Capacity for parasitism of Trichogramma spp. in tomato fruit borer under different temperatures

Abstract:

The parasitoid in eggs of Trichogramma genre is the most studied in the world, being bred widely and used for flooding releases. This study aimed to evaluate the capacity for parasitism of Trichogramma galloi Zucchi, 1988 (Hymenoptera: Trichogrammatidae) in Neoleucinodes elegantalis (Guenée) (Lepidoptera: Crambidae) eggs at different temperatures. The experiment was developed at the Nucleus for Scientific and Technological Development in Phytosanitary Management (NUDEMAFI) in which the daily and accumulated biological parameters were assessed, as well as total parasitized eggs by female, sexual ratio (number of females/number of males + females), viability of the eggs (number of eggs with orifice/number of parasitized eggs x 100) and number of individuals per egg at temperatures 18, 21, 24, 27 and 30 °C. The eggs of the fruit borer were offered daily to each T. galloi female at each temperature until the death of the parasitoid could be verified. The larger number of parasitized eggs was found in the first 24h, at temperatures 24 and 27 °C with 17 parasitized eggs. The accumulated parasitism in N. elegantalis eggs reached 80% of total parasitized eggs for each thermal range (18, 21, 24, 27 and 30 °C) on the 2nd, 3rd, 3rd, 1st and 2nd days respectively. The ideal parasitism conditions for this lineage vary between 24 and 27°C. Therefore, it is concluded that the studied T. galloi lineage has adequate biological parameters in N. elegantalis eggs, demonstrating promise in phytosanitary management of this pest.

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Keywords: Egg parasitoid. Neoleucinodes elegantalis, Phytosanitary management, Tomato fruit, Oviposition, Trichogrammatidae.

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1. INTRODUCTION

Among the pests that attack the tomato culture, the tomato fruit borer Neoleucinodes elegantalis (Guenée) (Lepidoptera: Crambidae) is considered one of the main pests for its preference for this culture and the damages caused directly in the fruit, making them inadequate for consumption and industrial processing, with significant loss (Miranda et al., 2005; Picanço et al., 2007; Fornazier et al., 2010; Pratissoli, 2015; Carvalho et al.; 2017; Silva et al., 2017; Moraes & Foerster, 2015).

- 24 Since this is a culture of high risk, with high intensity for pest attack, it is important to
- 25 implement practices that aim to manage these pests. Among management methods, biological
- control is a viable technique, released in about 500,000 hectars every year in sugar cane
- 27 especially when using parasitoids of the *Trichogramma* genre since it acts on the eggs
- 28 avoiding the larvae to penetrate the fruit, reducing the loss caused by caterpillar feeding in its
- interior (Plaza et al., 1992, Oliveira et al., 2017).
- 30 The egg parasitoid Trichogramma is the most often studied in the world, being greatly bred
- 31 and used in flooding releases (Hassan, 1997; Davies et al., 2009). The advantage of its use is
- 32 its capacity to control pests from different cultures. Moreover, they are highly specialized and
- 33 efficient (Haji et al., 2002; Wang et al., 2007; Wang et al, 2018; Arruda et al. 2014).
- 34 In Brazil, studies aiming at the use of Trichogramma were initiated over 30 years ago, with
- 35 excellent results in many cultures, more recently Trichogramma galloi Zucchi, 1988
- 36 (Hymenoptera: Trichogrammatidae) being the most often used species, released in about
- 37 500,000 hectares every year in sugar cane to control the cane borer *Diatraea saccharalis*
- 38 Fabricius, 1794 (Lepidoptera: Crambidae) (Parra, 2010, Geremias, Parra, 2014, Arruda et al.
- 39 2014, Zago et al. 2007, Oliveira et al., 2017).
- 40 N. elegantalis studies have demonstrated its potential to use Trichogramma in its
- 41 management (Blackmer et al., 2001). Nonetheless, other studies must be conducted for better
- 42 reliability on the use of these parasitoids in the management of *N. elegantalis*. These studies
- 43 must involve the efficacy of the species, biological characteristics, thermal demands, ideal
- 44 release numbers and dispersion, breeding of the alternative host Anagasta kuehniella Zeller
- 45 (Lepidoptera capacity (Oliveira et al., 2017).
- 46 Thus, the aim of this study was to evaluate the potential for parasitism of *T. galloi* in *N.*
- 47 *elegantalis* eggs at different temperatures.

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2. MATERIAL AND METHODS RELEASE NUMBERS AND DIPERSION CAPACITY

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- The experiment was conducted in the Entomology Department at the Nucleus for Scientific
- and Technological Development in Phytosanitary Management (NUDEMAFI) at the Agronomic
- 53 Sciences Center at the Federal University of Espirito Santo (CCAE-UFES), Alegre, ES
- 54 (Brazil). A lineage Tg1of *T. galloi* species was used, provide by BUG Biological Agents.

