# Original Research Article

# Phytochemical Composition of Ethanol Extract of a cocktail Herbal Mixture (Aju Mbaise)

4 5 6

1

2

3

## ABSTRACT

7 8 *A* 

- Aim: This study was carried out to determine the phytochemical constituent of ethanol extract of
- 9 Aju Mbaise herbal mixture.
- 10 Study design: In the course of the experiment, fresh samples of the plants that make up Aju Mbaise
- were collected and identified as Cnestis ferruginea, Xylopia aethiopica, Uvaria chamae, Palisota
- 12 hirsuta, Scleria sp., Napoleona imperialis, Dialium guineense, Combretum racemosun, and Heterotis
- 13 rotundifolia respectively. The fresh plants were air-dried, cut into small pieces and blended before
- the extraction process. Ethanol was used as the extraction solvent.
- 15 Place and Duration of Study: The study was carried out in the Research Laboratory of the
- Department of Biochemistry, Faculty of Science, University of Port Harcourt, in July 2018.
- 17 Methodology: The qualitative phytochemical analysis was determined by Standard methods for
- 18 testing alkaloids, flavonoids, glycosides, phenols, saponins, steroids, tannins, and terpenoids, while
- 19 the quantitative phytochemical was estimated by spectrophotometric method.
- 20 **Results:** The phytochemical result showed the presence of alkaloids (8.69%), flavonoids (19.10%),
- 21 glycosides (6.86%), hydrogen cyanide (0.92%), phenols (31.56%), saponins (0.83%), steroids
- 22 (0.94%), tannins (16.80%), and terpenoids (14.31%).
- 23 Conclusion: The study showed that ethanol extract of Aju Mbaise herbal mixture contains
- tremendous amount of phytochemicals.
- 25 Keywords:
- 26 Aju Mbaise, Extraction, Phytochemical, Spectrophotometric and *Xylopia aethiopica*.

27 Introduction

- 28 Medicinal plants, also known as medicinal herbs, have been revealed and used in traditional
- 29 medicine practices since ancient times. They are used to attempt to maintain good health, whether
- 30 in modern medicine or in traditional medicine [1]. According to [2], plant's medicinal properties
- are dependent on the plant secondary metabolites contained in them. These metabolites that possess
- medicinal properties are found only in a few species of plants [2]. However, development of plants
- 33 or extracts having potential medicinal uses is blunted by weak scientific evidence, poor practices in
- 34 the process of drug development, and insufficient financing. Some other functions of these
- 35 secondary metabolites include; serving as defensive compounds against herbivores and pathogens,
- 36 mechanical support to the plant, absorbing harmful ultraviolet radiation and reducing the growth of

Comment [U1]: the word and Xylopia aethiopica don't need to be written anymore, because Aju Mbaise has a mixture of these herbs

