

Anticancer and molecular docking studies of 1-(5-substituted phenyl) isoxazol-3-yl)-5-phenyl-1H-tetrazoles

Abstract:

Cancer a leading cause of human mortality worldwide, is characterized by unseemly growth of cellular mass and signaled through the enlargement of stress. Management of cancer treatment is still buried and has been recently alert the need to discover a drug molecule with lesser side effects. The objective of present study is to explore the anticancer activity and docking studies of 1-(5-substituted phenyl) isoxazol-3-yl)-5-phenyl-1H-tetrazole derivatives. The compounds were evaluated for *in-vitro* anticancer activity under the drug discovery program of National Cancer Institute (NCI), USA. Only seven compounds was selected and screened for anticancer activity at a single high dose (10^{-5} M) using NCI 60 cancer cell lines. Among all the selected compounds, **4b** and **4i** exhibited significant anticancer activity against Leukemia cell lines. Molecular docking studies for the 5-phenyl-1-(5-substituted phenylisoxazol-3-yl)-1H-tetrazole analogs was done by Schrodinger software. Docking results stated that the compounds **4b** and **4i** has good dock score among the other derivatives which shows good binding efficiency towards receptor.

Keywords: Sodium azide, isoxazole, tetrazole, renal cancer, breast cancer, molecular docking.

1. Introduction

Cancer is a disease of prominent significance in the world today. It is the second foremost cause of death in the world after cardiovascular diseases and is predictable to be the primary cause within the coming years (Gibbs, 2000; KSY et al., 2014). The identification of novel structures that can be potentially useful in development of new, potent, selective and less toxic anticancer agents is a major challenge to researchers till date (Heffeter et al., 2006, Lokhande et al., 2013). Although important advances achieved over recent decades in the development of various cancerostatic drugs, current antitumor chemotherapy still face two major limitations-the first one is lack of selectivity of conservative chemotherapeutic agents for cancer tissues, leading to undesired effects and the second is attainment of multiple-drug

33 resistance by cancer cells. Undesired effects of anticancer drugs could be overcome with the
34 agents capable of astute tumor cells from normal proliferative cells and the resistance is
35 minimized by using combined modality approach with different complementary mode of
36 action (Menta and Palumbo, 1997).

37 The current scenario highlights the need for the discovery and development of new lead
38 compounds of simple structure, exhibiting optimal in-vivo antitumor potency and new
39 mechanism of action. Recent advances in clinical techniques, including large co-operative
40 studies are allowing more rapid and reliable evaluation of new drugs. The combination of
41 these advantages with improved preliminary screening systems is enhancing the emergence
42 of newer and more potent compounds. In this regard, it should be emphasized that National
43 Cancer Institute (NCI) in-vitro primary anticancer drug screen represents a valuable research
44 tool to facilitate the drug discovery of new structural/ mechanistic types of antitumor agents
45 (Boyd and Paull, 1995).

46 Tetrazole derivatives are well known compounds with a high level of biological activity
47 (Schmidt and Schieffer, 2003) and screened for various biological activities such as
48 antibacterial, antifungal (Bhaskar and Mohite, 2010), anticancer (Dhayanithi et al., 2011),
49 analgesic (Bachar and Lahiri, 2004), anti-inflammatory (Mohite and Bhaskar, 2011), anti-
50 diabetic, anti-hyperlipidemic (Momose et al., 2002) and anti-tubercular agents (Adamec et
51 al., 2005). In drug design, tetrazoles are regarded as an isostere for the carboxylate group, and
52 extensive work on tetrazoles has been carried out in the field of medicinal chemistry
53 (Myznikov et al., 2007).

54 They are important ligands for many useful transformations and also precursors for a variety
55 of nitrogen-containing heterocycles (Faucher et al., 2004). It was also noticed that toxic
56 properties of a drug can decrease through the introduction of a tetrazole ring into the
57 molecule (Bekhit et al., 2004). The tetrazole moiety is also generally accepted to exhibit
58 stronger resistance to in vivo metabolism than the carboxylate group, thus conferring to
59 the corresponding drug with longer bioavailability (Herr, 2002). The isoxazole moiety act as
60 lead molecule in the drug development and has a broad range of biological activities like
61 antimicrobial (Gautam and Singh, 2013), anti-inflammatory, analgesic (Karabasanagouda et
62 al., 2009), antitubercular (Tangallapally et al., 2007), anticancer (Yong et al., 2015), anti-
63 hyperglycemic, lipid lowering activity (Kumar et al., 2009) and antihypertensive activity
64 (Carenzi et al., 1989). The designed compounds were also subjected to molecular docking
65 into the colchicines binding site of tubulin.

66 As part of our ongoing studies dealing with the synthesis of various derivatives of tetrazole
67 containing isoxazole moiety, we describe here the synthesis, molecular docking studies of
68 new derivatives and the outcome of preliminary evaluation of their anticancer activity.

69 **2. Experimental**

70 *2.1. Chemistry*

71 All of the reagents used were laboratory grade and purchased from commercial sources and
72 were used after being purified by standard procedures. Melting point was determined by open
73 capillary method and is uncorrected. The purity of synthesized compounds, commercial
74 reagents used and monitoring of chemical reaction was done by thin layer chromatography.
75 The spots were observed under iodine vapors and UV light. IR spectra of synthesized
76 compounds were recorded on Shimadzu FTIR-8400S by using KBr disk. ¹H-NMR spectra
77 were recorded on JEOL AL300 FTNMR 300 MHz spectrophotometer and tetramethylsilane
78 (TMS) is used as internal standard. Chemical shift (δ) values are given in ppm. Mass
79 spectrum was taken using Waters Micromass Q-ToF Micro. Mass spectrometer equipped with
80 electrospray ionization (ESI).

81 *2.2. General procedure for the synthesis of 5-phenyl tetrazole,1*

82 The synthetic procedure and spectral characterization of synthesized compounds described in
83 earlier reports by author (Kaushik et al., 2015, Kaushik et al., 2016,). The equal quantity of
84 sodium azide, ammonium chloride and benzonitrile were refluxed with dimethyl formamide
85 at 125 °C for 7-8 hours, reaction mixture was poured in 100 ml of water; a milky solution
86 was obtained which was acidified with concentrated hydrochloric acid to get the precipitate.
87 The solution was cooled to 5°C, filtered, dried and recrystallized by using ethanol to obtain
88 the 5-phenyl tetrazole (1).

