

Diagnosis of Therapy-related Acute Myeloid Leukemia with t(8;21)(q22;q22.1) After Treatment for Mantle Cell Lymphoma and Oral Squamous Cell Carcinoma

ABSTRACT

Aims: We report a rare case of therapy-related AML with t(8;21)(q22;q22.1) that occurred after treatment for mantle cell lymphoma (MCL) and oral squamous cell carcinoma (OSCC). **Presentation of case:** A 52 years-old male patient was diagnosed with MCL in leukemic phase. The treatment consisted in R-CHOP rituximab, cyclophosphamide, doxorubicin, vincristine and prednisone, then patient experienced remission. Three months later, he presented a lump that was diagnosed as OSCC, which was surgically removed and treated with cisplatin and radiotherapy. Then, the patient's hemogram presented 35.0% of blasts and, after morphologic, phenotypic and molecular analysis, it was classified as AML with t(8;21)(q22;q22.1). However, due to the previous historic of chemotherapy and radiotherapy, the final diagnosis was t-AML. **Discussion:** The correct diagnosis of therapy related malignancies is important due to its severity as they are very aggressive and, usually, considered incurable. t-AMLs with t(8;21)(q22;q22.1) is considered as favorable karyotype, still, it has a poorer outcome compared with its *de novo* counterpart. **Conclusion:** t-AML with t(8;21)(q22;q22.1) is rare and few cases are described in the literature. More reports are necessary to better elucidate the mechanisms involved in this disease to define better treatment strategies to prevent these events and to improve the poor outcomes.

Keywords: Therapy-related neoplasms, mantle cell lymphoma, oral squamous carcinoma, t-AML.

1. INTRODUCTION

According to the Classification of Tumours of Haematopoietic and Lymphoid Tissues by the World Health Organization (WHO), therapy-related myeloid neoplasms (t-MNs) are a distinct class of hematological malignancies that occur after cytotoxic chemotherapy and/or radiation therapy (RT) administered for a previous neoplastic disorder. t-MNs includes therapy-related acute myeloid leukemia (t-AML), myelodysplastic syndromes (t-MDS) and myelodysplastic/myeloproliferative neoplasms (t-MDS/MPN). These neoplasms carry high-risk karyotypes and have a significantly poorer outcome compared with *de novo* hematopoietic malignancies [1,2].

t-MNs are the consequence of mutations or changes in hematopoietic stem cells and/or in the bone marrow (BM) microenvironment induced by cytotoxic treatments or by the selection of a myeloid clone with a mutate phenotype. The most commonly cytotoxic agents implicated in t-MNs are alkylating agents (AA) (such as cyclophosphamide and cisplatin), topoisomerase II inhibitors (TPI) (like etoposide and doxorubicin), ionizing RT, antimetabolites and antitubulin agents (such as vincristine) [1].

35 t-AML with cytogenetic abnormalities associated with a favorable prognosis like
36 t(8;21), inv(16) and t(15;17) are uncommon and these cases represent approximately only
37 10% of all t-MNs [3]. AML with t(8;21)(q22;q22.1) usually has a favorable prognosis,
38 however, AMLs with favorable karyotypes have a slightly poorer outcome compared with
39 their *de novo* counterparts. Even so, these cases still have significantly better outcomes than
40 other t-MNs [2].