nearby competing plants. Secondary plant metabolites with reported medicinal properties include 37 alkaloids, terpenoids, saponins, polysaccharides, waxes and fatty acids, simple phenolics, 38 39 flavonoids and glycosides and their derivatives. According to [3], alkaloids are group of naturally occurring chemical compounds that contain mostly basic nitrogen atoms. It also includes some 40 related compounds with neutral and even weakly acidic properties. According to [4], about ninety-41 42 five percent (95%) of alkaloids taste bitter with high level of toxicity, and they are naturally synthesized by a large diversity of organisms including fungi, bacteria, plants, and animals. Some 43 of the pharmacological benefits 44 45 alkaloids include; antimalarial (e.g. quinine) [4], antiasthma (e.g. ephedrine), anticancer (e.g. homo 46 harringtonine) [5], cholinomimetic (e.g. galantamine) [6], vasodilatory(e.g. vincamine), antiarrhyth mic (e.g. quinidine, analgesic (e.g. morphine) [7], antibacterial (e.g. chelerythrine) [8], and 47 antihyperglycemic activities (e.g. piperine) [3]. Other alkaloids possess psychotropic (e.g. psilocin) 48 and stimulant activities (e.g. cocaine, caffeine, nicotine, theobromine) [9] and have been used in ent 49 50 heogenic rituals or as recreational drugs. Also according to [9], some alkaloids can be toxic too (e.g. atropine, tubocurarine). Flavonoids are the most common group of polyphenolic compounds in 51 52 the human diet and are found mainly in plants [10]. Its widespread distribution, varieties and 53 relatively low toxicity compared to other active plant compounds (for instance alkaloids), shows that many animals, including humans, ingest significant quantities in their diet. According to [11], 54 55 some foods with high flavonoid content include parsley, onions, blueberries and other berries, black tea, green tea and oolong tea, bananas, all citrus fruits. Flavonoids are classified into six major 56 classes, which are; flavones, flavonols, flavonols, flavonols (catechins), anthocyanidins and 57 58 isoflavones. The biological and pharmacological activities of flavonoids include anti-allergic [12], 59 anti-inflammatory [13], antioxidant [13], antibacterial [14, 15], antifungal [16, 17], antiviral [16, 60 17], anti-cancer [18] and anti-diarrheal activities [19]. According to [20], almost every group of 61 flavonoids is capable of acting as powerful antioxidants which can protect the human body from free radicals and reactive oxygen species. Glycosides are plant secondary metabolites composed of 62 two components, glycone (a carbohydrate component) and aglycone (a non-carbohydrate 63 64 component) [2]. According to [21], the glycone component usually consists of one or more sugar 65 moieties containing glucose, galactose, xylose, arabinose, rhamnose, or glucuronic acid is linked to a sapogenin (aglycone) to form a glycoside. Therapeutic activities of glycosides include, analgesic, 66 67 antipyretic, anti-inflammatory and laxative effects [22]. Saponins are group of secondary plant metabolites with foaming characteristics and a bitter taste. This phytochemical is widely found in 68 69 most vegetables, beans and herbs [23]. Its foaming ability is caused by the combination of a hydrophobic (fat-soluble) sapogenin and a hydrophilic (water-soluble) sugar part. Some saponins 70 are toxic and are known as sapotoxin. According to [24], saponins have been considered to have 71 important roles in plants defense against pathogens, pests and herbivores due to their antimicrobial, 72 73 antifungal, antiparasitic, insecticidal and anti-feedant properties. According to [25], saponins have 74 also been found to possess hypoglycemic properties, antivirals activity and used as adjuvants in development of vaccines [26], though there is no high-quality clinical evidence that they have any 75 beneficial effect on human health. According to [27], tannins are heterogeneous group of high 76 77 molecular weight polyphenolic compounds that have the capacity to form reversible and irreversible complexes with proteins, polysaccharides (especially cellulose, hemicellulose, pectin, 78 79 etc), alkaloids, nucleic acids, large molecular compounds, metallic ions, and minerals. Its 80 therapeutic properties include its use as astringents against diarrhea, as diuretics against stomach and duodenal tumours [28], as antiseptic, anti-inflammatory, antioxidant, antimicrobial, antitumor, and haemostatic pharmaceuticals. According to [29], it also possess superoxide anion scavenging and anti-plasmin inhibitory activities. Hydrogen cyanide also known as prussic acid, is a colourless, extremely poisonous and flammable chemical compound with the chemical formula HCN. It has a faint bitter almond-like odour that some people are unable to detect owing to a recessive genetic trait. It can be produced on an industrial scale and is a highly valuable precursor to many chemical compounds ranging from polymers to pharmaceuticals. The volatile compound has been used as inhalation rodenticide and human poison, as well as for killing whales [30]. HCN is obtainable from fruits that have a pit, such as cherries, apricots, apples, and bitter almonds, from which almond oil and flavoring are made. Phenols constitute probably the largest group of plant secondary metabolites, varying in size from a simple structure with an aromatic ring to complex ones such as lignins. Phenols are antioxidants in human and plants [31]. Phenolic compounds have antioxidant and antimicrobial properties [32]. Its antioxidant activity is due to the hydroxyl functional group. and other factors such as presence of electron withdrawing or releasing group in the aromatic ring having hydroxyl moiety which may increase or decrease the activity [33]. Steroid is a biologically active organic compound that functions as components of cell membranes which alter membrane fluidity; and as signalling molecules. Hundreds of steroids are found in plants, animals and fungi. All steroids are manufactured in cells from the sterols; lanosterol or cycloartenol, which are derived from the cyclization of the triterpene squalene. Steroids play critical roles in a number of disorders, including malignancies like prostate cancer, where steroid production inside and outside the tumour promotes cancer cell aggressiveness [34]. Terpenoids also called isoprenoids, are a large and diverse class of naturally occurring organic chemicals derived from terpenes. About 60% of known natural products are terpenoids [35]. Plant terpenoids are used for their aromatic qualities and play a role in traditional herbal remedies. Terpenoids contribute to the scent of eucalyptus, the flavours of cinnamon, cloves, and ginger, the yellow colour in sunflowers, and the red colour in tomatoes [36]. The resource plant Aju Mbaise is a traditional medicine, composed of combination of leaves, roots, and trunk of medicinal tree wrapped together commonly used by the people of Mbaise in Igboland, to help detoxify, cleanse and sanitize the womb after child delivery. The bioactive compounds are not known and claims associated with the use are yet to be scientifically substantiated, though aged women who deal in this herb, have tested and proven its efficacy. According to the herbalists, this decoction gets rid of the excess water, stale and bad blood in the womb, and every post-natal substance that may be left hence allowing the stomach to return to its normal size in good time. Other claimed benefits of Aju Mbaise decoction include enhancement of ovulation and fertility, prevents halitosis (mouth odour that comes out from the stomach), stops painful and scanty menstruation, and detoxification of dead particles left after miscarriage, anti-malaria, antitumor and anti-inflammatory. [37], reported that the decoction contains bioactive compounds believed to be responsible for the observed antibacterial activities. According to [37], intake of adequate amounts of the decoction can make some contributions to the macro- and micro-mineral value of lactating mothers towards achieving the Recommended Nutrient Intake (RNI) for these minerals. The ability of this plant to demonstrate such quality is dependent on the accumulated natural products, biologically active materials and ingredients found in them. Thus, the need to determine the phytochemical composition of this herbal mixture.

**Comment [U2]:** what is the relationship between diuretics against stomach and duodenal tumors

Materials and Methods

81

82

83

84

85 86

87

88 89

90

91

92

93 94

95 96

97

98 99

100

101102

103

104

105

106

107108

109

110111

112113

114

115

116117

118

119

120 121

122

123

124

**Comment [U3]:** move to the next page, the title should not be separated from the contents

# **Collection of Plant Samples**

125

133 134

141

142143

144

147

148

155

156

161

162

167

168

- 126 Fresh samples of the plants that make up Aju Mbaise were collected at Obodo Ujichi, Ahiazu and
- 127 Amuzi, Ahiara Towns, both in Aboh Mbaise L.G.A, of Imo State, Nigeria. The plants were identified
- 128 as Cnestis ferruginea, Xylopia aethiopica, Uvaria chamae, Palisota hirsuta, Scleria sp., Napoleona
- imperialis, Dialium guineense, Combretum racemosun, and Heterotis rotundifolia, respectively by
- 130 Dr. Chimezie Ekeke of the Department of Plant Science and Biotechnology, University of Port
- 131 Harcourt. The fresh plants after collection were air-dried, cut into small pieces and blended before
- the extraction process. The extraction was done with ethanol as the solvent.