89 White solid, Yield: 82%; m.p.(210-212°C).IR (KBr disk, cm⁻¹): ν 3348 (NH), 3062 (Ar-CH),
90 1628 (C=N), 1292 (N-N=N-). ¹H-NMR (300 MHz, CDCl₃, ppm) δ : 8.72(s, 1H, NH), 7.56-
91 7.18 (m, 5H, Ar-H).

92 *2.3. General procedure for the synthesis of 5-phenyl-1-acetyl tetrazole,2*

93 5-phenyl tetrazole (1), (10 mmol) was mixed with acetic anhydride (10 mmol) and few drops
94 of concentrated sulphuric acid was added to the solution and warmed this solution at 60-70
95 °C for 15-20 minutes on water bath. The reaction mixture was cooled to room temperature
96 and poured into ice cold water. The white precipitate was filtered, washed, dried and
97 recrystallized from ethanol.

98 White solid, Yield: 78%; m.p. (218-220°C).IR (KBr disk, cm^{-1}):v3058 (Ar-CH), 1730 (C=O),
99 1638 (C=N), 1285 (N-N=N-). $^1\text{H-NMR}$ (300 MHz, CDCl_3 , ppm) δ : 7.51-7.28 (m, 5H, Ar-H),
100 2.28 (s, 3H, CH_3).

101 *2.4. General procedure for the synthesis of 3-(substituted phenyl)-1-(5-phenyl-1H-tetrazol-1-*
102 *yl) prop-2-en-1-one, 3a-3j*

103 5-phenyl-1-acetyl tetrazole (2), (10 mmol) was mixed with different substituted aromatic
104 aldehyde (10 mmol) in presence of ethanol (20 ml), the reaction mixture was cooled to 5 to
105 10 °C in an ice bath. The ice cooled solution was treated with 40% sodium hydroxide
106 solution. The reaction mixture was magnetically stirred for 30 minutes and left over night in
107 the refrigerator. The resulting dark solution was diluted with ice cold water and acidified by
108 hydrochloric acid. The solution was filtered, washed with water and recrystallized with
109 ethanol.

110 *2.4.1. Synthesis of 3-(3-bromo-4-nitrophenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-*
111 *one, 3a*

112 Brown solid, Yield: 86%; m.p.(260-262°C).IR (KBr disk, cm^{-1}):v3034 (Ar-CH), 1746 (C=O),
113 1618 (C=C), 1593 (C=N), 1570 (NO_2), 1284 (N-N=N-), 646 (C-Br). $^1\text{H-NMR}$ (300 MHz,
114 CDCl_3 , ppm) δ : 7.94 (d, 1H, Ar-H), 7.53 (s, 1H, Ar-H), 7.47 (d, 1H, Ar-H), 7.40-7.28 (m, 5H,
115 Ar-H), 7.23 (d, 1H, CH), 6.80 (d, 1H, CH). Anal. Calcd. (%) for $\text{C}_{16}\text{H}_{10}\text{BrN}_5\text{O}_3$: C, 48.02; H,
116 2.52; Br, 19.97; N, 17.50; O, 11.99.

117 *2.4.2. Synthesis of 3-(3, 4, 5-trimethoxyphenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-*
118 *one. 3b*

119 Yellowish Brown solid, Yield: 82%; m.p.(230-232°C).IR (KBr disk, cm^{-1}):v3054 (Ar-CH),
120 1740 (C=O), 1630 (C=C), 1600 (C=N), 1248 (N-N=N-), 1221 (OCH_3). $^1\text{H-NMR}$ (300 MHz,
121 CDCl_3 , ppm) δ : 7.47 (d, 1H, CH), 7.45-7.28 (m, 5H, Ar-H), 6.56 (s, 2H, Ar-H), 6.40 (d, 1H,
122 CH), 3.66 (s, 9H, OCH_3). Anal. Calcd. (%) for $\text{C}_{19}\text{H}_{18}\text{N}_4\text{O}_4$: C, 62.29; H, 4.95; N, 15.29; O,
123 17.47.

124 *2.4.3. Synthesis of 3-(2, 4-difluorophenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-one, 3c*

125 Creamy solid, Yield: 79%; m.p.(234-236°C).IR (KBr disk, cm^{-1}):v3059 (Ar-CH), 1712
126 (C=O), 1614 (C=C), 1608 (C=N), 1268 (N-N=N-), 1163 (C-F). $^1\text{H-NMR}$ (300 MHz, CDCl_3 ,
127 ppm) δ : 7.68 (d, 1H, CH), 7.51-7.34 (m, 5H, Ar-H), 7.18 (d, 1H, Ar-H), 6.77 (s, 1H, Ar-H),

128 6.58 (s, 1H, Ar-H), 6.43 (d, 1H, CH). Anal. Calcd. (%) for C₁₆H₁₀F₂N₄O: C, 61.54; H, 3.23;
129 F, 12.17; N, 17.94; O, 5.12.

130 *2.4.4. Synthesis of 3-(4-fluorophenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-one, 3d*

131 Brown solid, Yield: 68%; m.p.(224-226°C).IR (KBr disk, cm⁻¹):v3060 (Ar-CH),1764 (C=O),
132 1620 (C=C), 1606 (C=N), 1310 (N-N=N-), 1166 (C-F). ¹H-NMR (300 MHz, CDCl₃, ppm) δ:
133 7.48 (d, 1H, CH), 7.42-7.29 (m, 5H, Ar-H), 7.21 (d, 2H, Ar-H), 7.04 (d, 2H, Ar-H), 6.56 (d,
134 1H, CH). Anal. Calcd. (%) for C₁₆H₁₁FN₄O: C, 65.30; H, 3.77; F, 6.46; N, 19.04; O, 5.44.

