

## Original Research Article

# Physiological Maturity and determination of the harvest time of *Vigna unguiculata* L. Walp.

### ABSTRACT

The maturation process of seeds is genetically controlled and involves an organized sequence of physiological changes from the fertilization until the complete independence from the plant. It is recommended that the harvest occurs in the ideal moment, this way the seeds can express their full potential, with maximum dry matter accumulation, reaching high potential of germination and vigor. The objective of this study was to determine the physiological maturity point of cowpea bean seeds (*Vigna unguiculata* L. Walp.), cv. Corujinha, aiming to indicate the best harvesting period, in order to guarantee greater germination potential and seed vigor. Cowpea pods were harvest from the third until the twenty-first day after anthesis, with intervals of three days between the harvests. After each harvest, the following evaluations were carried out: fruit and seed color, number of seeds per pod, pod and seed biometry, pod and seed moisture, water content, germination, germination speed index, length and dry matter of the shoots and roots. At 15 DAA, the seeds and fruits presented light green coloration, with maximum values of length, width, thickness, dry matter, germination percentage and germination speed index, at a vigor level. There was a gradual reduction of water content in the seeds and number of seeds up to 21 DAA. The highest values for shoot and root length were observed at 18 DAA, when seeds and fruits showed light brown color and for shoot and root dry matter at 21 DAA, with brown color. The physiological maturity of cowpea seeds was rapid and occurred between 15 and 21 DAA. The harvest is recommended at 15 days after anthesis, when the seeds present high germination and vigor.

**Comment [AL1]:** Please use one name. I suggest \*pod\* and change the \*fruit\* into \*pod\*. The description must be precise. In one place is under biometry, in another fruit biometry. Please check and correct the description for other alternative names.

**Keywords:** Cowpean bean, physiological quality, germination, vigor.

### 1. INTRODUCTION

The cowpea (*Vigna unguiculata* L. Walp.) is cultivated throughout the North and Northeast of Brazil, where it is considered the main component of the agricultural production of these regions, constituting an important source of income and subsistence for small farmers who practice agriculture. In addition, this crop is used as a staple food for the population, which consumes it in the form of green and dry grains, being very appreciated due to the fast cooking and nutritional aspects, such as the quantity of proteins [1].

The difficulty of obtaining seeds of good physiological quality is among the limiting factors in the production of cowpea, since the seeds are one of the main inputs of the agricultural production, where the quality is an important factor to obtain stands of uniform and vigorous plants, directly reflecting the yield [2].

To express its full potential, it is essential that the harvest occurs at the ideal moment, with maximum dry matter accumulation, reaching high germination and vigor potential [3]. Therefore, the study of the physiological maturation process of seeds is very important to

27 determine the ideal harvest time and, consequently, obtain seeds of high physiological  
28 quality [4].

29 The seed maturation process is genetically controlled and involves an organized sequence  
30 of physical, biochemical, physiological and morphological changes, from the fertilization until  
31 its independence from the plant, these changes also include a set of preparatory steps for  
32 the process of germination, which are characterized by the synthesis and accumulation of  
33 nutrient reserves [5].

34 Several studies on the influence of physiological maturity on the seed quality and  
35 productivity of several crops have been carried out, such as the studies with pepper seeds  
36 (*Capsicum annuum* L.) [6] common bean (*Phaseolus vulgaris* L.) [7], ginger (*Sesamum*  
37 *indicum* L.) [8] and pumpkin (*Curcubita moschata* Duch) [9]. However, currently, for cowpea,  
38 there is little information on the maturation and the ideal harvest period of the seeds,  
39 justifying the need to perform this evaluation [2].

40 The objective of this study was to determine the physiological maturity point of cowpea  
41 seeds (*Vigna unguiculata* L. Walp.), cv. Corujinha, aiming to indicate the best **harvest** time,  
42 in order to guarantee greater germination potential and seed **vigor**.

## 43 2. MATERIAL AND METHODS

### 44 2.1 Experimental Location

45 The field experiment was performed with cowpea bean seeds, *Vigna unguiculata* cv.  
46 Corujinha, between September 2015 and January 2016 at the Chã de Jardim Experimental  
47 Farm of the Centro de Ciências Agrárias of the Universidade Federal da Paraíba (CCA-  
48 UFPB), in Areia-Paraíba, located in the micro-region of the Paraíba, under the geographic  
49 coordinates 6°58'12 "S and 35°42'15" W.

