

1 **Mycoflora, Proximate Composition and Mineral Analysis** 2 **during the Storage of Smoked Dried Crayfish (*Penaeus*** 3 ***natialis* - Shrimps)**

6 **Abstract**

7 This study was carried out to assess the changes in proximate composition, mineral content and mycoflora
8 associated with smoked dried crayfish *Penaeus natialis* (shrimps) stored for twenty weeks. Smoked dried
9 crayfish *Penaeus natialis* (shrimps) were purchased at Igbokoda, Ilaje Local Government Market, Ondo State,
10 Nigeria. They were studied under storage for twenty weeks (6 months) and the proximate, mineral and
11 mycofloral analyses were carried out at four weeks interval. The mycoflora were isolated using direct plating
12 and dilution methods on Potato Dextrose Agar (PDA), Saboraud Dextrose Agar (SDA) and Malt Extract Agar
13 (MEA) and identified using their cultural and morphological features with reference to standard procedures
14 accordingly. The fungi isolated using direct plating methods and dilution methods were *Aspergillus niger*,
15 *Aspergillus flavus*, *Aspergillus fumigates*, *Rhizopus* sp., *Phytophthora siskiyouensis*, *Penicillium* sp. and *Mucor*
16 sp. The proximate analysis result showed a decrease in Ash, fat, and crude fibre content while moisture, crude
17 protein and carbohydrate content increased respectively during the twenty four weeks storage. The mineral
18 analysis result of the smoked dried crayfish *Penaeus natialis* (shrimps) showed a decrease in Sodium,
19 Potassium, Calcium, Magnesium, Zinc, Iron, Copper, Manganese, Cadmium and Phosphorous respectively. This
20 study showed that the smoked dried crayfish *Penaeus natialis* (shrimps) were contaminated by fungi; which is
21 an indication that the market places where these products were displayed for sale were not hygienic coupled
22 with leaving the products in open air without coverage which could allow products contamination with fungal
23 spores leading to fungal spores germination, deterioration and spoilage of products during storage. Good
24 hygiene, constant product checking and sensitization of the products processors, handlers and sellers will
25 minimize exposure to fungal spores' contamination while mitigating deterioration and spoilage of the products
26 during storage.

27 **Keywords**

28 **Storage, Mycoflora, Proximate, Minerals, Shrimps**

31 **1. Introduction**

32 Crayfish *Penaeus natialis* (shrimps) is an important flavour ingredient in many Nigerian local preparations.
33 Crayfish are eaten worldwide like other edible crustaceans, only a small portion of the body of a crayfish is
34 eaten in most prepared dishes, such as soups, bisques, only the tail portion is served [1]. Crayfish processing has

35 become a large part of the crawfish industry. Crawfish processing is a modern industry that produces a high
36 quality product available for consumption world-wide [2,3].

37

38 Preservation of crayfish *Penaeus natialis* (shrimps) is very important because it is easily susceptible to
39 deterioration immediately after harvest and to prevent economic losses. The development of machinery that
40 could be employed for effective handling, harvesting, processing and storage of sea foods such as fish and
41 crayfish cannot be over-emphasized especially when aquaculture is growing fast in Nigeria [4]. The use of
42 smoke in local fish preservation was reported by Eyo [5] and the implication of poor postharvest handling of
43 crayfish has also been reported Kumolu-Johnson *et al* [6]. Smoke drying is done to partially cook, remove
44 water, obtain brown colour, improve organoleptic flavor and control microbial and enzymatic actions that may
45 cause spoilage. Preservation effects of smoke derived from the antioxidant and antimicrobial properties of its
46 phenolic compound have been reported by Shehu *et al* [7] and Abou-zaid and Mohammed [8]. In local markets,
47 crayfish is retailed open as small heaps on tables to attract consumers and information on duration of
48 effectiveness of smoke drying on crayfish quality is scarce. The essence of processing is to preserve and stop
49 microbial deterioration action on food and to retain the quality of the food [9]. However, there is little or no
50 adequate information on the effectiveness of smoke drying on crayfish quality; hence this study is aimed at
51 studying the changes in proximate composition, mineral content and mycoflora associated with smoked dried
52 crayfish *Penaeus natialis* (shrimps) during twenty four weeks storage.

