

Original Research Article

Multi-drug resistance pattern of *Staphylococcus aureus* from Paediatric ward, General Hospital, Ikot-Ekpaw, Mkpato Enin LGA, Akwa-Ibom State, Nigeria.

ABSTRACT

AIM: This study evaluated the Multi-drug resistance (MDR) pattern of *Staphylococcus aureus* from a paediatric ward and was conducted using standard clinical microbiological procedures.

LOCATION AND DURATION OF STUDY: The study was carried out on infant samples collected from the Paediatric ward, General Hospital, Ikot-Ekpaw, Mkpato Enin LGA, Akwa-Ibom State, Nigeria, over three (3) months period.

DESIGN OF STUDY: Hundred swab-samples were inoculated on Mannitol salt agar. Positive growths were further biochemically confirmed for *Staphylococcus aureus*. Confirmed isolates were then used for MDR evaluation.

RESULT AND INTERPRETATION: Of the 100 samples from skin, wound, ear, throat and nose swabs, 28 isolates were confirmed as *S. aureus* and were subjected to a range of selected commercially available antibiotics like: amoxicillin, ampiclox, chloramphenicol, ciprofloxacin, erythromycin, gentamicin, levofloxacin, norfloxacin, rifampicin and streptomycin, to evaluate their susceptibilities. The wound swabs gave the highest isolate percentage yield (32%) followed by skin swabs (29%). Susceptibility results showed that amoxicillin and ampiclox were more resisted by the isolates, while ciprofloxacin, levofloxacin and norfloxacin were more effective against the isolates. The Multiple antibiotics resistance (MAR) indices showed that 85.7% of the isolates had confirmed multi-drug resistance status, with 60.7% of the isolates showing resistance to between four or more of the tested antimicrobials. MAR indices revealed that 96.4% of the isolates had 0.3, indicating that the resistance resulted from isolates that adapted to the tested drugs due to some form of abuse. Restricted use of these drugs would help curtail the high resistance currently observed amongst microorganisms.

Key words: multi-drug resistance, *Staphylococcus aureus*, multiple antibiotics resistance index, susceptibility test, nosocomial infections

INTRODUCTION

Microbes do manifest themselves in three ways, through substance spoilage, fermentation of organic and inorganic matters and causation of ailments. Different microorganisms, with their different mode of aetiology, causing different types of ailments, will require different methods

36 and effective drugs for treatments. Continuous deployment of antimicrobial drugs in treating
37 microbial infections has led to the emergence of resistance amongst various strains of
38 microorganisms [1, 2]. Multi-drug resistance (MDR) literally means ‘being resistant to more than
39 one antimicrobial agent’, although a standardized definition has not yet been agreed upon by the
40 medical community. There are currently other definitions that are used to characterize multidrug
41 resistance. The most practical definition used for Gram-positive and Gram-negative bacteria is
42 ‘resistance to three or more antimicrobial classes according to Magiorakos *et al.* [3]. MDR could
43 also be defined as the insensitivity or resistance of a microorganism to administered
44 antimicrobial medicines (which are structurally unrelated and have different molecular targets)
45 despite earlier sensitivity to the same medicines [4].

46 According to Nikaido [5], multidrug resistance in bacteria cells come about by their
47 accumulation of resistance (R) plasmids or transposons, or genes, with each coding for resistance
48 to a specific agent, and/or by the action of multidrug efflux pumps, each of which can pump out
49 more than one drug type. This MDR abilities lead to ineffective ailment treatment, resulting in its
50 persistence, infections spread and high cost [2, 6].