50 According to Graussem's bioclimatic classification, the predominant bioclimate in the area is  
51 the sub-dry Northeastern 3dfh with annual rainfall of approximately 1,400 mm. According to  
52 Köppen's classification, the climate is characterized as warm and humid, with autumn-winter  
53 rains. The average annual temperature ranges from 22 to 26 °C and relative humidity  
54 between 75 and 87% [10]. During the conduction of the experiment the minimum  
55 temperature was 20.3 °C and the maximum was 28.5 °C, with average relative humidity of  
56 76.4%. According to Embrapa [11], the soil of the experimental area is classified as a typical  
57 Psamitic Regolithic Neosols, of medium texture.

### 58 2.2 Experimental Design

59 For the soil preparation the area was cleaned with garden hoes and pits at a depth of 4 cm,  
60 spaced 0.30 m between plants and 1.0 m between rows were opened. Three seeds/pit were  
61 sown, after thinning, one plant/pit was left, the plants were monitored periodically to follow  
62 the flowering stage, while the cultural treatments were recommended for the crop.

63 Fifty-four days after sowing, when approximately 70% of the plants started the anthesis they  
64 were identified using wool yarns. The plants were monitored until fruiting and, every 3 days  
65 were harvested, with a total of seven harvests, manually performed, mechanical injuries in  
66 the pods and seeds were **avoided**. After **harvested**, the pods were packed in plastic bags,  
67 identified and sent to the laboratory.

Comment [AL2]: You have to write what indicators have been used to achieve this goal. This is what we know in the results.

## 68 2.3 Evaluated Parameters

69 The pod and seed biometry, and also their physiological quality, were evaluated in the  
70 Laboratório de Análise de Sementes, also located in the previously mentioned Center.

71 After each **harvest**, four replicates of 15 pods and 25 seeds were submitted to direct  
72 measurements with the aid of a digital **caliper**, in which measurements of length, width and  
73 thickness were performed, the results were expressed in millimeters, only the length of the  
74 fruit was expressed in centimeters.

Comment [AL3]: Give accuracy; 0.01 g?

75 After each **harvest**, by using a sample of 40 pods the number of seeds per pod was  
76 determined by manual counting, and the results were expressed as number of seeds per  
77 pod<sup>-1</sup>.

78 The water content of the pods and seeds were obtained by the stove method at 105 °C for  
79 24 hours [12], using four replicates of 25 seeds and four replicates of 5 pods at each **harvest**  
80 period, the results were expressed in **percentage**.

Comment [AL4]: Did you weigh the samples. On what type of weight and with what accuracy. What mass had the sample. These percentages are not clear.

81 **The samples were placed in a stove at 105 ± 3 °C for 24 hours [12], after that, the dry matter**  
82 **of the pods and seeds were determined together with the water content, at all harvest time.**  
83 **The results were expressed in grams.**

Comment [AL5]: Why this procedure is described separately. After all, when determining the moisture content, you have the same data. Why do you enter the dry matter content in grams. It does not make sense. The dry mass is given in percent. It should be a relative value due to the initial mass.

84 The germination test was performed following the requirements of the Rules for Seed  
85 Analysis - RSA [12], using 200 seeds per treatment, distributed in four replicates of 50  
86 seeds, placed in a paper towel substrate (**germitest**<sup>®</sup>) moistened with sterilized distilled water  
87 in a quantity equivalent to 2.5 times the dry paper weight, distributed on two sheets of paper,  
88 covered by a third and organized in the form of rolls, which were packed in transparent  
89 plastic bags to avoid the loss of water by evaporation. The rolls were placed in germination  
90 chamber of the Biological Oxygen Demand type (B.O.D.) regulated at a constant  
91 temperature of 25 °C. The counting was performed five to eight days after the test,  
92 considering the normal **seedlings were considered**, characterized by having a long, thin  
93 primary root coated with absorbent hairs along the entire surface, well defined lateral roots  
94 and well developed shoot, presenting the potential to continue its development and give rise  
95 to normal plants, the results were expressed in percentage.

96 The first germination counting was carried out concurrently with the germination test, the  
97 germinated **seeds were counted** on the 5th day after sowing [12].

