

# Original Research Article

## **Effects of using different types of organic wastes for the mass culture of *Moina macrocopa***

### **ABSTRACT**

*Moina macrocopa* was cultured with different animal manures (chicken manure, Pig manure and cow manure) and food waste to determine the impact of these food sources on its mass production. All diets were provided at five different concentrations: 500, 1000, 2500, 5000 and 10000 ppm. Gross and net reproductive rates were higher in 1000 ppm concentration of food waste medium and the highest average population growth was obtained of about 9 individual *M. macrocopa* per ml, whereas pig manure treatment showed the lowest among all the culture medium. Highest population density was observed in low concentration treatments, on the contrary higher concentrations showed adverse effect on *M. macrocopa* cultivation. The results of this study suggest that 1000 ppm concentration of food waste produces better results than other animal manures which showed the highest population density and exhibited comparatively higher percentage of highly unsaturated fatty acids than the other treatments and could be an inexpensive and sustainable cultivation approach of *Moina macrocopa*.

**Keywords:** *Moina macrocopa*, Animal manure, Food waste, Vial test, Life table demography

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### **1. INTRODUCTION**

To feed the increasing human population, it is therefore imperious to upgrade aquaculture, including fish farming whose development goes essentially through the success of larval rearing which requires the availability of zooplankton [1,2,3,4]. Yet, the most used zooplankton for the feeding of the fish larvae was *Artemia* [5,6]. But, the utilization, mostly in developing countries is difficult because of these cysts hatching conditions, high cost and low availability on the local market [7]. It is then important to make an intensive production of zooplankton at low cost for the expansion of fish farming. *M. macrocopa* is increasingly used as food for larval and post larval rearing of crustaceans [8] and teleost

29 fish in culture [9,10,11,12]. It is a superior live food compared to *Artemia* due to its relatively high  
30 protein and nutrient content [13]. Although its culture technique is relatively simple but the specific  
31 production and fed technique knowledge for the commercial scale production was incipient in spite of  
32 its wide distribution from temperate to the tropical region.

33 Food resources play an important role in the production of *M. macrocopa* in natural systems [14,15]. In  
34 natural habitats, biotic and abiotic parameters such as water quality, quantity, quality level of food  
35 available and population density are one of the most important factors that interact in the population  
36 growth of the zooplankton. Among these factors, population density and food availability are the  
37 predominant factors affecting the growth of *M. macrocopa* [16]. Conventional food sources of *M.*  
38 *macrocopa* are very expensive and contains very low nutritive value. So, the scientists are now trying  
39 to find an alternative food source of *M. macrocopa* at low cost with high nutritive value.

40 Hence, relatively large amount of *M. macrocopa* that are required for fish larvae cultivation can be  
41 produced from inexpensive, renewable waste materials. However, different types of wastes generated  
42 day by day in extensive quantities, creating a significant problem in its management and disposal.  
43 Besides, domestic policy of South Korea totally banned the ocean dumping of all wastes from 2014,  
44 following this banning; Korean policy and industry have been tending to convert the waste into  
45 resources [17]. Animal manures have a long history of use as a source of soluble phosphorous,  
46 nitrogen and carbon for natural food production [18]. Animal manure used as organic matter supplied  
47 to ponds can stimulate the phytoplankton growth and also increases the biomass of zooplankton  
48 [19,20]. Animal wastes using for fertilization practices are popular in many countries to sustain  
49 productivity at low cost [21, 22]. But, only limited information is available on the utilization prospects of  
50 food waste as an alternative or additional protein source of *M. macrocopa*, which can lower the cost of  
51 fish farming and at the same time, conserve the ecological value of fish ponds.

52 When Cladocera is used as a food for larval fish, nutrient enrichment is necessary as is the case with  
53 *Artemia* [23]. Because the ingredients that compose their body change according to the food they  
54 intakes [24]. Some quantitative data are available on the fatty acid profiles of rotifers, copepods and  
55 cladocerans using algae as food, but no data are available on the fatty acid profiles of *M. macrocopa*  
56 using organic wastes as food. *Chlorella vulgaris* is commonly used in *M. macrocopa* culture [41]. But it  
57 needs to be enriched by a commercial enrichment diet before feeding to the fish larvae [25]. However,

58 it is necessary to improve the fatty acid composition of *M. macrocopa* in natural way by switching its  
59 diet to organic wastes. Highly unsaturated fatty acids (HUFA) enhance the essential lipid levels and  
60 these essential fatty acids promote the growth of *M. macrocopa* [26]. So, it need to be investigated  
61 because the synthesis and accumulation of fatty acids in zooplankton are related to the stage of the  
62 individual and the frequency of reproduction among others.

63 The present study was designed to test the effect of different animal manures and food waste in mass  
64 culture of *M. macrocopa* with a view of investigating quality and quantity required for maximizing  
65 production. Determination the optimum concentration of each organic waste for the culture of *M.*  
66 *macrocopa* are important for its mass cultivation. Development of a suitable culture media for  
67 commercial production of *M. macrocopa* will be an inexpensive alternative approach to live feeds  
68 needed for fish rearing.

