

***In vitro* study on anti-Salmonella activities of Boerhaavia diffusa (L. syn) Leaf extract**

Abstract

Various strategies have been employed in the treatment and management of *Salmonella* infection however, *Salmonella* strains have gained resistance to antibiotics. This study was to determine *in vitro* anti-Salmonella activity of *Boerhaavia diffusa* leaf extract against clinical isolate of *Salmonella typhi* and *Salmonella typhi* ATCC 14028. The aqueous and ethanol extracts of *B. diffusa* was studied for their antibacterial activity against pathogenic *Salmonella typhi*. This study was carried out between April and September, 2018. The *in vitro* antibacterial activity was performed by agar well diffusion method and broth dilution using spectrophotometric method and the results were expressed as the average diameter of zone of inhibition of bacterial growth around the well and optical density respectively. It was observed that aqueous extract exerted slightly higher activity than ethanolic extract as revealed by mean diameter of zone of inhibitions at concentration of 200 mg/ml, aqueous extract had 35.21±0.47 mm (*Salmonella typhi* ATCC 14028) compared with ethanol extract 26.41±0.32 mm (clinical). However, in broth dilution method, ethanol extract significantly ($p=0.05$) reduced the cell, at 48 hours, the optical density of clinical isolate of *S. typhi* treated at concentration of 200 mg/ml of extract was 0.47±0.02 nm while at the same concentration of extract, aqueous extract had optical density of 0.52±0.11 nm respectively. Phytochemical assay revealed that tannin (5.18±0.02 mg/g) and quinone (8.45±0.13 mg/g) in ethanol extract was significantly ($p=0.05$) higher than aqueous extract while saponin (14.18±0.06 mg/g) was higher in aqueous extract. The ethanol and aqueous extracts of leaves of *B. diffusa* whole plant exhibited significant antibacterial activity against both clinical and typed *Salmonella typhi*. Therefore, the plant extract could be used for the treatment of Salmonellosis, however, the *in vivo* studies is needed to ascertain the safety of the extract.

Key words: Anti-*Salmonella* activity, plant extracts, agar well diffusion, broth dilution, *Salmonella* strains

32 **Introduction**

33 The bacterium *Salmonella typhi* causes typhoid fever [1, 2]. The bacterium is a gram-negative,
34 motile, non-sporing, non-capsulated bacillus that can be contracted through contaminated water,
35 milk, food or fruits and vegetables or via convalescent or chronic carriers [3]. It has also been
36 linked with zoonotic transmission via reptiles and common domestic pets [4]. *Salmonella*
37 *enterica*, which is a group of Gram-negative bacterial pathogens capable of infecting humans and
38 animals, cause significant morbidity and mortality worldwide [5]. Certain serotypes adapted to
39 human, such as *Salmonella typhi* (*S. typhi*) and *Salmonella paratyphi* (*S. paratyphi*), usually
40 cause severe diseases in humans, such as enteric fevers (typhoid and paratyphoid fevers). In most
41 endemic areas like Africa, Asia, and Latin America, approximately 90% of enteric fever is
42 typhoid. This disease is an important global health problem with an estimated 16 million cases
43 and 600 000 deaths each year.

44 Various strategies have been employed in the treatment and management of *Salmonella*
45 infection. Fluoroquinolones and tetracyclines are most commonly used to treat *Salmonella*
46 infections. However, *Salmonella* strains resistant to these antibiotics have been reported in Korea
47 and other countries [6]. One major concern to public health has been the global dissemination of
48 *S. typhimurium* Definitive Type 104, which is resistant to cotrimoxazole, nalidixic acid and
49 ampicillin [7]. The rise in antibiotic-resistant strains has led to increased interest in use of plant
50 materials to develop new effective drugs [6]. Moreover, conventional antityphoid drugs are
51 becoming more and more unavailable to the common man in Africa due to increased cost [8].

52 The rise in antibiotic-resistant strains has led to increased interest in use of plant materials to
53 develop new effective drugs [6]. It has been reported that 80% of the world population are rural
54 dwellers and rely on medicinal plants for their daily medications, also, plants have been
55 reported to have minimal or no side effects compared to antibiotics [9, 10]. *Boerhaavia diffusa*
56 (Spreading Hogweed in English), belonging to the family of the Nyctaginaceae, is mainly a
57 diffused perennial herbaceous creeping weed of India (known also under its traditional name as
58 *Punarnava*). *Boerhaavia diffusa* is traditionally known in Nigeria as *Etiponla* in Yoruba,
59 *Azeigwe* in Igbo and *Babba-juju* in Hausa. *B. diffusa* is a perennial creeping weed, prostrate or
60 ascending herb, up to 1 m long or more, having spreading branches [11].

61 The root, leaves, aerial parts and the whole plant of *B. diffusa* (L. syn) are used worldwide for
62 the treatment of a number of disorders e.g. liver complaints, kidney disorders, rheumatism e.t.c.

