^b	<mark>26.55°</mark>	14.83 ^a		
Fresh weight (g)	Leaf fresh weight	13.75 ^a	13.85 ^a	13.97 ^a	13.32ª	12.73 ^a		
	Stem fresh weight	14.20 ^a	<mark>16.93a</mark>	15.27 ^a	19.68 ^b	13.53ª		
	Root dry weight	<mark>2.92^ª</mark>	<mark>4.17^a</mark>	<mark>3.52^a</mark>	5.68 ^b	2.69 ^ª		
<mark>Dry weight</mark> (g)	Leaf dry weight	2.20 ^ª	2.45 ^ª	<mark>2.46^a</mark>	2.42ª	1.88ª		
	Stem dry weight	<mark>2.41^ª</mark>	3.23 ^{ab}	2.73 ^{ab}	3.75 ^b	2.22ª		

Comment [ASP2]: Table format is different

Each value is a mean \pm S.E of 6 replicates. For each value, means with the same letter(s) in superscript on the same row are not significantly different at P \ge 0.05 (Tukey HSD test)

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224	Table 4. shows the effect of beauty salon wastewater on the total biomass of the two
225	vegetables. The total biomass is the total sum of the root and shoot dry mass. The total biomass
226	increased upon irrigation with beauty salon waste water in comparison with the control. (Also see
227	Figures 1 and 2).

Table 4. Dry mass, Root: shoot ratio of *Abelmoschus esculentus* and *Telfairiaoccidentalis* grown in
 soil irrigated with beauty salon wastewater.

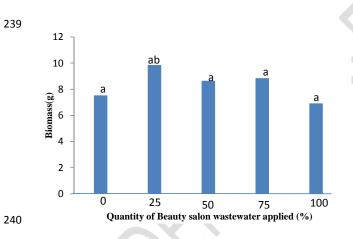
	Vegetable species	Concentration of beauty salon wastewater applied (%)						
		0	25	50	75	100		
Root dry mass (g)	Telfairiaoccidentalis	2.92 ^a	4.17 ^a	3.52 ^a	5.68 ^a	2.69 ^a		
	Abelmoschusesculentus	2.49 ^{ab}	3.29 ^c	2.70 ^{abc}	2.88 ^{bc}	2.03 ^a		
Shoot dry mass (g)	Telfairiaoccidentalis	4.3 ^b	5.06 ^a	5.75 ^a	6.17 ^a	6.63 ^a		
	Abelmoschusesculentus	3.3 ^{bc}	4.13 ^b	5.86 ^b	7.86 ^a	7.73 ^a		
Total biomass (g)	Telfairiaoccidentalis	6.43 ^b	7.29 ^{ab}	8.08 ^{ab}	8.53 ^a	9.19 ^a		
	Abelmoschusesculentus	5.8°	7.03 ^{ab}	8.89 ^{ab}	10.99 ^a	9.59ª		

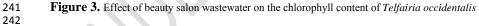
Each value is a mean± S.E of 6 replicates. For each value, means with the same letter(s) in superscript

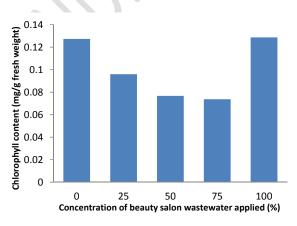
231 on the same row are not significantly different at P≥0.05 (Tukey HSD test)



238 FIGURE 2. Effect of beauty salon wastewater on the biomass of *Telfairia occidentalis*.



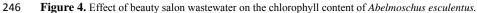






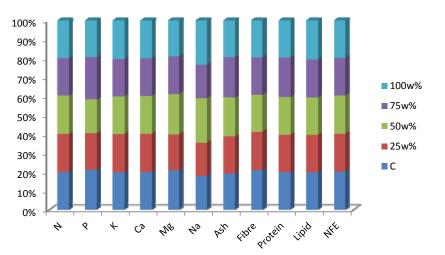
234 FIGURE 1. Effect of beauty salon wastewater on biomass of Abelmoschus esculentus.