#### Preparation of Extract

- The whole plants parts (leaves and stem) were washed, air dried and blended to a powdered form.
- 136 Powdered sample weighing 1,000g was soaked in 3,000ml of 95% ethanol for 48 hours after which
- it was sieved using a muslin cloth and afterwards filtered through a Whatmann filter paper No. 1.
- 138 The filtrate was concentrated using a rotary evaporator at 45° C and afterwards placed on a
- thermostatic water bath for further drying. The concentrate (paste) was collected, weighed, kept in
- sterile bottles and stored at 4° C until usage.

## Phytochemical Screening

#### Qualitative phytochemical screening

- Phytochemical screenings were carried out on the powdered sample using standard procedures to
- confirm the presence of alkaloids, flavonoids, saponins, tannins, steroids, cardiac glycosides,
- terpenoids, and total phenolic compounds, as described by [38], [39] and [40].

## Test for Alkaloids

- To 0.5g of pulverized plant sample was added 5 ml of 1% HC1 and boiled for 5 mins in a steam
- bath. This was filtered and 1 ml of the filtrate was individually treated in various test tubes with a
- 151 few drops of Draggendorf's reagent, Wagner's reagent and Mayers reagent respectively. The
- 152 formation of red, reddish-brown and creamy white precipitates respectively indicates the presence
- 153 of alkaloids.

#### **Test for Cyanide**

- A volume of 15 ml dd.  $H_20$  was added to 0.1 g of the extract in a test tube. An alkaline picrate
- 158 paper was suspended over the mixture and held in place by rubber bung. The arrangement was
- allowed to stand for 18 hr at room temperature. Colour change from yellow to orange indicated the
- presence of cyanide.

## Test for Flavonoids

- 163 The pulverized plant samples weighing 0.2g were respectively heated with 10ml of ethylacetate in
- boiling water bath for 3 mins. The mixture was filtered, after which 4 ml of the filtrate was
- vigorously shaken with 1ml of 1% aluminium chloride solution. A yellowish coloration in the layer
- of the ethylacetate indicates the presence of flavonoids.

## Test for Glycosides

**Comment [U4]:** what is the mesh size of the powder

**Comment [U5]:** move to the next page, the title should not be separated from the contents

To 0.5g of respective pulverized plant sample was added 10 ml of distilled water and boiled for 5 mins. This was filtered and about 2 ml of the respective filtrate hydrolyzed with a few drops of concentrated HCI and the solution turned alkaline with a few drops of ammonia solution, Furthermore, 5 drops of the resultant solution was added to 2 ml of Benedict's qualitative reagent and boiled. The precipitation of a reddish-brown colour indicates the presence of glycosides.

## **Test for Phenols**

The test sample weighing 0.1 g was added to 10 ml of distilled water. The solution was heated in a boiling water bath for about 3 mins and filtered. A 2 ml aliquot of the filtrate was placed in each of 3 test tubes. The filtrate in one of the test tubes was diluted with distilled water in the ratio 1:4. A blue or greenish colour indicated the presence of phenols.

#### **Test for Saponins**

About 2g of the pulverized plant samples was respectively boiled with 20 ml of distilled water in a water bath and filtered after which 10 ml of the filtrates were respectively mixed with 5 ml of distilled water in a test tube and vigorously shaken to obtain a stable persistent froth, which was then mixed with 3 drops of olive oil and observed for the formation of emulsion, which indicates the presence of saponins.

#### **Test for Steroids**

To 9 ml of ethanol, was added 1g of pulverised plant sample and refluxed for a few minutes. The filtrate was concentrated to 2.5 ml in a boiling water bath after which 5 ml of hot water was added. The resultant mixture was allowed to stand for 1 hour and the waxy matter filtered off. The filtrate was extracted with chloroform (2.5 ml) using separation funnel. Thereafter, 1ml of 0.5 ml of concentrated sulphuric acid was added to the ethanol extract in a test tube to form a lower layer. A reddish brown interface indicates the presence of steroids.

#### **Test for Tannins**

Each plant was tested for tannins by weighing the respective pulverized samples (0.5g) and boiled in 20 ml of distilled water in a test tube, then filtered with Whatman No. I filter paper. Then to the filtrates, was added 0.1 % FeCl<sub>3</sub> and observed for brownish green or a blue black colouration, which indicates the presence of tannins.

#### **Test for Terpenoids**

To 1g of the extract, 9 ml of ethanol was added and refluxed for a few minutes and filtered. The filtrate was concentrated down to 2.5 ml in a boiling water bath. Hot distilled water of volume 5ml was added to the concentrated solution; the mixture was allowed to stand for 1 hour and the waxy substance was filtered off. The filtrate was extracted with 2.5 ml of chloroform using a separating funnel. The chloroform extract was evaporated to dryness in a water bath and dissolved in 3 ml of concentrated sulphuric acid and then heated for 10 mins in a water bath. A grey colour indicated the presence of terpenoids.

## Quantitative Phytochemical analysis

Comment [U6]: check this method again, extracts only given water can be colored? Test for PhenoI: Ferric Chloride Test: About 0.2 g of plant extract was weighed and treated with 5% ferric chloride and observed for the formation of deep blue color which indicates the presence of phenoI.

**Comment [U7]:** move to the next page, the title should not be separated from the contents

#### **Estimation of Alkaloid content**

The extract (1 g) was macerated with 20 ml of ethanol and 20% H<sub>2</sub>SO<sub>4</sub> (1:1 v/v). The filtrate (1 ml) 214 215 was added to 5 ml of 60% sulphuric acid. After 5 mins, 5 ml of 0.5% formaldehyde in 60% sulphuric acid was mixed with the mixture and allowed to stand for 3 hr. The absorbance was read 216 217

at 565 nm. What standards are used for calculating alkaloid content?