53

54 **2. Materials and Methods**

55 **2.1. Collection of Samples**

56 Samples of Crayfish namely *Penaeus natialis* (shrimps) were randomly purchased at Igbokoda, Ilaje Local
57 Government Market, Ondo State, Nigeria. The dried crayfish *Penaeus natialis* (shrimps) samples were clearly-
58 labeled, stored at room temperature in a sterile airtight container, and kept in a well-ventilated laboratory for a
59 period of twenty four weeks (6 months) under investigation.

60 **2.2. Mycoflora Isolation from the Stored Smoked Dried Crayfish** 61 ***Penaeus natialis* (Shrimps)**

62 The mycoflora associated with smoked dried crayfish *Penaeus natialis* (shrimps) during storage were isolated
63 using the methods described below:

64 **2.2.1. Direct plating method**

65 Visible mouldy sundried crayfish *Penaeus natialis* (shrimps) were examined and randomly selected from the
66 stored samples for mycofloral isolation using the method described by Amusa [10]. The sample surfaces were
67 sterilized with ethanol and washed in sterile distilled water. The sterilized samples were aseptically placed on
68 Potato Dextrose Agar plates with sterilized spatula and incubated at 28°C for 5days. The hyphae tips of each
69 fungal growth were successively sub-cultured on freshly prepared Potato Dextrose Agar plates until pure
70 colonies were obtained [11]. The cultures were examined microscopically to assess the fungi present.

71 **2.2.2. Dilution plate method**

72 The dilution plate method was done by placing 1g of smoked dried crayfish *Penaeus natialis* (shrimps) in sterile
73 distilled water and shaken thoroughly. One ml each of the standardized sample was pipette into 9ml of sterile
74 distilled water in test tube; and serially diluted in series of test tubes containing sterile distilled water. One ml
75 each of aliquots of 10^{-2} and 10^{-3} was introduced into molten Potato Dextrose Agar (PDA) plates in duplicates for
76 each isolate and incubated at 28°C for 5days. The fungal growths were observed every 24hours for the fruiting
77 bodies and hyphae tips of each fungus were sub-cultured successively until pure cultures were obtained [11].
78 The cultures were examined microscopically to assess the fungi present.

79 **2.3. Identification of Mycoflora**

80 The mycoflora isolated from the stored smoked dried crayfish *Penaeus natialis* (shrimps) were identified by
81 their gross cultural and morphological features. The mycoflora were examined under bright day light for colour
82 of the culture and further examination were carried out using Needle mount preparation method as described by
83 Tuite [12], Crowley *et al.* [13] and Egbebi *et al.* [11] and Slide culture technique method as described by
84 Fagbohun *et al.* [14].

85 **2.4. Nutrient Analysis**

86 **2.4.1. Proximate Analysis**

87 Samples of the stored smoked dried crayfish *Penaeus natialis* (shrimps) were analysed for the for ash, crude
88 fiber, moisture and fat contents according to the methods described by Pearson [15] and A.O.A.C. [16]. The
89 nitrogen was determined by Micro-Kjeldahl method as described by Pearson [15] and the percentage nitrogen
90 was converted to crude protein by multiplying 6.25. The carbohydrate content was estimated by the difference
91 in value obtained when all the chemical composition values were subtracted from 100%. All determinations
92 were in triplicates and values of each constituent were expressed in percentage.

93 **2.4.2. Mineral Analysis**

94 The stored smoked dried crayfish *Penaeus natialis* (shrimps) were analysed for the minerals using the solution
95 obtained by dry ashing the sample at 550°C and dissolving it in 10% HCL (25ml) and 5% lanthanum chloride
96 (2ml), boiling, filtering and making up to standard volume with deionized water. Mn, Cu, Co, Zn, Fe, Mg, Na,
97 and Ca were determined with a Buck Atomic Absorption Spectrometer (Buck Scientific, Model 200A/200, Inc.
98 East Norwalk, Connecticut, U.S.A). Sodium was measured with a Corning 405 flame photometer (Corning
99 Halstead, Essex, UK, Model 405) (AOAC) [16]. The detection limits had precisely been determined using the
100 methods of Varian Techtron [17] as Mn 0.01, Cu 0.005, Co 0.05, Zn 0.005, Fe 0.02, Mg 0.002, Ca 0.04, Na
101 0.001, ppm (all for aqueous solutions). The optimum analytical range was 0.5 to 10 absorbance units with
102 coefficient of variation of 0.05-0.40%. Phosphovanadomolybdate method using a spectronic 20 colorimeter
103 (Galenkamp, London, UK) (AOAC) [16]. All chemicals were BDH analytical grade