51 The hospital environment has been said to be an active reservoir for infectious microorganisms,
52 being the meeting point for people with diverse disease etiological agents and susceptible
53 individuals [7, 8, 9]. Nikaido [5] mentioned that *Staphylococcus aureus* has a known
54 nosocomial, multi-drug-resistant strain referred to as the methicillin-resistant *Staphylococcus*
55 *aureus* (MRSA). MRSA was initially controlled but currently is also resistant to other
56 antimicrobials like the aminoglycosides, chloramphenicol, lincosamides, macrolides and
57 tetracycline [1]. This study was conducted as part of evidence to buttress the efficacy of
58 Staphylococcal infections in young children and the scale to which MDR pathogens are
59 becoming threats to the health of the younger generation amongst the Mkpato Enin, Akwa-Ibom
60 State populace.

61

62 MATERIALS AND METHODS

63 Study Facility, Group and Sample Collection

64 The study facility is in a growing town and services a couple of adjoining communities, with a
65 number of established institutions, stable commerce and ever growing population. The General

66 Hospital, Ikot-Ekpaw, in Mkpat Enin LGA, Akwa-Ibom State, South-South Nigeria, has Out-
67 patient department, Post-natal ward, Paediatric ward and servicing laboratories. The study
68 focused on children aged 1-15 years.

69 One hundred (100) samples from skin, nostrils, wound, throat and ear were collected over a
70 period of three months using sterile swabs. Once gotten, the swabs were labelled, placed in an
71 ice pack and taken to the Microbiology laboratory, Akwa-Ibom State University.

72 **Sample Analysis**

73 The samples were dipped into peptone water and incubated at 37°C for 24 hours. A loopful from
74 each sample was streaked on separate Mannitol salt agar (MSA) plates and incubated at 37°C for
75 24 hours. Discrete golden yellow colonies were subcultured, purified and preserved. Only Gram
76 positive cocci bacterial colonies were further tested for catalase and coagulase.

77 **Antibiotics Susceptibility Test**

78 Confirmed *Staphylococcus aureus* isolates were tested for their susceptibilities to various
79 selected commercial antibiotic drugs like Ciprofloxacin, Erythromycin, Levofloxacin,
80 Gentamicin, Ampiclox, Rifampicin, Amoxicillin, Streptomycin, Norfloxacin and
81 Chloramphenicol. Overnight cultures using Kirby-Bauer method [10] were inoculated on
82 Mueller-Hinton agar (Oxoid, Uk), cultures adjusted to 0.5 McFarland standard. After pre-
83 diffusion, the plates were incubated at 37°C for 24 hours. Diameter of zones of inhibition (IZDs)
84 produced by the antibiotics were measured and recorded in millimeter. Thereafter, the Multiple
85 antibiotics resistance (MAR) index was determined for each isolate using the formula $MAR =$
86 x/y , where x is the number of antibiotics to which test isolate displayed resistance and y is the
87 total number of antibiotics to which the test organism has been evaluated for sensitivity [11, 12].

88

89 **RESULTS AND DISCUSSIONS**

90 Result for the prevalence of *S. aureus* is as shown in Table 1. The result table shows that of the
91 28 samples that yielded *S. aureus*, wound samples had the highest number, which was followed
92 by samples from children skin swabs. The least number of confirmed *Staphylococcus aureus*
93 isolates were from the ear swabs.

94 The susceptibility pattern of the 28 confirmed test isolates to the selected commercially available
95 drugs (amoxicillin, ampiclox, chloramphenicol, ciprofloxacin, erythromycin, gentamicin,
96 levofloxacin, norfloxacin, rifampicin and streptomycin) is as shown in Figure 1. Ciprofloxacin

97 was the most effective drugs against the test organism, followed by Levofloxacin and
 98 Norfloxacin. Confirmed *Staphylococcus aureus* isolates had high resistance for Amoxicillin,
 99 closely followed by their resistance for Ampiclox.