218 219

213

## **Estimation of Cyanide content**

The extract weighing 1 g was macerated with 50 ml of distilled water and then filtered. To 1 ml of the filtrate, 4 ml of alkaline picrate solution was added. The mixture was boiled for 5 mins, and allowed to cool. The absorbance was measured at 490 nm. What standards are used for calculating cyanide content?

223 224 225

226

227

228 229

230

231

232

234

235

236

220 221

222

#### **Estimation of Flavonoid content**

Flavonoid content was determined in accordance with the method described by [41] with minimal modifications [42]. About 100µl of plant extracts in ethanol (10 mgml<sup>-1</sup>) was mixed with 100µl of 20% aluminium trichloride, with a drop of acetic acid, and then diluted with ethanol to 5ml. The absorbance was read after 40 mins at 415nm. Blank samples were prepared from 100µl of plant extracts with a drop of acetic acid, and then diluted to 5ml with ethanol. The absorption of standard rutin solution (0.5mgml<sup>-1</sup>) in ethanol was measured under the same conditions. The amount of flavonoids in the plant extracts in rutin equivalents (RE) was calculated by the following formula:

233 Flavonoid content =

$$\frac{A \times m_0}{A_0 \times m}$$

where A is the absorption of plant extract solution, A<sub>0</sub> is the absorption of standard rutin solution, m is the weight of plant extract, mg and  $m_0$  is the weight of rutin in the solution, mg. The flavonoid content was expressed in mg rutin equivalents/mg plant extract.

237 238 239

240

241

## **Estimation of Glycoside content**

The extract weighing 1 g was macerated with 50 ml of distilled water and filtered. To the filtrate (1 ml), 4 ml of alkaline pirate solution was added. The mixture was boiled for 5 mins and allowed to cool. The absorbance was read at 490 nm. What standards are used for calculating glycoside content?

242 243 244

245

246

247 248

249

250

## **Estimation of Saponin content**

The extract weighing 1g was macerated with 10 ml of petroleum ether and decanted into a beaker. Another 10 ml of the petroleum ether was added into the beaker and the filtrate was evaporated to dryness. The residue was dissolved in 6 ml of ethanol. The solution (2 ml) was put in a test tube and 2 ml of chromagen solution added into it. It was left to stand for 30 mins and the absorbance was read at 550 nm. What standards are used for calculating saponin content?

251 252 253

# Estimation of Steroid content

Comment [U8]: move to the next page, the title should not be separated from the contents

The extract weighing 1 g was macerated with 20 ml of ethanol and filtered. To the filtrate (2 ml), 2 ml of chromagen solution was added and the solution was left to stand for 30 mins. The absorbance was read at 550 nm. What standards are used for calculating steroid content?

#### **Estimation of total Phenolic content**

 The total phenolic content of extract was measured using Folin-Ciocalteu reagent. The extract weighing 1 g was macerated with 20 ml of 80% ethanol and then filtered. The filtrate (5 ml) was added to 0.5 ml of Folin-Ciocalteu reagent and allowed to stand for 30 mins. Then 2 ml of 20% sodium carbonate was added and absorbance measured at 650 nm. Total phenolic content was estimated using gallic acid as standard [43].

## **Estimation of Tannin content**

The determination of tannin content in each sample was carried out using insoluble polyviny1-polypirrolidone (PVPP), which binds tannins as described by [44]. About, 1ml of extract dissolved in ethanol (1mgml<sup>-1</sup>), in which the total phenolics were determined, was mixed with 100mg PVPP, vortexed, allowed to stand for 15 mins at 4°C and then centrifuged for 10mins at 3000 rpm using a Sorvall Scientific centrifuge. In the clear supernatant, the non-tannin phenolics were determined the same way as the total phenolics content was calculated as a difference between total and non-tannin phenolic content.

#### **Estimation of Terpenoid content**

The extract weighing 1 g was macerated with 50 ml of ethanol and filtered. To the filtrate (2.5 ml), was added 2.5 ml of 5% aqueous phosphomolybdic acid solution and 2.5 ml of concentrated sulphuric acid and mixed. The mixture was left to stand for 30 mins and then made up to 12.5 ml with ethanol. The absorbance was read at 700 nm. What standards are used for calculating terpenoid content?

#### Statistical Analysis

Data were presented as Mean  $\pm$  standard deviation.

#### Results

The result obtained in the qualitative analysis carried out on the plant extract is presented in Table 1 below. Results obtained showed the presence of some important phytochemicals. From the results, it was observed that phenols, flavonoids, tannins, alkaloids, saponins, steroids, glycoside, and terpenoids were present in the plant extract. It also showed the presence of hydrogen cyanide (HCN).

The result obtained in the quantitative analysis carried out on the plant extract is presented in Table 2. Results obtained showed that Aju Mbaise plant extract contains alkaloid (348.56mg), phenols (1265.23mg), flavonoids (765.94mg), tannins (673.67mg), glycosides (274.87mg), and terpenoids (573.63mg) in 100g of the plant extract. These are relatively higher than saponins (33.20mg), steroids (37.6mg), and cyanide (36.8mg) which are also present in the plant extract.