98 For the germination speed index, daily countings were performed, five to eight days after the  
99 test, and the index was determined according to the equation proposed by Maguire [13].

100 At the end of the germination test, the normal seedlings of each replicate were measured  
101 with a ruler graduated in centimeters, the length of the seedlings were measured, and the  
102 results were expressed in centimeters per seedlings. The seedlings previously measured  
103 were packed in Kraft paper bags type, taken to a stove regulated at 80 °C for 24 hours and,  
104 after that period, weighed in an analytical scale with an accuracy of 0.001 g, the results were  
105 expressed in g.plantula<sup>-1</sup>.

Comment [AL6]: And what do they express?

## 106 2.4 Data Analysis

107 The experimental design used in the field was a randomized block, and completely  
108 randomized at the laboratory, the results were submitted to analysis of variance and

109 polynomial regression to evaluate the characteristics described previously, the linear and  
110 quadratic model were tested, where the significant model of higher order was selected to  
111 express the results. The program Sisvar 5.0 was used to perform the statistics analysis at  
112 the significance level of 5% probability ( $P = .05$ ). [14].

Comment [AL7]: I suggest you do a correlation analysis between the indicators.

### 113 3. RESULTS AND DISCUSSION

114 Changes in the coloration of the pods and seeds were observed during the maturation  
115 process (Table 1) and varied from dark green to brown with small dark brown dots.  
116 According to Lopez et al. [4], the color of the pods and seeds has been used as a good  
117 indicator of the harvest point, however, environmental factors must be observed since the  
118 differences in coloring can also be caused by its influence.

Comment [AL8]: Is this an objective or subjective evaluation? The method should be described in Material and methods if the result is described in this chapter.

119 **Table 1. Color of the pods and seeds of cowpea (*Vigna unguiculata* L. Walp.), cv.**  
120 **Corujinha, at different times of harvest.**  
121

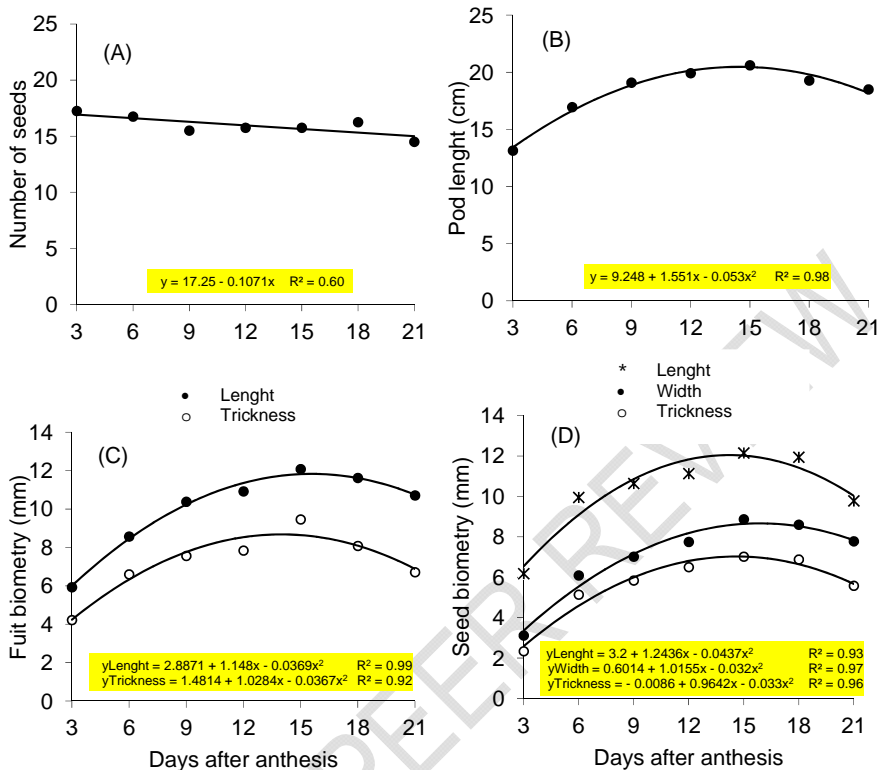
Harvest time	Days after anthesis	Color	
		Pods	Seeds
1 <sup>st</sup>	3	Dark green	Dark green
2 <sup>nd</sup>	6	Dark green	Dark green
3 <sup>rd</sup>	9	Light green	Dark green
4 <sup>th</sup>	12	Light green	Light green
5 <sup>th</sup>	15	Light green	Light green
6 <sup>th</sup>	18	Light brown	Light brown
7 <sup>th</sup>	21	Brown (dots)	Brown