63 [12]. The quest to identify and isolate novel phytochemicals from *B. diffusa* has led many
64 researchers to discover various compounds such as flavonoids, alkaloids, glycosides, steroids,
65 triterpenoids, lipids, lignans, carbohydrates, proteins, and glycoproteins from its leaves, stems,
66 seeds and roots [13]. Sourav [14], explored the Anti bacterial from *Boerhaavia diffusa*. In his
67 study, the chloroform and alcohol extracts of the plant were screened against six bacteria viz
68 *Staphylococcus aureus*, *Escherichia coli*, *Proteus mirabilis*, *Salmonella typhimurium*,
69 *Pseudomonas aeruginosa* and *Klebsiella aerogenosa*. Chloroform extract showed activity
70 against *E. coli*, *S. typhimurium* and *P. aeruginosa* while the alcohol extract was active against *P.*
71 *mirabilis* and *S. typhimurium*. The present study was undertaken to further investigate
72 antibacterial activity of *Boerhaavia diffusa* on typed and clinical strains of *Salmonella typhi*
73 with the view to provide scientific evidence for its application as a medicinal plant.

74 **Materials and methods**

75 **Collection of leaves of *Boerhaavia***

76 Fresh leaves of *Boerhaavia diffusa* (Plate 1) were collected from the School of Health
77 Technology, Oda Road, Akure, and identified in the Department of Crop, Soil and Pest
78 Management, Federal University of Technology, Akure Ondo State.



79
80 **Plate 1: Photograph of *Boerhaavia diffusa* leaf**

81 82 **Preparation of plant extract**

83 **Aqueous extraction**

84 The aqueous extractions of the water soluble ingredient were carried out using the filter method.
85 A 2 g of each of the grounded leaves were extracted by successive soaking for 2 days using 50
86 ml of distilled water in a 250 ml sterile conical flask. The extracts were concentrated in vacuum
87 at 60 °C and stored in universal bottles and refrigerated at 4 °C prior to use [15].

88 **Ethanol extraction**

89 The organic solvent leaf extract was prepared by 2 g of plant mixture with ethanol and kept for
90 two days. The extract was concentrated to one-fifth volume, filter sterilized and stored at 4 °C
91 [15].

92 **Test organism**

93 The clinical bacterial strains were obtained from the Department of Microbiology, Federal
94 University of Technology Akure. Clinical *Salmonella typhi* and typed (ATCC 14028) *Salmonella*
95 *typhi* were used. The isolates were confirmed based on cultural, morphological and biochemical
96 characteristics following standard methods of identifying *Salmonella typhi* [16]. The bacterial
97 strain was grown in nutrient broth for 12-18 hours at 37 °C on rotary shaker. Cells were grown at
98 37 °C for 18 hours and cultures were kept at 4 °C.

99 **Antimicrobial susceptibility tests**

100 **Standardization of the inoculum**

101 The inoculum was prepared by inoculating colonies of fresh test cultures into sterile
102 distilled water. The turbidity was compared to 0.5McFarland standard prepared according to
103 method of Cheesbrough [16]

104 **Antibacterial susceptibility assay**

105 The extracts were dissolved and diluted using 50 % v/v dimethylsulphoxide (DMSO) to
106 obtain different concentrations (50, 100 and 200 mg) in 1 mL. The 50 mg/ml, 100 mg/ml and
107 200 mg/ml of the extracts of *B. diffusa* leaves were introduced into the wells of Muller Hinton
108 agar plate. The plates were incubated aerobically at 37 °C and examined after 24 hours. The
109 plates were examined for microbial growth inhibition and the Inhibition Zone Diameter (IZD)
110 was measured to the nearest millimeter and compared with those produced by the commercial
111 antibiotic ciprofloxacin which was used as control. Effect of extract on anti-*Salmonella* efficacy
112 of the extract in broth was also assayed using spectrophotometric method, the absorbance of the
113 tube was read at 620 nm [16, 17].

114 **Antibiotics sensitivity test using commercial**

115 Antibiotics sensitivity test of the bacterial isolates were determined by disc diffusion
116 method as described by Cheesbrough [16]. Standard inoculum of 18 hours broth was spread on
117 Muller Hinton agar using sterile swab in triplicate. The antibiotic discs were placed on the plate
118 at equidistance. The plates were then incubated for 24 hours at 37°C and diameter of zone of
119 inhibition were measured and recorded. The commercial antibiotics discs (Fondoz Laboratories
120 Ltd, Nigeria) used were; Chloramphenicol (CH) 30 µg, Sparfloxacin (SP) 25 µg, Ciprofloxacin
121 (CPX) 10µg, Amoxicillin (AM) 25µg, Augmentin (AU) 30µg, Gentamycin (CN) 10µg,
122 Pefloxacin (PEF) 5µg, Ofloxacin (OFX) 5µg, Streptomycin (S) 10 µg and Septra (SXT) 30µg.