- Breeding of the alternative host Anagasta kuehniella Zeller (Lepidoptera: Pyralidae)
- 57 The alternative host *Anagasta kuehniella* Zeller (Lepidoptera: Pyralidae) was bred in the
- homogenized diet of whole wheat flour (60%), corn (37%) and beer yeast (3%). The diet was
- 59 disposed into plastic bins (30 x 25 x 10 cm) with corrugated cardboard strips (25 x 2 cm) on
- 60 the inside, with the host eggs randomly selected for the diet. The adults were collected daily,

with an adapted vacuum and transferred into PVC tubes (150 mm diameter x 25 cm height) with nylon strips folded in its interior for oviposition (Pratissoli, 2010, Zuim et al., 2017).

Breeding of T. galloi

For the maintenance of parasitoids, *Anagasta kuehniella* eggs were invisibilized in germicide lamp during 50 minutes and fixated in rectangles of sky blue cardboard (8.0 x 2.0 cm), with Arabic gum diluted to 20%. Those cards were inserted in glass tubes (8.5 x 2.4 cm), containing adult parasitoids recently emerged. Furthermore, the tubes were sealed with PVC plastic film to avoid parasitoid escape. The cards were kept in the tubes for 24 hours and later stored in clean glass tubes (9 x 3 cm) in an acclimatized room at 25 \pm 1 °C, relative humidity 70 \pm 10% and photophase of 14h (Zuim et al., 2017).

Breeding of N. elegantalis

Breeding of pests was conducted in an acclimatized room (25 \pm 2 $^{\circ}$ C, RH 70 \pm 10% and photophase of 12h). Adults were kept in acrylic cages and fed with a solution of 10% honey. For oviposition, tomato fruit from the F1 wire was conditioned in the cages. Daily, the eggs were removed from the tomato fruit and distributed in African eggplant fruit (mean 5 eggs/fruit) which remained in plastic containers covered in non-woven fabric serving as places for pupation of caterpillars. Once this phase is finished, pupae were transferred into plastic containers or Petri dishes and stowed in acclimatized chambers in the above-mentioned conditions until adults emerge, then again taken to the acrylic cages.

Capacity of parasitism

- *N. elegantalis* eggs with up to 12h of age were collected from the tomato fruit with the help of a scalpel and glued to sky blue cardboard (0.5 x 2.0 cm) with a brush and Arabic gum at 20%. For each temperature of the study, 20 recently emerged females were isolated in *Eppendorf* tubes (2.0 ml), containing drops of honey for feeding and sealed with the tubes' own lid. The cards with the 20 tomato fruit borer eggs were offered daily to each one of the *T. galloi* females at each temperature (18, 21, 24, 27 and 30 °C) until the death of the parasitoid was confirmed. The cards removed daily were identified and bagged (23.0 x 4.0 cm) and kept at its respective temperatures.
- The following biological parameters were assessed: daily and accumulated parasitism, total parasitized eggs per female, sexual ratio (number of females/number of males + number of females), viability of the eggs (number of eggs with orifice/number of parasitized eggs x 100) and number of individuals per egg at different temperatures.
- The experiment was conducted with a completely casual design, with five treatments (temperatures) and 20 repetitions, each repetition represented by a *T. galloi* female. For data analysis, a regression with test F was used at 5% probability level.

3. RESULTS 99 100 101 Daily parasitism decreased at all temperatures with the advance of age of the *T. galloi* female. 102 At the different studied temperatures, higher rates of parasitism occurred in the first 24 hours, 103 presenting between 13 and 17 parasitized eggs. The higher rates of parasitism were at 104 temperatures 24 °C and 27 °C with around 17 eggs parasitized (Figure 1). 105 In terms of longevity of the females, it was noted that lower temperatures (18 °C to 24°C) 106 females were able to live longer due to a reduction in the physiological activity of females 107 when exposed to lower temperatures. At higher temperatures (27 °C and 30 °C), there is 108 higher energy expenditure and, consequently, females lived for a shorter period of time 109 (Figure 1). 110 The parasitism (parasitism) period for T. galloi females was increased in the thermal range, of 111 18 to 24°C (5, 7 and 8 days) and in the 27 to the 30°C range there was a decrease (5 and 4 112 days). Therefore, the ideal conditions for the survival of T. galloi vary between 24 °C and 27 °C 113 where better performance was observed. 114 Accumulated parasitism in N. elegantalis eggs in the studied thermal range reached 80% of 115 total parasitized eggs in a maximum of three days. At extreme temperatures (18 and 30°C) 116 this condition was reached in two days. At milder temperatures (21 and 24°C) the 117 accumulated parasitism reached 80% in three days and at 27 °C was reached in the first day 118 of parasitism (Figure 1). Due to the 80% parasitism, it was noted that the potential for 119 parasitism of this lineage occurs within the first days of parasitism, independent from 120 temperature. 121 For this lineage, the 24°C temperature highlights the total amount of eggs parasitized per 122 female reaching an average of 30 eggs. In terms of viability, as the temperature increased 123 there was a reduction in viability, coming to 50% at 30°C. For sexual ratio, it was observed 124 that when temperature increased there was a higher number of males in the population, but 125 the number of individuals per egg was constant (Figure 2). 126 127 128