122

123 For the number of seeds per fruit, a decreasing linear behavior is observed as a function of  
124 the harvesting time (Figure 1A) and, in relation to the size of the pods and the seeds, the  
125 data were adjusted to the quadratic model, with maximum length of (20.6 cm), width (8.8  
126 mm) and thickness (11.8 mm) of the pods obtained at 15 days after anthesis (Figures 1A  
127 and B). For the seeds, the maximum length (12 mm), width (8.6 mm) and thickness (7.0 mm)  
128 were also verified at 15 days after anthesis (Figure 1D).

129 Similar results were found by Botelho et al. [7] when studying the ideal harvest time for  
130 beans (*Phaseolus vulgaris* L.) where was verified a direct relation between seed size and  
131 physiological quality, in which seeds of lower size negatively influenced the seed quality of  
132 the lot.

133 Padua et al. [15] also verified that larger seeds originated higher soybean plants than plants  
134 originated from smaller seeds. According to Carvalho and Nakagawa [16], larger seeds were  
135 better nourished during their development, have well-formed embryos and a greater amount  
136 of reserves, with greater potential for germination and more vigorous plants when compared  
137 to smaller seeds.



138

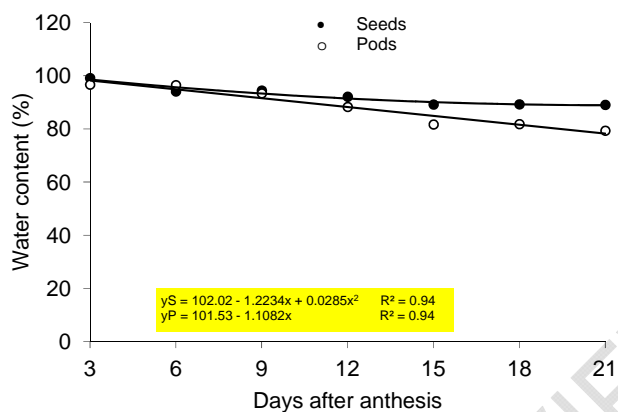
139

140 **Figure 1. Number of seeds (A), pod length (B), Fruit biometry (C) and biometry of the**  
 141 **seeds (D) of Cowpea bean (*Vigna unguiculata* L. Walp.), cv Corujinha, at different**  
 142 **times of harvest.**

143 The water content of the pods presented a linear behavior and the seed water content  
 144 presented a quadratic behavior according to the harvest times, in which, in the first  
 145 harvesting, at three days after the anthesis, the water content was high in the pods (96.7%)  
 146 and seeds (99.0%). Then, there was a gradual decrease until the last harvest, 21 days after  
 147 anthesis (18% for pods and 10% for seeds) (Figure 2). Botelho et al. [7] found similar results,  
 148 and verified a decrease in water content of common bean seeds during the physiological  
 149 maturation process.

150 However, the water content at the time of harvesting was high and this permanence for a  
 151 long period can negatively affect the storage and commercialization of the seeds, which can  
 152 result in the reduction of the physiological quality, cause deformations and favor conditions  
 153 for the development of fungi, which are factors responsible that accelerate the deterioration  
 154 process [3].