Beauty salon wastewater at all treatment levels(25-100%) caused an increase in the nutrient composition of the fruits of *Abelmoschus esculentus* and leaves of *Telfairia occidentalis* when compared with the control. N, K, Ca and Na composition of the two vegetables increased in comparison with the control. The result also shows increase in the percentage ash and protein content of the plants. Lipid content increased while the fibre content decreased in *T. occidentalis* whereas in *A. esculentus* fibre content increased while the lipid content decreased in comparison to the control.

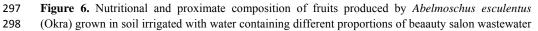
Figure 5: Nutritional and proximate composition of leaves produced by *Telfairia occidentalis* (Fluted
 pumpkin)grown in soil irrigated with water containing different proportions of beauty salon
 wastewater under screen house condition



267 268 269 **Table 5:** proximate and nutritional composition of *Telfairia occidentalis*

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uantity of B	eauty Salon wa	stewater applie	d (%)	270
0	25	50		75 271
				272
				273
1.64	1.66	1.67	1.65	^{1.65} 274
0.13	0.12	0.11	0.14	0.12
0.85	0.88	0.87	0.86	275 0.89
0.67	0.69	0.68	0.68	0.2 3 6
0.28	0.26	0.29	0.27	0 2%7
0.03	0.03	0.04	0.03	^{0.04} 278
4.43	4.57	4.75	5.00	4.50
9.37	9.10	8.78	8.88	279 8.86
10.25	10.38	10.44	10.86	1280
1.56	1.59	1.57	1.61	2851
74.39	74.36	74.35	73.95	73.98 282
	0 1.64 0.13 0.85 0.67 0.28 0.03 4.43 9.37 10.25 1.56	0 25 1.64 1.66 0.13 0.12 0.85 0.88 0.67 0.69 0.28 0.26 0.03 0.03 4.43 4.57 9.37 9.10 10.25 10.38 1.56 1.59	0 25 50 1.64 1.66 1.67 0.13 0.12 0.11 0.85 0.88 0.87 0.67 0.69 0.68 0.28 0.26 0.29 0.03 0.03 0.04 4.43 4.57 4.75 9.37 9.10 8.78 10.25 10.38 10.44 1.56 1.59 1.57	1.64 1.66 1.67 1.65 0.13 0.12 0.11 0.14 0.85 0.88 0.87 0.86 0.67 0.69 0.68 0.68 0.28 0.26 0.29 0.27 0.03 0.03 0.04 0.03 4.43 4.57 4.75 5.00 9.37 9.10 8.78 8.88 10.25 10.38 10.44 10.86 1.56 1.59 1.57 1.61



299 under screen house condition

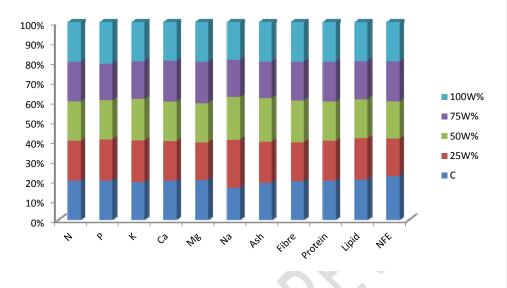




Table 6: Proximate and nutritional composition of Abelmoschus esculentus

	Concentratio	n of Dooute: Co	lon wostoweter	r applied (9/)		306
	Concentratio	n of Beauty Sa	lon wastewater	r applied (%)		307
Nutritional and	0	25	50	75	100	308
Nutritional and	0	25	50	15	100	309
Proximate						310
Composition						311
Composition						312
N (%)	2.46	2.52	2.49	2.50	2.51	313 314
P (%)	0.17	0.18	0.17	0.16	0.18	315
17 (0/)	0.01	1.02	1.01	0.02	0.05	316
K (%)	0.91	1.02	1.01	0.92	0.95	317
Ca (%)	0.86	0.87	0.87	0.90	0.85	318
$M = \langle 0 \rangle$	0.22	0.21	0.22	0.24	0.22	319
Mg (%)	0.33	0.31	0.33	0.34	0.33	320
Na (%)	0.06	0.09	0.08	0.07	0.07	321 322
Ash (%)	7.90	8.96	9.45	7.86	8.50	323
						324
Fibre (%)	17.74	17.94	19.27	17.86	18.19	325
Protein (%)	15.38	15.75	15.56	15.63	15.69	326
T::::1(0/)	11.22	11.74	11.07	10.05	10.07	327
Lipid (%)	11.33	11.74	11.07	10.85	10.97	328
NFE (%)	52.35	44.61	44.65	47.80	46.65	329 330
						331
						221