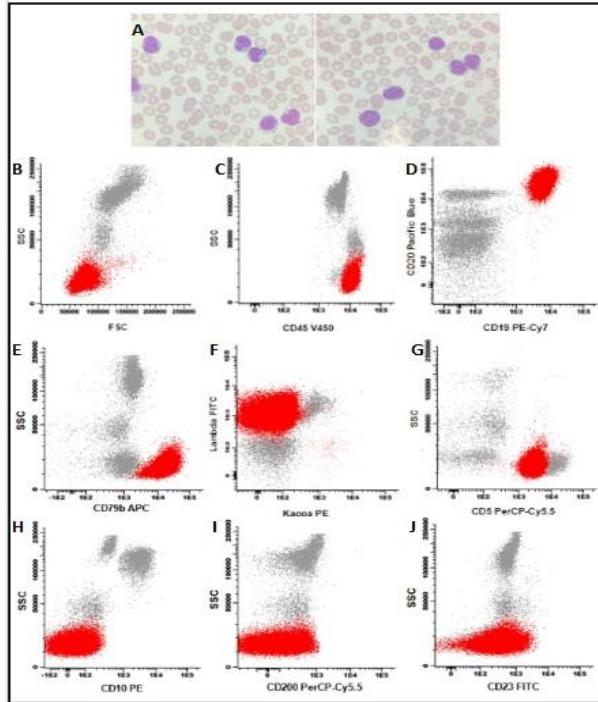
41 The incidence of t-MNs is expected to raise due to the increased survival rates of
42 cancer patients. In fact, there are nearly 12 million cancer survivors today only in the United
43 States [4]. Therefore, considering the poor outcome of t-MNs and the increasing cancer
44 survival rates, the correct diagnosis of these malignant disorders is crucial to better assist
45 these patients.

46 In this study we report a rare case of t-AML with t(8;21)(q22;q22.1) that occurred after
47 treatment for mantle cell lymphoma (MCL) and oral squamous cell carcinoma (OSCC). The
48 Research Ethics Committee of the Federal University of Santa Catarina approved this study
49 (CAAE: 61598816.7.0000.0121).

51 2. PRESENTATION OF CASE

52
53 A 52 years-old male patient, smoker, diagnosed with MCL was admitted to the
54 University Hospital Professor Polydoro Ernani de São Thiago (HU-UFSC) for lymphoma
55 staging and to start chemotherapy treatment.

56 The blood smear analysis presented a predominance of small cells with a high
57 nucleus-cytoplasm ratio and slight nuclear chromatin condensation, whereas some of these
58 cells also presented cleaved nucleus (Figure 1A). In order to confirm a possible peripheral
59 blood (PB) involvement, immunophenotyping by flow cytometry was required. The analysis
60 presented 78.1% of lymphoid B (CD19+) mature (CD20+, CD45++) cells, with low FSC and
61 SSC, an aberrant expression of CD5 and no expression of CD10, CD23 and CD200. Among
62 these cells, 99% presented lambda light chain restriction. The phenotype of these
63 pathological cells was suggestive of MCL with PB involvement, which characterizes MCL in
64 leukemic phase (Figure 1B-J). The treatment consisted in 8 cycles of R-CHOP rituximab
65 (600mg), cyclophosphamide (1.230 mg), doxorubicin (82 mg), vincristine (1 mg) and
66 prednisone (20 mg) over 5 months, then, the patient experienced remission.



67
 68 Figure 1 - A) Morphology of PB smear presenting small cells with large nucleus,
 69 slight nuclear chromatin condensation, abnormal segmentation and some cleaved
 70 nucleus. B-J) Demonstrative dot plots of pathological cells immunophenotyping (red
 71 population): B) Demonstration of small cells with low FSC x SSC. C) Expression of
 72 CD45. D) Expression of CD19 and CD20. E) Expression of CD79b. F)
 73 Demonstration of Lambda restriction. G) Expression of CD5. H) No expression of
 74 CD10. I) No expression of CD200. J) No expression of CD23.
 75

76 About three months after lymphoma remission, the patient presented a 1.5 cm tumor
 77 in the mucosa of left molar trigone region. The tumor was biopsied and diagnosed as OSCC,
 78 which was surgically removed and treated with cisplatin and RT at the Oncology Research
 79 Center (CEPON).