## 69 **2. MATERIALS AND METHODS**

### 70 **2.1 Source of *M. macrocopa***

71 *M. macrocopa* were collected from a pond near Tongyeong, South Korea and the sample was taken to  
72 the laboratory immediately. *M. macrocopa* species were isolated from the collected sample by the  
73 micropipette and placed individually in petri dishes filled with dechlorinated tap water (10ml/plate) for  
74 breeding. Mature Moina started breeding overnight and baker's yeast was added at 1g/l to the petri  
75 dishes as a food source during the breeding period. Newborn *M. macrocopa* were collected for  
76 subsequent experiments.

### 77 **2.2 Source of Organic Wastes**

78 Three different types of animal manures such as pig manure, cow manure and poultry manure were  
79 sourced from the pig, cow and poultry production institute in Goseong, South Korea. The food wastes  
80 used in the present study included food processing waste (e.g., various types of fruit peels and leafy  
81 vegetables, rice bran, and soybean meal) and post-consumption waste (e.g., rice grain, spaghetti, beef,  
82 pork, and chicken) collected from local hotels and restaurants. The collected food wastes were  
83 transferred to the laboratory, for further processing. The food wastes were mixing together in a mixer  
84 machine, diced into small pieces, and excessive water was squeezed out by waste compressing  
85 equipment. Then the final leachate was used in this experiment.

86 **2.3 Experimental design**

87 **2.3.1 Population growth experiment**

88 The experiment was conducted in total 60 tanks with a water volume of 40 L. Three replicates were  
89 used for each treatment. The tanks were cleaned and dried for two days and filled with tap water and  
90 left for one day with aeration for dechlorination. Water temperature in the tanks was maintained at  
91 25°C. The temperature of water reservoir was regulated by a thermostat, which controlled the on / off  
92 switch of a 2000-W electric heater. Four treatments with five different concentrations: chicken manure,  
93 pig manure, cow manure, and food waste of 500, 1000, 2500, 5000, and 10000 ppm were assigned in  
94 the experiment. Twenty healthy individuals *M. macrocopa* were individually introduced into the tanks.  
95 The trial has repeated a total of three times and data pooled at the end of the period for each  
96 treatment.

97 Following initiation of different growth experiments, the number of living individuals of each tank was  
98 counted daily. The population of *M. macrocopa* was recorded by using the Sedgewick-Rafter counter  
99 cell which is 50 mm long, 20 mm wide and 1 mm deep. *M. macrocopa* cultured in each experimental  
100 tanks was recorded by using a tally counter under a dissecting microscope (10X to 40X magnification).  
101 The number (no./mL) was calculated according to the formula outlined by Boyd and Lichtoppler [27] :

$$\text{Number of } Moina \text{ macrocopa /ml} = \frac{T \times 1000}{A \times N \times \text{Vol. of concentrate in ml/Vol. of sample}}$$

103 Where,

104 T = Total number of *M. macrocopa* counted

105 A = Area of grid in mm<sup>2</sup>

106 N = Number of grids counted

107 1000 = Area of counting chambers in mm<sup>2</sup>

108 **2.3.2 Water quality parameters**

109 Dissolved oxygen (mg/L), pH and salinity(%) were measured by dipping into the water surface.  
110 Ammonia was measured by Palintest compact ammonia duo meter. Recordings were taken after tank  
111 inoculation and thereafter every 24 hours.

### 112 **2.3.3 Vial test**

113 Twenty vials of 50mL size were selected for this test and each was replicated three times. Four  
114 treatments with five different concentrations: chicken manure, pig manure, cow manure, and food  
115 waste of 500, 1000, 2500, 5000, and 10000 ppm were assigned in each vial. One healthy neonate of  
116 less than 24hrs old was transferred in each vial and the alive individual and offspring produced from it  
117 were quantified every 24hrs. The neonates produced by *M. macrocopa* were collected gently and  
118 transferred into a culture dish for quantification. This counting was also carried out using a tally counter.  
119 Then it transferred to the new test jars with appropriate culture medium and the dead adults and  
120 neonates were removed. Each vial test was discontinued after the last adult in each vial was died.  
121 Mortality and fecundity were recorded to calculate the life table demography of *M. macrocopa*.

### 122 **2.3.4 Life table demography**

123 Life table demographics is an important tool for describing the life cycle of zooplankton under  
124 continuously changing environmental conditions. The survival period, initial age of reproduction,  
125 average longevity, gross reproduction rate, net reproduction rate, rate of increase, and generation time  
126 were selected for life history variables for this study [28]. The following definitions apply: initial age of  
127 reproduction = the time when a female started to produce her first batch of offspring (number of days);  
128 longevity = the average number of days the female survived. The following formulae were used [29].