104

105 **3.0. Results and Discussion**

106 The proximate content of smoked dried crayfish *Penaeus natialis* (shrimps) during twenty four weeks storage is
107 shown in Table 1 below

108

109 **Table 1: Results of proximate analysis of smoked dried crayfish *Penaeus natialis* (shrimps) during 24 weeks storage (g/100g)**

Weeks of Storage	Ash	MC	CP	FAT	CF	CHO
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Fresh	13.45±0.07 ^E	6.40±0.14 ^C	68.46±0.79 ^A	5.10±0.28 ^C	ND	6.40±0.57 ^A
4	13.38±0.04 ^E	6.39±0.02 ^{BC}	68.19±0.03 ^A	5.40±0.02 ^C	ND	6.66±0.04 ^A
8	13.42±0.01 ^E	6.36±0.01 ^B	68.26±0.01 ^A	5.36±0.01 ^C	ND	6.62±0.02 ^A
12	13.24±0.02 ^D	6.42±0.02 ^{BC}	68.36±0.04 ^A	5.32±0.01 ^C	ND	6.63±0.05 ^A
16	12.25±0.01 ^C	6.52±0.02 ^C	68.13±0.04 ^A	4.26±0.02 ^B	ND	8.86±0.01 ^B
20	11.68±0.01 ^B	6.36±0.01 ^B	68.21±0.01 ^A	3.99±0.03 ^{AB}	ND	9.79±0.00 ^C
24	10.29±0.01 ^A	6.21±0.01 ^A	68.07±0.01 ^A	3.89±0.16 ^{AB}	ND	11.53±0.16 ^D

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111 MC: Moisture content, CP: Crude protein, CF: Crude Fiber, CHO: Carbohydrate, ND: Not Detected. Means for each treatment with the
 112 same alphabet in each row are not significantly different at 5% level of significance ($p < 0.05$), while different alphabets in each row are
 113 significantly different at 5% level.

114 The mineral content of smoked dried crayfish *Penaeus natialis* (shrimps) during twenty four weeks storage is
 115 shown in Table 2.

116

117 **Table 2: Results of mineral analysis of smoked dried crayfish *Penaeus natialis* (shrimps) during twenty four weeks storage**
 118 **(mg/100g)**

Weeks of storage	Na	K	Ca	Mg	Zn	Fe	CU	Mn	CD	P
Fresh	58.90±0.14 ^F	66.30±0.14 ^A	86.65±0.01 ^A	49.90±0.14 ^E	0.63±0.01 ^D	6.33±0.11 ^C	0.28±0.04 ^E	0.99±0.01 ^E	0.23±0.04 ^{EF}	106.10±7.35 ^{AB}
4	57.89±0.03 ^D	65.40±0.04 ^A	76.90±14.15 ^A	49.29±0.08 ^D	0.62±0.01 ^C	6.62±0.01 ^E	0.24±0.01 ^{DE}	0.91±0.01 ^D	0.24±0.02 ^F	113.63±3.55 ^{BC}
8	58.12±0.02 ^E	65.49±0.02 ^A	87.03±0.03 ^A	49.43±0.04 ^D	0.68±0.02 ^D	6.55±0.01 ^b	0.19±0.01 ^C	1.12±0.02 ^F	0.19±0.02 ^{DE}	116.22±0.02 ^C
12	57.97±0.05 ^D	60.46±0.09 ^A	85.67±0.03 ^A	49.35±0.02 ^D	0.61±0.01 ^C	6.49±0.01 ^D	0.15±0.02 ^C	0.99±0.02 ^E	0.16±0.01 ^{CD}	115.14±0.02 ^C
16	56.88±0.04 ^B	64.81±0.38 ^A	83.63±0.01 ^A	48.78±0.33 ^C	0.54±0.01 ^B	5.68±0.01 ^B	0.11±0.01 ^A	0.59±0.01 ^C	0.12±0.01 ^{BC}	112.68±0.01 ^{BC}
20	55.65±0.21 ^A	60.39±0.02 ^A	81.52±0.02 ^A	47.39±0.01 ^B	0.48±0.04 ^A	5.52±0.01 ^A	0.07±0.01 ^A	0.50±0.00 ^A	0.08±0.01 ^{AB}	100.98±0.15 ^A
24	56.56±0.02 ^B	60.39±0.01 ^A	81.29±0.03 ^A	45.80±0.01 ^A	0.48±0.04 ^A	5.55±0.01 ^A	0.09±0.08 ^A	0.54±0.01 ^B	0.06±0.01 ^A	100.89±0.01 ^A

119

120 Na: Sodium, K: Potassium, Ca: Calcium, Mg: Magnesium, Zn: Zinc, Fe: Iron, Cu: Copper, Mn: Manganese, CD: Cadmium, P: Phosphorus.
 121 Means for each treatment with the same alphabet in each row are not significantly different at 5% level of significance ($p < 0.05$), while
 122 different alphabets in each row are significantly different at 5% level.

