

Multi-drug resistance pattern of *Staphylococcus aureus* from Paediatric ward, General Hospital, Ikot-Ekpaw, Mkpato Enin LGA, Akwa-Ibom State, Nigeria.

ABSTRACT

The evaluation of the Multi-drug resistance pattern of *Staphylococcus aureus* from a paediatric ward, in Akwa-Ibom State was conducted using standard clinical microbiological procedures. Of the 100 samples from skin, wound, ear, throat and nose swabs, 28 isolates were confirmed as *S. aureus* and were subjected to a range of selected commercially available antibiotics like: amoxicillin, ampiclox, chloramphenicol, ciprofloxacin, erythromycin, gentamicin, levofloxacin, norfloxacin, rifampicin and streptomycin, to evaluate their susceptibilities. The wound swabs gave the highest isolate percentage yield (32%) followed by skin swabs (29%). While susceptibility results showed that amoxicillin and ampiclox were more resisted by the isolates, while ciprofloxacin, levofloxacin and norfloxacin were more effective against the isolates. The MAR indices showed that 85.7% of the isolates had confirmed multi-drug resistance status, with 60.7% of the isolates having resistance for between four or more the tested antimicrobials. MAR indices revealed that 96.4% of the isolates had 0.3, indicating that the resistance resulted from isolates that adapted to the tested drugs due to some form of abuse. Restricted use of these drugs would help curtail the high resistance currently experienced amongst microorganisms.

Key words: multi-drug resistance, *Staphylococcus aureus*, multiple antibiotics resistance index, susceptibility test, nosocomial infections

INTRODUCTION

Microbes do manifest themselves in three ways, through substance spoilage, fermentation of organic and inorganic matters and causation of ailments. Different microorganisms, with their different mode of aetiology, causing different types of ailments, will require different methods and capable drugs for treatments. Continuous deployment of antimicrobial drugs in treating microbial infections has led to the emergence of resistance amongst various strains of microorganisms (McIntosh, 2018; Tanwar *et al.*, 2014). MDR literally means ‘being resistant to more than one antimicrobial agent’, although a standardized definition has not yet been agreed upon by the medical community. There are currently other definitions that are used to characterize patterns multidrug resistance. The most practical definition used for Gram-positive and Gram-negative bacteria is ‘resistance to three or more antimicrobial classes according to Magiorakos *et al.* (2012). MDR could also be defined as the insensitivity or resistance of a

36 microorganism to administered antimicrobial medicines (which are structurally unrelated and
37 have different molecular targets) despite earlier sensitivity to the same medicines (Singh, 2013).

38 According to Nikaido (2009), multidrug resistance in bacteria cells come about by their
39 accumulation, on resistance (R) plasmids or transposons, of genes, with each coding for
40 resistance to a specific agent, and/or by the action of multidrug efflux pumps, each of which can
41 pump out more than one drug type. This MDR abilities lead to ineffective ailment treatment,
42 resulting in its persistence, infection's spread and high cost (Tanwar *et al.*, 2014; WHO, 2014).

43 The hospital environment have been said to be a active reservoir for infectious microorganisms,
44 being the meeting point for people with diverse disease etiological agents and susceptible
45 individuals (Zhanel *et al.*, 2008; Rhomberg *et al.*, 2006; O'Brien *et al.*, 1999). Nikaido (2009)
46 mentioned that *Staphylococcus aureus* has a known nosocomial, multi-drug-resistant strain
47 referred to as the methicillin-resistant *Staphylococcus aureus* (MRSA). MRSA was initially
48 controlled but currently is also resistant to other antimicrobials like the aminoglycosides,
49 chloramphenicol, lincosamides, macrolides and tetracycline (McIntosh, 2018). This study was
50 conducted as part of evidence to buttress the efficacy of *Staphylococcal* infections in young
51 children and the scale to which MDR pathogens are becoming threats to the health of the
52 younger generation amongst the Mkpát Enin, Akwa-Ibom State populace.