337 4. Discussion

338 The effect of wastewater on yield and nutritional value of crops has been demonstrated and 339 discussed extensively by many authors. (Day et al., 1981) reported that using municipal wastewater 340 diluted with groundwater at 50:50 mixtures improved Gossypium spp yield when compared to 341 groundwater alone from wells in Arizona. Kiziloglu et al. (2008) showed that wastewater irrigation 342 treatments increased the availability of N, P, K, Ca, Mg, Na, Fe, Mn, Zn and Cu, to plant which led to increase of red cabbage yields. Gatta et al. (2015) observed that the source of irrigation water did 343 344 not affect significantly tomato yield traits except tomato quality. Results of this experiment show the 345 effect of beauty salon wastewater on the yield and nutritional value of Telfairia occidentalis(Fluted 346 Pumpkin) and Abelmoschus esculentus (Okra).

347 Table 1. shows that plant irrigated with 25-75% concentrations of beauty salon wastewater 348 had higher yield than the control. This concur with the previous finding of Bedbabis et al. (2015) that 349 found that wastewater irrigation of olive trees resulted in significant yield increase when compared to 350 yields from plot using well water. Qaryouti et al. (2015) concluded that, raw wastewater irrigation 351 increased significantly tomato crop parameters, cucumber plant height and fruit yield and average 352 fruit weight, and tomato leaf area and plant dry weight. Day et al. (1974) also compared the effect of 353 irrigation with wastewater and pump water on wheat. They concluded that wastewater irrigation 354 produced taller plants, heavier seeds and higher grain yields than pump water. Wastewater has the 355 potential to increase plant yield than the control. Similar results were also recorded by Juwarkar et al. 356 (1990) in Arachis hypogea. Nissim et al. (2015), showed that, irrigation with wastewater had a 357 positive effect on willow growth and biomass yield. Jiménez et al. (1999) concluded that, reuse wastewater increased significantly crop productivity to five crops/year of alfalfa, fodder oats, tomato, 358 359 barley and maize and the yield was higher than those obtained with rain. Golchin et al. (2013) indicated that use of wastewater could improve morphological characters, yield and yield components 360 361 of alfalfa as compared to control treatment. Increasing wastewater concentration more than 45 % 362 caused poisoning effects on plants which decreased biological yield.

363 Higher concentration of beauty salon wastewater decreased the dry weight of the root, stem 364 and leaf of A. esculentus and T. occidentalis. The reduction in the dry weight might be due to the poor growth of the seedlings under effluent stress. According to El Youssfi et al. (2012) studied the effect 365 of wastewater irrigation on three varieties of quinoa. The salinity caused the depression of plant's 366 height, and reduced fresh and dry weights of different parts of three varieties of plants tested. The 367 plant biomass of the two vegetables increased at 25-75% treatment levels of the wastewater. It was 368 369 reported by Misra et al. (2009) that Solanum lycoperscum irrigated with greywater obtained higher 370 nutrient uptake and biomass at the flowering stage when compared to tap water. Also, Gupta et al. 371 (2015) reported that plant irrigated with wastewater resulted in significant increase in plant height, 372 number of leaves per plant, leaf area index, leaf to stem (green and dry) biomass and green fodder 373 yield of fodder sorghum and significant decrease in dry matter content as compared to well water. 374 Zema et al. (2012) investigated the biomass yield of T. latifolia which increased by irrigation with 375 wastewater. Ntzala et al. (2015) found that the treated wastewater affected significantly the dry matter 376 yield and non-significantly the plant height on Lactuca sativa L. crop.