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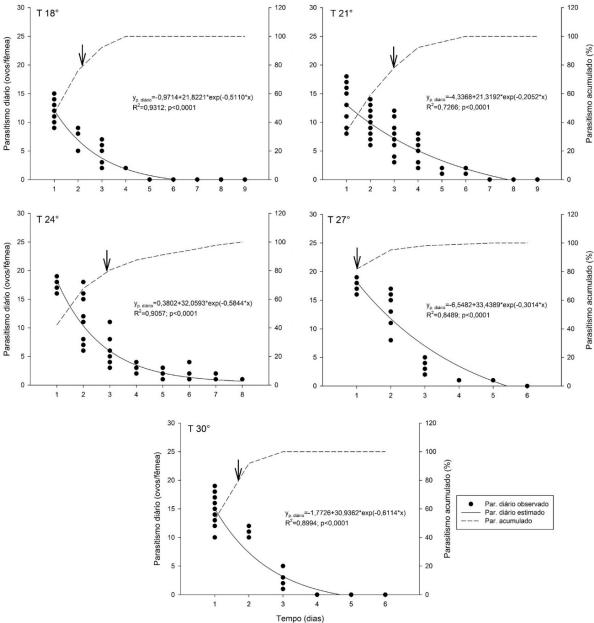


Figure 1 – Daily and accumulated parasitism for *T. galloi* in *N. elegantalis* eggs at different temperatures.

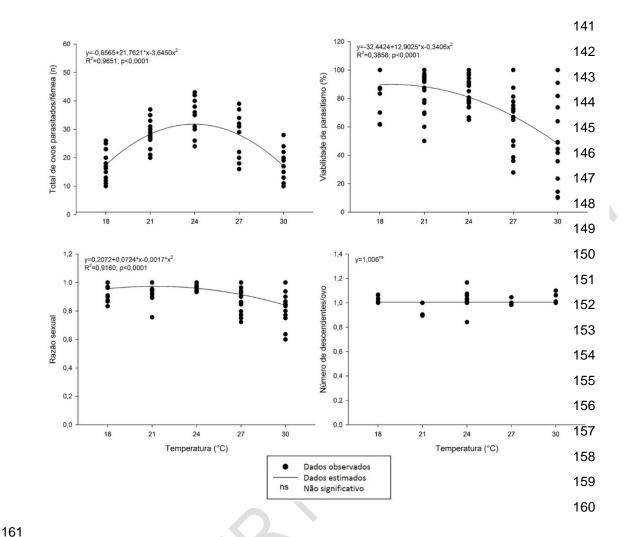


Figure 2 – Total *N. elegantalis* eggs parasitized by *T. galloi*, the viability of parasitism, sexual ratio and number of descendants per eggs at different temperatures.

4. DISCUSSION

We verified that the temperature interferes in the potential for parasitism and biological characteristics of *T. galloi*.

Among the main factors affecting biological characteristics of species in the *Trichogramma* genre, temperature is highlighted since with its increase, there is lower performance by females causing metabolism to increase and, therefore, reducing parasitism (Hansen & Jensen, 2002; Pratissoli et al., 2003; Rahimi-Kaldeh et al 2018, Pratissoli et al. 2004). This was demonstrated in the present study since extreme temperatures showed a decrease in parasitism with only 11 eggs parasitized on average at 18°C, 10 eggs at 21°C and 14 eggs parasitized at 30°C (Figure 2).

