INFLUENCE OF ETHANOLIC EXTRACTS OF SPONDIAS MOMBIN LEAVES ON PITUITARY-GONADAL AXIS OF MALE WISTAR RATS.

Original Research Article

10 **ABSTRACT**

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Background: Spondias mombin is one of the tropical plants used locally to treat various kinds of ailment, its use as an anti-conceptive remedy in our locality had been reported.

Objective: The aim of this study was to establish a dose-dependent or duration effect of ethanolic leaf extract of Spondias mombin on the anterior pituitary cells, testes, epididymides of Wistar rats.

Materials and methods: A total of thirty (30) matured male Wistar rats were randomly divided into five groups (n=5). Group 1 animals served as control and received vehicle (distilled water). Groups 2 and 3 were administered with 250mgkg⁻¹ body weight of extract for 4 and 6 weeks respectively, while groups 4 and 5 received 500mgkg⁻¹ body weight of extract for 4 and 6 weeks also. Animals were anaesthetized with chloroform and sacrificed at the end of administration. Body weight, weights of reproductive organs and vital organs were evaluated. Blood was taken from the animals for haematological and biochemical analysis. The pituitary gland, male reproductive and accessory glands were excised and fixed in 10% formalin for routine histological examination.

Results: The influence of ethanolic extract of Spondias mombin leaves on the pituitary cells and reproductive organs of male Wistar rats given 250 and 500mgkg⁻¹ body weight for 4 and 6 weeks showed loss of cytoplasmic contents and free spaces of pituitary cells, desquamation of seminiferous epithelial cells, degradation of seminiferous tubules and reduction in cells. The epididymis of the test groups showed abundant immature cells and cell debris in their lumen. The accessory glands showed homogenous pinkish fluid, glandular degeneration of the prostate and seminal vesicles with decreased structural integrity. The organ weights of the experimental animals were not significantly affected,

however, a significant (P<0.05) decrease in reproductive organ weights was recorded. Ethanolic extract of Spondias mombin on liver enzymes showed a significant protection against hepatobiliary damage. **Conclusion**: These results suggest that Spondias mombin has a dose dependent and duration deleterious effect on the pituitary and reproductive organs at their cellular levels rather than on the tissue as a whole.

Keywords: Accessory glands, Cells, Degradation, Epithelium, Pituitary, Reproduction

1. INTRODUCTION

Medicinal plants still play major roles in health Worldwide irrespective of the advances recorded in orthodox 19 20 medicine. Interest in drugs derived from higher plants has increased considerably in the past few years with 21 about 20-25% of modern drugs being derived from plants [1-2]. An expression in the biodiversity in number 22 had been carried out with an estimate of 10-100 million species, out of which 2.5-7.5 million to be attributed to 23 plants [3]; 5-105 of these plants have been scientifically evaluated for their therapeutic medicinal properties [4]. 24 Plants have been used and are still in use in most developing Countries and advanced Countries as their main 25 source of health care [5-6]. These plants are assumed to be safe and free from side effects since they are 26 naturally occurring [7-8]. However, studies have shown that medicinal plants may be toxic [9-12]. These adverse effects are however less frequent when used properly in comparison to synthetic medicines [13]. A lot 27 28 of these herbal plants used to treat or cure diseases locally have been found to be anticancer [14-16], anti-29 malarial [17-18], anti-diabetic [19-20] antihypertensive [21-22], antibacterial [23], antimicrobial [24-25], 30 antifertility [26-27], abortifacient [28-30] etc. These claims were first made by rural users which have been 31 authenticated by biological research. Spondias mombin is one of such plants used locally to treat various kinds 32 of ailment which biological research has supported [31-37]. We had earlier reported on scientific findings on 33 Spondias mombin [38-40]. The present study was carried out to further investigate the effect of Spondias 34 mombin on pituitary cells and reproductive organs of male Wistar rats based on duration of administration.