Table 1: Qualitative phytochemical constituents of Aju Mbaise Plant Extract

	<b>Phytochemical Constituent</b>	Relative amount
305	Alkaloids	++
306	Flavonoids	+++
307	Glycosides	++
308	Hydrogen Cyanide	+
309	Phenols	+++
310	Saponins	+
311	Steroids	+
312	Tannins	+++
313	Terpenoids	+++
	Key: + = Present in trace amount	
	++ = Present in average amount	
	= Present in high amount	

+++ = Present in high amount

Table 2: Quantitative phytochemical constituents of Aju Mbaise Plant Extract

315		
	<b>Phytochemical Constituent</b>	Relative amount (mg/100g)
316	Alkaloids	348.56±7.00
317	Flavonoids	765.94±19.82
318	Glycosides	274.87±28.00
319	Hydrogen Cyanide	36.80±7.07
320	Phenols	1,265.23±67.69
321	Saponins	33.20±33.60
322	Steroids	37.60±4.65
323	Tannins	673.67±26.40
324	Terpenoids	573.63±29.16

Values represent Mean  $\pm$  Standard Deviation. n = 3

327 Discussion

304

314

325 326

328 329

330

331 332

333

The use of plant materials including herbal or natural health products with supposed health benefits, is increasing in developed countries, and thus brings attendant risks of toxicity and other effects on human health, despite the safe image of herbal remedies [44]. According to [2], plant's medicinal properties are dependent on the plant secondary metabolites contained in them, and these metabolites that possess medicinal properties are found only in a few species of plants. Our resource plant Aju Mbaise was not an exception, as its constituent plants possess many therapeutic Comment [U9]: The letters in a table are usually smaller than text

Comment [U10]: The letters in a table are usually smaller than text

properties which are dependent on the secondary metabolites contained in them. The present study showed that there are many plants' secondary metabolite found in our resource plant. From the qualitative phytochemical analyses, it was observed that the ethanolic extract of cocktail of Aju Mbaise herbal mixture contains alkaloids (8.69%), flavonoids (19.10%), glycosides (6.86%), hydrogen cyanide (0.92%), phenols (31.56%), saponins (0.83%), steroids (0.94%), tannins (16.80%) and terpenoids (14.31%). This is consonant with the report of [45], that phytochemical screening of Aju Mbaise contained appreciable amount of alkaloids, tannins, flavonoids, cyanogenic glycoside, and saponin. This corresponds with a previous report by [46] who stated that plants contained active components with numerous therapeutic potentials. According to [47], tannins, saponins, terpenes, and alkaloids exist in stem bark of Sphenocentrum jollyanum which is one the plants found in the cocktail herbal mixture of Aju Mbaise. [48], also reported the presence of cardiac glycosides, flavonoids, trihydroxyl phenol, anthraquinones, tannins and polyphenolic compounds, such as flavone glycosides in *Cnestis ferruginea*, another plant found in the herbal mixture. Alkaloids, steroids, cardiac glycosides, saponins and tannins were also seen in the preliminary phytochemical screening of Combretum racemosum extracts [49], which is a constituent plant of Aju Mbaise. High tannin content was seen in Dialium guineense [50], which is also a constituent plant of Aju Mbaise. Other constituent plants of Aju Mbaise herbal mixture includes Heterotis rotundifolia which has high amount of phenolic and flavonoic compounds [51]; Napoleonaea imperialis leaves with high amount of tannins, glycosides, saponins and proteins [52]; Palisota hirsuta leaf extract showed high presence of flavonoids, tannins, terpenoids and alkaloids [53, 54]; Uvaria chamae contains medically active compounds such as oleo-resin, alkaloids, and tannins [55]; and also Xylopia aethiopica which contains alkaloids, glycosides, saponnis, tanins, and stereols [56]. These plants metabolites are known for their various benefits, and have been found to possess a wide range of therapeutic activities, which include protection against chronic diseases. For example, alkaloids protect against chronic diseases. Saponins protect against hypercholesterolemia and antibiotic properties. Steroids and triterpenoids possess analgesic properties. According to [57], plants containing alkaloids have been known to possess antidiarrheal activities and are known to be the largest groups of secondary metabolites in plants. Pure plant isolated alkaloids can also be used as basic medicinal agents for analgesic, antispasmodic and bactericidal effects [58]. Flavonoids are known to be antioxidants and free radical scavengers which prevent oxidative cell damage, and it has strong anticancer activity and protects the cells against all stages of carcinogenesis [59]. Flavonoids in the intestinal tract lower the risk of heart disease [60]. It has been discovered in various studies that flavonoids exhibited hypoglycaemic and hypolipidemic potential [61]. Tannins have been reported to possess astringent properties that hasten the healing of wound and inflamed mucus membranes [62]. According to [63], tannins form irreversible complexes with prolin-rich protein and this results in the inhibition of cell protein synthesis that helps in the treatment of inflamed/ulcerated tissues [64]. Plants that contain tannins as major constituents are used for the treatment of intestinal disorders like diarrhoea and dysentery [65]. According to [66], the steroid, phytosterols are currently used for treating symptoms of uterine cramps, abdominal colic and menstrual irregularity, while topical progesterone in pharmacological doses is used to treat a variety of conditions including premenstrual syndrome, anovulatory cycles, dysfunctional uterine bleeding, and menopausal symptoms. Phenolic compounds are synthesized in plants as secondary metabolites. They have several biological activities which include anti-oxidant, anti-inflammatory, anti-aging and inhibitory properties. These secondary metabolites play a vital

334 335

336

337338

339

340

341 342

343

344

345

346 347

348 349

350

351

352

353 354

355

356

357

358

359

360 361

362

363 364

365 366

367

368

369 370

371

372

373 374

375

- 378 role in reproduction and growth. These compounds also provide protection against harmful
- 379 pathogenic microbes and predators [67]. The plant derived polyphenolic compounds are promising
- nutraceuticals for control of various disorders such as cardiovascular, neurological and neoplastic
- 381 disease [68]. According to [69], phenolic compounds have the ability to reduce risk for
- development or treatment of cancers, cardiovascular disorders, obesity, diabetes, aging-diseases,
- 383 urinary tract infections, and periodontal disease. [67], also reported that the richness of the
- polyphenolic contents of green tea and red wine has made them popular choices for associated
- polyphononic contents of green tea and red who has made ment popular choices for associated
- anticancer and cardiovascular health benefits. [60], reported the hypoglycaemic potentials, wound
- healing properties, and haemolytic activities of saponins.