123 **Minimum Inhibitory Concentration (MIC) and Minimum Bacteriocidal Concentration** 124 **(MBC) of *Boerhaavia diffusa* Extracts**

125 The Minimum Inhibitory Concentration (MIC) and Minimum Bacteriocidal Concentration
126 (MBC) of the extracts were determined using the broth (tube) dilution technique [18]. Dilutions
127 of the extract in Mueller Hinton broth were prepared in tubes. The concentration of inoculum
128 was also standardized to 0.5 McFarland's turbidity, The Mueller Hinton broth in tubes
129 containing the different concentration of plant extract, 50, 100, and 200 mg/ml were then
130 inoculated with 0.5 ml of the standardized culture. The tubes were then incubated at 37 °C for 24
131 hours. MIC and MBC values were recorded.

132 **Screening of phytochemical compounds**

133 The various solvent extracts of the powder of leaves of *Boerhaavia diffusa* were subjected to
134 phytochemical tests for the identification of various action constituents using the method of
135 Marcelin *et al.* [17]. The following major pharmaceutical valuable phytochemical compounds
136 were analyzed qualitatively and quantitatively; alkaloids, phenols, tannins, flavonoids, quinones,
137 saponins, terpenoids, sterols and cardiac glycosides.

138 **Statistical analysis of data**

139 Data obtained were subjected to analysis of variance and means were compared using Duncan's
140 New Multiple Range Test (DNMRT) with the aid of SPSS software at $p \leq 0.05$ level of
141 significance.

142 **Results and Discussion**

143 *Salmonellosis* and enteric fever are always a public health concern in most developing countries,
144 which are mostly low or middle-income countries with inadequate sanitation and hygiene,
145 particularly, regarding food, water and disposal of human excreta [17]. Different plants and their

146 parts (flowers, buds, leaves, stem, bark, fruits, skin, pulp and root) have been used for thousands
147 of years to enhance the flavour and aroma of food. In addition, plants are rich in a wide variety
148 of second metabolites such as Alkaloids, Flavonoids, Phenols, which were found in vitro to have
149 antimicrobial properties [17, 19].

150 In this study, extracts of *Boerhaavia diffusa* leaves were investigated for antibacterial activity
151 against *Salmonella typhi*. Plant extracts were used to investigate antibacterial activity against
152 two bacterial strains (Clinical *Salmonella typhi* and *Salmonella typhi* ATCC 14028). In this
153 study, antibacterial activity of *B. diffusa* leaf extracts was compared against the test bacteria
154 with activities of model antibiotics.

155 The test organisms used for this study were identified based on biochemical characteristics
156 common to *Salmonella typhi*. The result is presented in Table 1. The antibiotic sensitivity
157 patterns of commercial antibiotics on the two strains of *S. typhi* are presented in Figure 1. The
158 result revealed that the zones of inhibition of antibiotics against typed isolates was higher than
159 that of clinical isolates however, chloramphenicol had highest inhibition against the isolates
160 (STC=24.30±0.42 mm, STT=24.36±0.07 mm). The higher antibacterial activity of model
161 antibiotics is not surprising, since the antibiotics are in a refined state. The standard antibiotics
162 (ampicillin, amoxycillin, ciprofloxacin, ofloxacin, chloramphenicol) used in this study are first
163 line drugs employed in the treatment of typhoid fever [1].

164 The results of antibacterial activity of both water and ethanol crude extracts of *B. diffusa*
165 showed anti*Samonella* activity on the two strains of *S. typhi* tested at different concentrations,
166 with aqueous extract exerting slightly higher activity than ethanolic extract as revealed by mean
167 diameter of zone of inhibitions, 200 mg/ml of aqueous extract had highest (35.21±0.47) zone of
168 inhibition (Figure 2). Minimum inhibitory concentration (MIC) and minimum bactericidal
169 concentrations (MBC) of the extracts is shown in Tables 2. The ethanol and aqueous extract had
170 the same MIC (100 mg/ml) on typed isolate, also, there was no difference in the MIC and MBC
171 of ethanol and aqueous extract on typed isolate. The aqueous and ethanol extracts exhibited
172 different zones of inhibition against the isolates, however, aqueous extract had higher zones of
173 inhibition than ethanol extract. Antimicrobial action may be due to the synergistic action of
174 different chemical constituents, some of which probably are lost upon extraction with solvent
175 [15, 17, 20]. Water could be a better extraction solvent than ethanol for *B. diffusa* leaf, also, the
176 demonstration of higher activity by the aqueous solvent may be an indication that the

177 phytoconstituents in the plant leaves are more soluble in water than the organic solvent [21].
178 The antimicrobial potential of *B. diffusa* and other plants sourced from traditional healers
179 through an ethnobotanical survey of anti-infective plants in Egbado South in Ogun State,
180 Nigeria was previously reported by Abo and Ashidi [22]. This study also corroborates the
181 findings of Madani and Jain [23] who reported higher anti-*Salmonella* activity in aqueous
182 extract of *Terminalia belerica* than chloroform and acetone extracts. It has been reported that
183 different phytoconstituents have different degrees of solubility in different types of solvents
184 depending on their polarity. In a traditional setting, water is the solvent largely used to prepare
185 these concoctions.