36 2. MATERIAL AND METHODS

38 The harvesting and extraction of plant material had earlier been reported in our previous work [40]. A total of 39 thirty (30) matured male Wistar rats were randomly divided into five groups (n=5). Rats were kept in 40 temperature controlled room of 25 ± 2°C with a 12-hour light/dark cycle under hygienic conditions and had free 41 access to water & rat chow. The animals were acclimatized for seven days before experimental use. Ethics on 42 the use of laboratory animals was applied and care of the animals was in accordance to the International 43 guidelines for animal research. The methodology was approved by the Department of Human Anatomy ethical 44 committee. Group 1 animals served as control and received vehicle (distilled water). Groups 2 and 3 were administered with 250mgkg⁻¹ body weight of extract for 4 and 6 weeks respectively, while groups 4 and 5 45 received 500mgkg⁻¹ body weight of extract for 4 and 6 weeks also. Animals were anaesthetized with 46 47 chloroform and sacrificed at the end of administration. Body weight, weights of reproductive organs and vital 48 organs were evaluated. Blood was taken from the animals for haematological and biochemical analysis. The 49 pituitary gland, male reproductive and accessory glands were excised and fixed in 10% formalin for routine 50 histological examination. The pituitary gland was double stained using the bromine alcian blue-orange fuschin 51 green (Br.AB-OFG) method of Slidders [41] to demonstrate anterior pituitary cells.

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53 3. RESULTS AND DISCUSSION

The mean vital organs weight was not affected by the administration of leaf extract of SpM for 4 and 6 55 weeks (Table 1). However, the reproductive organ weights were significantly (P<0.05) reduced at 6 weeks in 56 animals administered with 250 and 500 mgkg⁻¹, irrespective of a non- significant change in body weights 57 58 across the five groups (Table 2). The identification of the possible harmful effects of chemical and drugs is the 59 analysis of organ weight [42]. In this study, the organ weights and body weights of the experimental groups 60 were not different from that of the control which points to earlier findings of the safety use of SpM extracts [43]. 61 However, reproductive organ weights were affected by extract of SpM which is indicative of the shrunken 62 characteristics observed on histopathological examination of the tissues. The red blood cell counts were increased significantly (P<0.05) in groups 3 and 5 which received 250 and 500 mgkg⁻¹ for 6 weeks. Similarly, 63 values for haemoglobin also significantly increased (P<0.05) in these groups. Values of ALP was reduced in 64

65 groups 3 and 5 (Table 3). Enzymes have been reported to be found in tissues and blood as a result of insult to 66 the cell or from degraded cells [44]. Activities of liver enzymes are determined in serum as indicators of 67 biochemical changes which occur in response to treatment [45]. It had been stated that aminotransferases 68 (ALT and AST) are indicators of hepatotoxicity and hepatocellular damage, while ALP is used in diagnosing hepatobiliary or cholestatic obstruction [46]. ALP is cardinally involved in the transport of metabolites across 69 70 cell membranes, synthesize protein, secretory activities and glycogen metabolism [47]. The significant 71 (P<0.05) decrease observed in ALP activity may imply protection against hepatobiliary damage, since most 72 enzymes measured as indices of drug metabolism are released into bloodstream when cells are damaged or 73 their functions are disrupted.

74 Pituitary cells of control animals were well stained, normal and numerous on histological examination, 75 whereas cells of experimental animals were sparse with loss of cytoplasmic contents. The effect was more in 76 groups treated for 6 weeks having more loss of cytoplasmic contents and free spaces (Figure 1a-e). The testes 77 on histopathological examination showed seminiferous tubules of control possessing epithelia with well-defined 78 Sertoli cells and germ cells at various stages of spermatogenesis. Sertoli cells showed distinct granular 79 cytoplasm and irregular nuclei. Lumen of seminiferous tubules contained matured sperm cells and numerous 80 Leydig cells in the interstitium (Figure 2a). Groups 2-5 animals showed dose and time duration dependent 81 alteration on the testes evidenced by distortion of tubular cells, prominent spaces and severe structural 82 disorganization (Figure 2b-e).

83 Epididymis of experimental groups showed thinness of epididymal epithelial lining compared to control 84 with their lumen showing cell debris, large number of immature cells and degenerated cells (Figure 3a-e). The 85 lumen of the prostate of experimental animals showed pinkish fluid and inflammatory cells (Figure 4a-e) while 86 seminal vesicles of experimental animals exhibited glandular degeneration and increase fibrosis of interstitium 87 (Figure 5a-e). The presence of debris in the lumen of the epididymis may be a reflection of degenerated 88 testicular assault observed in the treated rats. This lesion may probably have been passed to the epididymis. 89 Thus, it is safe to deduce that extract of SpM has defective effect on the germ cells. The observed effect of the 90 extract on the accessory sex gland may also be as a result of its destructive tendency on testicular tissue that 91 led to a decrease in testosterone production [48]. Since decrease in testosterone production has been 92 observed to have negating effect on accessory sex glands [49]. Therefore, it is safe to state that the low

93 testosterone reported in our earlier work [48] may be responsible for the effect of the extract on the accessory

94 sex glands since male accessory sex glands are known to depend on male sex hormone for development and

95 secretory activity [50].

