Original Research Article

In-Silico Structural Annotation of Hypothetical Protein Zm00014a_031618 (Zea mays)

ABSTRACT

Aims: The aim of this study is to use *In-Silico* studies for structural annotation of hypothetical Protein Zm00014a_031618 (*Zea mays*) retrieved from NCBI.

Study design: The use of bioinformatics *In-Silico* studies for the structural annotation of a hypothetical protein.

Place and Duration of Study: The research was conducted at the Bioinformatics Laboratory, Chevron Biotechnology Centre, Modibbo Adama University of Yola, Nigeria. Between June 2018 to July 2018.

Methodology: The hypothetical protein was retrieved from NCBI, physical and chemical parameters was calculated using expasy's ProtParam server, the server SOPMA was used for secondary structure analysis (helix, sheets, and coils) and I-Tasser I was use to obtain the 3D structure.

Results: Expasy's ProtParam server computated the various physical and chemical parameters such as molecular weight (MW) 30117.97, total number of positively (+R) 27, negatively charged residues (-R) 30, theoretical isoelectric point (pl) 5.96, aliphatic index (AI) 103.67 and grand average hydropathy (GRAVY) 0.293. The SOPMA server was used for calculating the secondary structural features of protein sequences as Alpha helix 39.16%, 310 helix 0.00%, Pi helix 0.00%, Beta bridge 0.00%, Extended strand 14.69%, Beta turn 6.64%, Bend region 0.00%, Random coil 39.51%, Ambiguous states 0.00% and Other states 0.00%. I-Tasser was used for predicting the 3D structure 2qttA from PDB as template.

Conclusion: This study has help in understanding the structural analysis of the hypothetical Protein Zm00014a_031618 (Zea mays)

Keywords: [Zea mays, Hypothetical protein, structural annotation and Intraspecific }

1. INTRODUCTION

Zea mays for more than hundreds years has been a subject of genetics studies (1). It is one of the most extensively studied plant species in genetics and it is also been use as research model for genome evolution and genetic diversity (2,3). The genome is made up of 10 chromosomes with its size approximately 2.3 to 2.7 Gb and the genome is also diploid genetically (4,5,6,7,). Just like other larger genome of plant species, the *Z. mays* genome is typically made up of nongenic, low-copy DNA that harbor single genes or repetitive fraction punctuated by islands of unique. The repetitive elements is highly responsible for the wide range of diversity within the species which includes ribosomal DNA (rDNA), transposable elements (TEs) and high-copy short-tandem repeats mostly present at the centromeres, telomeres, and heterochromatin knobs (8,9,10,11). *Z. mays* plant has an extraordinary levels of genomic diversity, phenotypic (12) and transcriptomic (13,14,15). Looking at the genomic level *Z. mays* exhibits a high level of INDEL Polymorphisms (16) and (17) and Single Nucleotide Polymorphisms (18). Averagely the frequency of single nucleotide polymorphism (SNP) between two maize inbreds is said to be approximately 1 substitution per 100 bases (19,20). Recent studies using sequencing data have shown that maize genome exhibits rather variable levels of naturally occurring genetic diversity which depends on the lines involved in the comparison (21,22).

Intraspecific genome variation has been long attributed to changes in size of heterochromatic DNA outside coding sequences that expanded and contracted the chromosomes (98). Intraspecific variations which are approximately 38.8% from the average of 5.5 pg/2n nucleus have been reported in Zea mays (23,24,25,26,27,28). *Z. mays* is known to have large amount of intraspecific sequence variation (29,30) in form of deletion/insertion and single nucleotide polymorphisms. The main mechanism which have effect in the generation of intraspecific genome diversity and in the evolution of the maize genome, segmental duplications and whole genome duplications (polyploidization), retrotransposition and DNA transposition, expansion/contraction of simple sequence repeats (SSRs) and single base mutations and translocation of genes or gene segments by transposons and capture (31,32). Intraspecific allelic variation is mostly as a result of qualitative changes that change the nature of the gene products and quantitative changes it also alter the amount of the gene product produced. Quantitative changes in gene expression can be as a result of cis- or trans variations in gene regulation (33).

The present study focused on the *In Silico* Structural Annotation of Hypothetical Protein Zm00014a_031618 (*Zea mays*)

2. MATERIAL AND METHODS

2.1 Sequence Retrieval

Amino acid sequence of hypothetical Protein Zm00014a_031618 (Zea mays) was retrieved from NCBI database (www.ncbi.nlm.nih.gov/protein/1394916517).