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54 **MATERIALS AND METHODS**

55 **Study Facility, Group and Sample Collection**

56 The study facility is in a growing town and services a couple of adjoining communities, with a
57 number of established institutions, stable commerce and ever growing population. The General
58 Hospital, Ikot-Ekpaw, in Mkpát Enin LGA, Akwa-Ibom State, South-South Nigeria, has Out-
59 patient department, Post-natal ward, Paediatric ward and servicing laboratories. The study
60 focused on children aged 1-15 years.

61 A hundred (100) sterile swabs samples from skin, nostrils, wound, throat and ear was sourced
62 over a period of three months. Once gotten, the swabs were labelled, placed in an ice pack and
63 taken to the Microbiology laboratory, Akwa-Ibom State University.

64 **Sample Analysis**

65 The sterile swab sticks were depth into peptone water and incubated at 37°C for 24 hours. A
66 loopful from each sample was streaked on separate Mannitol salt agar (MSA) plates and
67 incubated at 37°C for 24 hours. Discrete golden yellow colonies were subcultured, purified and
68 preserved. Only Gram positive cocci bacterial colonies were further tested for catalase and
69 coagulase.

70 **Antibiotics Susceptibility Test**

71 Confirmed *Staphylococcus aureus* isolates were tested for their susceptibilities to various
72 selected commercial antibiotic drugs like Ciprofloxacin, Erythromycin, Levofloxacin,
73 Gentamicin, Ampiclox, Rifampicin, Amoxicillin, Streptomycin, Norfloxacin and
74 Chloramphenicol. Overnight cultures using Kirby-Bauer method (Hudzicki, 2009) were
75 inoculated on Mueller-Hinton agar (Oxoid, Uk), cultures adjusted to 0.5 McFarland standard.
76 After pre-diffusion, the plates were inoculated at 37°C for 24 hours. Diameter of zones of
77 inhibition (IZDs) produced by the antibiotics were measured and recorded in millimeter.
78 Thereafter, the Multiple antibiotics resistance (MAR) index was determine for each isolate using
79 a formula $MAR = x/y$, where x is the number of antibiotics to which test isolate displayed
80 resistance and y is the total number of antibiotics to which the test organism has been evaluated
81 for sensitivity (Tula-Sanchez *et al.*, 2013; Olayinka *et al.*, 2004).

82 **RESULTS**

83 Result for the prevalence of *S. aureus* is as shown in Table 1. The result table shows that of the
84 28 confirmed, wound samples had the highest number, which was followed by samples from
85 children skin swabs. The least number of confirmed *Staphylococcus aureus* isolates were from
86 the ear swabs.

87 The susceptibility pattern of the 28 confirmed test isolates to the selected commercially available
88 drugs (amoxicillin, ampiclox, chloramphenicol, ciprofloxacin, erythromycin, gentamicin,
89 levofloxacin, norfloxacin, rifampicin and streptomycin) is as shown in Figure 1. Ciprofloxacin
90 was the most effective drugs against the test organism, followed by Levofloxacin and
91 Norfloxacin. Confirmed *Staphylococcus aureus* isolates had high resistance for Amoxicillin,
92 closely followed by their resistance for Ampiclox.

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Table 1: Prevalence of *Staphylococcus aureus* from clinical samples

Type of Specimen	Sample Size	Number of <i>S. aureus</i> isolated	Total percentage
Skin	20	8	29
Nose	20	5	18
Wound	20	9	32
Throat	20	4	14
Ear	20	2	7