135 *2.4.5. Synthesis of 3-(2-hydroxy-5-nitrophenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-*
136 *one, 3e*

137 Brown solid, Yield: 63%; m.p.(252-254°C).IR (KBr disk, cm⁻¹):v3583 (OH), 3055 (Ar-CH),
138 1680 (C=O), 1630 (C=C), 1610 (C=N), 1564 (NO₂), 1344 (N-N=N-). ¹H-NMR (300 MHz,
139 CDCl₃, ppm) δ: 7.76 (s, 1H, Ar-H), 7.57 (d, 1H, CH), 7.54 (d, 1H, Ar-H), 7.48-7.25 (m, 5H,
140 Ar-H), 6.81 (d, 1H, Ar-H), 6.76 (d, 1H, CH), 4.92 (s, 1H, OH). Anal. Calcd. (%) for
141 C₁₆H₁₁N₅O₄: C, 56.98; H, 3.29; N, 20.76; O, 18.97.

142 *2.4.6. Synthesis of 3-(2-chloro-4-fluorophenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-*
143 *one, 3f*

144 Reddish brown solid, Yield: 74%; m.p.(220-222°C).IR (KBr disk, cm⁻¹):v3054 (Ar-CH),
145 1735 (C=O), 1630 (C=C), 1608 (C=N), 1285 (N-N=N-), 1178 (C-F), 786 (C-Cl). ¹H-NMR
146 (300 MHz, CDCl₃, ppm) δ: 7.72 (d, 1H, CH), 7.44-7.28 (m, 5H, Ar-H), 7.17 (d, 1H, Ar-H),
147 7.06 (s, 1H, Ar-H), 6.74 (d, 1H, Ar-H), 6.37 (d, 1H, CH). Anal. Calcd. (%) for
148 C₁₆H₁₀ClFN₄O: C, 58.46; H, 3.07; Cl, 10.78; F, 5.78; N, 17.04; O, 4.87.

149 *2.4.7. Synthesis of 3-(2, 4-dimethoxyphenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-*
150 *one, 3g*

151 Brown solid, Yield: 78%; m.p.(246-248°C).IR (KBr disk, cm⁻¹):v3054 (Ar-CH), 1666 (C=O),
152 1640 (C=C), 1606 (C=N), 1275 (N-N=N-), 1248 (OCH₃). ¹H-NMR (300 MHz, CDCl₃, ppm)
153 δ: 7.62 (d, 1H, CH), 7.41-7.30 (m, 5H, Ar-H), 7.16 (d, 1H, Ar-H), 6.71 (d, 1H, CH), 6.53 (d,
154 1H, Ar-H), 6.47 (d, 1H, Ar-H), 3.68 (s, 6H, OCH₃). Anal. Calcd. (%) for C₁₈H₁₆N₄O₃: C,
155 64.28; H, 4.79; N, 16.66; O, 14.27.

156 *2.4.8. Synthesis of 3-(2, 4, 6-trimethoxyphenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-*
157 *one, 3h*

158 Yellowish brown solid, Yield: 82%; m.p.(212-214°C).IR (KBr disk, cm^{-1}):v3064 (Ar-CH),
159 1666 (C=O), 1652 (C=C), 1606 (C=N), 1290 (N-N=N-), 1186 (OCH₃). ¹H-NMR (300 MHz,
160 CDCl₃, ppm) δ : 7.63 (d, 1H, CH), 7.45-7.30 (m, 5H, Ar-H), 6.67 (s, 2H, Ar-H), 6.63 (d, 1H,
161 CH), 3.66 (s, 9H, OCH₃). Anal. Calcd. (%) for C₁₉H₁₈N₄O₄; C, 62.29; H, 4.95; N, 15.29; O,
162 17.47.

163 *2.4.9 Synthesis of 3-(2-nitrophenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-one,3i*

164 Yellow solid, Yield: 87%; m.p.(228-230°C).IR (KBr disk, cm^{-1}):v3074 (Ar-CH), 1726
165 (C=O), 1620 (C=C), 1608 (C=N), 1578 (NO₂), 1248 (N-N=N-). ¹H-NMR (300 MHz, CDCl₃,
166 ppm) δ : 7.93 (d, 1H, CH), 7.78 (d, 1H, Ar-H), 7.58 (t, 1H, Ar-H), 7.61 (d, 1H, Ar-H), 7.53-
167 7.32 (m, 5H, Ar-H), 7.28 (t, 1H, Ar-H),6.65 (d, 1H, CH). Anal. Calcd. (%) for C₁₆H₁₁N₅O₃:
168 C, 59.81; H, 3.45; N, 21.80; O, 14.94.

169 *2.4.10.Synthesis of 3-(3-bromo-4-methoxyphenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-*
170 *one,3j*

171 Yellowish brown solid, Yield: 78%; m.p.(164-166°C).IR (KBr disk, cm^{-1}):v3034 (Ar-CH),
172 1660 (C=O), 1615 (C=C), 1608 (C=N), 1255 (N-N=N-), 1170 (OCH₃), 640 (C-Br); ¹H-NMR
173 (300 MHz, CDCl₃, ppm) δ : 7.47 (d, 1H, CH), 7.41-7.29 (m, 5H, Ar-H), 7.27 (s, 1H, Ar-H),
174 7.16 (d, 1H, Ar-H), 6.58 (d, 1H, CH), 6.64 (d, 1H, Ar-H); 3.77 (s, 3H, OCH₃). Anal. Calcd.
175 (%) for C₁₇H₁₃BrN₄O₂: C, 53.00; H, 3.40; Br, 20.74; N, 14.54; O, 8.31.

176 *2.5. General procedure for the synthesis of 1-(5-substituted phenyl) isoxazol-3-yl)-5-phenyl-*
177 *1H-tetrazole,4a-4j*

178 A mixture of 3-(substituted phenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-one (3a-3j, 10
179 mmol), hydroxylamine hydrochloride (10 mmol) and 40% potassium hydroxide in ethanol
180 were refluxed on water bath for 4-5 hours. After that, reaction mixture was cooled at room
181 temperature and poured into crushed ice to form the precipitate which was filtered, dried and
182 recrystallized from ethanol.