387388 Conclusion

391 392

395 396

This study has shown that the cocktail herbal mixture of Aju Mbaise contains tremendous amount of phytochemicals that possess numerous therapeutic potentials.

393 Competing Interests

394 Authors have declared that no competing interests exist.

397 References

- Smith-Hall, C., Larsen, H. O. & Pouliot, M. (2012). People, plants and health: a conceptual framework for assessing changes in medicinal plant consumption. *Journal of Ethnobiology and Ethnomedicine*, 8(1), 43.
- Heinrich, M., Barnes, J., Gibbon, S. and Williamson, E. M. (2004). Fundamentals of Pharmacognosy and Phytotherapy. In Kingdom A.D. (Ed.). Churchil Livingstone, (2<sup>nd</sup> Ed., pp. 211-219). Elsevier Science Ltd., UK.
- Shi, Q. I. U., Hui, S. U. N., ZHANG, A. H., Hong-Ying, X. U., Guang-Li, Y. A. N., Ying, H.
   A. N., & Xi-Jun, W. A. N. G. (2014). Natural alkaloids: basic aspects, biological roles, and future perspectives. *Chinese Journal of Natural Medicines*, 12(6), 401-406.
- Harbourne, J.B. (1998). Phytochemical Methods: A Guide to Modern Technique of Plant
   Analysis. 2nd edition London: Chapman and Hall Ltd.Pp. 282.
- Kittakoop, P., Mahidol, C., & Ruchirawat, S. (2014). Alkaloids as important scaffolds in
   therapeutic drugs for the treatments of cancer, tuberculosis, and smoking cessation. *Current topics in medicinal chemistry*, 14(2), 239-252.
- 412 6. Russo, P., Frustaci, A., Del Bufalo, A., Fini, M. & Cesario, A. (2013). Multitarget drugs of plants origin acting on Alzheimer's disease. *Current Medicinal Chemistry*, 20(13), 1686–93.
- Sinatra, R. S., Jahr, J. S., & Watkins-Pitchford, J. M. (Eds.). (2010). *The essence of analgesia and analgesics*. Cambridge University Press. pp. 82–90.
- 416 8. Cushnie, T. T., Cushnie, B. & Lamb, A. J. (2014). Alkaloids: an overview of their antibacterial,
- antibiotic-enhancing and antivirulence activities. *International Journal of Antimicrobial*Agents, 44(5), 377-386.
- 419 9. Robbers, J. E., Speedie, M. K., & Tyler, V. E. (1996). *Pharmacognosy and pharmacobiotechnology*. Williams & Wilkins. pp. 143–185.

- 421 10. Spencer, J. P. E. (2008). Flavonoids: modulators of brain function. *British Journal of Nutrition*, 99, 60–77.
- 423 11. Kyle, J., Butchart, C., McNeill, G., Corley, J., Gow, A. J., Starr, J. M., & Deary, I. J. (2011).
- Flavonoid intake in relation to cognitive function in later life in the Lothian Birth Cohort 1936.
- 425 British Journal of Nutrition, 106(1), 141-148.
- 426 12. Yamamoto, Y. and Gaynor, R. B. (2001). Therapeutic potential of inhibition of the NF-κB
- pathway in the treatment of inflammation and cancer. *Journal of Clinical Investigation*, 107(2),
- 428 135–42.

- 429 13. Cazarolli, L. H., Zanatta, L., Alberton, E. H., Figueiredo, M. S., Folador, P., Damazio, R. G.,
- 430 Pizzolatti, M. G. & Silva, F. R. (2008). Flavonoids: Prospective Drug Candidates. *Mini-*
- 431 *Reviews in Medicinal Chemistry*, 8 (13), 1429–1440.
- 432 14. Cushnie, T. P. T. & Lamb, A. J. (2011). Recent advances in understanding the antibacterial
- properties of flavonoids. *International Journal of Antimicrobial Agents*, 38 (2), 99–107.
- 434 15. Manner, S., Skogman, M., Goeres, D., Vuorela, P. & Fallarero, A. (2013). Systematic exploration of natural and synthetic flavonoids for the inhibition of Staphylococcus aureus
- biofilms. International Journal of Molecular Sciences, 14 (10), 19434–19451.
- 437 16. Cushnie, T. P. & Lamb, A. J. (2005). Antimicrobial activity of flavonoids. *International Journal of Antimicrobial Agents*, 26(5), 343 356.
- 439 17. Friedman, M. (2007). Overview of antibacterial, antitoxin, antiviral, and antifungal activities of tea flavonoids and teas. *Molecular Nutrition and Food Research*, 51(1), 116–134.
- 441 18. Ruela de Sousa, R. R., Queiroz, K. C. S., Souza, A. C. S., Gurgueira, S. A., Augusto, A. C.,
  - Miranda, M. A. & Aoyama, H. (2007). Phosphoprotein levels, MAPK activities and NFκB
- expression are affected by fisetin. Journal of Enzyme Inhibition and Medicinal
- 444 *Chemistry*, 22(4), 439-444.
- 445 19. Schuier, M., Sies, H., Billek, B. and Fischer, H. (2005). Cocoa-related flavonoids inhibit
- 446 CFTR-mediated chloride transport across T84 human colon epithelia. *Journal of Nutrition*, 447 35(10), 2320-2325.
- 448 20. Manach, C., Scalbert, A., Morand, C., Rémésy, C., & Jiménez, L. (2004). Polyphenols: food sources and bioavailability. *The American journal of clinical nutrition*, 79(5), 727-747.
- 450 21. Gurib-Fakim, A. (2006). Medicinal plants: Traditions of yesterday and drugs of tomorrow.
- 451 *Molecular Aspects of Medicine*, 27, 1 93.
- 452 22. Brito-Arias, M. (2007). Hydrolysis of glycosides. *Synthesis and Characterization of Glycosides*, 304-313.
- 454 23. Riguera, R. (1997). Isolating bioactive compounds from marine organisms. *Journal of Marine Biotechnology*, 5(4), 187–193.
- 456 24. Lacaille-Dubois, M. A., & Wagner, H. (2000). Bioactive saponins from plants: an update.
- In Studies in natural products chemistry (Vol. 21, pp. 633-687). Elsevier.
- 458 25. Morrissey, J. P. & Osbourn, A. E. (1999). Fungal resistance to plant antibiotics as a mechanism
- of pathogenesis. Microbiological and Molecular Biological Reviews, 63(3), 708-724.
- 460 26. Sun, H. X., Wang, H., Xu, H. S., & Ni, Y. (2009). Novel polysaccharide adjuvant from the
- roots of Actinidia eriantha with dual Th1 and Th2 potentiating activity. *Vaccine*, 27(30), 3984-3991.
- 463 27. Schofield, P., Mbugua, D. M. & Pell, A. N. (2001). Analysis of condensed tannins: A review.