100

101

Table 1: Prevalence of *Staphylococcus aureus* from clinical samples

Type of Specimen	Sample Size	Number of <i>S. aureus</i> isolated	Total percentage
Skin	20	8	29
Nose	20	5	18
Wound	20	9	32
Throat	20	4	14
Ear	20	2	7

102

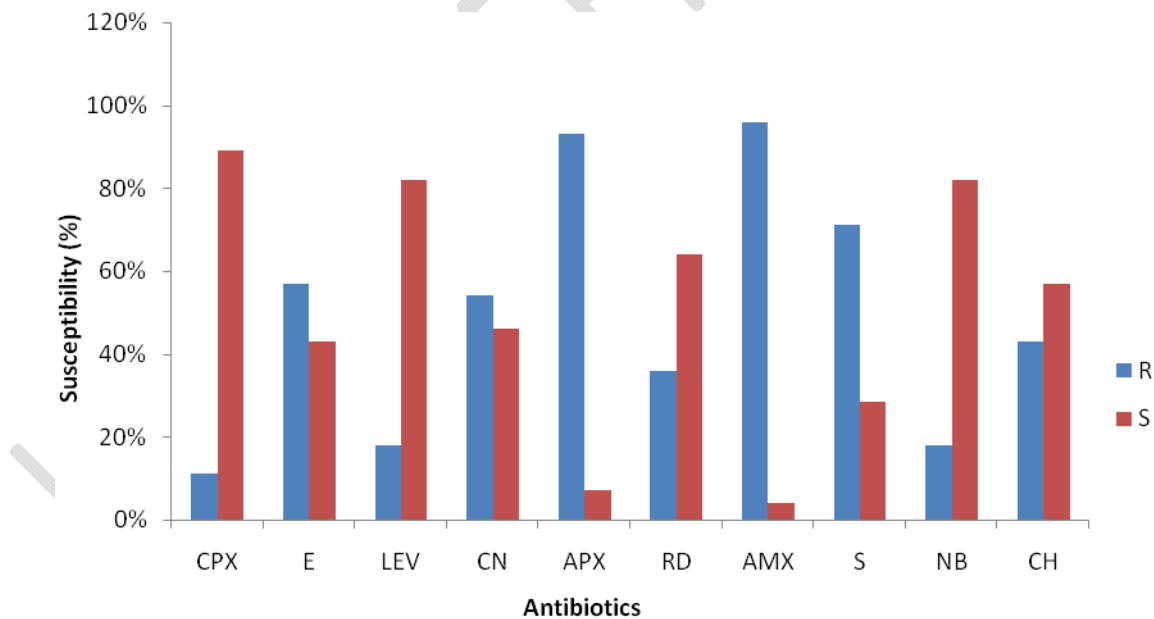


Figure 1: Antibiotics susceptibility pattern of *Staphylococcus* strains from clinical sample

103

104 **KEY:** R= Resistant, S= Sensitive, CPX= Ciprofloxacin, E= Erythromycin, LEV= Levofloxacin, CN= Gentamicin,
 105 APX= Ampiclox, RD= Rifampicin, AMX= Amoxicillin, S= Streptomycin, NB= Norfloxacin, CH=
 106 Chloramphenicol

107

108 Table 2 has the multiple antibiotics resistance (MAR) index result which shows that 85.7% of the
 109 confirmed test isolates were multi-drug resistant (showing resistance to three or more classes of
 110 antibiotics). Only 14.3% of the isolates showed resistance to only two classes of antibiotics.
 111 Results from this study show that 60.7% of test isolates had resistance for four or more
 112 antibiotics drugs. MAR index value for 96.4% of the test isolates reveal cases of source drug
 113 abuse.