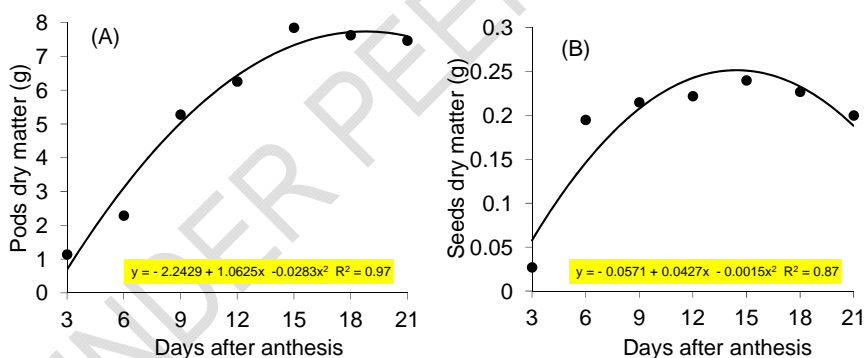
**Comment [AL9]:** Are all regression coefficients in the quadratic model statistically significant? I suggest that the functions for indicators should have specific designations, eg. instead of everywhere y write a designation for length  $l$ ; length seed -  $l_s$ , pod length -  $l_p$  (correct the fruit for the pod), etc. for other physical quantities. For time (day)  $t$ . Constants in equations should have such accuracy, as is the accuracy of measurement, eg. 2.8871 is too accurate, just 2.89. There is an error on the ordinate (C). Instead of (mm) it should be (cm). This is the method and it corresponds to reality. It's good in the text, see Line 125. Correct errors in length and thickness.



155

156 **Figure 2. Water content of the pods and seeds of Cowpea bean (*Vigna unguiculata* L.**  
 157 **Walp.), cv Corujinha, at different times of harvest.**

158 For the pods and seeds dry matter, data were adjusted to quadratic models, with maximum  
 159 values of 7.85 and 0.240 g, respectively, reached at 15 days after anthesis (Figure 3A and  
 160 3B). In the same harvesting time (15 days after anthesis) the maximum dry matter was  
 161 observed in the pods and seeds, the water content of the seeds was high, above 80%, and  
 162 the germination percentage reached the maximum values.



163

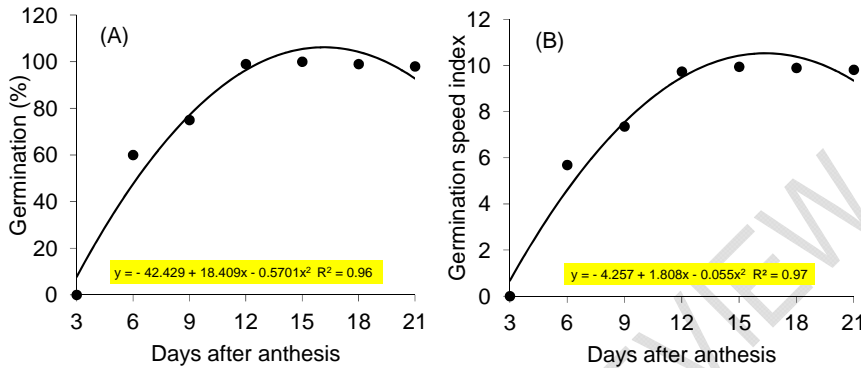
164 **Figure 3. Dry matter of the pods (A) and seeds (B) of Cowpea bean (*Vigna unguiculata***  
 165 **L. Walp.), cv Corujinha, at different times of harvest.**

166 A similar behavior was described by Eskandari [17] in seeds of *Vigna sinensis*, Botelho et al.  
 167 [7] and Bolina et al. [18] in common bean seeds and Nogueira et al. [2] in cowpea seeds.

168 The germination percentage and germination speed index were adjusted to the quadratic  
 169 model, with the highest values observed at 15 days after anthesis (100% and 9.9,  
 170 respectively), remaining high until the last day of evaluation (21 days after anthesis) (Figure  
 171 4A and B). Nogueira et al. [2] evaluated the development and physiological quality of  
 172 cowpea seeds, cv. BRS Guariba, during the maturation process, observed that at 14 days

**Comment [AL10]:** If the dry mass will be in percent, then please prepare one Figure with two graphs. In Fig. (B), the ordinate has commas. Please, change to dots. This is English and a separator for decimal numbers in the form of dots is required. Please change in the system settings to draw such coordinates.

173 after anthesis, the seeds reached their highest percentage of germination and germination  
 174 speed index, remaining stable until the last harvest, corroborating with the results obtained in  
 175 this work.