183 *2.5.1. Synthesis of 1-(5-(3-bromo-4-nitrophenyl) isoxazol-3-yl)-5-phenyl-1H-tetarzole,4a*

184 Yellow solid, Yield: 76%; m.p.(158-160°C). IR (KBr disk, cm^{-1}):v 3072 (Ar-CH), 2896
185 (CH), 1625 (C=N), 1560 (NO₂), 1502 (C=C), 1438 (N-O), 1261 (N-N=N-), 617 (C-Br). ¹H-
186 NMR (300 MHz, DMSO,ppm) δ : 7.92 (d, 1H, Ar-H), 7.83 (s, 1H, Ar-H), 7.71 (d, 1H, Ar-H),
187 7.56-7.28 (m, 5H, Ar-H), 7.04 (s, 1H, isoxazole). ¹³C NMR (125 MHz, DMSO-d₆) δ 170.4,
188 153.7, 152.4, 136.2, 134.8, 131.5, 130.4, 128.6, 122.7, 102.6. MS m/z: 411 [M⁺], Anal.

189 Calcd. (%) for C₁₆H₉BrN₆O₃: C, 46.51; H, 2.20; Br, 19.34; N, 20.54; O, 11.62%. Found: C,
190 46.63; H, 2.23; N, 20.48.

191 *2.5.2. Synthesis of 1-(5-(3, 4, 5-trimethoxyphenyl)-isoxazol-3-yl)-5-phenyl-1H-tetrazole, 4b*

192 Yellow brown solid, Yield: 78%; m.p.(136-138°C).IR (KBr disk, cm⁻¹):ν3051 (Ar-CH), 2918
193 (CH), 1691 (C=N), 1564 (C=C), 1485 (N-O), 1288 (N-N=N-), 1163 (OCH₃). ¹H-NMR (300
194 MHz, DMSO, ppm) δ: 7.59-7.25 (m, 5H, Ar-H), 7.04 (s, 1H, isoxazole), 6.82 (d, 2H, Ar-H)
195 3.68 (s, 9H, OCH₃). ¹³C NMR (125 MHz, DMSO-d₆) δ 170.6, 152.2, 150.4, 140.7, 132.7,
196 130.2, 127.5, 126.3, 125.8, 102.7, 101.4, 57.3. MS m/z: 379 [M⁺], Anal. Calcd. (%) for
197 C₁₉H₁₇N₅O₄: C, 60.15; H, 4.52; N, 18.46; O, 16.87%. Found: C, 60.28; H, 4.56; N, 18.52.

198 *2.5.3. Synthesis of 1-(5-(2, 4-difluorophenyl)-isoxazol-3-yl)-5-phenyl-1H-tetrazole, 4c*

199 Brown solid, Yield: 68%; m.p.(144-146°C).IR (KBr disk, cm⁻¹):ν3093 (Ar-CH), 2921 (CH),
200 1610 (C=N), 1561 (C=C), 1474 (N-O), 1282 (N-N=N-), 1118 (C-F). ¹H-NMR (300 MHz,
201 DMSO, ppm) δ: 7.48 (d, 1H, Ar-H), 7.41-7.29 (m, 5H, Ar-H), 6.97 (s, 1H, isoxazole), 6.91
202 (d, 1H, Ar-H), 6.80 (s, 1H, Ar-H). ¹³C NMR (125 MHz, DMSO-d₆) δ 170.3, 165.5, 162.5,
203 152.5, 131.8, 130.7, 128.5, 127.4, 120.5, 113.6, 102.1, 101.2. MS m/z: 325 [M⁺], Anal.
204 Calcd. (%) for C₁₆H₉F₂N₅O: C, 59.08; H, 2.79; F, 11.68; N, 21.53; O, 4.92%. Found: C,
205 59.20; H, 2.83; N, 21.48.

206 *2.5.4. Synthesis of 1-(5-(4-fluorophenyl)-isoxazol-3-yl)-5-phenyl-1H-tetrazole, 4d*

207 Yellowish white solid, Yield: 74%; m.p.(153-155°C).IR (KBr disk, cm⁻¹):ν3063 (Ar-CH),
208 2896 (CH), 1682 (C=N), 1544 (C=C), 1485 (N-O), 1288 (N-N=N-), 1163 (C-F). ¹H-NMR
209 (300 MHz, DMSO, ppm) δ: 7.52 (d, 2H, Ar-H), 7.42-7.26 (m, 5H, Ar-H), 7.11 (d, 2H, Ar-H),
210 6.98 (s, 1H, isoxazole). ¹³C NMR (125 MHz, DMSO-d₆) δ 170.8, 163.7, 151.2, 132.5, 130.2,
211 129.6, 128.1, 115.2, 101.5. MS m/z: 308 [M⁺], Anal. Calcd. (%) for C₁₆H₁₀FN₅O: C, 62.54;
212 H, 3.28; F, 06.18; N, 22.79; O, 4.92.

213 *2.5.5. Synthesis of 4-nitro-2-(3-(5-phenyl-1H-tetrazol-1-yl)-isoxazol-5-yl) phenol, 4e*

214 Reddish brown solid, Yield: 73%; m.p.(139-141°C).IR (KBr disk, cm⁻¹):ν3577 (OH), 3053
215 (Ar-CH), 2902 (CH), 1689 (C=N), 1564 (NO₂), 1514 (C=C), 1485 (N-O), 1284 (N-N=N-).
216 ¹H-NMR (300 MHz, DMSO, ppm) δ: 7.92 (s, 1H, Ar-H), 7.74 (d, 1H, Ar-H), 7.56-7.27 (m,
217 5H, Ar-H), 7.13 (d, 1H, Ar-H), 7.05 (s, 1H, isoxazole), 4.92 (s, 1H, OH). ¹³C NMR (125
218 MHz, DMSO-d₆) δ 170.7, 163.8, 152.5, 140.6, 132.2, 130.8, 128.6, 127.8, 122.4, 120.9,

219 101.2. MS m/z: 350 [M⁺], Anal. Calcd. (%) for C₁₆H₁₀N₆O₄: C, 54.86; H, 2.88; N, 23.99; O,
220 18.27.