Table 2: Antibiotic Resistance Pattern and MAR index of *Staphylococcus aureus*

S/N	ISOLATE CODE	Antibiotic Resistant Pattern	MARI	Antibiotic Resistant Class
1	S9	RD, S	0.2	RIF, AMG
2	S8	APX, AMX, CH	0.3	PEN, CHL
3	N3	LEV, APX, AMX	0.3	QUI, PEN
4	W1	CN, AMX, CH	0.3	AMG, PEN, CHL
5	W9	APX, AMX, S	0.3	PEN, AMG
6	S10	CN, APX, RD, AMX	0.4	AMG, PEN, RIF
7	T6	E, APX, AMX, NB	0.4	MAC, PEN, QUI
8	T8	CPX, E, APX, AMX	0.4	QUI, MAC, PEN
9	S2	E, APX, RD, AMX, S	0.5	MAC, PEN, RIF, AMG
10	S3	E, CN, APX, AMX, S	0.5	MAC, AMG, PEN
11	N1	E, CN, APX, AMX, CH	0.5	MAC, AMG, PEN, CHL
12	N2	CN, APX, AMX, S, CH	0.5	AMG, PEN, CHL
13	W2	E, LEV, APX, AMX, S	0.5	MAC, QUI, PEN, AMG
14	W5	E, APX, AMX, S, CH	0.5	MAC, PEN, AMG, CHL
15	W6	CN, APX, AMX, S, CH	0.5	AMG, PEN, CHL
16	W7	APX, AMX, S, NB, CH	0.5	PEN, AMG, QUI, CHL
17	W10	E, LEV, APX, AMX, S	0.5	MAC, QUI, PEN, AMG
18	T3	E, APX, AMX, S, CH	0.5	MAC, PEN, AMG, CHL
19	E5	E, CN, APX, RD, AMX	0.5	MAC, AMG, PEN, RIF
20	S1	E, CN, APX, RD, AMX, S	0.6	MAC, AMG, PEN, RIF
21	S5	E, LEV, APX, RD, AMX, S	0.6	MAC, QUI, PEN, RIF, AMG
22	S6	CN, APX, RD, AMX, S, CH	0.6	AMG, PEN, RIF, CHL
23	N5	CPX, CN, APX, AMX, S, CH	0.6	QUI, AMG, PEN, CHL
24	W8	E, CN, APX, AMX, S, NB	0.6	MAC, AMG, PEN, QUI
25	N8	E, CN, APX, RD, AMX, S, NB	0.7	MAC, AMG, PEN, RIF, QUI

26	W3	LEV, CN, APX, RD, AMX, S, NB	0.7	QUI, AMG, PEN, RIF
27	T2	E, CN, APX, RD, AMX, S, CH	0.7	MAC, AMG, PEN, RIF, CHL
28	E9	CPX, E, CN, APX, AMX, S, CH	0.7	QUI, MAC, AMG, PEN, CHL

KEY: CPX= Ciprofloxacin, E= Erythromycin, LEV= Levofloxacin, CN= Gentamicin, APX= Ampiclox, RD= Rifampicin, AMX= Amoxicillin, S= Streptomycin, NB= Norfloxacin, CH= Chloramphenicol, MARI= Multiple antibiotic resistance index, RIF= Rifamycins, AMG= Aminoglycosides, PEN= Penicillins, CHL= Chloramphenicol, MAC= Macrolides, QUI= Quinolones

114 Data for isolate occurrence and confirmation showed that there were more confirmed
 115 *Staphylococcus* isolates from wound swab-samples, followed by skin **swab-samples**. Parta *et al.*
 116 [13] also recorded very high *Staphylococcus* number from wound swabs. This high isolate-
 117 numericals is suggestive of the exposed nature of the sampling points. This is supported by
 118 findings presented by Nimmo *et al.* [14], who found more *Staphylococcus* isolates on exposed
 119 body surfaces than the internal parts. While uncovered wounds have sticky surfaces and the skin
 120 is continuously exposed, it is therefore easy for such high microbial numbers to be recorded.

121 Susceptibility data showed that all the confirmed and tested isolates resisted two or more
 122 antimicrobials. Qureshi *et al.* [15] also isolated MRSA that resisted multiple anti-microbials from
 123 hospital specimens. This study's result showed a higher MDR percentage than the "nearly half"
 124 proportion reported by Nimmo *et al.* [14].

