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2 **INVIVO STUDY ON ORGANOMETALLIC COMPOUNDS AS**  
3 **ANTICANCER AGENTS**

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**ABSTRACT:**

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This study aims to study the invivo anticancer effect of the synthesized copper complexes of 2,3- 6  
dihydroxy benzaldehyde thiosemicarbazone (3a,b), followed by evaluating their antioxidant activity. 7  
*Materials and methods:* A total number of 80 adult female swiss albino mice weighing 20-25 gm were 8  
divided into 8 groups (10 mice /each group). The acute toxicity was estimated by intraperitoneal 9  
injection of the compounds (3a, b). *Results:* We found that, 5 mg /kg and 10 mg /kg were considered 10  
to be the most effective dose of compounds 3a & 3b; respectively. The mean volume of EAC in the 11  
positive control group was found to be 4.2 ±0.5 (mL), this value was significantly decreased by 100%, 12  
(p<0.001) for 3a & 3b treated groups; respectively. 13

**Keywords:** anticancer, copper complexes,2,3-dihydroxy benzaldehyde thiosemicarbazone, EAC,swiss 14  
albino mice 15

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## 1. INTRODUCTION:

Cancer is a disease characterized by failure of tissue growth regulation when the genes that regulate cell growth and differentiation are altered. Most cancers have multiple causes, only a small minority of cancer is due to inherited genetic mutations whereas the vast majority is non-hereditary epigenetic mutations that are caused by various agents (environmental factors, physical factors and hormones). Thus, although there are some genetic predispositions in a small fraction of cancers, the major fraction is due to a set of new genetic mutations (called "epigenetic" mutations) [1].

Cancer is a group of diseases involving abnormal cell growth with the potential to invade or spread to other parts of the body. Estimates are that in 2018, 18.1 million new cases of cancer and 9.6 million deaths occur globally [2]. Cancer is considered one of the major causes of mortality in the world. The recent advances in science, cancer have not been cured yet. It is estimated that by 2020 there will be 16 million new cancer cases every year [3]. It is, therefore, essential that new therapeutic options are needed for cancer therapy with attention to toxicity and side effects, besides the major treatment modalities including surgery, immunotherapy and radiotherapy [4].

The human genome is composed of deoxyribonucleic acid (DNA), which is the heritable macromolecule that carries the information essential for life. As a chemical, DNA is susceptible to changes that affect its capacity to perform this role. Cells use highly regulated biochemical pathways to replicate DNA, detect if it is modified, and repair modifications as they arise. Many processes

are required to prevent change and to transfer the genome to daughter cells: replication must be accurate, chromosomes must be distributed correctly during cell division, and damage to DNA must be detected and repaired. The fate of a cell, be it healthy or cancerous, is dependent upon the integrity of the genome and its ability to maintain this integrity. By preventing change to DNA, healthy cells ensure their viability and the delivery of a copy of their genetic material to the next generation [5].

Anticancer activity of thiosemicarbazone complexes is mainly attributed to inhibition of RR activity, Topo- II activity and generation of ROS, but there are other possible targets as well which need to be explored. In many cases, in vitro ribonucleotide inhibitors have been found to be poor proliferation inhibitors on whole cells. Another area which needs attention is metal/ion sequestering since thiosemicarbazones are versatile chelators, they sometimes deprive the cell of essential metal ions by forming stable chelates with them. On the other hand, the fact cannot be overruled that metal-ligand complexes are more active than pure ligands. The redox capability of transition metals like copper play an important role in activity enhancement but it can also trigger off Fenton's reactions producing significant amount of  $\text{OH}^\bullet$  radicals that can create hindrance in normal cell functions. It has also been observed that some of the ligands are more active while others are inactive for the same cell lines, hence questioning the simple diffusion hypothesis. Likewise the interaction of one metal with another can also be explored taking synergistic effect into consideration. Not only this, whether the complex acts in unison or metal and ligand act independently inside the body needs a greater depth of understanding by bridging the gap between chemistry and molecular biology [6].

This study aims to evaluate the *in vivo* anticancer effect as well as the antioxidant activity of the synthesized of copper complexes of 2,3-dihydroxy benzaldehyde thiosemicarbazone (3a,b).