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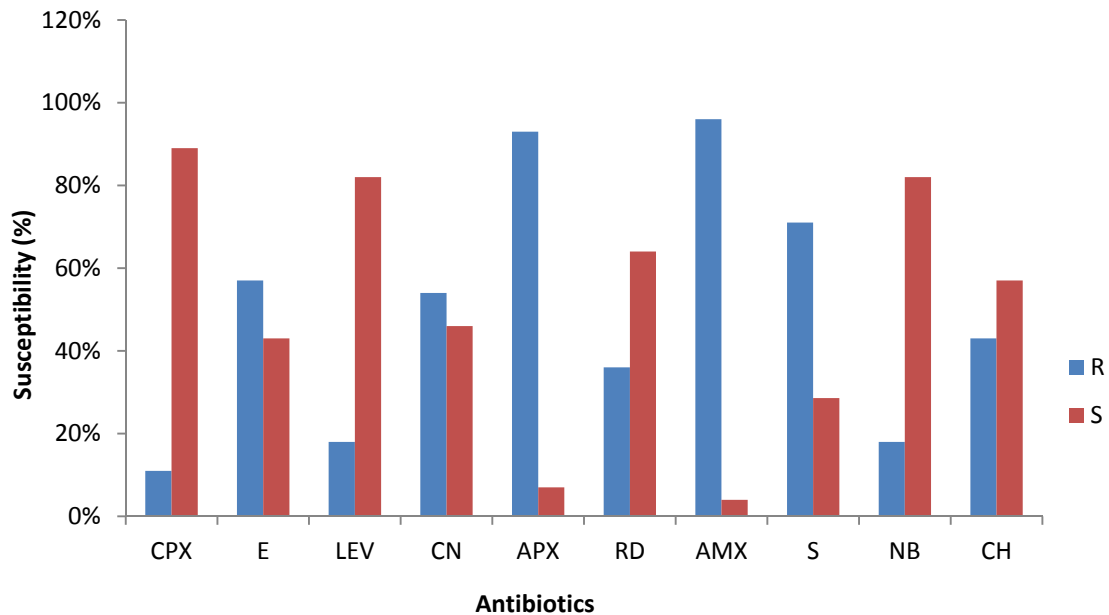


Figure 1: Antibiotics susceptibility pattern of *Staphylococcus* strains from clinical sample

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97 **KEY:** R= Resistant, S= Sensitive, CPX= Ciprofloxacin, E= Erythromycin, LEV= Levofloxacin, CN= Gentamicin,
98 APX= Ampiclox, RD= Rifampicin, AMX= Amoxicillin, S= Streptomycin, NB= Norfloxacin, CH=
99 Chloramphenicol

100

101 Table 2 has the multiple antibiotics resistance (MAR) index result which shows that 85.7% of the
102 confirmed test isolates were multi-drug resistant (showing resistance to three or more classes of
103 antibiotics). Only 14.3% of the isolates showed resistance to only two classes of antibiotics.
104 Results from this study show that 60.7% of test isolates had resistance for four or more

105 antibiotics drugs. MAR index value for 96.4% of the test isolates reveal cases of source drug
 106 abuse.

Table 2: Antibiotic Resistance Pattern and MAR index of *Staphylococcus aureus*

S/N	ISOLATE CODE	Antibiotic Resistant Pattern	MARI	Antibiotic Resistant Class
1	S9	RD, S	0.2	RIF, AMG
2	S8	APX, AMX, CH	0.3	PEN, CHL
3	N3	LEV, APX, AMX	0.3	QUI, PEN
4	W1	CN, AMX, CH	0.3	AMG, PEN, CHL
5	W9	APX, AMX, S	0.3	PEN, AMG
6	S10	CN, APX, RD, AMX	0.4	AMG, PEN, RIF
7	T6	E, APX, AMX, NB	0.4	MAC, PEN, QUI
8	T8	CPX, E, APX, AMX	0.4	QUI, MAC, PEN
9	S2	E, APX, RD, AMX, S	0.5	MAC, PEN, RIF, AMG
10	S3	E, CN, APX, AMX, S	0.5	MAC, AMG, PEN
11	N1	E, CN, APX, AMX, CH	0.5	MAC, AMG, PEN, CHL
12	N2	CN, APX, AMX, S, CH	0.5	AMG, PEN, CHL
13	W2	E, LEV, APX, AMX, S	0.5	MAC, QUI, PEN, AMG
14	W5	E, APX, AMX, S, CH	0.5	MAC, PEN, AMG, CHL
15	W6	CN, APX, AMX, S, CH	0.5	AMG, PEN, CHL
16	W7	APX, AMX, S, NB, CH	0.5	PEN, AMG, QUI, CHL
17	W10	E, LEV, APX, AMX, S	0.5	MAC, QUI, PEN, AMG
18	T3	E, APX, AMX, S, CH	0.5	MAC, PEN, AMG, CHL
19	E5	E, CN, APX, RD, AMX	0.5	MAC, AMG, PEN, RIF
20	S1	E, CN, APX, RD, AMX, S	0.6	MAC, AMG, PEN, RIF
21	S5	E, LEV, APX, RD, AMX, S	0.6	MAC, QUI, PEN, RIF, AMG
22	S6	CN, APX, RD, AMX, S, CH	0.6	AMG, PEN, RIF, CHL
23	N5	CPX, CN, APX, AMX, S, CH	0.6	QUI, AMG, PEN, CHL
24	W8	E, CN, APX, AMX, S, NB	0.6	MAC, AMG, PEN, QUI
25	N8	E, CN, APX, RD, AMX, S, NB	0.7	MAC, AMG, PEN, RIF, QUI
26	W3	LEV, CN, APX, RD, AMX, S, NB	0.7	QUI, AMG, PEN, RIF
27	T2	E, CN, APX, RD, AMX, S, CH	0.7	MAC, AMG, PEN, RIF, CHL
28	E9	CPX, E, CN, APX, AMX, S, CH	0.7	QUI, MAC, AMG, PEN, CHL