125 More than 96.4% of the MAR indices were 0.3 from this study evaluation. This assertion is
 126 indicative that resistance to these multiple drugs come from over exposure of the isolates to
 127 drugs, making them adapt or resistant to them with recurrent treatments. The high case of MDR
 128 amongst microorganisms can drastically be reduced by mere restricting the indiscriminate and
 129 readily availability of these drugs over the counter [14].

130

131 CONCLUSION

132 **In conclusion**, many studies have shown that prolonged stays in hospitals increases the risk for
 133 **the** colonization or infection with MDR *Staphylococcus aureus*. This reflects an inherent risk in
 134 acquiring MDR organisms through environmental contaminations and hospital stay conditions.
 135 With infant patients, another potential transmission route is through infected staff members
 136 handling and this calls for special care [16, 17].

137

138 The isolation of MDR *Staphylococcus aureus* from infant patients is a call for the proper
139 implementation of contact precautions during hospitalization especially in health facilities across
140 developing countries [16, 18].

141 **Ethical consideration**

142 The study was approved by the ministry of health, Akwa Ibom State. Permission was obtained
143 from General Administration of the General hospital prior to collecting any data. Participants'
144 privacy and confidentiality have been assured (no names have been used, only numbers were
145 used) and all data and results have been handled and treated confidentially.

146

147 **REFERENCES**

148 1 McIntosh, J. (2018). Antibiotic resistance: What you need to know. Medical news today,
149 (<https://www.medicalnewstoday.com/articles/283963.php>).

150 2 Tanwar, J., Das, S., Zeeshan, F. and Hameed, S. (2014). Multidrug Resistance: An Emerging
151 Crisis. *Interdisciplinary Perspectives on Infectious Diseases*, 7 pages.

152 3 Magiorakos, A. P., Srinivasan, A., Carey, R. B., Carmeli, Y., Falagas, M. E., Giske, C. G.,
153 Harbarth, S., Hindler, J. F., Kahlmeter, G., Olsson-Liljequist, B., Paterson, D. L., Rice, I. B.,
154 Stelling, J., Struelens, M. J., Vatopoulos, A., Weber J. T. and Monnet, D. L. (2012). Multidrug-
155 resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert
156 proposal for interim standard definitions for acquired resistance. *Clinical Microbiology and*
157 *Infection*, 18, 268–281.

158 4 Singh, V. (2013). Antimicrobial resistance *In: Microbial Pathogens and Strategies for*
159 *Combating Them: Science, Technology and Education*, Formatex Research Center, 1, 291-296.

160 5 Nikaido, H. (2009). Multidrug Resistance in Bacteria. *Annual Review of Biochemistry*, 78,
161 119–146.

162 6 WHO (2014). *Antimicrobial Resistance Global Report on Surveillance*. Geneva, Switzerland:
163 World Health Organization.

164 7 Zhanel, G. G., DeCorby, M., Laing, N., Weshnoweski, B., Vashisht, R., Tailor, F., Nichol, K.
165 A., Wierzbowski, A., Baudry, P. J., Karlowsky, J. A., Lagacé-Wiens, P., Walkty, A.,
166 McCracken, M., Mulvey, M. R. and Johnson, J. (2008). The Canadian Antimicrobial Resistance
167 Alliance (CARA), and Hoban DJ (2008). Antimicrobial-resistant pathogens in intensive care
168 units in Canada: results of the Canadian National Intensive Care Unit (CAN-ICU) study, 2005-
169 2006. *Antimicrobial Agents and Chemotherapy*, 52, 1430-1437.

170 8 Rhomberg, P. R., Fristoche, T. R., Sader, H. S. and Jones, R. N. (2006). Antimicrobial
171 susceptibility pattern comparisons among intensive care unit and general ward Gram negative
172 isolates from meropenem yearly susceptibility test Information programme (USA). *Diagnostic*
173 *Microbial Infectious Disease*, 56, 57-62.