## 2. MATERIALS & METHODS:

### 2.1 Materials:

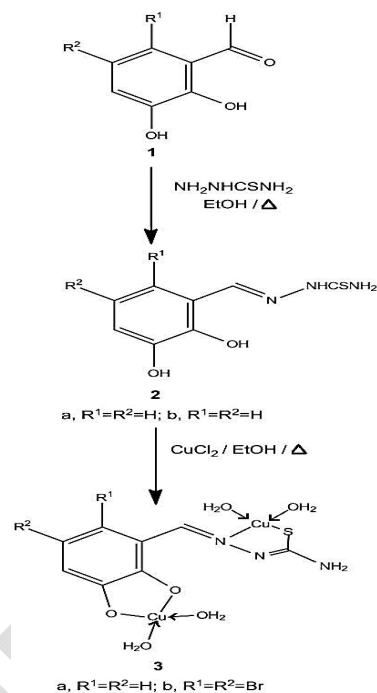
Chemicals for synthesis of copper complexes of 2,3-dihydroxybenzaldehyde thiosemicarbazones (2a,b) : 2,3-dihydroxybenzaldehyde; 5,6-dibromo-2,3-dihydroxybenzaldehyde; thiosemicarbazide; Copper chloride; Ammonium hydroxide (10%) and ethanol.

Ehrlich ascites carcinoma (EAC):

EAC cells were initially supplied from the National Cancer Institute, Cairo, Egypt (only for the first transplantation), and maintained in female Swiss albino mice through serial intraperitoneal (I.P.) injection at 8 or 10 day intervals in our laboratory in a liquid form.

### 2.3 Methods:

2,3-dihydroxybenzaldehyde thiosemicarbazones derivatives (2a, b) were obtained via the condensation of aromatic aldehydes (namely, 2,3-dihydroxybenzaldehyde & 5,6-dibromo-2,3-dihydroxybenzaldehyde) with thiosemicarbazide in ethanol under reflux. The copper complexes of 2,3-dihydroxybenzaldehyde thiosemicarbazones derivatives (3a, b) were prepared from the reaction of thiosemicarbazone derivatives (2a,b) with two mole of copper chloride in ethanol under reflux (scheme 1).



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- Determination median lethal dose (LD 50) of synthetic compounds: Approximate LD 50 of synthetic compounds were determined according to published method [7].

- Determination response curve: Dose response curve was determined according to published method [8]. Studies carried out for determination of the most effective dose.

- Viability and Counting of EAC cells: The viability of EAC cells was determined by the trypan blue exclusion method [9].

### 2.2 Groups of the study:

Adult female Swiss albino mice weigh (20-25 g) purchased from breeding unit of the Egyptian Stock Holding Company for Biological Products of Vaccines, Sera & Drugs (VACSERA) were used throughout this study. The animals were housed in steel mesh cages (animal house, faculty of Science, Zagazig University) and maintained on a commercial pellet diet and water for one week before starting

the experiment as an acclimatization period. A total number of 80 adult female swiss albino mice weighing 20-25 gm were divided randomly into 8 groups (10 mice /each group) as following:

**Group (1):** Negative Control: This group received sterile saline solution (0.9 % NaCl) day after day for 9 days.

**Group (2):** Positive Control: This group received Ehrlich ascites carcinoma (EAC), ( $2.5 \times 10^6$  cells/ 0.3 ml/mouse) by (I.P) injection once at the first day.

**Group (3):** Drug group I: This group were injected I.P. with compound 3a (5 mg/Kg) at 1, 3, 5, 7, 9 days for 10 days (day after day).

**Group (4):** Preventive group I: (EAC + compound 3a): This group were injected I.P. with compound 3a (5 mg/Kg) in the day before EAC injection ( $2.5 \times 10^6$  cells/mouse), followed by I.P. injection of compound 3a at 3, 5, 7, 9 days of EAC injection for 10 days (day after day).

**Group (5):** Therapeutic group I: (EAC + compound 3a): This group were injected I.P. with compound 3a (5 mg/Kg) in the day after EAC injection ( $2.5 \times 10^6$  cells/mouse), followed by I.P. injection of compound 3a at 3, 5, 7, 9 days of EAC injection for 10 days (day after day).

**Group (6):** Drug group II: This group were injected I.P. with compound 3b (10 mg/Kg) at 1, 3, 5, 7, 9 days for 10 days (day after day).

**Group (7):** Preventive Group II: (EAC + compound 3b): This were injected I.P. with compound 3b (10 mg/Kg) in the day before EAC injection ( $2.5 \times 10^6$  cells/mouse), followed by I.P. injection of compound 3b at 3, 5, 7, 9 days of EAC injection for 10 days (day after day).