80 One month after the end of OSCC treatment, in a following medical appointment, the
 81 patient's hemogram showed 4400 leucocytes/mm³, 35.0% of blast cells, hemoglobin of 6.9
 82 g/dL and a platelet count of 2000 /mm³. The immature cells presented large size, basophilic
 83 cytoplasm, slight nuclear chromatin condensation and visible nuclei; besides, some
 84 granulocytic cells showed abnormal nuclear segmentation (pseudo-Pelger-Huët nuclei)
 85 (Figure 2B). The immunophenotyping of PB showed 38.60% of blasts (CD34++, CD45+),
 86 medium to large sized, committed with the myeloid lineage (MPO+, CD13+, CD33+,
 87 CD117+, HLA-DR+) and with aberrant expression of CD19 (70%), CD79a (30%) e CD56
 88 (80%) (Figure 2D-L). The aberrant expression of CD19 in myeloid blasts suggests the
 89 presence of t(8;21)(q22;q22.1), which was confirmed by karyotype and Nested RT-PCR
 90 (Figure 2C).

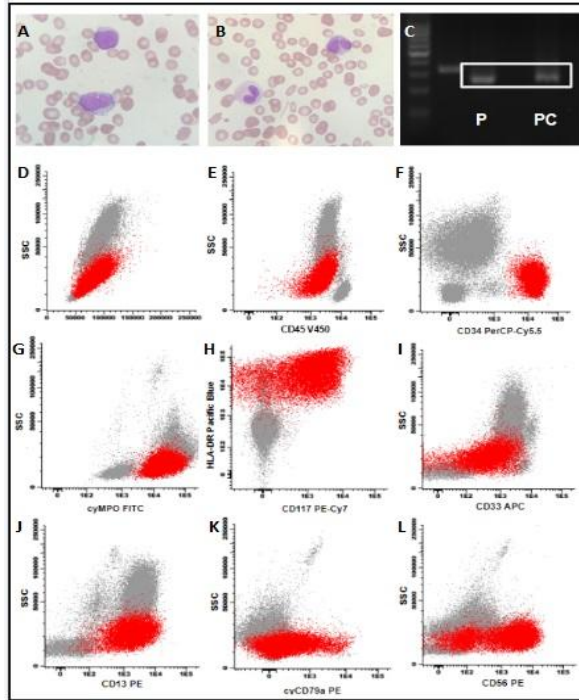


Figure 2. A) Large blasts with high cytoplasm/nucleus ratio, visible nuclei and basophilic cytoplasm. B) Granulocytic cells presenting abnormal segmentation (pseudo-Pelger-Hüet nuclei). C) Nested RT-PCR for t(8;21): P: patient's PB band, PC: positive control band. D-L) Demonstrative dot plots of blasts immunophenotyping (red population): D) Demonstration of large cells with high FSC x SSC. E) Expression of CD45. F) Expression of CD34. G) Expression of cyMPO. H) Expression of CD117 and HLA-DR. I) Weak expression of CD33. J) Expression of CD13. K) Aberrant expression of CD19 and CD79a. L) Aberrant expression of CD56.

91
92
93
94
95
96
97
98
99
100
101
102
103
104
105
106
107
108
109
110
111
112
113
114
115
116
117
118
119
120
121

After morphologic, phenotypic and molecular analysis, the disease was classified as AML with t(8;21)(q22;q22.1); however, due to the previous historic of chemotherapy and radiotherapy, the correct diagnosis was t-AML, which has a worse prognosis.

3. DISCUSSION

MCL is a mature B cell neoplasm characterized by an advanced stage at the diagnosis and most patients present PB or BM involvement. The correct diagnosis of MCL is important due to its severity as it has been considered incurable, very aggressive and associated with a poor prognosis. The laboratorial diagnosis of MCL is established by WHO and, overall, the histological confirmation is mandatory. However, the variant morphology observed in MCL can difficult the differential morphological diagnosis between MCL and other B cell neoplasms. Thereby, the immunophenotyping of neoplastic cells is important to differentiate the lymphoma subtypes. Additionally, cytogenetics has a key role in MCL diagnosis by detecting t(11;14), the molecular hallmark of MCL, which is found in more than 95% of cases [1].