123

124 The mycoflora isolated from smoked dried crayfish *Penaeus natialis* (shrimps) during twenty four weeks
 125 storage is shown in Table 3.

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135 **Table 3: Mycoflora isolated from smoked dried crayfish *Penaeus natialis* (shrimps) during twenty four weeks storage (mg/100g)**

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Mycoflora	Week of storage													
	0	0	4	4	8	8	12	12	16	16	20	20	24	24
<i>Aspergillus niger</i>	1	2	1	2	1	2	1	2	1	2	1	2	1	2
	+	+	+	+	+	+	+	+	-	-	-	-	-	-

<i>Aspergillus fumigatus</i>	-	-	+	+	+	+	-	-	-	-	-	-	-
<i>Aspergillus flavus</i>	-	-	-	-	-	-	+	+	+	+	-	-	-
<i>Rhizopus</i> sp.	+	+	+	+	+	+	-	-	-	-	-	-	-
<i>Phytophthora siskiyouensis</i>	-	-	-	-	-	-	+	+	+	+	+	-	-
<i>Penicillium</i> sp.	-	-	-	-	-	-	-	-	+	+	+	-	-
<i>Mucor</i> sp.	-	-	-	-	-	-	-	-	-	-	-	+	+

137 1: Dilution method, 2: Direct plating method, (+): isolated, (-): not isolated

138 3.1. Proximate Analysis

139 There was a significant decrease in ash content (13.45-10.29 g/100g), fat (5.10-3.89 g/100g), moisture content
140 (6.40-6.21 g/100g), crude protein (68.46-68.07 g/100g) and there was an increase in carbohydrate (6.40-11.53
141 g/100g) while crude fibre was not detected during the twentieth four weeks storage of smoked dried crayfish
142 *Penaeus natialis* (shrimps) as shown in Table 1. This result is in agreement with the findings of Girard [18] who
143 reported a significant reduction in ash content of cattle hide from (1.67-0.83) mg/100g after storage for months.
144 Decrease in ash content indicates loss of nutrients as the storage progressed. Ash content in food contributes to
145 the residue remaining after all the moisture has been removed as well as the organic material (fat, protein,
146 carbohydrates, vitamins, organic acid etc.) have been incinerated at a temperature of about 500°C. Ash content
147 is generally taken to be a measure of the mineral content of the original food [19]. However, this result
148 contradicts that of Oladejo and Adebayo-Tayo [20] who reported an increase in crude protein (21.68-54.16)
149 mg/100g of “Banda” dried meat during storage and Rodolfo *et al.* [21] who found out that fungi increase the
150 protein content of the samples on which they grow. This result is also different from the findings of Lawal *et al.*
151 [22] who reported a decrease in the proximate content such as carbohydrate content of sundried coco yam chips
152 during storage. Crude fiber was not detected in stored smoked dried crayfish (shrimps) which is similar to that
153 of Eleazu [23] who reported that crude fiber was not found in the 10%, 30%, or 40% NRCRI cassava bread
154 samples or in the 100% wheat bread. There was a reduction in the moisture content from 6.40 – 6.21 (g/100g).
155 This result is in agreement with the work of Ajai *et al* [24] who reported a decrease in the moisture contents of
156 milk samples after storage from (8.32-7.51) g/100g. It is known that products that have low fat values normally
157 have high moisture contents. Decrease in water content in this study could be attributed to the fact that infecting
158 fungus utilizes the moisture content for its survival and growth. The shelf life of any product is influenced by
159 the amount of water present in it [25]. Moisture content is a widely used parameter in the processing and testing
160 of food. It is an index of water activity of many foods and determines the shelf life or keeping quality of the
161 food. The observed value in this study implies that smoked dried crayfish (shrimp) will have a long shelf life
162 because of the low moisture content.