- 175 The potential for parasitism in the first days may be directly connected to the instinct of animal
- 176 preservation once all species in the *Trichogramma* genre present this behavior (Pratissoli at
- al., 2004; Zago et al., 2007; Paes et al., 2018). This behavior may be related to the parasitism
- 178 of 80% of the eggs as studies have confirmed this rate to be, in most cases, in the first few
- days of life in females (Pratissoli at al., 2004; Zago et al., 2007).
- 180 Parasitism period may vary according to temperature and within each temperature. This fact
- may be related to the capacity of adaptability in each species and/or lineage of Trichogramma
- to the habitat in which it was collected (Hansen & Jensen, 2002; Pratissoli at al., 2004; Zago et
- 183 al.,2007; Arruda et al., 2014; Paes et al., 2018).
- 184 Accumulated parasitism is another factor that may be related to the capacity of adaptability of
- each species and/or lineage of *Trichogramma* to the habitat in which it was collected since the
- necessary time to reach total percentage is variable (Pratissoli at al., 2004; Zago et al., 2007).
- 187 The range of temperature in which species and/or lineage of Trichogramma present their
- higher potential for parasitism (number of parasitized eggs) is between 24 and 27°C (Hansen
- 4 Lensen, 2002; Pratissoli et al., 2004; Zago et al., 2007; Arruda et al., 2014).
- 190 Through viability, there seems to also be direct interference from temperature. It is possible to
- 191 verify that there is an inverse relationship between the percentage of the emergence of
- 192 descendants and the increase in temperature. However, this was not found in any other
- 193 studies.
- 194 The variation in sexual ration has been reported as influenced especially by temperature
- 195 (Vinson, 1997, Rahimi-Kaldeh, et al, 2018). This was verified in extreme temperatures once
- 196 the humidity, female age, and host were constant for all temperatures.
- 197 On the number of descendants per egg, it was verified that it was constant, that is, one
- 198 individual per egg independent from temperature. The variation in this factor is directly related
- 199 to nutritional and morphological characteristics of the egg such as size, shape, thickness,
- 200 corion stiffness and lay behavior (Hassan, 1997; Bakthavatsalam, et al., 2013, Paes et al.,
- 201 2018).
- 202 It was verified that the lineage studied for T. galloi holds true the adequate biological
- 203 parameters for parasitism in N. elegantalis eggs, proving to be promising in phytosanitary
- 204 management of this pest.

2062075. CONCLUSION

- The studied *T. galloi* strain presents the appropriate biological parameters for parasitism in *N.*
- 209 *elegantalis* eggs, showing promise in the phytosanitary management of this pest.

212 **COMPETING INTERESTS** 213 214 "Authors have declared that no competing interests exist" 215 216 217 REFERENCES 218 Arruda LA, Leite R C, Tonquelski GV, Leal AF. Borges FSP, Rodrigues LA. Eficiência do 219 parasitismo de três espécies de Trichogramma (T. galloi, T.atopovirilia e T. bruni) sobre ovos 220 da praga Datraea saccharalis. Global Science Technology. 2014;07:67 – 75. 221 Bakthavatsalam N, Tandon PL, Bhagat D. Trichogrammatids: Behavioural Ecology In. 222 Sithanantem CR. Ballal, Jajali SK, Bakthavatsalam N. (Ed). Biological Control of Insects Pests 223 using Egg Parasitoids. 2013;77-103. New Delhi-India: Springer 224 Blackmer JL, Eiras AE, Souza CLM. Oviposition preference of Neoleucinodes elegantalis 225 (Guenee) (Lepidoptera: Crambidae) and rates of parasitism by Trichogramma pretiosum Riley 226 (Hymenoptera: Trichogrammatidae) on Lycopersicon esculentum in São José de Ubá, RJ, 227 Brazil. Neotropical Entomology. 2001;30:89-95. 228 Carvalho GS, Silva LB, Reis SS, Veras MS, Carneiro E, Almeida MLS, Silva AF, Lopes GN. 229 Biological parameters and thermal requirements of *Trichogramma pretiosum* reared on 230 Helicoverpa armigera eggs. Pesquisa agropecuária brasileira. 2017;52:961-968. 231 Davies AP, Pufke US, Zalucki MP. Trichogramma (Hymenoptera: Trichogrammatidae) 232 Ecology in a Tropical Bt Transgenic Cotton Cropping System: 18 Sampling to Improve 233 Seasonal Pest Impact Estimates in the Ord River Irrigation Area, Australia. Journal Economic 234 Entomological. 2019;102:1018-1031. 235 Fornazier M, Pratissoli D, Martins D S. Principais pragas da cultura do tomatoiro estaqueado 236 na região das montanhas do Espírito Santo. In: Incaper(Ed.). Tomato; 2010;185-226. Vitória: 237 Incaper. 238 Geremias LD, Parra JRP. Dispersal of Trichogramma galloi in corn for the control of Diatraea 239 saccharalis. Biocontrol science and technology. 2014;24:751-762. 240 Haji FNP, Prezotti L, Carneiro JS, Alencar JA Trichogramma pretiosum para controle de 241 pragas no tomatoiro industrial, Vol. 1: Controle biológico no Brasil: Parasitoides e predadores 242 (ed. Parra, JRP, Botelho, SM, Ferreira, BSC, Bento JMS) Manole, São Paulo, SP, Brazil, 243 2002, pp. 477-494. 244 Hassan, AS. Seleção de espécies de Trichogramma para o uso em programas de controle 245 biológico., Vol. 1: Trichogramma e o Controle Biológico Aplicado (ed. Parra JRP & Zucchi 246 RA) FEALQ, Piracicaba, São Paulo, Brazil, pp. 183-205, 1997.

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