129 Average longevity =  $\sum n_x/n$ ;

130 Gross Reproduction rate =  $\sum m_x$ ;

131 Net reproduction rate (Ro) =  $\sum l_x m_x$ ;

132 Generation time (T) =  $\sum l_x m_x X / Ro$

133 Where,

134  $n_x$  = Number of individuals alive for each age class

135  $m$ =The age specific fecundity (number of neonates produced per surviving female at age X)

136  $l_x$ =The proportion of individuals surviving to age  $x$

137  $n$  = The number of replicates

138 The final rate of population increase ( $r$ ) was calculated over the 21 days experimental period. The rate  
139 of population increase ( $r$ ) was derived using the following equation;  $r = (\ln N_t - \ln N_0)/t$ , where  $N_0$ =initial  
140 population density and  $N_t$ =population density after time  $t$  [29].

#### 141 **2.3.5 Population density of *M. macrocopa***

142 Twenty vials of 50 mL size were assigned with four treatments of five different concentrations: chicken  
143 manure, pig manure, cow manure, and food waste of 500; 1,000; 2,500; 5,000; and 10,000 ppm in  
144 each vial. One healthy neonate of less than 24hrs old was transferred in each vial and the alive  
145 individual and offspring produced from it were quantified every 24hrs, which was carried out until  
146 finishing of this experiment. *M. macrocopa* were transferred into different culture dishes for  
147 quantification and after quantification, live *M. macrocopa* were returned to the culture vial, and the  
148 dead organisms were discarded. This experiment was carried out for 21 days.

#### 149 **2.3.6 Fatty acid analysis of *M. macrocopa* cultured in different organic wastes**

150 Total lipids of *M. macrocopa* were extracted according to the Bligh and Dyer method [30] by using  
151 solvent mixture consisting of chloroform and methanol (2:1, v/v). After phase equilibration, the lower  
152 chloroform layer was removed and total lipids were extracted by removing solvent using a rotary  
153 evaporator (R-114, BUCHI, Swiss) at 38 °C. 100 mg of extracted total lipid were put into a capped tube  
154 and added 1.5 ml 0.5 N NaOH-methanol solution. The sample was mixed by vortex and heated 100 °C  
155 for 8 minutes for saponification. After cooling, methylation was done by using a fatty acid methyl ester  
156 (FAME) with BF<sub>3</sub>-methanol. Then the sample was dissolved into 2 ml iso-octane and fatty acids were  
157 analyzed using gas chromatography (Clarus 600, Perkin Elmer, USA) equipped with capillary column  
158 (Omegawax-320, 30 m × 0.25 mm I.D., Supelco Co., Bellefonte, PA, USA). The operating parameters  
159 were as follows: carrier gas =helium; detector (FID) temperature =270 °C; injection temperature =  
160 250 °C; column temperature =180 °C for 8 min, programmed to increase at 3 °C/min up to 230 °C with a  
161 final holding time of 10 min; split injection at 1:50 ratio. Menhaden oil was used as standard. Each of

162 the specific fatty acid methyl ester peaks was identified by determining its equivalent chain length with  
163 reference to the known standard.

164

## 165 **2.4 Statistical Analysis**

166 The statistical analysis were carried out to evaluate the differences in the means of the derived  
167 individual number of *M. macrocopa* and environmental parameters of different treatments by using  
168 one-way ANOVA. Statistical significance among the different treatments was accepted at  $p < 0.05$  and  
169 the statistical package of SPSS- 16 (SYSTA, USA) was used to express the result.

170

## 171 **3. RESULTS**

172

### 173 **3.1 Water quality**

174 Table 1 shows the mean pH, DO and ammonia content of five various concentrations of four different  
175 culture medium over three weeks experimental period. The highest pH was recorded in the pig manure  
176 treatment which was in the range between 7.33 to 7.72 and the lowest was recorded in the food waste  
177 which was in the range of 5.50 to 6.46. pH increased with the increase concentration of animal  
178 manures, but in case of food waste it decreased. Food waste treated media showed significantly  
179 lowest ( $P < 0.05$ ) DO level throughout the culture period, which were found to be in the range of 0.4 to  
180 1.08 mg/l. As the temperature was fixed from the beginning of the experiment, there is no significant  
181 difference was observed between the treatments. Ammonia contents increased with the increasing  
182 concentrations of organic wastes but showed no significant difference ( $P > 0.05$ ) among the treatments  
183 of all concentration.