KEY: CPX= Ciprofloxacin, E= Erythromycin, LEV= Levofloxacin, CN= Gentamicin, APX= Ampiclox, RD= Rifampicin, AMX= Amoxicillin, S= Streptomycin, NB= Norfloxacin, CH= Chloramphenicol, MARI= Multiple antibiotic resistance index, RIF= Rifamycins, AMG= Aminoglycosides, PEN= Penicillins, CHL= Chloramphenicol, MAC= Macrolides, QUI= Quinolones

107 **DISCUSSIONS AND CONCLUSION**

108 Data for isolate occurrence and confirmation showed that there were more confirmed
109 *staphylococcus* isolates from wound swab-samples, followed by skin sample-swabs. Parta *et al.*
110 (2009) also recorded very high *Staphylococcus* number from wound swabs. This high isolate-
111 numericals is suggestive of the exposed nature of the sampling points. This is supported by
112 findings presented by Nimmo *et al.* (2009), who found more *Staphylococcus* isolates on exposed
113 body surfaces than the internal parts. While uncovered wounds have sticky surfaces and the skin
114 is continuously exposed, it is therefore easy for such high microbial numbers to be recorded.

115 Susceptibility data showed that all the confirmed and tested isolates resisted two or more
116 antimicrobials. Qureshi *et al.* (2004) also isolated MRSA that resisted multiple anti-microbials
117 from hospital specimens. This study's result showed a higher MDR percentage than the "nearly
118 half" proportion reported by Nimmo *et al.* (2009).

119 More than 96.4% of the MAR indices were 0.3 from this study evaluation. This assertion is
120 indicative that resistance to these multiple drugs come from over exposure of the isolates to
121 drugs, making them adapt or resistant to them with recurrent treatments. The high case of MDR
122 amongst microorganisms can drastically be reduced by mere restricting the indiscriminate and
123 readily availability of these drugs over the counter (Nimmo *et al.*, 2009).

124 Many studies have shown that prolonged stays in hospitals increases the risk for colonization or
125 infection with MDR *Staphylococcus aureus*. This reflects an inherent risk in acquiring MDR
126 organisms through environmental contaminations and hospital stay conditions. With infant
127 patients, another potential transmission route is through infected staff members handling and this
128 calls for special care (Buke *et al.*, 2007; Maamar *et al.*, 2016).

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130 The isolation of MDR *Staphylococcus aureus* from infant patients is a call for the proper
131 implementation of contact precautions during hospitalization especially in developing countries
132 (Harbarth *et al.*, 2006; Buke *et al.*, 2007).

133 **Ethical consideration**

134 The study was approved by the ministry of health, Akwa Ibom State. Permission was obtained
135 from General Administration of the General hospital prior to collecting any data. Participants'

136 privacy and confidentiality have been assured (no names have been used, only numbers were
137 used) and all data and results have been handled and treated confidentially.
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