221 *2.5.6. Synthesis of 1-(5-(2-chloro-4-fluorophenyl)-isoxazol-3-yl)-5-phenyl-1H-tetrazol, 4f*

222 White solid, Yield: 68%; m.p.(145-147°C).IR (KBr disk, cm⁻¹):v3095 (Ar-CH), 2848 (CH),
223 1610 (C=N), 1508 (C=C), 1465 (N-O), 1288 (N-N=N-), 1120 (C-F), 727 (C-Cl). ¹H-NMR
224 (300 MHz, DMSO, ppm) δ:7.51 (d, 1H, Ar-H), 7.43-7.25 (m, 5H, Ar-H), 7.18 (d, 1H, Ar-H),
225 7.02 (s, 1H, Ar-H), 6.95 (s, 1H, isoxazole). ¹³C NMR (125 MHz, DMSO-d₆) δ 168.7, 165.8,
226 153.5, 135.2, 134.0, 132.5, 130.4, 131.7, 128.1, 115.7, 102.6. MS m/z: 341 [M⁺], Anal.
227 Calcd. (%) for C₁₆H₉ClFN₅O: C, 56.24; H, 2.65; Cl, 10.37; F, 5.56; N, 20.49.

228 *2.5.7. Synthesis of 1-(5-(2, 4-dimethoxyphenyl)-isoxazol-3-yl)-5-phenyl-1H-tetrazol, 4g*

229 Brown solid, Yield: 72%; m.p.(172-174°C).IR (KBr disk, cm⁻¹):v3056 (Ar-CH), 2880 (CH),
230 1608 (C=N), 1560 (C=C), 1485 (N-O), 1288 (N-N=N-), 1163 (OCH₃). ¹H-NMR (300 MHz,
231 DMSO, ppm) δ: 7.57-7.28 (m, 5H, Ar-H), 7.18(d, 1H, Ar-H), 6.84 (s, 1H, isoxazole), 6.77 (d,
232 1H, Ar-H), 6.56 (s, 1H, Ar-H), 3.78 (s, 6H, OCH₃). ¹³C NMR (125 MHz, DMSO-d₆) δ 172.6,
233 163.2, 160.3, 157.4, 153.2, 132.4, 130.5, 128.5, 127.2, 109.2, 106.8, 102.6, 56.4. MS m/z:
234 349 [M⁺], Anal. Calcd. (%) for C₁₈H₁₅N₅O₃: C, 61.89; H, 4.33; N, 20.05; O, 13.74.

235 *2.5.8. Synthesis of 1-(5-(2, 4, 6-trimethoxyphenyl)-isoxazol-3-yl)-5-phenyl-1H-tetrazole, 4h*

236 Brown solid, Yield: 76%; m.p.(131-133°C).IR (KBr disk, cm⁻¹):v3055 (Ar-CH), 2877 (CH),
237 1606 (C=N), 1564 (C=C), 1485 (N-O), 1288 (N-N=N-), 1163 (OCH₃). ¹H-NMR (300 MHz,
238 DMSO, ppm) δ: 7.42-7.29 (m, 5H, Ar-H), 6.89 (s, 1H, isoxazole), 6.82 (s, 2H, Ar-H), 3.65 (s,
239 9H, OCH₃). ¹³C NMR (125 MHz, DMSO-d₆) δ 172.8, 164.5, 152.2, 132.4, 131.2, 129.1,
240 128.7, 102.3, 94.2, 57.5. MS m/z: 379 [M⁺], Anal. Calcd. (%) for C₁₉H₁₇N₅O₄: C, 60.15; H,
241 4.52; N, 18.46; O, 16.87.

242 *2.5.9. Synthesis of 1-(5-(2-nitrophenyl)-isoxazol-3-yl)-5-phenyl-1H-tetrazole, 4i*

243 Yellowish brown solid, Yield: 68%; m.p.(174-176°C).IR (KBr disk, cm⁻¹):v3086 (Ar-CH),
244 2893 (CH), 1623 (C=N), 1548 (NO₂), 1517 (C=C), 1485 (N-O), 1276 (N-N=N-). ¹H-NMR
245 (300 MHz, DMSO, ppm) δ: 7.73-7.58 (m, 4H, Ar-H), 7.50-7.27 (m, 5H, Ar-H), 7.08 (s, 1H,
246 isoxazole). ¹³C NMR (125 MHz, DMSO-d₆) δ 170.2, 152.6, 146.8, 136.4, 132.4, 131.3,
247 130.5, 129.2, 128.4, 127.3, 120.6, 103.2. MS m/z: 334 [M⁺], Anal. Calcd. (%) for
248 C₁₆H₁₀N₆O₃: C, 57.49; H, 3.02; N, 25.14; O, 14.36.

249 2.5.10. Synthesis of 1- (5- (3-bromo-4-methoxyphenyl)-isoxazol-3-yl)-5-phenyl-1H-tetrazole,
250 4j

251 Creamy white solid, Yield: 73%; m.p.(136-138°C).IR (KBr disk, cm^{-1}): ν 3066 (Ar-CH), 2877
252 (CH) 1602 (C=N), 1564 (C=C), 1465 (N-O), 1261 (N-N=N-), 1114 (OCH₃), 676 (C-Br). ¹H-
253 NMR (300 MHz, DMSO, ppm) δ :7.48 (s, 1H, Ar-H), 7.42-7.25 (m, 5H, Ar-H), 7.21 (d, 1H,
254 Ar-H), 7.12(s, 1H, isoxazole), 6.91 (d, 1H, Ar-H), 3.68 (s, 3H, OCH₃). ¹³C NMR (125 MHz,
255 DMSO-d₆) δ 170.2, 158.9, 151.7, 135.2, 132.2, 129.8, 128.2, 127.2, 125.4, 118.0, 101.4, 56.4.
256 MS m/z: 397 [M⁺], Anal. Calcd. (%) for C₁₇H₁₂BrN₅O₂: C, 51.27; H, 3.04; Br, 20.07; N,
257 17.59; O, 08.04.

258 2.6. Anticancer activity

259 Preliminary anticancer assay was performed as per the protocol of National Cancer Institute,
260 USA (Bekircan et al., 2006). The synthesized compounds were evaluated at single
261 concentration of 10⁻⁵ M towards the panel of 60 cell lines derived from different cancer types
262 such as: leukemia, lung, colon, CNS, melanoma, ovarian, renal, prostate and breast cancers.
263 All synthesized compounds were registered on its website and seven compounds 4b, 4c, 4d,
264 4f, 4g, 4h, and 4i were selected. All the selected compounds were added to a previously
265 prepared cell culture at a single concentration. The cell culture was incubated for 48 h. End
266 point was determined by a protein binding dye, sulforhodamine B (SRB). The result of
267 anticancer activity of each compound was reported as the percent growth of treated cell lines
268 when compared to untreated control cells (Zahera et al., 2017, Lokhande et al., 2013).