186 The anti-*Salmonella* efficacy of *Boerhaavia diffusa* extracts in broth was assayed and were
187 shown in Figure 3, 4, 5 and 6. The result presented in Figure 3 revealed the effect of ethanol
188 extract on clinical isolate of *S. typhi*, it was noted that the extract significantly ($p < 0.05$) reduced
189 the cell, at 48 hours, the optical density of clinical isolate of *S. typhi* treated with 50, 100, 200
190 mg/ml of extract were 0.52 ± 0.03 , 0.50 ± 0.10 , 0.47 ± 0.02 nm respectively while at the same
191 concentration of extract, aqueous extract had optical density of 0.64 ± 0.21 , 0.54 ± 0.03 , 0.52 ± 0.11
192 nm respectively (Figure 4). Also, the anti-*Salmonella* efficacy of *B. diffusa* ethanol extracts on
193 typed isolate of *S. typhi* is shown in Figure 5. It was observed that the extract significantly
194 ($p < 0.05$) reduced the cell, at 48 hours, the optical density were of typed isolate of *S. typhi* treated
195 with 50, 100, 200 mg/ml of extract were 0.49 ± 0.00 , 0.48 ± 0.01 and 0.37 ± 0.12 while at the same
196 concentration, aqueous extract had optical density of 0.62 ± 0.03 , 0.53 ± 0.11 and 0.49 ± 0.21 nm
197 respectively (Figure 6). It was noted from this study that plant extracts tested by microdilution
198 technique and the optical density was measured after 48 hours showed that ethanol extract had
199 higher anti-*Salmonella* activity compared to aqueous extract which was higher in values obtained
200 from agar well diffusion technique. It could be that the bioactive components in ethanol extract
201 did not diffuse into agar in agar well but was able to inhibit microbial cells directly in broth. This
202 was previously reported by other findings that the active components of the extract does not
203 diffuse into Muller Hinton agar, however, they were able to cause inhibition of microbial cells in
204 broth microdilution [6, 24].

205 Both plant extracts (ethanolic and aqueous) were subjected to preliminary qualitative
206 phytochemical evaluation. The phytochemical profiles of the two solvent extracts from plant
207 used in this study are presented in Table 2. The analysis revealed the presence of alkaloids,

208 phenol, glycosides, steroids, carboxylic acid, reducing sugar, flavonoids, saponins, tannins,
 209 proteins, triterpenoids, quinines, carbohydrates and sterols. Also, tannin (5.18 ± 0.02 mg/g) and
 210 quinone (8.45 ± 0.13 mg/g) in ethanol extract was significantly ($p < 0.05$) higher than aqueous
 211 extract while saponin (14.18 ± 0.06 mg/g) was higher in aqueous extract. The preliminary
 212 qualitative phytochemical screening carried out showed that the leaf extracts of *B. diffusa*
 213 contain vital secondary metabolites such as alkaloids, saponins, tannins and glycosides. The
 214 bioactive compounds in medicinal plants have been reported to be the active principles
 215 responsible for the pharmacological potentials of medicinal plants [25]. The presence of these
 216 chemicals in the leaves and root of these plants justifies the local uses of these plants for the
 217 treatment of various ill conditions. Phytoconstituents such as saponins, phenolic compounds and
 218 glycosides have been reported to inhibit bacterial growth and to be protective to plants against
 219 bacterial and fungal infections [6, 26]. Ethanol extract of *Boerhavia diffusa* leaves possess some
 220 phytochemicals like Alkaloids, Anthraquinone, Glycoside, Flavanoids and Tannins. Saponins are
 221 natural glycosides that act as hypoglycemic, antifungal and serum cholesterol lowering agents
 222 in animals [27]. Saponins are essential elements in ensuring hormonal balance and synthesis of
 223 sex hormones [28]. Tannins are bitter polyphenolic compounds that hasten the healing of
 224 wounds. They also possess anti-diuretic and anti-diarrhea properties [28]. Terpenoids was
 225 present in both ethanolic extract of and aqueous extracts of AOU and AFU. Terpenoids have
 226 been found to be useful in the prevention and therapy of several diseases, including cancer.
 227 Terpenoids are also known to possess antimicrobial, antifungal, anti-parasitic, antiviral, anti-
 228 allergenic, antispasmodic, antihyperglycemic, anti-inflammatory and immunomodulatory
 229 properties [29]. The presence of these compounds promises its potential application in the
 230 treatment of microbial ailment. However, tannins were present in aqueous extract of but not in
 231 the ethanolic extract. Saponin and flavanoid is higher in the aqueous extract of the leaf (14.18
 232 and 11.26 mg/g) than the ethanolic extract (6.36 and 9.98 mg/g).