97 Table 1: Weight of vital organs of control and experimental SpM extract treated rats.

Parameters	Groups				
(g)	1	2	3	4	5
Brain	1.85±0.25	1.82±0.05	1.77±0.14	1.73±0.23	1.72±0.28
Heart	0.82±0.05	0.76±0.02	0.72±0.03	0.70±0.06	0.68±0.24
Lungs	1.56±0.30	1.54±0.35	1.48±0.20	1.49±0.30	1.46±0.28
Thyroid	0.060±0.002	0.060±0.002	0.050±0.002	0.048±0.006	0.046±0.006
Kidney	1.80±0.28	1.78±0.22	1.72±0.29	1.70±0.20	1.70±0.24
Liver	8.26±0.80	8.00±0.78	7.95±0.80	7.64±0.75	7.62±0.82
Adrenal	0.180±0.002	0.178±0.002	0.178±0.002	0.176±0.002	0.173±0.002
Spleen	0.62±0.20	0.60±0.27	0.60±0.18	0.58±0.22	0.55±0.21

Values are Mean ± SEM, n=5. Extract had no significant effect on the weights of vital organs.

Table 2: Body and reproductive organ weights in control and treated rats.

Parameters	Groups				
(g)	1	2	3	4	5
Body weight	210±2.42	208.7±3.42	214.0±2.07	204.5±3.57	210.0±1.82
Testis	3.20±0.75	2.75±0.43	1.43±0.36*	2.44±0.23*	1.26±0.15*
Epididymis	2.75±1.36	2.55±1.80	1.89±0.82*	2.24±1.08*	1.54±0.62*
Seminalvesicle	3.75±1.10	2.92±0.34	1.83±0.33*	1.72±0.64*	1.46±0.73*
Prostate	1.92±0.79	1.63±0.64	1.42±0.64*	1.26±0.48*	1.08±0.84*

 Values are Meant \pm SEM, n=5. P<0.05. Extract showed a significant effect on the weights of reproductive organs compared to body weights of animals were no significant effect was recorded.

Table 3: Haematological and biochemical parameters of control and SpM extract treated rats.

Parameters	Groups					
	1	2	3	4	5	
RBC (10⁴/µL)	7.44±0,38	7.93±0.22	8.82±0.12*	8.72±0.10*	8.85±0.23*	
WBC (10 ³ /µL)	19.58±3.50	18.78±1.40	16.64±2.10	19.32±1.96	18.42±2.46	
HB (g/dL)	11.68±0.59	12.20±0.27	15.42±0.21*	13.28±0.26	18.38±1.05*	
PCV (%)	45.78±2.64	49.74±1.78	47.16±0.62	54.30±1.45	47.58±0.94	
ALP (µL)	27.05±1.77	17.70±0.36*	0.66±0.03*	12.96±1.06*	0.19±0.07*	
AST (µL)	373.42±47.45	294.88±17.07	447.01±8.05	218.41±50.03	442.67±14.75	
ALT (µL)	170.76±9.66	154.15±6.31	171.56±6.95	151.78±4.17	171.62±2.14	

TP(mg/L)	0.51±0.13	0.40±0.06	0.64±0.08	0.27±0.03	0.59±0.13
ALB (mg/L)	5.31±0.74	3.90±1.58	1.79±0.34*	2.73±0.93	1.58±0.48*
CRT (µmol/l)	2.06±0.28	2.92±0.59	2.77±0.79*	2.74±0.64	3.95±0.73*
UREA (mg/L)	32.08±0.93	29.72±1.42	16.03±0.71*	34.86±1.87	14.50±3.83*
Values are Mean ± SEM, n=5. P<0.05					

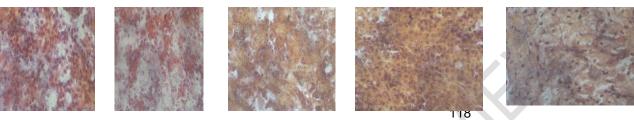
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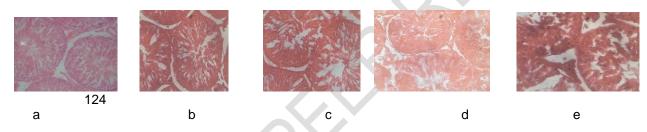