269 Using the seven absorbance measurements [time zero, (Tz), control growth, (C), and test
270 growth in the presence of drug at the five concentration levels (Ti)], the percentage growth
271 was calculated at each of the drug concentration levels.

272 Percentage growth inhibition is calculated as:

273 For concentrations when $T_i > / = T_z$:

$$274 \quad [(T_i - T_z) / (C - T_z)] \times 100$$

275 For concentrations when $T_i < T_z$:

$$276 \quad [(T_i - T_z) / T_z] \times 100$$

277 2.7. Molecular Docking studies

278 Molecular docking studies were done to check out interaction between the synthesized
279 compounds and the active site of the receptor. The computation was carried out using
280 Schrodinger molecular modeling software package. Docking was performed by using the
281 Glide integrated with Maestro (Schrodinger, LLC, 2011) interface on the Linux operating
282 system. The starting coordinates of the human Tubulin-Colchicine: Stathmin-Like Domain
283 Complex [PDB:1SA0] was taken from the Protein Data Bank (www.rcsb.org) and further
284 modified for docking calculations. A compound library of synthesized 1-(5-substituted
285 phenyl) isoxazol-3-yl)-5-phenyl-1H-tetrazole derivatives was built on Maestro build panel
286 and minimized in Schrodinger and optimized by means of the OPLS molecular mechanics
287 force field using default settings. For Glide (Schrodinger) calculations, the protein Tubulin-
288 Colchicine: Stathmin-Like Domain Complex [PDB: 1SA0] was optimized with the “protein
289 preparation wizard” workflow by subjecting a cycle of constrained minimization steps
290 allowing a maximum root mean square deviation (RMSD) from the original structure. For
291 Glide (Schrodinger) calculations, Tubulin-Colchicine: Stathmin-Like Domain Complex was
292 imported to Maestro (Schrodinger), the co-crystallized ligands were identified and removed
293 from the structure. Docking was performed using Glide.

294

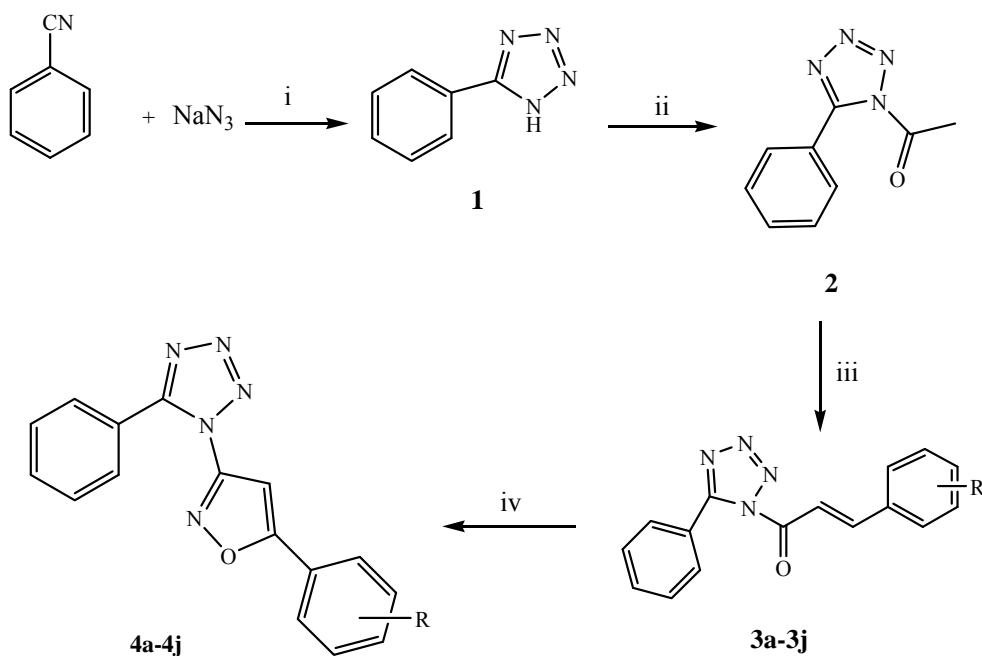
295 **3. Results and discussions**

296 *3.1. Chemistry*

297 The reaction sequences used for the synthesis of titled compounds are shown in Scheme 1.
298 The equal stoichiometric amount of sodium azide, benzonitrile, ammonium chloride were
299 refluxed with dimethylformamide to obtain 5-phenyltetrazole 1, which after acetylation
300 yield 5-phenyl-1-acetyl tetrazole 2. The compound 2 was treated with substituted aromatic
301 aldehyde in ethanol and sodium hydroxide under ice cooled condition then acidified to form
302 3-(substituted phenyl)-1-(5-phenyl-1H-tetrazol-1-yl) prop-2-en-1-one 3a-3j. The compounds
303 3a-3j was refluxed with hydrazine hydrate and acetic acid to form titled compounds 4a-4j.
304 Structures of all the synthesized compounds were characterized by their spectral data
305 interpretation. The IR spectral data of compounds 4a-4j showed characteristic absorption
306 band of methene group of isoxazole ring at 2877-2921 cm^{-1} . The absorption bands of the
307 C=N group and C=C group appeared at 1602-1691 cm^{-1} and 1502-1564 cm^{-1} respectively.
308 The presence of N-O group and N=N- group was confirmed by characteristic absorption band
309 at 1438-1485 cm^{-1} and 1261-1288 cm^{-1} . In $^1\text{H-NMR}$ spectra of compounds 4a-4j, the one

310 proton of isoxazole ring appeared as singlet in the range of δ 6.84-7.12 ppm. Protons of
311 methoxy group showed as singlet at δ 3.68-3.78 ppm. The one proton of OH group of
312 compound 4e showed a singlet at δ 4.92. All the aromatic protons were observed in the
313 expected regions. Mass spectra of the compounds showed M+1 in agreement with their
314 molecular formula.