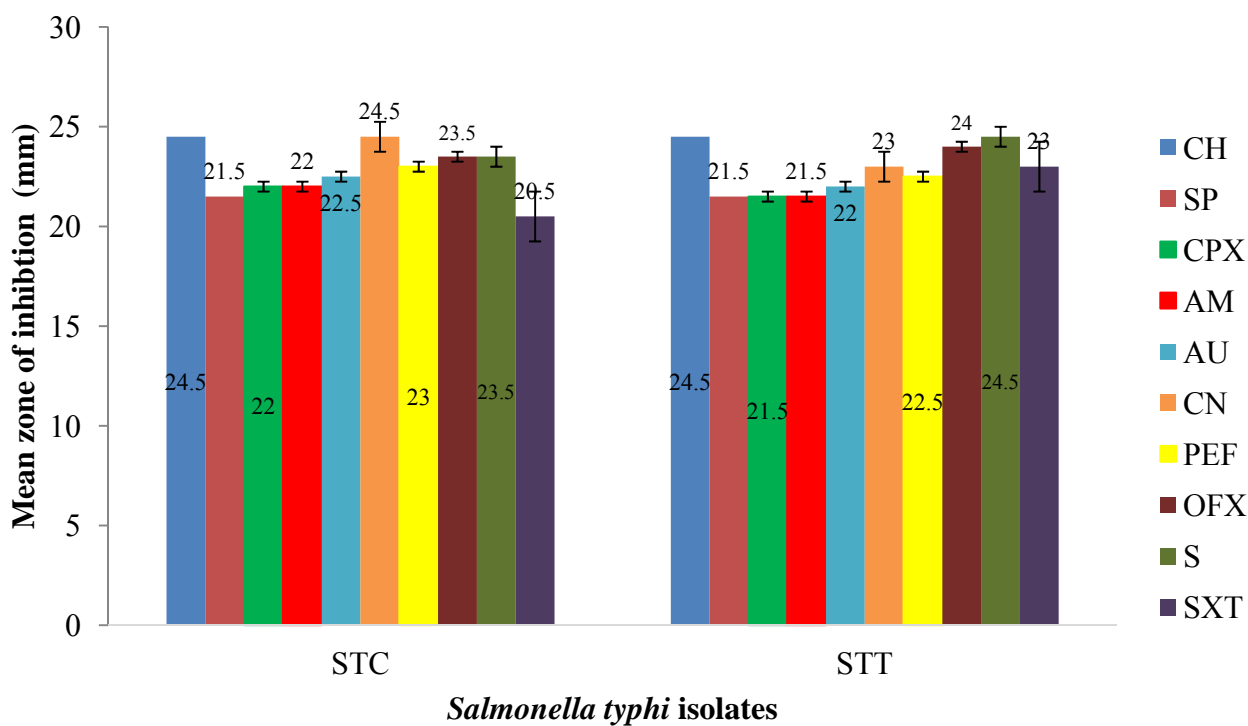
233 **Table 1: Biochemical characteristics of *Salmonella* strains**

Biochemical characteristics	<i>Salmonella typhi</i> (Clinical isolate)	<i>Salmonella typhi</i> (ATCC 14028)
Gram reaction	-ve	-ve
Shape	Rod	Rod
Motility	+ve	+ve
Catalase	+ve	+ve
Coagulase	-ve	-ve

Citrate	+ve	+ve
H ₂ S	+ve	+ve
Lactose	-ve	-ve
Glucose	+ve	+ve
Fructose	+ve	+ve
Sucrose	-ve	-ve
Galactose	+ve	+ve
Indole	-ve	-ve
Methyl red	+ve	+ve
Voges-Proskauer	-ve	-ve
Oxidase	-ve	-ve