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119 b d а е Fig 1: Photomicrograph of the anterior pituitary of control (a) animals and experimental animals (b-e) Br.AB-120

121 OFG X 100.

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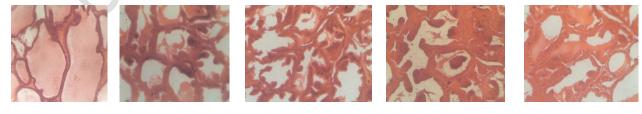


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127 Fig 2: Photomicrograph of the testis of control (a) animals and experimental animals (b-e) H & E X 400. 128 129

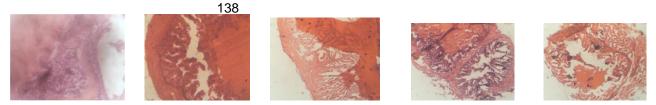


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- 132 133
- b d е С а Fig 3: Photomicrograph of the epididymis of control (a) and experimental animals (b-e) H & E X 400.



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b d а С е Fig 4: Photomicrograph of the prostate of control (a) animals and experimental animals (b-e) H & E X 400. 136



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- 140 141
- 142abcde143Fig 5: Photomicrograph of the seminal vesicle of control (a) animals and experimental animals (b-e) H & E X144400.
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146 **4. CONCLUSION**

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148 In conclusion, we propose that the effect of extract of SpM is dose and duration dependent with its effect
149 localized to the pituitary and male reproductive system which supports its use locally to stall conception. The
150 mechanism through which this is mediated is not known. Further research will be based on the mechanism
151 through which SpM mediate this action.

153154 COMPETING INTERESTS

- 155156 Authors have declared that no competing interests exist.
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159 ETHICAL APPROVAL

- 161 Approval was given by the Faculty of Basic Medical Sciences Committee on animal use and care, Universiity
- 162 of Calabar to carry out this research work following laid down rules and guidelines of the institution in the use
- 163 of medicinal plants and animal models.

164 **REFERENCES**

- Veeresham C. Natural products derived from plants as a source of drugs. *J Adv Pharm Technol Res* 2012; 3(4): 200-207.
- Newman DJ, Cragg GM, Snader KM. The influence of natural products upon drug discovery. *Nat Prod Reports* 2000; **17**: 215-234.
- 170 3. Pimm SL, Russell GJ. Gittlemqn JL, Brooks TM. The future of biodiversity. *Science* 1995; **5** 347-350.
- Verpoorte R. Exploration of nature's chemodiversity: the role of secondary metabolites as lead drugs for drug development. *Drug Dev Today* 1998; **3**: 232-238.
- Kumar S, Kumar R, Khan A. Medicinal plant resources: manifestation and prospects of life-sustaining healthcare system. *Cont J Biol Sci* 2001; **4** (1): 19-29.