**Group (8):** Therapeutic Group II: (EAC + compound 3b): This group were injected

198 I.P. with compound 3b (10 mg/Kg) in the 242  
 199 day after EAC injection ( $2.5 \times 10^6$  243  
 200 cells/mouse), followed by I.P. injection 244  
 201 of compound 3b at 3, 5, 7, 9 days of EAC 245  
 202 injection for 10 days (day after day). 246  
 203 - ~~247~~ , EAC and tissue sampling:  
 204 ~~248~~ At the end of the experiment, the blood  
 205 ~~249~~ samples were collected from the retro-  
 206 ~~250~~ peritoneal venous plexus under light ether  
 207 ~~251~~ anesthesia divided to 2 parts to obtain  
 208 ~~252~~ serum and plasma. Serum was prepared by  
 209 ~~253~~ centrifuging blood at 3000 r.p.m for 10  
 210 ~~254~~ minutes. Serum samples were aliquoted  
 211 ~~255~~ and stored at  $-20^\circ\text{C}$  until biochemical  
 212 ~~256~~ analysis [10].  
 213 - ~~257~~ Oxidant assays:  
 214 ~~258~~ **Plasma malondialdehyde: (MDA)** was  
 215 ~~259~~ determined by using Biodiagnostic kit  
 216 ~~260~~ (Biodiagnostic company, Dokki, Giza,  
 217 ~~261~~ Egypt), according to the published method  
 218 ~~262~~ [10].  
 219 ~~263~~ Determination of **catalase enzyme**  
 220 ~~264~~ **activity (CAT):** was measured in plasma  
 221 ~~265~~ and tissues. Catalase reacts with a known  
 222 ~~266~~ quantity of  $\text{H}_2\text{O}_2$ . The reaction is stopped  
 223 ~~267~~ exactly one minute with catalase  
 224 ~~268~~ inhibitor. Catalase converts  $\text{H}_2\text{O}_2$  to  $\text{H}_2\text{O}$   
 225 ~~269~~ and  $\text{O}_2$ . According to published method  
 226 ~~270~~ [10].  
 227 ~~271~~ Determination of **glutathione reductase**  
 228 ~~272~~ **activity:** This assay is based on the  
 229 ~~273~~ oxidation of NADPH to  $\text{NADP}^+$  catalyzed by  
 230 ~~274~~ limiting concentration of glutathione  
 231 ~~275~~ reductase. One GR activity unit is defined  
 232 ~~276~~ as the amount of enzyme catalyzing the  
 233 ~~277~~ reduction of one micromole of GSSG per  
 234 ~~278~~ minute at pH 7.6 and  $25^\circ\text{C}$ . One molecule  
 235 ~~279~~ of NADPH is consumed for each molecule  
 236 ~~280~~ of GSSG reduced. Therefore, the reduction  
 237 ~~281~~ of GSSG is determined indirectly by the  
 238 ~~282~~ measurement of the consumption of  
 239 ~~283~~ NADPH, as demonstrated by a decrease in  
 240 ~~284~~ absorbance at 340 nm ( $A_{340}$ ) as a function  
 241 ~~285~~ of time.

## 2.2.3 Statistical Analysis:

All statistical analyses were done by a statistical for social science package "SPSS" 20.0 for Microsoft Windows, SPSS Inc and considered statistically significant at a two-tailed  $P < 0.05$ . Numerical data were expressed as mean  $\pm$  SD. The levels of markers were analyzed by ANOVA. The correlations between serum biochemical data in different studied groups were evaluated by Pearson's correlation coefficient, to quantify the relationship between the studied parameters. P value  $< 0.01$  was considered significant [13].

## 3.1 RESULTS:

The most effective doses were found to be "5 mg /kg" and "10 mg /kg" for compounds 3a and 3b; respectively.

The mean volume of EAC in the positive control group was found to be 4.2  $\pm$  0.5 (mL), this value was significantly increased by 100%, ( $p < 0.001$ ) for 3a & 3b treated groups; respectively as no detectable EAC cells were found in the treated groups.

The anti-oxidant effect of compounds (3a, 3b), was evaluated through the stimulation of MDA, CAT and G-reductase activities. The mean values of MDA concentration in EAC cells in positive control group were found, that found to be  $30.66 \pm 5.86$  (nmol/g.tissue). 3a and 3b treated groups showed a significant increase to  $35.79 \pm 6.58$  &  $35.46 \pm 3.27$  (nmol/g.tissue) respectively ( $p < 0.001$ ); compared to the positive control group .