377 The proximate analysis of Telfairia occidentalis shows that Beauty salon wastewater 378 increased leaf N, K, and Ca while other nutrients were not affected. Similarly, percentage ash, lipid 379 and protein increased in the leaves of beauty salon waste treated plants (Figure 3). This finding is in 380 accordance with Babyshakila et al. (2009) that biochemical content of lipid, ash and protein increased 381 at 50 and 75% concentrations of wastewater in the leaf samples of Vigna radiate. The Fibre and 382 carbohydrate contents decreased relative to the control. The proximate analysis of Abelmoschus esculentus shows that Beauty salon wastewater increased the composition of N, K, Ca and Na at all 383 treatment levels in the fruits of Abelmoschus esculentus in comparison to the control (Figure 4). Al-384 385 Jaloud et al. (1995) reported elevated concentration of N, Ca, Mg, and Na in leaves of Sorghum when 386 the crop was irrigated with wastewater. Moreover, Vazquez-Montieletal.(1996) found that irrigation

of maize (Zea mays. L.) with treated wastewater resulted in increase in N, P, K and Mg concentration
in leaves. Fonseca *et al.*(2005a) also obtained similar results in a greenhouse experiment with maize.
Also, there was an increase in the percentage ash, fibre and protein at all treatment levels whereas the
lipid and carbohydrate contents decreased in comparison to the control.

391 Figure (3 and 4) show the effects of beauty salon wastewater on the chlorophyll content of T. 392 occidentalis and A. esculentus. Beauty salon wastewater treatments reduced the chlorophyll content of 393 Abelmoschus esculentus but at non-significant level when compared to the control. The Chlorophyll 394 content of Telfairia occidentalis reduced significantly with a significance difference at 25-75% 395 concentrations when compared to the control upon treatment with beauty salon wastewater. Sahai and 396 Singh (1977), Benergi and Kumar (1979) and Singh and Srivastava (1984) have reported adverse 397 effect of wastewater on chlorophyll content and metabolism. Agrawal et al. (1961), suggested that 398 heavy metals can inhibit chlorophyll formation by preventing magnesium uptake.

400 5. Conclusion

401 The use of beauty salon wastewater improved yields and nutritional values of A. esculentus 402 and T. occidentals when diluted with water at 25-75%. Consequently, beauty salon wastewater can 403 serve as an alternative liquid fertilizer in the production of A. esculentus and T. occidentalis if applied 404 to soil at levels not above 75% concentration of the wastewater. This study cofirms that high 405 concentration (>75% concentration) of beauty salon wastewater can have negative impact on soil 406 make it unfavourable for plant yields. Therefore, wastewater should not be used directly on crops 407 without sufficient treatment or dilution with water. There is however the need for furthe evaluation or 408 assessment of environmental health challenges associated with beauty salon waste disposal.

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410 6. Reference

- Raschid-Sally, L. and Jayakody, P. (2008). Drivers and characteristics of wastewater agriculture in
 developing countries: Results from a global assessment, Colombo, Sri Lanka. *IWMI Research Report, International Water Management Institute, Colombo.* 127.
- Farahat, E. and Linderholm, H.W. (2015). The effect of long-term wastewater irrigation on
 accumulation and transfer of heavy metals in Cupressus sempervirens leaves and adjacent
 soils. Science of the Total Environment, Vol: 512–513, 1-7.
- Heidarpour, M. Mostafazadeh-Fard, B. Abedi Koupai, J. Malekian, R. (2007). The effects of treated
 wastewater on soil chemical properties using subsurface and surface irrigation methods. *Agric. Water Manag.* 90:87-94.
- Singh, P.K. Deshbhratar, P.B. Ramteke, D.S. (2011). Effects of sewage wastewater irrigation on soil
 properties, crop yield and environment. *Agric. Water Manag.* 103:100–104.
- Rusan, M.J. Hinnawi, S. Rousan, L. (2007). Long term effect of wastewater irrigation of forage crops
 on soil and plant quality parameters. *Desalination*, 215:143-152.
- Ghanbari, A. Abedikoupai, J. TaieSemiromi, J. (2007). Effect of municipal wastewater irrigation on
 yield and quality of wheat and some soil properties in sistan zone. J. Sci. Technol. Agric. *Natural Recou.* 10:59-74.
- Mohammad, M.J. and Ayadi, M. (2004). Forage yield and nutrient uptake as influenced by Secondary
 treated wastewater. *Journal of Plant Nutrition*, 27(2):351-365.
- 429 Kiziloglu, F.M. Turan, M. Sahin, U. Angin, I. Anapali, O. and Okuroglu, M. (2007). Effect of
 430 wastewater irrigation on soil and cabbage-plant (*Brassica olerecea*) chemical properties.
 431 *Journal of Plant Nutrition and Soil Science*, 170: 166-172.
- 432 Siebe, C. (1998). Nutrient inputs to soils and their uptake by alfalfa through long-term irrigation with
 433 untreated sewage effluent in Mexico. *Soil Use Manage*. 13, 1-5
- Aghtape, A. Ghanbari, A. Sirousmehr, A. Siahsar, B. Asgharipour, M. Tavssol, A. (2011). Effect of
 irrigation with wastewater and foliar fertilizer application on some forage characteristics of