184 **Table 1. Water quality parameters for the *M. macrocopa* cultures at different concentrations of animal**  
185 **manures and food waste of the experimental duration**

Culture medium	Concentration (ppm)	pH	DO (mg/l)	Temperature (°C)	Ammonia (mg/l)	Salinity (PSU)
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Chicken manure	500	6.816±0.02	2.83±0.01	25±0.1	0.20±0.03	0.01±0.0
	1000	6.973±0.02	2.71±0.01	25±0.1	0.22±0.03	0.01±0.0
	25000	7.281±0.04	2.25±0.02	25±0.1	0.33±0.06	0.01±0.0
	5000	7.406±0.03	2.28±0.03	25±0.1	0.39±0.04	0.02±0.1
	10000	7.554±0.02	0.57±0.03	25±0.1	0.41±0.08	0.02±0.0
Pig manure	500	7.335±0.01	3.54±0.02	25±0.1	0.23±0.02	0.01±0.0
	1000	7.208±0.02	3.23±0.02	25±0.1	0.25±0.06	0.01±0.0
	25000	7.557±0.02	3.06±0.04	25±0.1	0.29±0.05	0.01±0.0
	5000	7.524± 0.03	2.86±0.07	25±0.1	0.41±0.03	0.02±0.0
	10000	7.729±0.06	1.03±0.03	25±0.1	0.44±0.05	0.03±0.0
Cow manure	500	6.719±0.02	3.45±0.01	25±0.1	0.19±0.05	0.01±0.0
	1000	7.083±0.01	3.11±0.03	25±0.1	0.22±0.04	0.01±0.0
	25000	7.159±0.03	1.76±0.02	25±0.1	0.26±0.04	0.01±0.0
	5000	7.230±0.02	1.53±0.05	25±0.1	0.36±0.03	0.01±0.0
	10000	7.592±0.04	0.61±0.03	25±0.1	0.37±0.08	0.03±0.1
Food waste	500	6.461±0.02	1.08±0.01	25±0.1	0.23±0.02	0.01±0.0
	1000	6.823±0.03	0.91±0.02	25±0.1	0.25±0.04	0.01±0.0
	25000	6.064±0.05	0.93±0.01	25±0.1	0.36±0.06	0.01±0.0
	5000	5.963±0.02	0.32±0.01	25±0.5	0.35±0.05	0.03±0.1
	10000	5.501±0.03	0.41±0.02	25±0.1	0.38±0.04	0.03±0.0

186

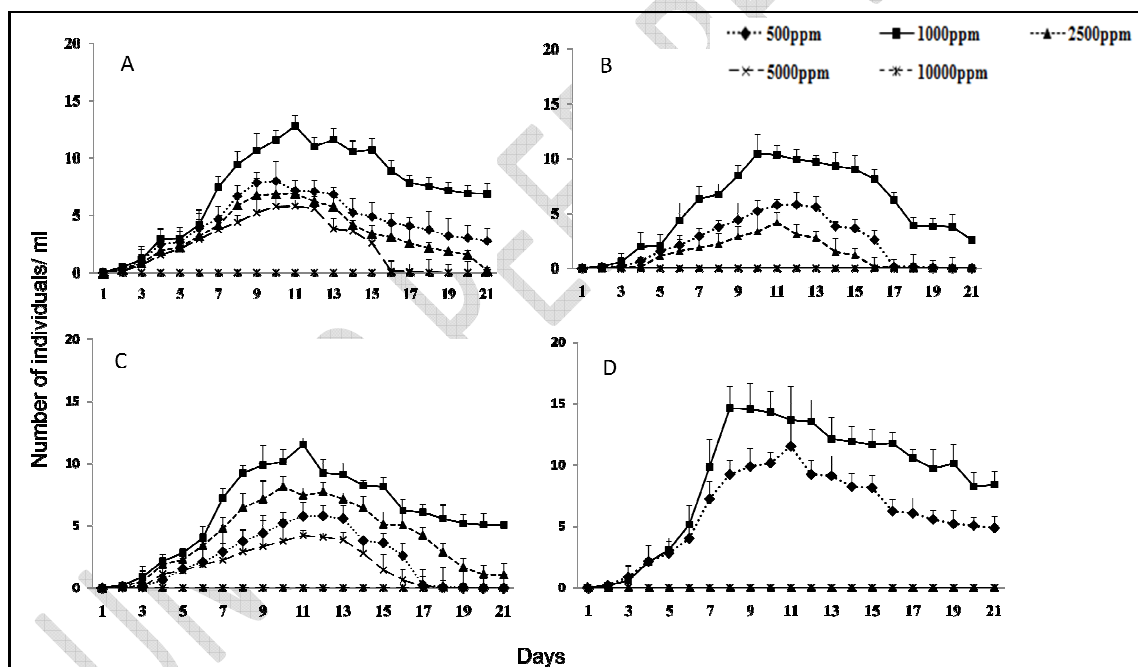
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188