On the other hand, CAT activity in positive control group was found to be  $308.14 \pm 19.66$  (U/g). CAT activity showed a significant increase in 3a treated group to  $323.16 \pm 38.78$  (U/g) ( $p < 0.001$ ); and to  $329.47 \pm 58.78$  in 3b treated group, compared to positive control, ( $p < 0.001$ ). Moreover, the mean value of G- Reductase activity in positive control group was found to be  $637.19 \pm 65.12$  (U/g). Compounds 3a & 3b treated groups showed a significant increase to  $920.1 \pm 246.89$  &  $1442.66 \pm 126.9$  (U/g) respectively; ( $p < 0.001$ )

compared to the positive control group.

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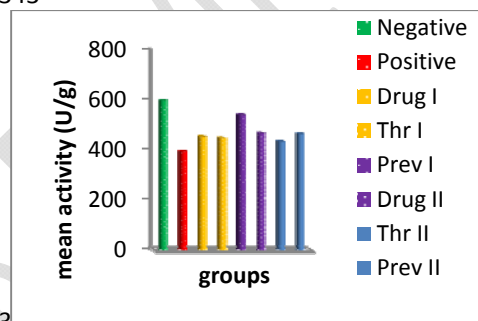
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**Table (1):** the effect of compounds (3a, 3b) on antioxidant catalase activity:

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Group	Mean $\pm$ SD
Negative	603.63 $\pm$ 58.67
Positive	398.14 $\pm$ 19.66
Drug I	458.97 $\pm$ 22.38
Thr I	453.16 $\pm$ 38.78
Prev I	546.58 $\pm$ 18.12
Drug II	472.23 $\pm$ 46.12
Thr II	438.47 $\pm$ 58.78
Prev II	469.82 $\pm$ 62.04

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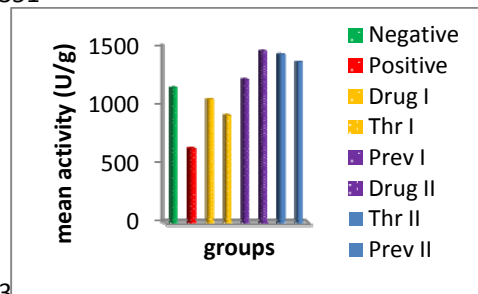
**Figure (1):** the effect of compounds (3a, 3b) on antioxidant catalase activity.

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**Table (2):** the effect of compounds (3a, 3b) on antioxidant G.reductase activity :

Group	Mean $\pm$ SD
Negative	1158.63 $\pm$ 6
Positive	637.19 $\pm$ 65.12
Drug I	1059.15 $\pm$ 132.9
Thr I	920.1 $\pm$ 246.88
Prev I	1229.3 $\pm$ 143.46
Drug II	1472.12 $\pm$ 140.01
Thr II	1442.66 $\pm$ 126.9
Prev II	1375.66 $\pm$ 227.73

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**Figure (2):** the effect of compounds (3a, 3b)  
effect on antioxidant G.reductase activity.  
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#### 4. DISCUSSION:

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Most cancer cells divide more often than  
normal cells and the process of cell division  
can be targeted to treat cancer patients. The  
aim of targeting cell proliferation is to arrest  
the cell cycle and/or cause cancer cell death  
using cytotoxic compounds (chemotherapy)  
or ionising radiation (radiation therapy). DNA  
is one of the main targets of these therapies  
because DNA replication is an essential phase  
of the cell cycle. Many of the cytotoxic agents  
commonly used to treat cancer patients  
cause high levels of DNA damage, that initiate  
cell cycle checkpoints, leading to cell cycle  
arrest and/or cell death [14]

The synthesis of new organometallic  
compounds and the development of  
combination therapies containing  
organometallic components have shown  
significant progress in utilization of transition  
metal complexes as anticancer agents [15]

Thiosemicarbazones have emerged as  
ligands of great biological activity. The ability  
of thiosemicarbazones to chelate metal ions  
has now been recognized as a major factor in  
their antiproliferative effects [16]

Coordination to copper increased the  
cytotoxic potential considerably when  
compared to that of free ligand. It is well-  
known that copper is an essential  
micronutrient and has important biological  
functions, such as cellular trafficking, redox  
regulation and angiogenesis modulation etc  
[17].

Only a limited number of in vivo studies  
have been done which indicate that some  
thiosemicarbazones show potential as  
chemotherapeutic agents [5].