2.2 Physiochemical Analysis of the Protein

The physiochemical properties of the hypothetical protein such as molecular weight, atomic composition, amino acid composition, theoretical pI, instability index, and grand average of hydrophobicity (GRAVY) using ProtParam tool (web.expasy.org/cgi-bin/protparam/protparam) (34).

2.3 Secondary Structure Analysis

The server SOPMA was use for secondary structure analysis (helix, sheets, and coils) of the hypothetical protein using SOPMA (https://npsa-prabi.ibcp.fr/cgi-bin/npsa automat.pl?page=npsa sopma.html) (35).

2.4 Sequence Analysis

BLASTP is mostly used for the identification of a query amino acid sequence and to find similar sequences in protein databases. The FASTA sequence of the Hypothetical Protein Zm00014a_031618 (*Zea mays*) was the query sequence and similar proteins in different databases were searched for using the BLASTP program (36).

2.5 3D Structural Prediction

The 3D structural was predicted with the use of I-Tasser (37)

3. RESULTS AND DISCUSSION

The present study focused on the *In Silico* Structural Annotation of a hypothetical Protein Zm00014a 031618 (*Zea mays*) from NCBI database.

The presented in table 1 showed the physicochemical characterization of Hypothetical Protein Zm00014a_031618 (Zea mays) with 286 amino acid sequence using Expasy's ProtParam server where the computation of various physical and chemical parameters was performed. Molecular weight (MW), total number of positively (+R), negatively charged residues (-R), theoretical isoelectric point (pl), extinction coefficient (EC), aliphatic index (Al) and grand average hydropathy (GRAVY).

Table 1: Physiochemical Analysis of the Hypothetical Protein

Molecular weight (Da)	pl	-R	+R	EC	II	Al	GRAVY
30117.97	5.96	30	27	13200	23.10	103.67	0.293

The results are presented in Table 2 showed the SOPMA which was used for calculating the secondary structural features of protein sequences such as Alpha helix, 310 helix, Pi helix, Beta bridge, Extended strand, Beta turn, Bend region, Random coil, Ambiguous states and Other states.

Table 2. SOPMA of the Hypothetical Protein

	97
	98
	99
1	00
1	01
1	02
1	03
1	04

Parameter	% content	Parameter	% content	
A lock on the Elec-	00.400/	Data tom	0.040/	
Alpha helix	39.16%	Beta turn	6.64%	
310 helix	0.00%	Bend region	0.00%	
Pi helix	0.00%	Random coil	39.51%	
Beta bridge	0.00%	Ambiguous states	0.00%	
Extended strand	14.69%	Other states	0.00%	

>PWZ58979.1 hypothetical protein Zm00014a_031618 [Zea mays]
MAAEAGPISKVLIVVGNPTPCCSLRPKALLCSVSRFAYSVGIGLCSGLDAAMQTEAMPLVHKFKLVEAPA
HESTFPKGAPWVRYHGNYKGLHIDLVLPGKDAVLGVDSVGTVSAALLTSFSIQTLKPDLIINAGTAGGFK
AKGASIGDVFLASDVSFHDRRIPIPVFDMYGIGARKTSAVPNILKELNLKIGKLSTGDSLDMSPQDEKVI
LSNDATVKDMEGAAVAYVADMFSTPAIFVKAVTDIVDGEKPTSEEFLQNLIAVTAALDLAVTKVVDFISG
KRISDL

Fig. 1. Faster sequence of the hypothetical Protein Zm00014a 031618 (Zea mays)

MAAEAGPISKVLIVVGNPTPCCSLRPKALLCSVSRFAYSVGIGLCSGLDAAMQTEAMPLVHKFKLVEAPA HESTFPKGAPWVRYHGNYKGLHIDLVLPGKDAVLGVDSVGTVSAALLTSFSIQTLKPDLIINAGTAGGFK AKGASIGDVFLASDVSFHDRRIPIPVFDMYGIGARKTSAVPNILKELNLKIGKLSTGDSLDMSPQDEKVI LSNDATVKDMEGAAVAYVADMFSTPAIFVKAVTDIVDGEKPTSEEFLQNLIAVTAALDLAVTKVVDFISG KRISDL ccchhh

Fig. 2. SOPMA of structural prediction of the hypothetical Protein

Fig. 3. The prediction 3D structural using I-Tasser of the Hypothetical Protein the temple used is 2qttA from PDB