234 Key: -ve = negative +ve = positive

235

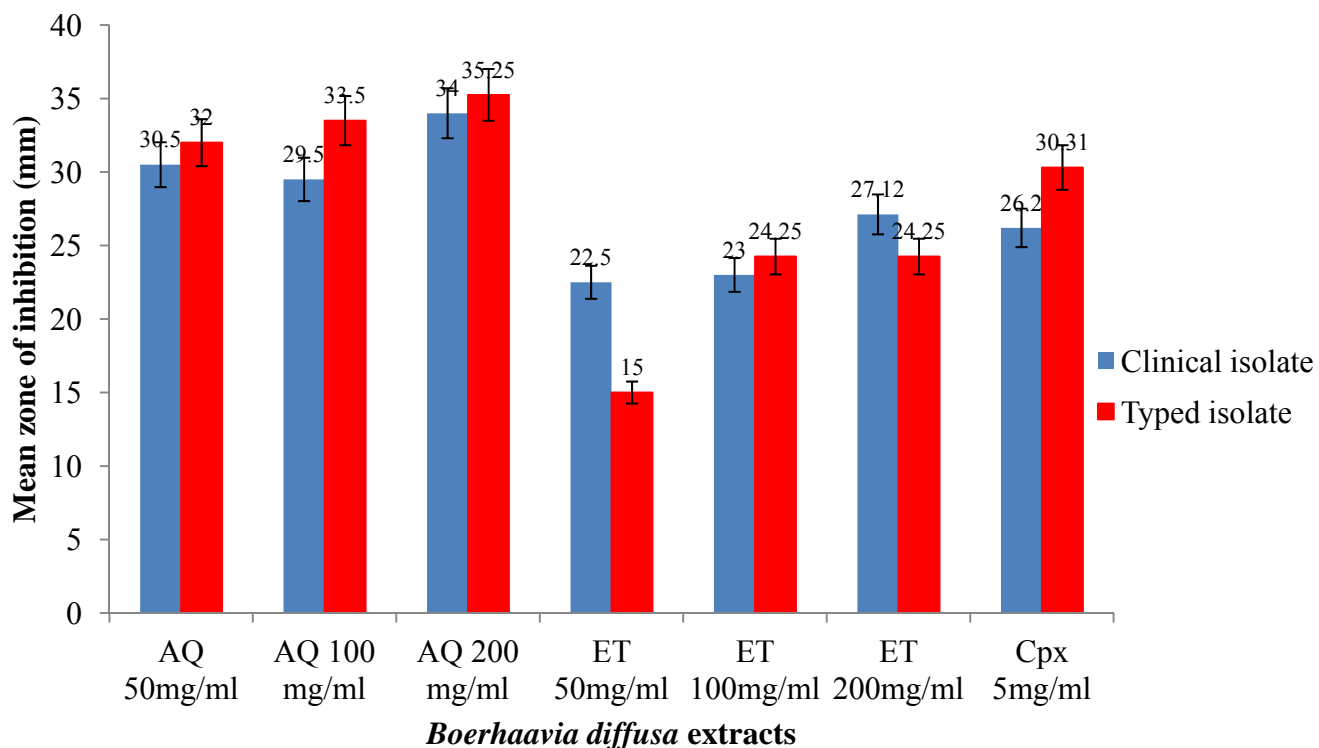


236

237 **Figure 1: Antibiotic sensitivity pattern of commercial antibiotic discs on *S. typhi* strains**

238 Key: STC – *Salmonella typhi* (clinical isolate), STT – *Salmonella typhi* (typed isolate),
 239 Chloramphenicol (CH) 30 µg, Sparfloxacin (SP) 25 µg, Ciprofloxacin (CPX) 10µg, Amoxicillin
 240 (AM) 25µg, Augmentin (AU) 30µg, Gentamycin (CN) 10µg, Pefloxacin (PEF) 5µg, Ofloxacin
 241 (OFX) 5µg, Streptomycin (S) 10 µg and Septra (SXT) 30µg.

242



244

245 **Figure 2: AntiSalmonella activity of *Boerhaavia diffusa* extracts**

246 **Key:** AQ = Aqueous extracts of *Boerhaavia diffusa*, ET= Ethanolic extracts of *Boerhaavia*
 247 *diffusa*, Cpx= Ciprofloxacin

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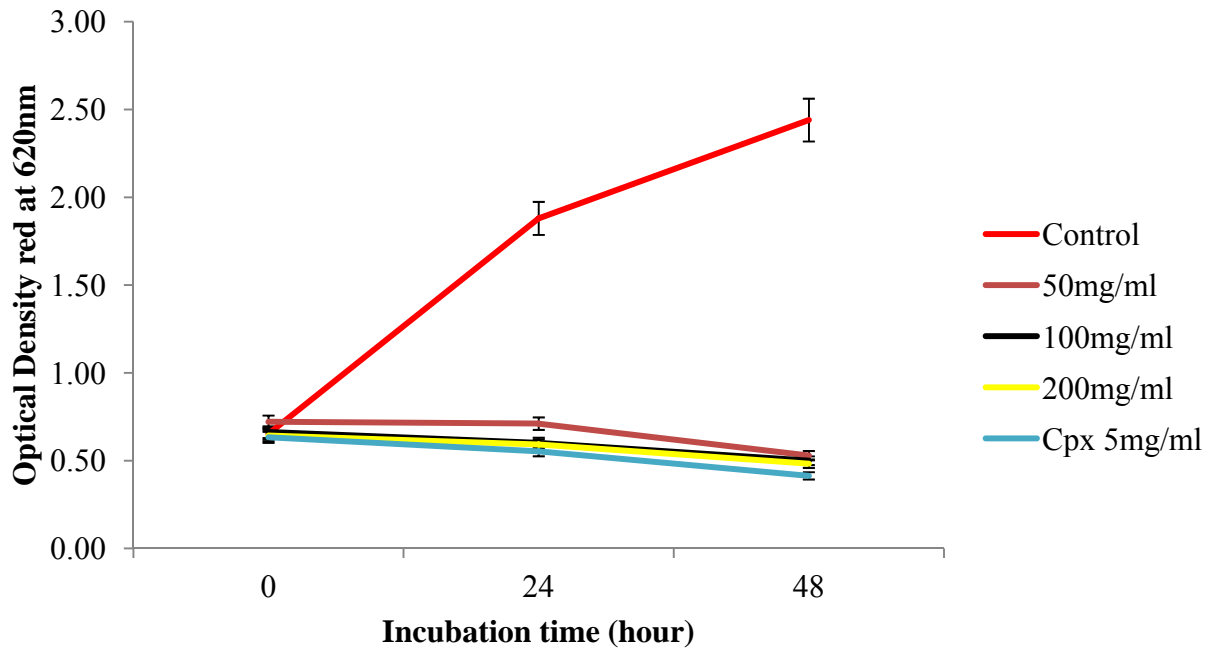
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250 **Table 2: Minimum Inhibitory Concentration (MIC) and Minimum Bacteriocidal**
 251 **Concentration (MBC) of *Boerhaavia diffusa* extracts on *Salmonella***