189 **3.2 Population growth of *M. macrocopa***

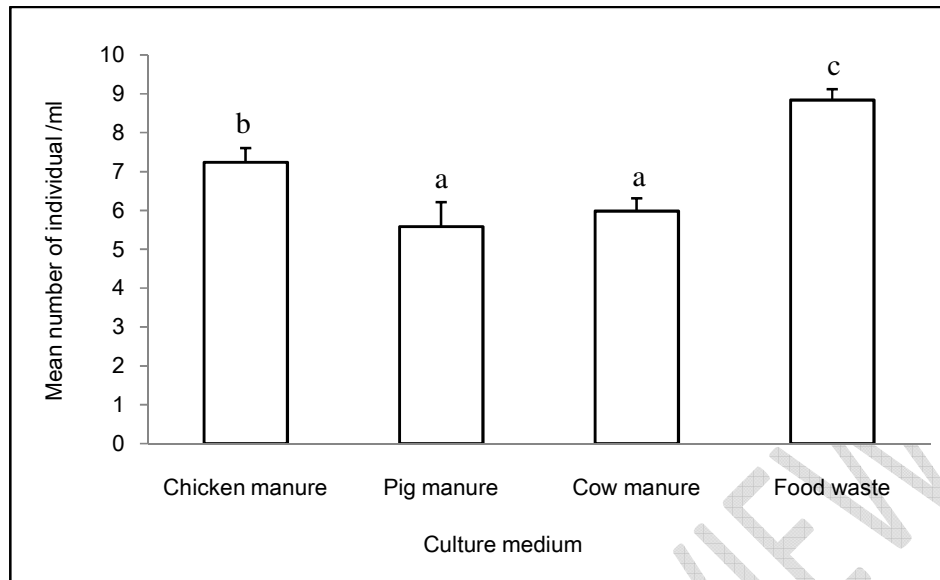
190 Figure 1 shows the population growth of *M. macrocopa* with five various concentrations of four  
191 different organic waste treated medium for 21 days experimental period. Population growth was  
192 significantly higher in the treatments with low concentrations, on the contrary there is no population  
193 growth was observed in 10,000 ppm; the highest concentration used in this experiment. The highest  
194 growth was recorded in 1,000 ppm concentration for four culture medium and among these four  
195 mediums, food waste showed the best growth rate. Figure 2 showed the mean population growth of *M.*  
196 *macrocopa* cultivated in 1,000 ppm concentration of four culture medium. Highest mean population  
197 was found in food waste medium which was followed by chicken manure medium and showed a  
198 significant difference ( $P<0.05$ ) among the other medium, where there is no significant ( $P<0.05$ )  
199 difference was observed between pig manure and cow manure treated treatment.



200

201 Fig. 1. Population growth of *M. macrocopa* in different concentrations of (A) chicken manure, (B) pig  
202 manure, (C) cow manure, & (D) food waste culture medium for 21 days experimental period. Error  
203 bars indicate means  $\pm$  standard deviation.

204



205

206 Fig. 2. Mean population growth of *M. macrocopa* cultured in 1000ppm concentration of four culture  
 207 medium. Values are the ( mean  $\pm$  SD ). Different subscripts denote significant differences at  $P < 0.05$   
 208 (a < b < c) .

209

### 210 **3.3 Life table demography**

211 Data on the selected life history variables (Table 2) of *M. macrocopa* showed that the average lifespan  
 212 was lowest in treatment with high concentration, about 1 to 2 days. The offspring production of *M.*  
 213 *macrocopa* in relation to the different concentrations of treatments showed a distinct shift towards early  
 214 reproduction with low concentration treatments. Gross and net reproductive rate also showed similar  
 215 trends. Large numbers of offspring were produced by *M. macrocopa* cultured in food waste (500;  
 216 1000ppm), chicken manure (500; 1000 ppm/l), cow manure (500; 1,000 ppm) and pig manure (500,  
 217 1,000 ppm) medium. Fecundity declined at high concentrations (2500 ppm and 5000 ppm), while the  
 218 highest 10,000 ppm concentration did not lead to the production of offspring for every treatments. The  
 219 rate of population increase was positive for all the culture medium, but 1000 ppm/l showed the  
 220 maximum. The highest rate of population increase ( $r$ ) calculated for this experiment was  $0.51 \pm 0.08$   
 221 obtained in the food waste treated treatment. This was followed by the chicken manure, cow manure  
 222 and pig manure medium with ' $r$ ' value of  $0.47 \pm 0.23$ ,  $0.43 \pm 0.16$ , and  $0.33 \pm 0.02$  respectively.