- foxtail millet (Setaria italica), *International Journal of Plant Physiology and Biochemistry*.
 3(3), pp. 34-42.
- Abu Nada Ziyad. (2009). Long term impact of wastewater irrigation on soil and crop quality
 parameters in Gaza strip. *Master Thesis, Islamic University of Gaza*.
- Khurana, M.P.S. Singh, P. (2012). Waste Water Use in Crop Production: A Review. *Resources and Environment.* 2(4): 116-131.
- Nadav, I. Tarchitzky, J. Chen, Y. (2013). Water repellency induced by organic matter (OM) in treated
 wastewater (TWW) infiltration ponds and irrigation. In: Jianming, X., Jianjun, W., Yan, H.
 (Eds.), Functions of Natural Organic Matter in Changing Environment. Springer, Netherlands,
 pp. 883–887.
- Zavadil, J. (2009). The effect of municipal wastewater irrigation on the yield and quality of
 vegetables and crops. *Soil and Water Res*ource, 4 (3): 91–103.
- Ogle, B.M. Johansson, M. and Tuyet, L. (2001). Evaluation of the significance of dietary foliage from
 wild vegetables in Vietnam. *Asia pacific Journal of clinical nutrition*, pp216-221
- Arai, S. (2002). Global view on functional foods: Asian perspectives. British Journal of Nutrition, 88:
 139-143.
- Arnon, D.T. (1949). Copper enzymes in isolated chloroplast: polyphenol oxidase in *Beta vulgaris*.
 Plant Physiology. 24: 1-15.
- Day, A.D. McFadyen, J.A. Tucker, T.C. and Cluff, C.B. (1981). Effects of municipal wastewater on
 the yield and quality of cotton. *Journal of Environmental Quality*, 10:47-49.
- 456 Kiziloglu, F.M. Turan, M. Sahin, U. Kuslu, Y. and Dursun, A. (2008). Effects of untreated and
 457 treated wastewater irrigation on some chemical properties of cauliflower (*Brassica olerecea*)
 458 grown on calcareous soil in Turkey. *Agricultural Water Management*, 95: 716-24.
- Gatta, G. Libutti, A. Gagliardi, A. Beneduce, L. Brusetti, L. Borruso, L. Disciglio, G. Tarantino, E.
 (2015). Treated agro-industrial wastewater irrigation of tomato crop: Effects on qualitative/quantitative characteristics of production and microbiological properties of the soil. Agricultural Water Management, Vol: 149, 33-43.
- Bedbabis, S. Trigui, D. Ahmed, C.B. Clodoveo, M.L. Camposeo, S. Vivaldi, G.A. Rouina, B.B.
 (2015). Long-terms effects of irrigation with treated municipal wastewater on soil, yield and
 olive oil quality. *Agricultural Water Management*, Vol: 160, 14-21.
- Qaryouti, M. Bani-Hani, N. Abu-Sharar, T.M. Shnikat, I. Hiari, M. Radiadeh, M. (2015). Effect of
 using raw waste water from food industry on soil fertility, cucumber and tomato growth, yield
 and fruit quality. *Scientia Horticulturae*. 193, 99–104.
- 469 Day, A.D. Rahman, A. Katterman, F.R. and Jensen, V. (1974). Effects of treated municipal
 470 wastewater and commercial fertilizer on growth, fibre, acid-soluble nucleotides, protein and
 471 amino acid content in wheat hay. *Journal of Enivonmental Quality*, 3: 17-19.
- Juwarkar, A.S. Juwarkar, A. Deshbratar, P.B. and Bal, A.S. (1990). "Exploration of nutrient
 potential of domestic and sludge through and land application". *RAPA Report*, pp178-201.
- Nissim, W.G. Jerbi, A. Lafleur, B. Fluet, R. Labrecque, M. (20150. Willows for the treatment of municipal wastewater: Performance under different irrigation rates. *Ecological Engineering*, Vol: 81, 395-404.
- Jiménez, B. Chávez, A. Hernández, C. (1999). Alternative Wastewater Treatment Intended for
 Agricultural Use. *Water Science and Technology*. 40(4-5) 355-362.
- Golchin, L. salmasi, S.Z. Shafagh-kolvanagh, J. Shahin oustan, Shokati, B, Hashemi-Amidi, N.
 Haghverdi, H. (2013). Effects of irrigation times and wastewater concentration of a leaven
 producing factory (Iran Mayeh) on some morphological characters of alfalfa. *Intl J Agri Crop Sci.* Vol., 5 (23), 2831-2836.
- El Youssfi, L. Choukr-Allah, R. Zaafrani, M. Mediouni, T. Ba Samba, M. Hirich, A. (2012). Effect of Domestic Treated Wastewater use on Three Varieties of Quinoa (Chenopodium quinoa) under Semi Arid Conditions. *International Science* Index Vol:6, No:8, 116-119.
 waset.org/Publication/8903.