- Sathiyaraj K, Sivaraj A, Thirumalai T, SenthilKumar, B. Ethnobotanical study of antifertility medicinal plants used by the local people in Kathiyavadi village, Vellore District, Tamilnadu, India. *Asian Pac J Trop Biomed* 2012; S1285-S1288.
- 178 7. Haq I Safety of medicinal plants. *Pak J Med Res* 2004,**43** (4): 203-210.
- 179 8. Nasri H, Shirzad H. Toxicity and safety of medicinal plants. *J Herb Med Pharmacol* 2013; **2**(2): 21-22.
- Cupp MJ. Toxicology and chemical pharmacology of herbal products. Totowa, NJ: Humaila Press.
 2000
- 182 10. Boullata JI, Nace AM. Safety issues with herbal medicine. *Pharmacotherapy* 2000; **20** (3): 257-269.
- 183
 11. Posadzki P, Watson LK, Ernst E. Adverse effect of herbal medicines: an overview of systematic review. *Clin Med* 2013; **13** (1): 7-12.
- 185
 12. Hussin AHJ. Adverse effects of herbs and drug-herbal interactions. *Malaysian J Pharm* 2001; 1(2):
 39-44.
- 13. Calixto JB. Efficacy, safety, quality control marketing and regulatory guidelines for herbal medicines
 (phytotherapeutic agents). *Brazilian J Med Biol Res* 2000; **33**: 179-189.
- 14. Cragg GM, Newman DJ. Plants as a source of anti-cancer agents. *J Ethnopharm* 2005; **100**(1-2): 72 79.
- 191 15. Soladoye MO, Amusa VA, RAji-Esan SO, Chukuma EC, Ayanbamiji AT. Ethnobotanical survey of anti-192 cancer plants in Ogun State, Nigeria. *Annals Biol Res* 2010; **1**(4): 261-273.
- 193 16. Mohan S, Bustamam A, Ibrahim S, Al-Zubain AS, Aspollah M. Anti-cancerous effect of *Tryphonium* 194 *flagelliforme* on human T4-lymphoblastoid cell line CEM-SS. *J Pharm Toxi*col 2008; **3**(6): 449-456.
- 195 17. Adebayo JO, Krettlu AO. Potential anti-malarials from Nigerian plants: A review. *J Ethnopharmacol* 2011; 133: 289-302.
- 197 18. Okpako LC, Ajsiyebo EO. In vitro and in vivo anti-malarial activities of *Striga hermonthiacca* and *Tapinanthus sessifolius* extracts. *Afr J Med Sci* 2004; **1**: 73-75.
- 19. Ezekwesili CN, Ogbunugafor HA, Ezekwesili-Ofili JO. Anti-diabetic activity of aqueous extracts of *Vites* doniana leaves and *Cinchona calisaya* bark in alloxan induced diabetic rats. Int *J Trop Dis Health* 201 2012; 2(4): 290-300.
- 202 20. Chauhan A, Sharma PK, Srivastava P, Kumar N, Dudhe R. Plants having potential anti-diabetic 203 activity: A review. *Der Pharmacie Lettre* 2010; **2**(3): 369-387.
- 204 21. Patel SS, Verma NK, Ravi V, GAuthaman K, Soni N. Anti-hypertensive effect of an aqueous extract of 205 Passiflora nepalensis wall. Int J Appl Res Nat Prod 2010; **3**(2): 22-27.
- 206 22. Iwalokun BA, Hodonu SA, Nwoke S, Ojo O, Agomo PU. Evaluation of the possible mechanisms of 207 anti-hypertensive activity of *Loranthus micranthus*: African mistletoe. *Biochem Res Int* 2011; **11**: 1-9.
- 208 23. Ekundayo EO, Ekekwe JN. Antibacterial activity of leaf extracts of *Jatropha curcas* and *Euphorbia* 209 *heterophylla*. *Afr J Microbiol Res* 2013; **7**(44): 5097-5100.
- 24. Oliveira AA, Segovia JFO, Sousa VYK, Mata ECG, Gonçalves MCA, Bezerra RM, Jumor POM,
 Kanzaki LIB. Anti-microbial activity of Amazonian medicinal plants. *Biomed Life Sci* 2013; 2: 371-376.