223

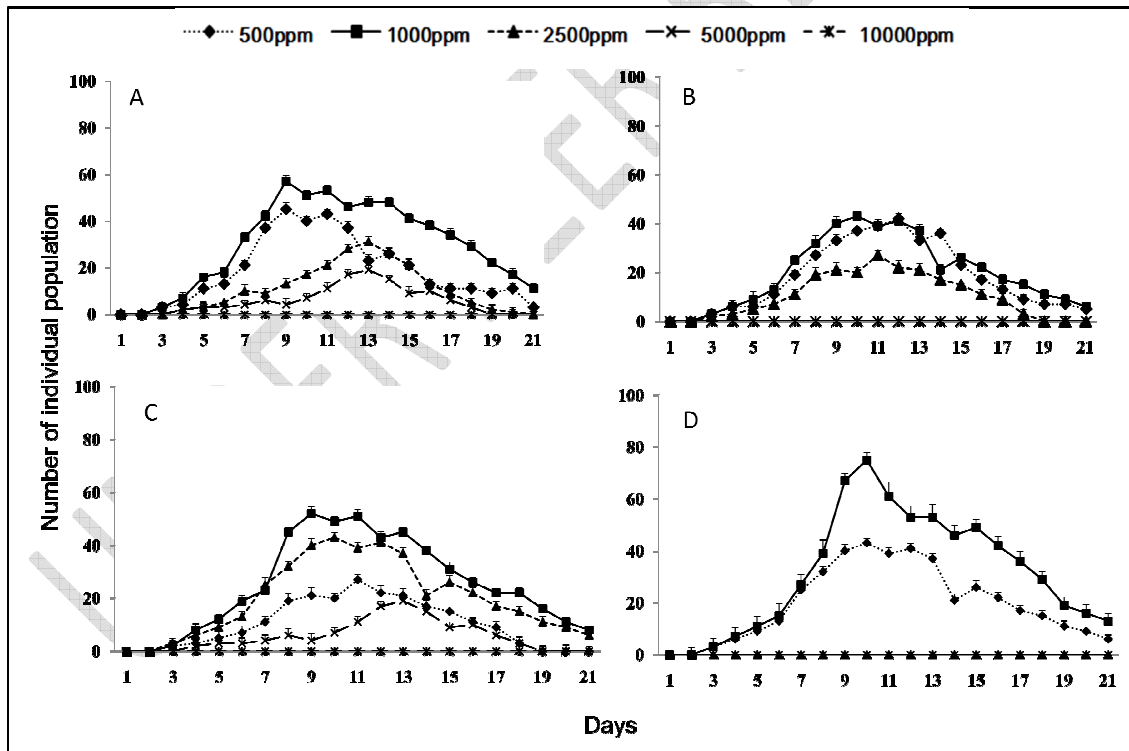
**Table 2. Life table of *M. macrocopa*, cultured with different organic wastes at different concentrations. Data are the means with the standard error of three replicates**

Medium types	Medium Conc. (ppm)	Initial age of reproduction	Longevity	Net reproduction rate	Gross reproduction rate	Generation time	Rate of population increase
Chicken manure	500	2.89±0.06	8.16±0.13	11.73±0.11	14.06±1.03	3.41±0.11	0.39
	1000	2.87±0.08	9.20±0.23	19.32±0.08	23.42±0.91	3.00±0.36	0.47
	2500	3.00±0.08	6.27±0.18	9.17±0.13	11.42±0.85	3.16±0.25	0.36
	5000	3.07±0.05	5.63±0.20	5.21±0.16	5.21±1.14	3.27±0.25	0.32
	10000	-	2.86±0.41	-	-	-	-
Pig manure	500	3.06±0.03	5.63±0.13	8.25±0.21	10.14±0.21	4.00±0.17	0.24
	1000	3.08±0.08	7.74±0.11	11.69±0.17	13.32±0.16	3.53±0.26	0.33
	2500	3.00±0.07	6.75±0.18	7.32±0.17	9.11±0.20	3.60±0.25	0.27
	5000	-	1.00±0.20	-	-	-	-
	10000	-	1.00±0.41	-	-	-	-
Cow manure	500	3.06±0.08	7.63±0.11	10.21±0.21	10.53±1.11	3.23±0.11	0.33
	1000	3.06±0.05	9.14±0.16	15.57±0.18	17.71±1.31	3.10±0.36	0.43
	2500	3.26±0.08	9.17±0.28	15.42±0.16	17.24±0.93	3.00±0.25	0.43
	5000	3.33±0.05	5.00±1.22	4.16 ±0.16	4.23±1.14	3.27±0.25	0.32
	10000	-	3.75±0.41	-	-	-	-
Food waste	500	3.00±0.07	8.71±0.11	12.42±0.18	15.35±1.00	3.00±0.10	0.43
	1000	2.88±0.06	9.82±0.26	28.16±0.18	33.71±0.21	3.00±0.16	0.51
	2500	-	1.75±0.08	-	-	-	-
	5000	-	1.00±0.01	-	-	-	-
	10000	-	1.00±0.00	-	-	-	-

Dash (-) indicates no offspring was produced

227 **3.4 Populations density of *M. macrocopa***

228 Populations from a single neonate of *M. macrocopa* showed average growth rate until the first week in  
229 various concentrations of four different culture medium (Figure 3). It was growing continuously until the  
230 second week and after that, it started to decline and this same trend was observed in all treatments.  
231 Among all the concentrations highest population was found in 1,000 ppm concentration of all the  
232 culture medium. In case of food waste medium neonate is died in 2500, 5000 and 10000 ppm  
233 concentrations within 24hrs and 75 numbers of individual *M. macrocopa* were counted on day 10, in  
234 1,000 ppm concentration, which was its peak population. Vials containing chicken manure, cow  
235 manure and pig manure medium showed moderate production with a peak population of 57, 52 and 43  
236 individual on day 9 in 1,000 ppm concentration. After that the decreasing trend was started, which  
237 means that these peak populations are the maximum density of *M. macrocopa* for different culture  
238 medium.