In the present study, we aimed to  
evaluate the anti-tumor and antioxidant  
properties of recently developed synthetic

copper complexes of 2,3-dihydroxy  
benzaldehyde thiosemi carbazone (3a,b), as  
anticancer agents. 403

The acute toxicity was estimated by  
intraperitoneal injection of the compounds  
(3a, b) to assess the dose response curve. We  
found that, 5 mg /kg and 10 mg /kg were  
considered to be the most effective dose of  
compounds 3a & 3b; respectively

The mean volume of EAC in the  
positive control group was found to be 4.2  
±0.5 (mL), this value was significantly  
decreased by 100%, (p<0.001) for 3a & 3b  
treated groups; respectively. As compared to  
*Sathisha et al., 2010*, Who studied the  
effect of thiosemicarbazide metal complexes  
on Ehrlich ascites carcinoma (EAC), the  
results show that the copper (II) complex  
showed more than 85% reduction in the  
growth of tumor cells. This confirm the in vivo  
antitumor activity against EAC of the studied  
compunds [18].

Lipid peroxidation/oxidation process  
plays a key role in tumor growth  
invasiveness. ROS exhibit multiple  
functions and are involved in tumor  
initiation and progression. MDA, a free  
oxygen radical product formed during  
oxidative degeneration of cancerous  
tissues and as the end product of lipid  
peroxidation, is a biomarker of oxidative  
stress that has been reported to be  
exhibited at higher levels in cancer tissues  
than in non-diseased organs [19].  
Antioxidants with free radical scavenging  
activities may have great relevance in the  
prevention and therapeutics of diseases in  
which oxidants or free radicals are  
implicated such as cancer [20]. Catalase is  
a hemoprotein and it protects cells from  
the accumulation of H<sub>2</sub>O<sub>2</sub> and able to  
prevent the tissue from reactive free  
oxygen and hydroxyl radicals, by  
catalysing the reduction of H<sub>2</sub>O<sub>2</sub> to form  
H<sub>2</sub>O and O<sub>2</sub>. Catalase protects the tissue  
from highly reactive hydroxyl radicals by  
decomposing the hydrogen peroxide. So  
reduced levels of catalase may indicate

the toxic effects on the tissue [21]. Glutathione reductase is a widely occurring enzyme and has been studied from several sources including *Plasmodium falciparum*, and most thoroughly from human erythrocytes and *E. coli*. It is one of a chain of enzymes which serves to maintain glutathione in the reduced form. It catalyzes the NADPH-driven reduction of GSSG (Oxidized glutathione) to GSH (reduced glutathione). GSH helps detoxify reactive oxygen species by donating reducing equivalents to glutathione peroxidase and detoxifies electrophilic xenobiotics with glutathione S-transferase [22].

The anti-oxidant effect of compounds **3a** and **3b** were evaluated in the present study, through the estimation of MDA, CAT, and G. Reductase in EAC cells.

Our results found that, the mean values of MDA concentration in EAC cells in positive control group were found to be  $80.66 \pm 5.86$  (nmol/g.tissue). **3a** and **3b** treated groups showed a significant decrease to  $35.79 \pm 6.58$  &  $35.46 \pm 3.27$  (nmol/g.tissue) respectively ( $p < 0.001$ ) compared to the positive control group.

On the other hand, CAT activity in positive control group was found to be  $398.14 \pm 19.66$  (U/g). CAT activity showed a significantly increase in **3a** treated group to  $453.16 \pm 38.78$  (U/g) ( $p < 0.001$ ); and to  $438.47 \pm 58.78$  in **3b** treated group compared to positive control, ( $p < 0.001$ ). Moreover, the mean value of G-Reductase activity in positive control group was found to be  $637.19 \pm 65.12$  (U/g). Compounds **3a** and **3b** treated groups showed a significant increase to  $920.1 \pm 246.89$  &  $1442.66 \pm 126.9$  (U/g) respectively; ( $p < 0.001$ ) compared to the positive control group. All these findings ensure the anti-oxidant activity of the studied compounds, and in agreement with Thanh and Hoai, (2012) who found that some copper thiosemicarbazone

complex derivatives caused inhibition of lipid peroxidation [23].

## 5. CONCLUSION

The compounds (**3a** & **3b**) revealed significant anticancer activity towards Ehrlich ascites carcinoma (EAC) cells by significant reduction of its volume and the cell count in treated groups; respectively compared to the positive control group. It turned out that they reduced cell viability of cancer cells in a time and concentration dependent manner in vivo studies. The synthesized compounds have potent antioxidant activity.

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