- 212 25. Aladesanmi AJ, Iwalewa EO, Adebajo AC, Akinkunmi EO, TAiwo BJ, Olorunmola FO, Lamikanra A.
 213 Anti-microbial and anti-oxidant activities of some Nigerian medicinal plants. *Afr J Trad Comp Alt Med* 214 2007; 4(2): 173-184.
- 215 26. Raj A, Singh A, Sharma A, Singh N, Kumar P, Bhatia V. Antifertility activity of medicinal plants on 216 reproductive system of female rat. *Int J Bio-Eng Sci Tech* 2011; **2**(3): 44-50.
- 217 27. Joshi SC, Sharma A, Chaturvedi M. Antifertility potential of some medicinal plants in males: An
 218 overview. Int J Pharm Pharmaceut Sci 2011; 3(5): 204-217.
- 219 28. Akah PA. Abortifacient activity of some Nigerian medicinal plants. *Phytother Res* 1994; **8**(2): 106-108.
- 220 29. Sethi N, Nath D, Shukla Sc, Dyal R. Abortifacient activity of a medicinal plant '*Moringa olifera*' in rats.
 221 Ancient Sci Life 1988; 7(3-4): 172-174.
- 30. Yakubu MT, Bukoye BB. Abortifacient potentials of the aqueous extract of *Bambusa vulgaris* leaves in pregnant Dutch rabbits. *Contraception* 2009; **80**(2009): 308-313.
- 31. Ayoka AO, Akomolafe RO, Iwalewa EO, Ukponmwan OE. Studies on the anxiolytic effect of Spondias
 mombin L (Anacardiaceae) extracts. Afr J Trad Compl Alt Med 2005; 2(2): 153-165.
- 32. Ajao AO, Shonukan O, Femi-Onadeko B. Anti-bacterial effect of aqueous and alcohol extracts of Spondias mombin and Alchonea cordifolia: two local antimicrobial remedies. Int J Crude Drug Res 1985; 23: 67-72.
- 33. Iweala EEJ, Oludare FD. Hypoglycaemic effect, biochemical and histological changes of *Spondias mombin* and *Parinari polyandra* Benth Seeds ethanolic extracts in alloxan induced diabetic rats. J
 Pharm Toxicol 2011; 6 2): 101-112.
- 34. Uchendu CN, ISek T. Antifertility activity of aqueous ethanolic extract of Spondias mombin (Anacardiaceae) in rats. Afr Health Sci 2008; 8(3): 163-167.
- 35. Abo KA, Ogunleye VO, Asindi JS. Antimicrobial potential of *Spondias mombin*, *Croton zambesicus* and *Zygotritonia crocea*. *Phytother Res* 1999; **13**: 494-497.
- 236
 236. Corthout J, Pieters LA, Claeys M, Vanden-Berghe DA, Viletinck AJ. Antibacterial and molluscicidal
 237 phenolic acid from *Spondias mombin. Planta Med* 1994; **60**: 460-463.
- 37. Goncalves JL, Lopez RC, Oliviera DB, Costa SS, Miranda MM, Romanos MT, Santos NS, Wigg MD.
 In vitro anti- rotavirus activity of some medicinal plants used in Brazil against diarrhea. J
 Ethnopharmacol 2005; **99**(3): 403-407.
- 38. Asuquo OR, Udonwa UN, Eluwa MA, Ekanem TB. Effects of *Spondias mombin* leaf extract on the cytoarchitecture of the cerebral cortex and on learning and memory in Wistar rats. *Int J Sci Res* 2013;
 243 2(9): 5-8.
- 39. Asuquo OR, Ekanem TB, Udoh PB, Mesembe OE, Ebong PE. Haematinic potential of *Spondias mombin* leaf extract in Wistar rats. *Adv Biores* 2013; **4**(2): 53-56.
- 40. Asuquo OR, Fischer CE, Mesembe OE, Igiri AO, Ekom IJ. Comparative study of aqueous and ethanolic leaf extracts of *Spondias mombin* on neurobehaviour in male rats. *IOSR J Pharm Biol Scis* 2013; 5(2): 29-35.
- Slidders W. The OFG and BrAB-OFG methods for staining the adenohypohysis. *J Path Bacteriol* 1961;
 82: 532-534.

- 42. Bailey SA, Zidell RH, Perry RW. Relationship between organ weight and body/brain weight in the rat:
 what is the best analytical endpoint? *Toxicol pathol* 2004; **32**: 448-466.
- 43. Asuquo OR, Ekanem TB, Eluwa MA, Oko OO, Ikpi DE. Evaluation of toxicological effects of *Spondias mombin* in adult male Wistar rats. *J Nat Sci Res* 2012; **2**(7): 144-151.
- 44. Hurtuk BL, Krefetz RG. Enzymes: In Bishop MC, Duben-Engel Kirk, JL & Fody EP (Eds). Clinical chemistry, principles, procedures and correlations (2nd ed). Philadelphia, JB Lippincott Company,1992; PP 215-233.
- 45. Akpanabiatu MI, Umoh IB, Eyong EU, Udoh FV. Influence of *Nauclea latifolia* leaf extracts on some hepatic enzymes of rats fed on coconut oil and non-coconut oil meals. *Pharm Biol* 2005; **43**(2): 153-157.
- 46. Johnson DF, Fody EP. Liver function: In Bishop MC, Duben-Engel Kirk, JL & Fody EP (Eds). Clinical chemistry, principles, procedures and correlations (2nd ed). Philadelphia, JB Lippincott Company, 1992; PP 473-478.
- 47. Sharma A, Mathur R, Skukla S. Hepatoprotective action of a proprietary herbal preparation against carbon tetrachloride intoxication. *Indian Drugs* 1995; **32**: 120-124.
- 48. Asuquo OR, Ekanem TB, Udoh PB, Eluwa MA, Mesembe OE. Antigonadotrophic effect of *Spondias mombin* extract in adult male Wistar rats. *J Biol Agric Healthcare* 2012; 2(7): 14-17.
- 49. Kumara M, Singh P. Study of the reproductive organs and fertility of the male mice following administration of metronidazole. *Int J Fert Sterl* 2013; 7(3): 225-238.
- 50. Desjardins C. Endocrine regulation of reproductive development and function in the male. *J Ani Sci* 1978; **47**: 56-79.