  464 *Animal Feed Science Technology*, 91, 21-40.

- 465 28. De Bruyne, T., Pieters, L., Deelstra, H. & Vlietinck, A. (1999). Condensed vegetables tannins:
- Biodiversity in structure and biological activities. *Biochemical System Ecology*, 27, 445 459.
- 467 29. Okuda, T., & Ito, H. (2011). Tannins of constant structure in medicinal and food plants— 468 hydrolyzable tannins and polyphenols related to tannins. *Molecules*, 16(3), 2191-2217.
- 469 30. Gunasekar, P. G., Prabhakaran, K., Li, L., Zhang, L., Isom, G. E., & Borowitz, J. L. (2004).
- 470 Receptor mechanisms mediating cyanide generation in PC12 cells and rat brain. *Neuroscience* 471 *research*, 49(1), 13-18.
- 472 31. Dillard, C. J., & German, J. B. (2000). Phytochemicals: nutraceuticals and human health. *Journal of the Science of Food and Agriculture*, 80(12), 1744-1756.
- 474 32. Batawila, K., Kokou, K., Akpagana, K., Koumaglo, K., & Bouchet, P. (2002). Activité
- antifongique d'une espèce en voie de disparition de la flore togolaise: Conyza aegyptiaca (L.)
- Ait. var. lineariloba (DC.) O. Hoffm.(Asteraceae). Acta botanica gallica, 149(1), 41-48.
- 477 33. Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free radical biology and medicine*, 20(7), 933-956.
- 480 34. Lubik, A. A., Nouri, M., Truong, S., Ghaffari, M., Adomat, H. H., Corey, E., Cox, M. E., Li,
- N., Guns, E. S., Yenki, P., Buttyan, R. & Pham, S. (2017). Paracrine sonic hedgehog signaling
- contributes significantly to acquired steroidogenesis in the prostate tumor microenvironment. *International journal of cancer*, *140*(2), 358-369.
- 484 35. Firn, R. (2010). *Nature's chemicals: the natural products that shaped our world.* Oxford University Press on Demand.
- 486 36. Specter, M. (2009). A life of its own. The New Yorker, 28.
- 487 37. Ogueke, C. C., Owuamanam, C. I., Onyedinma, C., Iroanya, A., Bede, E. N., & Nwachukwu, I.
- 488 N. (2016). Antibacterial activity, phytochemical properties and mineral Content of "Aju
- Mbaise" decoction: A liquid extract administered to nursing mothers. *Nigerian Journal of Nutritional Sciences*, 37(1), 114-121.
- 491 38. Harborne, J. B. (2014). Introduction to Ecological Biochemistry. Academic press.
- 492 39. Trease, G.E. and Evans, W.C. (1985). *Pharmacognosy*. In: Pal, S.B. (Ed.). Pharmacognosy. 493 (11<sup>th</sup> Ed., pp. 60-75) Tindal LTD, London
- 494 40. Sofowora, A. (1993). Medicinal plants and medicine in Africa. *John Whilley Spectrum Books*,
   495 *Ibadan, Nigeria*, 120-123.
- 496 41. Kumaran, A. & Karunakaran, R. (2007). Activity-guided isolation and identification of free radical-scavenging components from an aqueous extract of *Coleus aromaticus*. *Food*
- 498 *Chemistry*, 100, 356 361.
- 499 42. Awah, F. M., Uzoegwu, P. N., Ifeonu, P., Oyugi, J. O., Rutherford, J., Yao, X. J., Fehrmann,
- 500 F., Fowke, K. R. & Eze, M. O. (2012). Free radical scavenging activity, phenolic content and cytotoxicity of selected Nigerian medicinal plants. *Food Chemistry*, 131(4), 1279 1286.
- 502 43. Madaan, R., Bansal, G., Kumar, S., & Sharma, A. (2011). Estimation of total phenols and
- flavonoids in extracts of Actaea spicata roots and antioxidant activity studies. *Indian journal of*
- 504 pharmaceutical sciences, 73(6), 666.
- Ekor, M. (2013). The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. *Frontiers in Pharmacology*, 4 (3): 202–4.