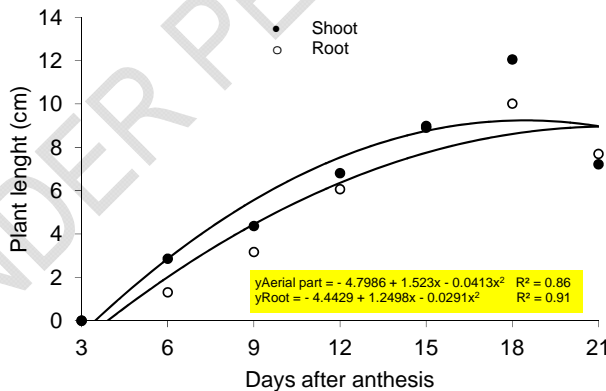


176

177 **Figure 4. Germination (A) and germination speed index (B) of seeds of Cowpea bean**  
 178 **(*Vigna unguiculata* L. Walp.), cv Corujinha, at different times of harvest.**

179 The shoot and root length data (Figure 5) were also adjusted to the quadratic model, where  
 180 a gradual increase was observed during the maturation process, with an estimated  
 181 maximum value for shoot (12.06 cm) and root (10.02 cm) at 18 days after the anthesis and,  
 182 with a subsequent small decrease.

Comment [AL11]: How is the difference between these indicators. What the new explains the second. Only one indicator is cleared.



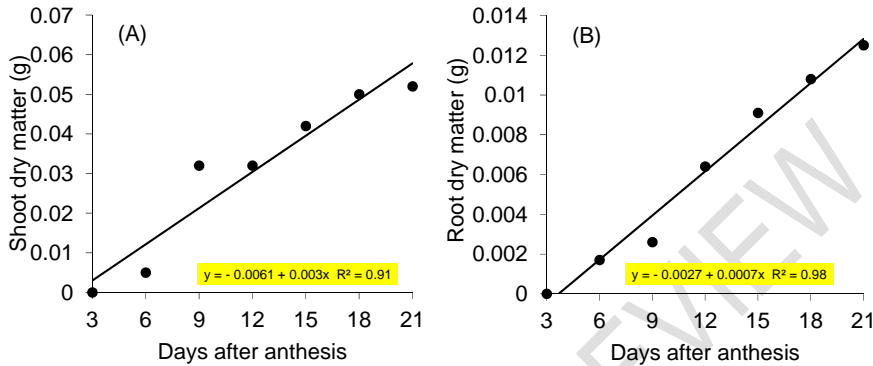
183

184 **Figure 5. Shoot and root length of seedlings of Cowpea (*Vigna unguiculata* L. Walp.),**  
 185 **cv Corujinha, at different times of harvest.**

186 For the shoot (Figure 6A) and root (Figure 6B) dry matter, a linear and increasing behavior  
 187 was observed as a function of the harvest time, reaching its maximum value (0.052 g for  
 188 shoot and 0.0125 g for root), at the last harvest, at 21 days after anthesis, which is due to

189 the metabolic and catabolic events of accumulation in the reserves tissue throughout the  
190 development of the seed.

191



192

193 **Figure 6. Shoot (A) and root (B) dry matter of seedlings of Cowpea bean (*Vigna***  
194 ***unguiculata* L. Walp.), cv Corujinha, at different times of harvest.**

Comment [AL12]: On the ordinate axes, replace the commas with dots as the decimal separator.

195 A direct relation between the seed size (Figure 1D) and physiological quality results could be  
196 verified, where the seeds of higher size were also those with higher percentage of  
197 germination (Figure 4A and 4B) and vigor (Figure 5 6A and 6B). According to Carvalho and  
198 Nakagawa [16], the size of the seeds may influence the germination and vigor, since larger  
199 seeds were well nourished during their development, usually have well-formed embryos,  
200 have larger amounts of reserves and are potentially more vigorous.

201 Similar results were also observed by Padua et al. [15], where they evaluated the influence  
202 of soybean seed size on the initial growth of plants and their effect on yield, they observed  
203 that larger seeds presented higher percentages of germination, vigor and produce plants  
204 with higher height at the **harvest** time, with higher yield, when compared to smaller seeds.

205 Therefore, it is important to harvest the seeds when they reach their maximum size,  
206 considering that it will result in higher seed quality, uniformity, more vigorous and productive  
207 plant stands.