315



Scheme 1 Synthetic routes to titled compounds (4a-4j).

Reagents and conditions: (i) NH_4Cl , DMF, 125°C , 7-8 h; (ii) $(\text{CH}_3\text{CO})_2\text{O}$, H_2SO_4 ; $60-70^\circ\text{C}$, 15-20 min; (iii) R-CHO/ NaOH , $\text{C}_2\text{H}_5\text{OH}$; (iv) $\text{NH}_2\text{OH}\cdot\text{HCl}$, KOH , $\text{C}_2\text{H}_5\text{OH}$, 130°C , 4-5h.

316
317

318 3.2. Anticancer activity

319 All the synthesized compounds were submitted to National Cancer Institute (NCI, USA), for
320 anticancer activity under the drug discovery program. Only seven compounds were selected
321 according to NCI protocol and screened for *in vitro* anticancer activity at a single high dose 10^{-5}
322 M in full NCI 60 cell lines panel. Anticancer activity data of synthesized compounds on NCI
323 cancer cell lines were presented in Table 1. The synthesized compounds displayed moderate to
324 low activity in the *in vitro* screening in all tested cancer cell line. The compound **4b** and **4i**
325 were found to be most active compounds by showing 65.50 and 58.95 growth percent and
326 highly active on MOLT-4 (Leukemia) and CCRF-CEM (Leukemia) cell line respectively while
327 rest of the compounds showed less activity.

328 The possible mechanism of action of synthesized compounds would be inhibition of non-
 329 covalent polymerization of tubulin into microtubules. Tubulin, the major structural component
 330 of microtubules, is a target for the development of anticancer agents. Microtubules are a key
 331 component of cytoskeleton, and they are involved in a wide range of cellular functions,
 332 including regulation of motility, cell division, organelle transport, maintenance of cell
 333 morphology, and signal transduction. The essential role of microtubules in mitotic spindle
 334 formation and proper chromosomal separation makes them one of the most attractive targets for
 335 the design and development of synthetic antitumor drugs.

Table 1. Anticancer activity of title compounds

Compound	60 cell line assay in one dose 10^{-5} M conc.				
	NSC code	Mean growth %	Range of growth %	The most sensitive cell line	Growth % of most sensitive cell line
4b	761443	95.42	65.50 - 152.35	MOLT-4 (Leukemia)	65.5
4c	778577	100.12	75.41 - 116.41	SR (Leukemia)	69.34
				A498 (Renal cancer)	75.41
4d	761444	95.48	73.81 - 131.39	HOP-92 (Non small cell lung cancer)	81.22
				HL-60 (Leukemia)	73.81
4f	778579	99.11	68.97-117.02	UO-31 (Renal cancer)	78.96
				BT-459 (Breast cancer)	68.97
4g	778578	100.95	80.52-121.84	UO-31 (Renal cancer)	80.52
4h	778580	101.81	79.84-118.88	A498 (Renal cancer)	84.9
				A498 (Renal Cancer)	79.84
4i	761445	93.18	58.95 - 119.00	BT-459 (Breast cancer)	80.62
				CCRF-CEM (Leukemia)	58.95
				SR (Leukemia)	74.13

336

337 3.3. Molecular Docking studies

338 To check the molecular interaction and affinity of binding of Tubulin-Colchicine: Stathmin-
 339 Like Domain Complex [PDB: 1SA0] of synthesized 1-(5-substituted phenyl) isoxazol-3-yl)-
 340 5-phenyl-1H-tetrazole derivatives, all the ligands were docked into the domain of Tubulin-
 341 Colchicine: Stathmin-Like Domain Complex [PDB: 1SA0]. Docking was done using Glide
 342 module of Schrodinger software. Docking results of these ligands are given in Table 2.
 343 Compounds **4b** and **4i** showed good interaction with Tubulin-Colchicine: Stathmin-Like

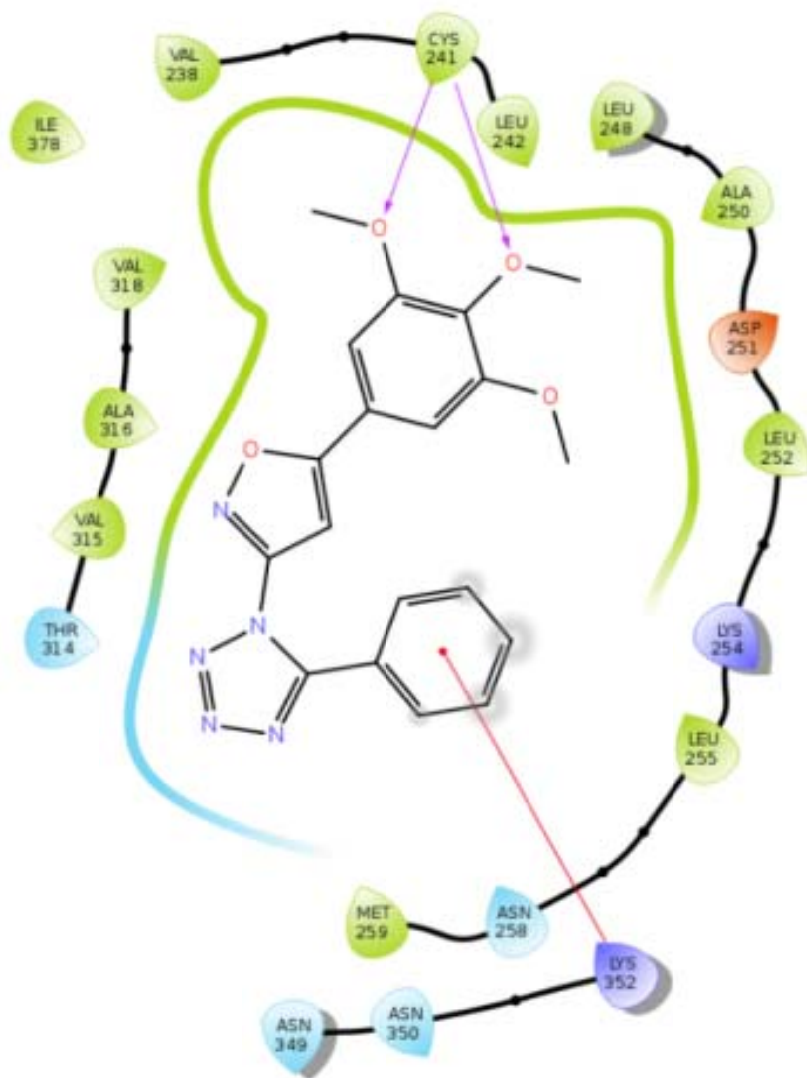
344 Domain Complex and these results matched with wet laboratory findings. Compound **4b** was
345 found to have a glide score of **-6.551** with two strong H bonds between Cys-241 and methoxy
346 groups and one Pi-cation bond with Lys-352 and compound **4i** was having have a glide score
347 of **-6.421** with strong H bond between Lys-352 and nitro group of the compound. They were
348 also found to have strong hydrophobic contacts with the residues of active site. **This signifies**
349 **a strong binding of the molecules to the receptor at the site LYS 352.** The ligand receptor
350 interactions of compound **4b** and **4i** are shown in Fig 1.
351