<i>Boerhaavia diffusa</i> extracts	Ethanolic extract		Aqueous extract	
	S1	S2	S1	S2
MIC (mg/ml)	100	100	50	100
MBC (mg/ml)	50	100	50	100

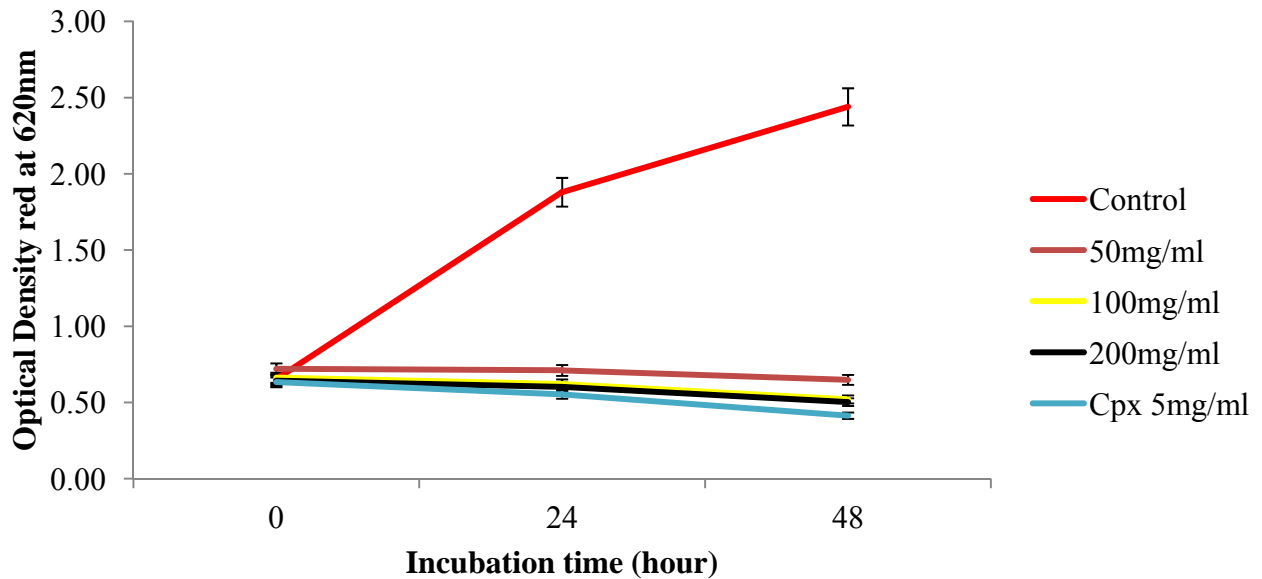
252 Key: S1 - *S. typhi* (Clinical isolate), S2 – *S. typhi* (ATCC 14028)

253



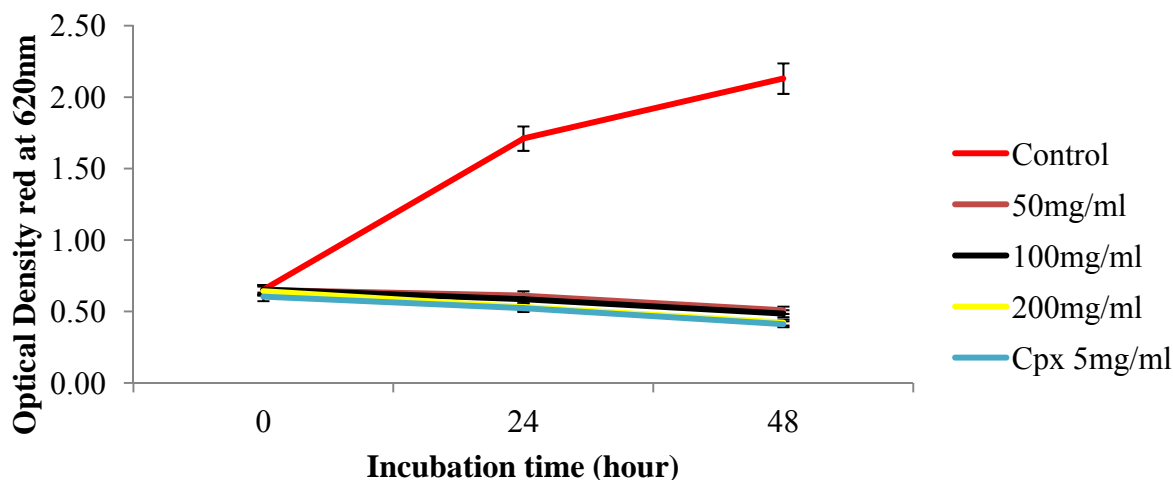
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255 **Figure 3: Effect of *Boerhaavia diffusa* ethanol extract on Clinical isolate**