- 507 45. Ezejindu, C. N. & Iro, O. K. (2017). Antibacterial activity, phytochemical properties and mineral content of "Aju Mbaise" decoction administered to nursing mothers. *Direct Research*
- Journal of Health and Pharmacology, **5**(3), 33-38.
- 510 46. Rakesh, D. D., Handa, S. S., & Vasisht, K. (2006). Compendium of medicinal and aromatic plants ASIA. *ICS UNIDO*. *Asia*, 2, 305.
- 512 47. Nia, R., Paper, D. H. & Essien, E. E. (2004). Evaluation of the anti-oxidant and anti-angiogenic
- effects of Sphenocentrum jollyanum Pierre. The African Journal of Biomedical Research, 7,
- 514 129–132.
- 515 48. Adisa, A. R., Farooq, A. D. & Iqbal, M. C. (2014). Protection of CCl<sub>4</sub>-induced liver and kidney
- damage by phenolic compounds in leaf extracts of *Cnestis ferruginea* (de Candolle).
- 517 Pharmacognosy Research, 6(1), 19–28.
- 518 49. Onocha, P. A., Audu, E.O., Ekundayo, O. & Dosumu, O. O. (2005). Phytochemical and
- antimicrobial properties of extracts of *Combretum racemosum*. *Acta Horticulturae*, 675, 97–
- 520 101.
- 521 50. Arogba, S. S., Ajiboro, A. & Odukwe, I. J. (2006). A physiochemical study Nigerian Velvet
- tamarind (Dialium guineense) fruit. The Journal of the Science of Food and Agriculture, 66,
- 523 533-534.
- 524 51. Etekpo, S. D., N'Gaman-Kouassi, C. C., Mamyrbekova-Békro, J. A. & Békro, Y. (2018).
- Antioxidant profiles of alcoholic tinctures from *Heterotis rotundifolia* (sm.) by DPPH radical
- trapping. European Journal of Biomedical and Pharmaceutical sciences, 5(10), 39-45.
- 527 52. Chah, K.F., C.A. Eze, C.E. Emuelosi & C.O. Esimone, (2006). Antibacterial and wound
- healing properties of methanolic extracts of some Nigerian medicinal plants. *Journal of*
- 529 *Ethnopharmacology*, 104, 164-167.
- 530 53. Kupeli, E. & Yesilada, E. (2007). Flavonoids with anti-inflammatory and antinociceptive
- activity from Cistus laurifolius L. leaves through bioassay-guided procedures. Journal of
- 532 *Ethnopharmacology*, 112(3), 524-530.
- 533 54. Clavin, M., Gorzalczany, S., Macho, A., Munoz, E., Ferraro, G., Acevedo, C., & Martino, V.
- 534 (2007). Anti-inflammatory activity of flavonoids from Eupatorium arnottianum. Journal of
- 535 Ethnopharmacology, 112(3), 585-589.
- 536 55. Achigan-Dako, E. G., Pasquini, M. W., Assogba Komlan, F., N'danikou, S., Yédomonhan, H.,
- 537 Dansi, A., & Ambrose-Oji, B. (2010). Traditional vegetables in Benin. *Institut National des*
- 538 Recherches Agricoles du Bénin, Imprimeries du CENAP, Cotonou.
- 539 56. Somova, L.I., Shode, F.O., Moodley, K. & Govender, Y. (2001). Cardiovascular and diuretic
- activity of kaurene derivatives of Xylopia aethiopica and Alepidea amatymbica. Journal of
- 541 *Ethnopharmacology*, 77, 165–74.
- 542 57. Obasi, N. B., Igboechi, A. C. & Anuforo, D. C. (1990). Studies of the antidiarrheal potentials
- of some composite plants. Medicinal plants in a developing economy. Proceeding of a
- workshop organized by the Nigeria society of Nigeria, 108-116.
- 545 58. Stray, F. (1998). The national guide to medicinal herbs and Plants. Tiger books international
- 546 London. Pp. 12-16.
- 547 59. Okwu, D. E. & Iroabuchi, F, (2004). Phytochemical analysis and antimicrobial activity
- 548 screening of aqueous and ethanolic root extracts of Uvaria chamae (Beuv) and Cnestis
- ferruginea. The Journal of Chemical Society of Nigeria, 29(2), 112-114.

- 550 60. Okwu, D. E. (2005). Phytochemicals, Vitamins and Mineral contents of two Nigeria Medicinal plants. *International Journal of Molecular Medicine and Advance Sciences*, 1(4), 375-381.
- 552 61. Narender, T., Khaliq, T., & Puri, A. (2006). Antidyslipidemic activity of furano-flavonoids 553 isolated from *Indigofera tinctoria*. *Bioorganic & Medicinal chemistry letters*, 16(13), 3411-554 3414.
- 555 62. Okwu, D. E. (1999). Flavouring properties of spices on cassava Fufu. *African Journal of Root* and Tuber Crops, 3(2), 19-21.
- 557 63. Shimada, T. (2006). Salivary proteins as a defense against dietary tannins. *Journal of Chemical Ecology*, *32*(6), 1149-1163.
- 559 64. Parekh, J. & Sumitra, C. V. (2007). In vitro antimicrobial activity and phytochemical analysis of some Indian medicinal plants. *Turkish Journal of Biology*, 31, 53-58.
- 561 65. Dharmanda, S. (2003). Gallnuts and the uses of tannins in Chinese Medicine-A paper presented at the Institute for Traditional Medicine. *Portlant, Oregon*, 3, 941-945.
- 563 66. Hudson, T. (1996). Townsend Letter for Doctors; 156.

- 67. Gautam, B., Vadivel, V., Stuetz, W., & Biesalski, H. K. (2012). Bioactive compounds extracted from Indian wild legume seeds: antioxidant and type II diabetes—related enzyme inhibition properties. *International journal of food sciences and nutrition*, 63(2), 242-245.
- 567 68. Ullah, M. F., & Khan, M. W. (2008). Food as medicine: potential therapeutic tendencies of 568 plant derived polyphenolic compounds. *The Asian Pacific Journal of Cancer Prevention*, 9(2), 569 187-196.
- 570 69. Yuan, W., Zhou, L., Deng, G., Wang, P., Creech, D., & Li, S. (2011). Anthocyanins, phenolics, and antioxidant capacity of *Vaccinium L*. in Texas, USA.