239  
240 Fig. 3. Population density from a single neonate of *M. macrocopa* in different concentrations of (A)  
241 chicken manure, (B) pig manure, (C) cow manure, & (D) food waste treated medium for 21 days  
242 experimental period. Error bars indicate means  $\pm$  standard deviation.

243

### 244 **3.5 Fatty acid composition of *M. macrocopa* cultured in different organic wastes**

245 Table 3 shows the average percentage of fatty acid composition of *M. macrocopa* cultured in different  
246 organic waste. Among the saturated fatty acids, 14:0, 16:0 and 18:0 comprised of about 33% of the  
247 total fatty acids and 16:1n-7, 18:1n-7, 18:2n-6 are the most dominant unsaturated fatty acids. The fatty  
248 acid composition of *M. macrocopa* cultured in cow manure, food waste, chicken manure and pig  
249 manure was dominated by, 16:00, 18:00, 18:1n-9, 18:2n-6. The level of EPA and DHA was  
250 significantly higher in *M. macrocopa* that cultured in food waste medium than those other wastes.

251

## 252 **4. DISCUSSION**

253

254 Among the cladocereans, *M. macrocopa* has been investigated most intensively with regard to the  
255 effects of food abundance on its growth and reproduction [10,31,32]. Quality and quantity of food are  
256 the most important factors in determining biomass production of *M. macrocopa* species. The results of  
257 this study indicate that the growth performance of *M. macrocopa* using various concentrations of  
258 different organic wastes as a culture medium are different. Low population growth was observed in  
259 high concentration of culture medium, while the highest concentration used in this study did not lead to  
260 the production of offspring in all the culture medium. This phenomenon has been attributed by various  
261 workers that the presence of high concentrations of animal manure significantly reduced the water  
262 quality, deplete the plankton population and cause adverse effect on the culture [4]. Nandini and  
263 Sarma [33], revealed that, the decline in neonate production that accompanied increasing  
264 concentrations of culture medium were presumably caused by the increased effort associated with  
265 food gathering due to active filtering of the food particles. In fact, high concentrations of all the diets  
266 produced suboptimal culture conditions. Burak [34] & Porter[35] described that, high concentration of  
267 particles can actually lead to starvation of cladocerans as they are unable to clean thoracic limbs that  
268 are clogged by high particulate concentrations. Savas [31] also found that, population of *M. macrocopa*  
269 declined in using high concentration of algal supplement. In this study, 1000 ppm concentration  
270 showed the optimal concentration of all the culture media in terms of growth and reproduction

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**Table 3. Fatty acids composition (%) of *M. macrocopa* cultured in different organic wastes**

Fatty acids	Chicken manure	Pig manure	Cow manure	Food waste
14:0	3.59±0.12	3.13±0.08	3.8±0.15	4.2±0.18
16:0	18.3±1.20	22.1±2.50	18.6±1.50	17.11±2.71
16:1n-7	6.9±0.03	14.12±0.11	9.8±0.08	6.83±0.09
18:0	12.88±0.51	10.6±0.40	8.8±0.72	11.94±0.26
18:1n-9	17.7±0.08	16±0.91	16.3±0.18	13.14±0.28
18:1n-7	20.9±2.11	11.3±0.57	8.3±1.20	9.6±0.90
18:2n-6	13.4±0.07	16.6±0.08	24.2±2.74	20.5±1.25
18:3n-3	3.2±0.04	3.8±0.01	4.6±0.24	7.3±0.14
18:4n-3	0.17±0.01	-	2.6±0.09	-
20:00	-	-	0.13±0.08	1.5±0.07
20:2n-6	0.03±0.01	0.03±0.01	0.07±0.01	0.11±0.05
20:3n-6	0.82±0.05	0.69±0.04	0.7±0.01	0.87±0.09
20:4n-6	0.69±0.03	0.82±0.09	0.54±0.08	0.87±0.04
20:3n-3	-	-	0.13±0.02	2.11±0.06
20:5n-3	1.06±0.06	0.33±0.04	0.29±0.08	1.8±0.07
22:5n-3	-	-	-	0.06±0.01
22:6n-3	0.26±0.09	0.13±0.03	0.08±0.05	0.81±0.01
ΣSFA	34.77	35.83	31.33	34.75
MUFA	45.5	41.42	34.4	29.57
PUFA	19.63	22.4	33.21	34.43

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Data of 1,000 ppm concentration of different organic waste are shown here.  
Hyphen (-) indicates non- detectable fatty acids.