163

164 3.2. Mineral Analysis

165 The summary of the mineral composition of smoked dried crayfish *penaeus natialis* (shrimps) during twenty
166 four weeks storage showed a decrease in Sodium (58.90-56.56mg/100g), Potassium (66.30-60.39mg/100g),
167 Calcium (86.65-81.29mg/100g), Magnesium (49.90-45.80mg/100g), Zinc (0.63-0.48mg/100g), Iron (6.33-
168 5.55mg/100g), Copper (0.28-0.09mg/100g), Manganese (0.99-0.54mg/100g), cadmium (0.23-0.06mg/100g) and
169 Phosphorous (106.10-100.89mg/100g) as shown in Table 2. This result supports the findings of Oladejo and
170 Adebayo-Tayo [20] who reported a reduction in Sodium (0.35-1.55) mg/100g in “Banda” dried meat during

171 storage. This result is in contrast to the work of Hassan *et al.*(2005), who reported an increase in sodium content
172 of *Vernonia amygdalina* leaf protein concentrates of (57.5±0.34 mg/100g). High sodium content in food is of
173 great concern for health because of its implication in high blood pressure [26]. The result of this study indicated
174 that eating of smoke dried crayfish (shrimp) could not lead to high in blood pressure. Low sodium content is
175 beneficial in the treatment of hypertension and renal diseases [27]. The manganese content of stored smoked
176 dried crayfish (shrimp) observed in this study significantly decreased from 0.99-0.54 mg/100g. The result of this
177 work is different from that of Mensah, [28], who reported a significant increase in Mn from (2.7 - 20.1) mg/kg
178 for meat hides. Thus, certain trace elements such as copper, iron and manganese constitute essential part of any
179 balanced diet. The RDA for manganese varies between 2.7mg/kg to 3.1mg/kg (RDA, 2001). However, the
180 manganese content observed in this study was low when compared to the RDA value for manganese.

181

182 **3.3. Mycoflora of smoked dried crayfish *Penaeus natialis* (Shrimps)**

183 The mycofloral associated with smoked dried crayfish *Penaeus monodon* (shrimps) during twenty four weeks
184 storage were *Aspergillus niger*, *Aspergillus fumigates*, *Aspergillus flavus*, *Rhizopus* sp., *Phytophthora*
185 *siskiyouensis*, *Penicillium* sp., and *Mucor* sp. This result supports that of Adebayo-Tayo *et al.* [29] who reported
186 the isolation of *Aspergillus flavus*, *Aspergillus tereus*, *Aspergillus fumigatus*, *Abisidia* sp., *Rhizopus* sp.,
187 *Aspergillus niger*, *Mucor* sp., *Cladosporium* sp., *Penicillium viridatus*, *Candida tropicalis* and *Fusarium*
188 *moniliformis* from selected smoked fish from different markets sites in Uyo, Akwa Ibom state. The implication
189 of mycofloral in these products could be attributed to the ever increasing demand for smoked dried crayfish
190 *Penaeus monodon* (shrimps) and in the quest of the retailers to meet this need the fish are overloaded on the
191 smoking kiln during processing; as a result they are exposed to a reduced intensity of heat for short period of
192 time. This leads to improper processing and vulnerability of the fish to fungal contamination [30]. The market
193 place where the smoked dried crayfish products are displayed for sale most times are not clean or hygienic, such
194 as in open trays without coverage Hassan *et al* [26] Fungi found in stored food are divided into two groups
195 namely the field fungi and the storage fungi. Most at times it is difficult to distinguish between the two as fungal
196 growth may start both in the field and during storage. Species of *Aspergillus*, *Rhizopus* and *Penicillium* have
197 been reported as storage fungi which infect crops on the field and may persist and proliferate in storage resulting
198 in increased fungal and mycotoxin contamination with increased duration of storage [31].

199

200 **4.0. Conclusion**

201 This current study indicated that the stored smoke dried crayfish (shrimps) were contaminated with fungal
202 species with significant loss of nutrients during the twenty four weeks storage. Therefore, special attention
203 should be paid to the microbial investigation to minimize the threats posed to public health. The crayfish
204 (shrimps) must be properly dried to reduce the moisture content before packaging to prevent fungal invasion and
205 enhance the good keeping and storage quality. Good sanitary practices including good storage practices must be
206 followed and microbiological standards must be adhered to by checking production procedures and handling
207 until the stored smoke dried crayfish (shrimps) reach the consumer's table. Stored smoke dried crayfish
208 (shrimps) sellers should be sensitized on the importance of good hygienic practices, good housekeeping and
209 proper storage conditions to prevent deterioration of their product

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