- 174 9 O'Brien, F. G., Pearman, J. W., Gracey, M., Riley, T. V. and Grubb, W. B. (1999). Community
175 strain of methicillin-resistant *Staphylococcus aureus* involved in a hospital outbreak. *Journal of*
176 *Clinical Microbiology*, 37(9):2858–2862.
- 177 10 Hudzicki, J. (2009). Kirby-Bauer disk diffusion susceptibility test protocol. *American Society*
178 *for Microbiology*, 23 pages.
- 179 11 Tula-Sanchez, A. A., Havas, A. P., Alonge, P. J., Klein, M. E., Doctor, S. R., Pinkston, W.,
180 Glinsmann-Gibson, B. J., Rimsza, L. M. and Smith, C. (2013). A model of sensitivity and
181 resistance to histone deacetylase inhibitors in diffuse large B cell lymphoma: Role of cyclin-
182 dependent kinase inhibitors. *Cancer Biology and Therapy*, 14(10): 949-961.
- 183 12 Olayinka, A. T., Onile, B. A. and Olayinka, B. O. (2004). Prevalence of multi-drug resistance
184 (MDR) *Pseudomonas aeruginosa* isolates in surgical units of Ahmadu Bello University
185 Teaching Hospital, Zaria, Nigeria: An indication for effective control measures. *Annals of*
186 *African Medicine*, 3(1): 13-16.
- 187 13 Parta, M., Goebel, M., Matloobi, M., Stager, C. and Musher, D. M. (2009). Identification of
188 Methicillin-Resistant or Methicillin-Susceptible *Staphylococcus aureus* in Blood Cultures
189 and Wound Swabs by GeneXpert. *Journal of Clinical Microbiology*, 47(5):1609–1610.
- 190 14 Nimmo, G. R., Pearson, J. C., Collignon, P. J., Christiansen, K. J., Coombs, G. W., Bell, J.
191 M., McLaws M. L. and the Australian Group on Antimicrobial Resistance (2011).
192 Antimicrobial susceptibility of *Staphylococcus aureus* isolated from hospital in-patient.
193 *Community Disease Intelligence*, 35(3):237–243.
- 194 15 Qureshi, A. H., Rafi, S., Qureshi, S. M. and Ali, A. M. (2004). The Current Susceptibility
195 Patterns of Methicillin Resistant *Staphylococcus aureus* to Conventional Anti-*Staphylococcus*
196 Antimicrobials at Rawalpindi. *Pakistan Journal of Medical Science*, 20(4): 361-364.
- 197 16 Buke, C., Armand-Lefevre, L., Lolom, I., Guerinot, W., Deblangy, C., Ruimy, R.,
198 Andremont, A. and Lucet J. C. (2007). Epidemiology of multidrug-resistant bacteria in patients
199 with long hospital stays. *Infection Control and Hospital Epidemiology*, 28(11):1255–1260.
- 200 17 Maamar, E., Ferjani, S., Jendoubi, A., Hammami, S., Hamzaoui, Z., Mayonnove-Coulange,
201 L., Saidani, M., Kammoun, A., Rehaïem, A., Ghedira, S., Houissa, M., Boutiba-Ben Boubaker,
202 I., Slim, A. and Dubois, V. (2016). High prevalence of gut microbiota colonization with broad-
203 spectrum cephalosporin resistant enterobacteriaceae in a tunisian intensive care unit. *Frontiers in*
204 *Microbiology*, 7, 1859.
- 205
- 206 18 Harbarth, S., Sax, H., Fankhauser-Rodriguez, C., Schrenzel, J., Agostinho, A. and Pittet, D.
207 (2006). Evaluating the probability of previously unknown carriage of MRSA at hospital
208 admission. *American Journal of Medicine* 119(3):275.e15-e23