277

278 performance. Among the culture medium, the highest population growth was observed in food waste  
279 medium. In this study pH increased with the high concentration of animal manures but food waste  
280 showed different trend where pH decreased with the increasing concentrations, might be the presence  
281 of higher amount longer chain fatty acids. Life table demography of *M. macrocopa* followed the same  
282 trend. Average lifespan was lowest in culture mediums with higher concentration. Food waste  
283 containing 1,000 ppm concentration showed the highest average lifespan and early reproduction ability.  
284 The present study showed that, *M. macrocopa* needs time to become sexually mature at high diet  
285 concentration. In contrast, Loh [32] reported that, initial age of reproduction of *M. macrocopa* is earlier  
286 in high concentration diet than in low concentration. Different results are observed in this test, which  
287 indicates that diet type and concentration play a significant role in determining the initial age of  
288 reproduction. Gross and net reproduction rates were generally higher at lower treatment  
289 concentrations and highest in food waste medium than other diets. Jana and Pal [36] revealed in their  
290 study, high fecundity and gross reproduction rates suggest that growth performance of any species is  
291 largely depends on the high carbon/nitrogen ratio in the food source. Which clearly indicates that food  
292 waste contained high C/N ratio than the other medium.

293 The highest population density was obtained in 1,000 ppm concentration of food waste medium. The  
294 good performance of this food source can be attributed to the feeding habit of *M. macrocopa* that tend  
295 to consume bacteria and filtered particles that are abundant in food waste , when other food sources  
296 has limited. In terms of time and efficiency, *M. macrocopa* cultured in food waste reached its peak  
297 population on day 10 which was about 75 individual. This also means that *M. macrocopa* could be  
298 harvested by food waste within a shorter period of time, thus allowing more number of cultivation  
299 batches per cycle which is important for commercial live feed producers. After reaching the peak  
300 population day within second week, the population began to decrease from the starting of third week.  
301 Which could be caused by insufficient of space, food availability, sexual transformation, and/or  
302 allelopathic effects [37,10,38,39,40]. Jiun [41] reported that, *M. macrocopa* has a higher density  
303 adaptation in a captive culture environment compared to *M. micrura*, because high stocking density  
304 may possibly lead to a population collapse. According to Jana and Pal [36], the growth performance of  
305 *M. macrocopa* was limited at the density of 4 ind. and 20 ind./ 50ml. Results of this study showed that,  
306 *M. macrocopa* had a better adaptation in food waste treatment at the density up to 75 individual per  
307 50mL which is more higher than that previous study.

308 Muller et al., [26] revealed that, population growth and reproduction of the species depends on the  
309 quantity of reserve lipids (14:0, 16:0 and 18:0). *M. macrocopa* cultured in this study comprised of 33%  
310 of saturated fatty acids in each treatment. *M. macrocopa* exhibited a fatty acid profile of poly  
311 unsaturated fatty acids (18:1n-9, 18:2n-6 and 18:3n-3) constituting 45.1%, 37.6%, 36.4% and 34.3%  
312 when grown in cow manure, food waste, pig manure and chicken manure treatment respectively. It  
313 could be that a decrease in PUFA, reduces the capacity of animals to withstand in the environment as  
314 reflected in adverse changes in demography [42]. Comparatively higher percentage of EPA and DHA  
315 was found in *M. macrocopa* grown in food waste treatment and the levels of EPA (1.8%) were  
316 comparable with those in rotifers or *Artemia* that were fed algal diets in another study [10]. This result  
317 suggests that *M. macrocopa*, when culture in food waste treatment, has the potential to be a suitable  
318 diet containing essential n-3 HUFAs for fish larvae.

319 Our study demonstrated that *M. macrocopa* can be cultured using animal manure and food waste.  
320 This is also in agreement with Nandini [33] and Golder [43]. However, Siebe C [44] reported that,  
321 *Moina* sp. cultivation using domestic wastes as a food source posing a high risk of pathogen  
322 contamination or toxicant pollution. But, in case of food waste this contamination possibilities is  
323 comparatively low. Studies has shown that food waste can replace part of the fish meal used in fish  
324 feeds to produce quality fish and no health risk was observed in the health risk assessment test [45].

## 325 **5. CONCLUSION**

326 In conclusion, the results of this experiment suggest that *M. macrocopa* could be cultivated using  
327 1000 ppm concentration of different animal manure and food waste. Results indicate that, the food  
328 waste appeared to be more effective compared to all other treatments for successful mass culture of *M.*  
329 *macrocopa* to high density and higher percentage of n-3 HUFA , which may serve as effective,  
330 inexpensive and sustainable food sources for *M. macrocopa* cultivation.

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### 334 **COMPETING INTERESTS**

335 The authors declare that they have no competing interests.

336

### 337 **ETHICS APPROVAL**

338 All experimental protocols followed the guidelines of the Institutional animal Care and Use Committee  
339 of the Gyeongsang National University.

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UNDER PEER REVIEW