118 119 120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

The instability index (II) was computed to be 23.10 which make the hypothetical protein classified as a stable protein because a protein whose instability index is less than 40 is said to be a stable protein (38). The protein was predicted to contain several helices which is consistent with the ProtParam results present in Figure 1 this makes the protein more flexible for folding which is likely to increase the protein interaction. The sequence of the hypothetical protein was found to be rich in alanine. The proteins with very high AI may show stability in a wide temperature range where lower AI proteins are not thermal stable and show more flexibility. The amino acid sequences which had most in number are alanine (32) followed by leucine and valine (28), glycine and serine (23) and while the least is Tryptophan (1). The hypothetical protein had a total number of 30 negatively charged residues (Asp + Glu) and total number of 27 positively charged residues (Arg + Lys). The molecular formula of the protein was found as C₁₃₅₄H₂₁₈₁N₃₄₉₀₄₀₁S₁₁. The GRAVY was shown to be 0.293GRAVY which shows a better interaction of protein and water is occurring in low GRAVY. (Ikai AJ. 1980). The secondary structure of the hypothetical protein was predicted by SOPMA serve showed the random coil was the most predominant (39.51%), followed by alpha helix (39.16%), followed extended strand (14.69%) and beta turn (6.64%) was the least. I-Tasser modeling server was used to get the 3D structure of the model with a C-score of 1.03 which is the highest score compare to the remaining four other 3D structure.

139

4. CONCLUSION

In silico studies helps in the annotation of protein structure with reduction in the cost of material and time. This study has help in understanding the structural analysis of the hypothetical Protein Zm00014a_031618 (*Zea mays*).

144 CONSENT

6 Not Applicable

149 ETHICAL APPROVAL

Not Applicable

REFERENCES

1. COE E, EAST E. THE BIRTH OF MAIZE GENETICS. IN MAIZE HANDBOOK.VOLUME II: 156 GENETICS AND GENOMICS. EDITED BY BENNETZEN JL, HAKE S. NEW YORK,USA: 157 SPRINGER; 2009:3–15.

2. HE E. "THE ORIGINS OF MAIZE GENETICS," NATURE REVIEWS GENETICS, L. 2001:2(11);898–905.

3. LB J. "THE FUTURE OFMAIZE," IN HANDBOOK OFMAIZE: GENETICS AND GENOMICS, J. L.
 BENNETZEN AND S. HAKE, EDS., SPRINGER, BERLIN, GERMANY, 2009. 771–779.

4. BENNETZEN JL. "MAIZE GENOME STRUCTURE AND EVOLUTION," IN HANDBOOK OF MAIZE: GENETICS AND GENOMICS, J. L. BENNETZEN AND S. HAKE, EDS., SPRINGER, BERLIN, GERMANY. 2009:179–199

169 5. SCHNABLE PS, WARE D, FULTON RS, et al., "THE B73 MAIZE GENOME: COMPLEXITY, 170 DIVERSITY, AND DYNAMICS," SCIENCE. 2009;326(5956):1112–1115.

6. RAYBURN AL, BIRADAR DB. BULLOCK DG, MCMURPHY LM. "NUCLEAR DNA CONTENT IN F1 HYBRIDS OF MAIZE," HEREDITY. 1993;70:294–300.

175 7. ZHOU S, WEI F, NGUYEN J. *et al.*, "A SINGLE MOLECULE SCAFFOLD FOR THE MAIZE GENOME," PLOS GENETICS. 2009;5(11).

8. MCCLINTOCK B. "THE ORDER OF GENES C, SH, AND WX IN ZEA MAYS WITH REFERENCE TO A CYTOLOGICAL KNOWN POINT ON THE CHROMOSOME," PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA. 1931;17(8)485–491.

9. PEACOCK WJ, DENNIS ES, RHOADES, PRYOR AJ. "HIGHLY REPEATED DNA SEQUENCE LIMITED TO KNOB HETEROCHROMATIN IN MAIZE," PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, 1981;78(7):4490-4494.

10. ANANIEV EV, PHILLIPS RL, RINES HW, "CHROMOSOMESPECIFIC MOLECULAR ORGANIZATION OF MAIZE (ZEA MAYS L.) CENTROMERIC REGIONS," PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA. 1998;95(22)13073–13078.

192 11. MORGANTE M., "PLANT GENOME ORGANISATION AND DIVERSITY: THE YEAR OF THE JUNK!," CURRENT OPINION IN BIOTECHNOLOGY. 2006;17(2)168–173.

195 12. BUCKLER ES, GAUT BS, MCMULLEN MD. MOLECULAR AND FUNCTIONAL DIVERSITY OF MAIZE. CURRENT OPINION IN PLANT BIOLOGY. 2006;9(2):172–6.