#### 208 4. CONCLUSION

209 **The physiological maturity of cowpea bean seeds, cv. Corujinha is fast and occurred**  
210 **between 15 and 21 days after the anthesis, the period that correspond to 15 days after the**  
211 **anthesis is the best period for the harvest of this cultivar, which coincides with a greater**  
212 **germination and vigor.**



213 **COMPETING INTERESTS**

214 Authors have declared that no competing interests exist.

215 **REFERENCES**

- 216 1. Lima JME, Fagundes GS, Smiderle OJ. Physiological quality of cowpea beans  
217 treated with diatomaceous soil and infested by weevils. Magazine in Agribusiness  
218 and Environment. 2014; 7 (3): 733-746.
- 219 2. Nogueira NW, Freitas RMO, Torres SB, Leal CCP. Physiological maturation of  
220 cowpea seeds. Journal of Seed Science. 2014; 36 (3): 312-317.
- 221 3. Birth WM, Freitas RA, Croda MD. Conservation of vegetable seeds in family  
222 agriculture. Embrapa (Technical Bulletin 54), 2008; 6p.
- 223 4. Lopes IS, Nóbrega AMF, Matos VP. Maturation and harvest of seed of *Amburana*  
224 *cearensis* (Allem.) A.C. Smith. Forest Science. 2014; 24 (3): 565-572.
- 225 5. Marcos Filho J. Seed physiology of cultivated plants. Piracicaba: FEALQ. 2015.  
226 495p.
- 227 6. Vidigal DS, DCFS Days, EVRV Pine, LAS Days. Physiological and enzymatic  
228 changes during pepper seeds (*Capsicum annuum* L.) maturation. Brazilian Journal  
229 of Seeds. 2009; 31 (2): 129-136.
- 230 7. Botelho FJE, Guimarães RM, Oliveira JA, Evangelista JRE, Eloi TA, Baliza DP.  
231 Physiological performance of bean (*Phaseolus vulgaris* L.) seeds harvested in  
232 different developmental periods. Science and Agrotechnology. 2010; 34 (4): 900-  
233 907.
- 234 8. Noble DAC, Trogello E, Morais DLB, Brandão Junior DS. Quality of black sesame  
235 seeds (*Sesamum indicum* L.) at different harvest times after sowing. Brazilian  
236 Journal of Medicinal Plants. 2013; 15 (4): 609-616.
- 237 9. Figueiredo Neto A, Almeida FAC, Dantas BF, Garrido MS, Aragão CA. Maturation of  
238 pumpkin seeds (*Curcubita moschata* Duch) produced in the semiarid. Comunicata  
239 Scientiae. 2014; 5(3): 302-310.
- 240 10. Brazil. Ministry of Agriculture. Pedology team and soil fertility. Division of Agrology -  
241 SUDENE. Exploratory survey: soil recognition of the state of Paraíba. Rio de  
242 Janeiro: MA / CONTA / USAID / SUDENE, 1972. 670p. (Technical Bulletin, 15,  
243 Pedology Series, 8).
- 244 11. Brazilian Agricultural Research Corporation - EMBRAPA. Brazilian system of soil  
245 classification. 3.ed. Brasília, DF: 2013.
- 246 12. Brazil. Ministry of Agriculture, Livestock and Supply. Rules for seed analysis.  
247 National Secretariat of Agricultural and Livestock Defense. Brasília: MAPA / ACS,  
248 2009. 395p.
- 249 13. Maguire JD. Speed of germination aid in selection and evaluation of seedling  
250 emergence and vigor. Crop Science. 1962; 2: 176-177.
- 251 14. Ferreira DF. Sisvar: a computer statistical analysis system. *Ciência e*  
252 *Agrotecnologia*. 2011; 35(6): 1039-1042.
- 253 15. Padua GP, Zito RK, Arantes NE, França Neto JB. Influence of seed size on  
254 physiological seed quality and soybean yield. Brazilian Journal of Seeds. 2010; 32  
255 (3): 009-016.
- 256 16. Carvalho NM, Nakagawa J. Seeds: science, technology and production. 5.ed.  
257 FUNEP: Jaboticabal, 2012. 590p.
- 258 17. Eskandari H. Seed quality changes in cowpea (*Vigna sinensis*) during seed  
259 development and maturation. Seed Science and Technology. 2012; 40 (1): 108-112.

260 18. Bolina CC. Physiological maturation of the seed and determination of the  
261 appropriate time of harvest of the bean (*Phaseolus vulgaris* L.). Linkania Master  
262 Scientific Journal. 2012; 2 (3).

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