- Misra, R.K. Patel, J.H. and Baxi, V.R. (2009). Removal of pollutants by tomato plants duringre-use of
 laundry greywater for irrigation. *International Conference on Food Security and Environmental Sustainibility (FSES)*, 6(2): 407-409.
- Gupta, S.P. Gajender, Yadav, R.K. Magan, S. Koushik, P. (2015). Effect of irrigation schedules of
 domestic waste water on growth and yield of fodder sorghum. *Indian Journal of Small Ruminants.* Vol.: 21:2, 257-263. Article DOI: 10.5958/0973-9718.2015.00073.2.
- Zema, D.A. Bombino, G. Andiloro, S. Zimbone, S.M. (2012). Irrigation of energy crops with urban
 wastewater: Effects on biomass yields, soils and heating values. Agricultural Water
 Management, 115, 55-65.
- 496 Ntzala, G. Kalavrouziotis, I.K. Koukoulakis, P.H. Papadopoulos, A.H. (2015). Impact of sludge and
 497 wastewater on Lactuca sativa L. Growth and on soil pollution. *Global Nest Journal*,17
 498 (1):148-161.
- Babyshakila, P. and Usha, K. (2009). Effect of diluted effluents on soil properties and plant growth. *Journal of Advanced studies in Biology*, 1(8): 391-398.

501 Al-Jaloud, A.A., Hussian, G., Al-Saati, A.J. and Karimulla, S. (1995). Effect of wastewater

- irrigation on mineral composition of corn and sorghum plants in a pot experiment. *Journal of Plant Nutrition*, 18: 1677-1692.
- Fonseca, A.F. Melfi, A.J. and Montes, C.R. (2005). Maize growth and changes in soil fertility after
 irrigation with treated sewage effluent, plant dry matter yield and soil nitrogen and
 phosphorus availability. *Journal of Soil Science*, 36: 1965-1981.
- Sahai, R. and Singh, S.P. (1977). Effect of domestic waste on the growth performance of
 Ceratophyllum demersum Linn. *Indian J. Ecol*, 4: 118-120.
- Banergi, D. and Kumar, N. (1979). The twin effect of growth and heavy metal accumulation in certain
 crop plants by polluted irrigation water. *Indian J. Ecol.*, 6: 82-87.
- 511 Singh, A. and Srivastava, O. N. (1984). Effect of pesticides on the growth of *Azolla pinnata R*.
 512 Brown. *Indian J. Ecol.*, 11: 12-14.
- Agarwal, S.C. Kumar, A. and Sharma, O.P. (1961). Effect of excess supply of heavy metals on barley
 during germination with special reference to catalase and peroxidase. Nature, 191: 720-727.
- 515 516