13. FLINT-GARCIA SA, THUILLET AC, YU J, PRESSOIR G, ROMERO S M, MITCHELL SE, DOEBLEY J, et al. MAIZE ASSOCIATION POPULATION: A HIGH-RESOLUTION PLATFORM FOR QUANTITATIVE TRAIT LOCUS DISSECTION. THE PLANT JOURNAL: FOR CELL AND MOLECULAR BIOLOGY. 2005;44(6):1054–64.

216

236 237

238

239

240

243

248

252

256

- 14. STUPAR RM, SPRINGER NM. CIS-TRANSCRIPTIONAL VARIATION IN MAIZE INBRED LINES
 B73 AND MO17 LEADS TO ADDITIVE EXPRESSION PATTERNS IN THE F1 HYBRID. GENETICS.
 2005;173(4):2199–210.
- 15. SWANSON-WAGNER R. A, JIA Y, DECOOK R, BORSUK LA, NETTLETON D, SCHNABLE PS.
 208 ALL POSSIBLE MODES OF GENE ACTION ARE OBSERVED IN A GLOBAL COMPARISON OF
 209 GENE EXPRESSION IN A MAIZE F1 HYBRID AND ITS INBRED PARENTS. PROCEEDINGS OF
 210 THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA.
 211 2006;103(18):6805–10.
 212
- 213 16. FU Y, WEN TJ, RONIN YI, CHEN HD, GUO L, MESTER DI, YANG Y, *et al.* GENETIC 214 DISSECTION OF INTERMATED RECOMBINANT INBRED LINES USING A NEW GENETIC MAP 215 OF MAIZE. GENETICS. 2006;174(3):1671–83.
- 17. MESSING J, DOONER HK. ORGANIZATION AND VARIABILITY OF THE MAIZE GENOME.
 218 CURRENT OPINION IN PLANT BIOLOGY. 2006;9(2):157–63.
 219
- 220 18. VROH BI, MCMULLEN I, SANCHEZ-VILLEDA MD, SCHROEDER H, GARDINER S, POLACCO
 221 J, SODERLUND M, et al. SINGLE NUCLEOTIDE POLYMORPHISMS AND INSERTION—
 222 DELETIONS FOR GENETIC MARKERS AND ANCHORING THE MAIZE FINGERPRINT CONTIG
 223 PHYSICAL MAP. CROP SCIENCE. 2006;46(1):12.
 224
- 19. TENAILLON M. I., M. C. SAWKINS, A. D. LONG, R. L. GAUT, J. F. DOEBLEY, AND B. S. GAUT,
 "PATTERNS OF DNA SEQUENCE POLYMORPHISM ALONG CHROMOSOME 1 OF MAIZE (ZEA
 MAYS SSP. MAYS L.)," PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE
 UNITED STATES OF AMERICA. 2001;98(16)9161–9166.
- 230 20. CHING A, CALDWELL KS, JUNG M. *et al.*, "SNP FREQUENCY, HAPLOTYPE STRUCTURE 231 AND LINKAGE DISEQUILIBRIUM IN ELITEMAIZE INBRED LINES," BMC GENETICS. 2002;3,(19) 232
- 233 21. A. RAFALSKI AND E. ANANIEV, "GENETIC DIVERSITY, LINKAGE DISEQUILIBRIUM AND ASSOCIATION MAPPING," IN HANDBOOK OF MAIZE: GENETICS AND GENOMICS, J. L. BENNETZEN AND S. HAKE, EDS., PP. 201–219, SPRINGER, BERLIN, GERMANY, 2009
 - 22. RAFALSKI A. MORGANTE M, "CORN AND HUMANS: RECOMBINATION AND LINKAGE DISEQUILIBRIUM IN TWO GENOMES OF SIMILAR SIZE," TRENDS IN GENETICS. 2004;20(2)103–111,
- 23. RAYBURN AL, PRICE HJ, SMITH JD, GOLD JR. "C-BAND HETEROCHROMATIN AND DNA CONTENT IN ZEA MAYS," AMERICAN JOURNAL OF BOTANY. 1985;72(10):1610–1617,.
- 24. RAYBURN AL, "FLOW CYTOMETRIC ASSESSMENT OF NUCLEOTIDE VARIABILITY AND ITS EVOLUTIONARY IMPLICATIONS," IN CLASSICAL AND MOLECULAR CYTOGENETIC ANALYSIS, W.J. RAUP AND B.S. GILL, EDS, KANSAS AGRICULTURAL EXPERIMENTAL STATION, MANHATTAN, KAN, USA, 1994;110–115
- 25. LEE JH, ARUMUGANATHAN K, KAEPPLER S. M. *ET AL*., "VARIABILITY OF CHROMOSOMAL DNA CONTENTS IN MAIZE (ZEA MAYS L.) INBRED AND HYBRID LINES," PLANTA, 2002;215(4)666–671,
- 26. LAURIE DA, BENNET MD. "NUCLEAR DNA CONTENT IN THE GENERA ZEA AND SORGHUM.
 INTERGENERIC, INTERSPECIFIC AND INTRASPECIFIC VARIATION," HEREDITY.
 1985;55(3);307–313.
- 257 27. BIRADAR DP, RAYBURN AL. "HETEROSIS AND NUCLEAR DNA CONTENT IN MAIZE," 258 HEREDITY. 1993;71(3)300–304,.
- 28. A. RAFALSKI AND E. ANANIEV, "GENETIC DIVERSITY, LINKAGE DISEQUILIBRIUM AND ASSOCIATION MAPPING," IN HANDBOOK OF MAIZE: GENETICS AND GENOMICS, J. L.
- 262 BENNETZEN AND S. HAKE, EDS., SPRINGER, BERLIN, GERMANY, 2009;201–219