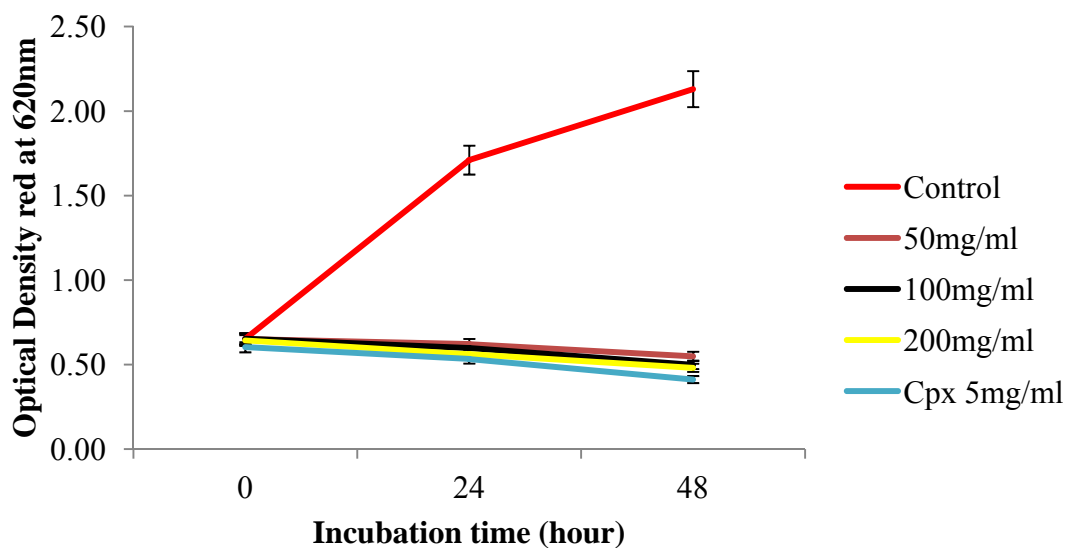
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257 **Figure 4: Effect of *Boerhaavia diffusa* aqueous extract on clinical isolate**



258

259 **Figure 5: Effect of *Boerhaavia diffusa* ethanol extract on typed isolate**



260

261 **Figure 6: Effect of *Boerhaavia diffusa* aqueous extract on typed isolate**

262 **Table 3: Qualitative analysis of phytochemicals in *Boerhavia diffusa* leaf extracts**

263	Phytochemical	Ethanolic extract	Aqueous extract
264	Alkaloids	-	+
265	Tannins	-	+
266	Flavonoids	+	+

267	Quinones	+	-
268	Saponins	+	+
269	Terpenoids	+	-
270	Sterols	+	-
271	Cardiac Glycosides	-	+
272	Phenols	+	+
273	Key : + = Present, - = Absent		

274
 275 **Table 4: Quantitative phytochemical screening of aqueous and ethanol extracts of *B.***
 276 ***diffusa***

Phytochemicals	Ethanol extract	Aqueous extract
Tannins (mg/g)	5.18±0.02 ^a	3.90±0.22 ^a
Quinones (mg/g)	8.45±0.13 ^b	6.60±0.31 ^a
Saponins (mg/g)	6.36±0.24 ^a	14.18±0.06 ^b
Triterpenoids (mg/g)	8.56±0.08 ^a	8.89±0.31 ^a
Steroid (mg/g)	9.03±0.11 ^a	6.73±0.14 ^a
Glycosides (mg/g)	30.39±0.06 ^b	28.29± 0.03 ^a
Flavonoids (mg/g)	9.98±0.61 ^a	11.26±0.33 ^a

277

278

279 **Conclusion**

280 Most of the antibiotics used nowadays have lost their effectiveness due to development of
 281 resistant genes in microbes. The antibiotics are sometimes associated with side effects such as
 282 hypersensitivity, immune suppression and allergic reaction.

283 More interest is being shown in developing alternative antimicrobial drugs for the treatment of
 284 infectious diseases without side effects. The results of our present study demonstrates anti-
 285 *Salmonella* activity of aqueous and ethanol extract of *Boerhavia diffusa*, tannin and quinone
 286 were higher in ethanol extract while saponin was higher in aqueous extract, using agar well
 287 diffusion, the aqueous extract showed higher anti-*Salmonella* efficacy while the broth
 288 microdilution examined by spectrophotometer revealed that ethanol extract had higher anti-
 289 *Salmonella* efficacy. In the present study, the anti-salmonella activity of *Boerhaavia diffusa*
 290 may be attributed to individual or synergistic effect of phytoconstituents present in it. The
 291 ethanol and aqueous extracts of leaves of *B. diffusa* whole plant exhibited significant
 292 antibacterial activity against both clinical and typed *Salmonella typhi*. Therefore, the plant

293 extract could be used for the treatment of Salmonellosis, however, the in vivo studies is needed
294 to ascertain the safety of the extract.

295 **Recommendation**

296 Based on our findings, it is therefore recommended that both agar well diffusion and broth
297 dilution method should be used to affirm the antimicrobial efficacy of the plant extracts.

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