Table 2 Docking results of title compound with tubulin-colchicine: stathmin-like domain complex [PDB: 1SA0]

Comp.	Glide score/docking score	Glide ligand efficiency	Glide rotatable bonds	Glide RMSD to input
4b	-6.551	-0.273	3	146.118
4c	-4.711	-0.168	6	148.736
4d	-4.623	-0.173	5	148.863
4f	-4.832	-0.165	6	147.362
4g	-4.594	-0.177	5	147.86
4h	-5.876	-0.21	6	148.944
4i	-6.421	-0.256	4	146.391

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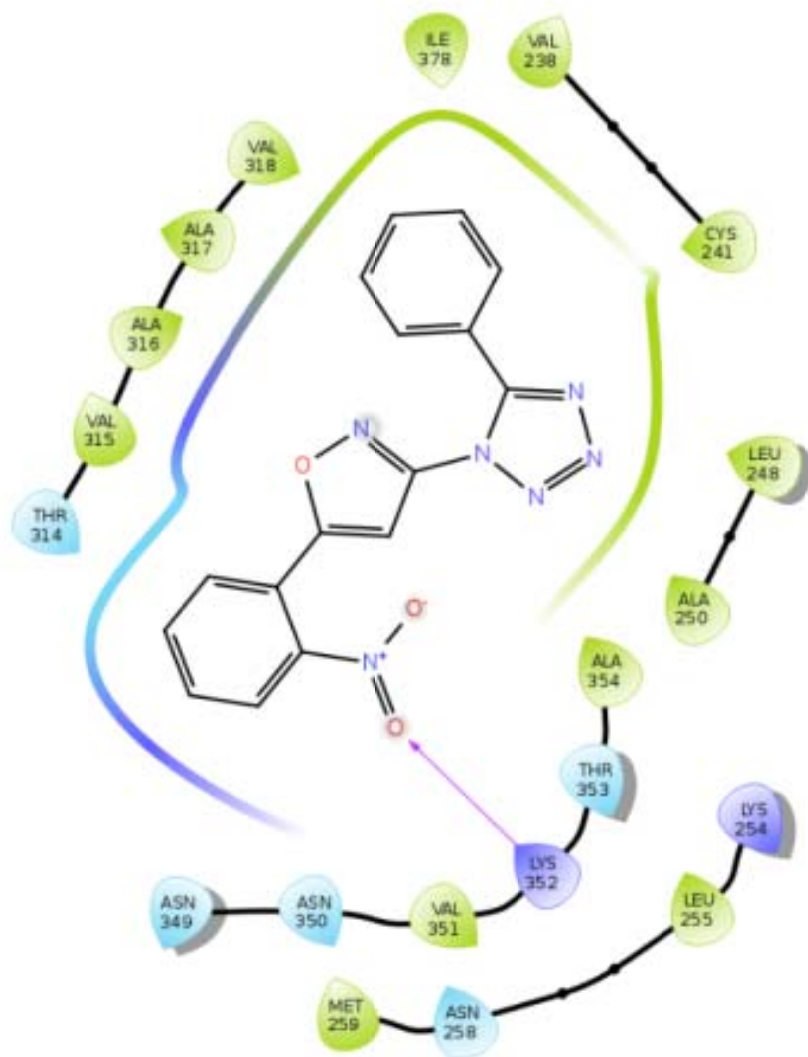
353



354

355

4b



356

4i

357 **Fig. 1** Schematic representation of interaction between compounds **4b** and **4i** with
 358 theTubulin-Colchicine: Stathmin-Like Domain Complex (1SA0), active site amino acid
 359 residues, i.e., Cys-241 and Lys-352.
 360

361 These compounds shows an increased anticancer activity and hence these are an ideally suited
 362 for further modifications to obtain more effective anticancer compounds. Hence, our study has
 363 identified some of 1-(5-substituted phenyl) isoxazol-3-yl)-5-phenyl-1H-tetrazole derivatives
 364 that are important for cytotoxic activity against a panel of human cancer cell lines, and these
 365 findings indicates the need for additional investigations with respect to some new 1-(5-
 366 substituted phenyl) isoxazol-3-yl)-5-phenyl-1H-tetrazole derivatives as anticancer agents.

367 **4. Conclusions**

368 In the present work, 1-(5-substituted phenyl) isoxazol-3-yl)-5-phenyl-1H-tetrazole derivatives
369 have been synthesized from sodium azide and benzonitrile as reported by author earlier
370 (Kaushik et al., 2015). All the synthesized compounds were characterized by different
371 spectral technique. The synthesized compounds were assayed for their in vitro anticancer
372 activity by the National Cancer Institute (NCI), USA under the drug discovery program. The
373 compound **4b** and **4i** were found to be most active compounds by showing 65.50 and 58.95
374 growth percent on MOLT-4 (Leukemia) and CCRF-CEM (Leukemia) cancer cell line
375 respectively. Molecular docking of these compounds was done by using Glide module of
376 Schrodinger software which state that compounds **4b** and **4i** showed good interaction with
377 Tubulin-Colchicine: Stathmin-Like Domain Complex and obtained results matched with wet
378 laboratory findings.

379
380

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