272

276

290

- 263
 264
 29. TENAILLON MI, SAWKINS MC, LONG AD, GAUT RL, DOEBLEY JF. et al., PATTERNS OF
 265
 DNA SEQUENCE POLYMORPHISM ALONG CHROMOSOME 1 OF MAIZE (ZEA MAYS SSP.
 266
 MAYS L). PROC. NATL. ACAD. SCI. 2001;98:9161–9166.
- 30. VROH BI, MCMULLEN M. D., SANCHEZ-VILLEDA H, SCHROEDER S, GARDINER J. et al., SINGLE NUCLEOTIDE POLYMORPHISMS AND INSERTION-DELETIONS FOR GENETIC MARKERS AND ANCHORING THE MAIZE FINGERPRINT CONTIG PHYSICAL MAP. CROP SCI. 271 2005;46:12-21
- 273 31. RAFALSKI A., ANANIEV E, "GENETIC DIVERSITY, LINKAGE DISEQUILIBRIUM AND 274 ASSOCIATION MAPPING," IN HANDBOOK OF MAIZE: GENETICS AND GENOMICS, J. L. 275 BENNETZEN AND S. HAKE, EDS., SPRINGER, BERLIN, GERMANY, 2009;201–219,
- 277 32. DOEBLEY J. "MOLECULAR EVIDENCE FOR GENE FLOW AMONG ZEA SPECIES,"
 278 BIOSCIENCE. 1990;40(6) 443–448.
 279
- 280 33. WITTKOPP PJ, HAERUM BK, CLARK AG. EVOLUTIONARY CHANGES IN CIS AND TRANS 281 GENE REGULATION. NATURE. 2004;430:85–88. 282
- 283 34. GASTEIGER E, HOOGLAND C, GATTIKER A, DUVAUD S, WILKINS MR, et al. PROTEIN 284 IDENTIFICATION AND ANALYSIS TOOLS ON THE EXPASY SERVER. THE PROTEOMICS 285 PROTOCOLS HANDBOOK. 2005;571-607 286
- 287 35. GEOURJON C, DELEAGE G SOPMA: SIGNIFICANT IMPROVEMENTS IN PROTEIN 288 SECONDARY STRUCTURE PREDICTION BY PREDICTION FROM MULTIPLE ALIGNMENTS. 289 COMPUT APPLIC BIOCI. 1995;11:681-684.
- 291 36. ALTSCHUL SF, GISH W, MILLER W, MYERS EW, LIPMAN DJ. BASICLOCAL ALIGNMENT SEARCH TOOL. J MOL BIOL. 1990;215:403—10 293
- 294 37. YANG Z. I-TASSER: FULLY AUTOMATED PROTEIN STRUCTURE PREDICTION IN CASP8. PROTEINS. 2009;77(9):100-113.
- 297 GURUPRASAD K. REDDY BV, PANDIT MW. PROTEIN ENG. 1990;4:155—61 298
- 38. IKAI AJ. THERMO STABILITY AND ALIPHATIC INDEX OF GLOBULAR PROTEINS. JOURNAL
 OF BIOCHEMISTRY. 1980;88:1895-1898