According to WHO, the characteristic MCL immunophenotype includes the expression of B-cell markers (CD19+, CD20+, CD22+, CD79a+, PAX5) and intense IgM/IgD, mostly with lambda light chain restriction. The commonest immunophenotypic markers are CD5+, FMC7+, CD45+, CD43+ and intranuclear cyclin D1+ [1]. In this clinical report, the neoplastic

122 cells presented a classic MCL morphology and immunophenotype (CD19+, CD20+,
123 CD45++, CD5+, CD23-, CD10- and CD200-) (Figure 1A-J).

124 The adopted treatment regimen was 8 cycles of R-CHOP. According to the literature,
125 cyclophosphamide is an AA and doxorubicin is a TPI, and they are both known to be
126 particularly mutagenic and have a strong leukemogenic potential [5]. Despite the advances
127 in cancer treatment, the currently available chemotherapy regimens, associated or not with
128 monoclonal antibodies, have potential to cause many side effects, including secondary
129 malignant neoplasms. There are many studies that investigate the influence of
130 chemotherapy on the development of leukemia and solid tumors [6,7]. However, the etiology
131 of t-AML and secondary solid cancers after administration of cyclophosphamide and
132 doxorubicin as well as rituximab has not been completely elucidated [5].

133 In this study, about three months after a lymphoma remission, the patient was
134 diagnosed with OSCC, a malignant neoplasm derived from the squamous epithelium of the
135 oral cavity [8]. The lesion was found in the mucosa of the left molar trigone and the diagnosis
136 was determined by histopathology.

137 For OSCC treatment, surgery remains the best option, but chemotherapy and
138 radiotherapy are also applied in combination to obtain a better response [8]. In this case, the
139 patient's treatment consisted of RT combined with cisplatin; and both methods have
140 potential for AML or SMD development [1]. However, one meta-analysis [9] found no
141 increased risk of secondary cancers associated with cisplatin compared with non-cisplatin-
142 based chemotherapy. Nevertheless, one month after the end of OSCC treatment, the patient
143 was diagnosed with t(8;21)(q22;q22.1) AML.

144 AML is a heterogeneous malignancy and cases with t(8;21)(q22;q22.1) represent a
145 group with specific clinical and biological characteristics. The diagnosis of AML with
146 t(8;21)(q22;q22.1) is based on cytomorphology, cytogenetics and immunophenotyping
147 according to the WHO classification. The commonest morphological features include the
148 presence of large blasts with abundant basophilic cytoplasm, sometimes containing
149 azurophilic granules and perinuclear clearing. Some blasts may contain pseudo-Chédiak-
150 Higashi large granules, suggesting the presence of the fused gene. Concomitant with the
151 large blasts, some smaller blasts with pseudo-Pelger-Huet abnormal nuclear segmentation
152 can also be found [1]. These abnormalities were observed in the patient's blood smear
153 (Figure 2B).

154 The PB immunophenotyping showed myeloid blasts with partial expression of CD19,
155 CD79a and CD56 (Figure 2J-L). This immunophenotype is suggestive of AML with
156 t(8;21)(q22;q22.1) according to the WHO classification, as its characteristic
157 immunophenotypic profile includes strong expression of CD34, HLA-DR, myeloperoxidase
158 (MPO) and CD13, and relatively weak expression of CD33. Furthermore, the presence of
159 lymphoid-associated markers like CD19, CD79a may also be observed and expression of
160 CD56 is associated with a poorer prognosis [1]. Molecular cytogenetic methods, such as
161 karyotype and PCR, are considered as the gold standard for the diagnosis by identifying the
162 t(8;21)(q22;q22.1). In this case report, this translocation was detected by nested RT-PCR
163 (Figure 2C).

164 Blasts of *de novo* AML with t(8;21)(q22;q22.1) and t-AML share morphological,
165 immunophenotypic, cytogenetic and molecular features, although t-AML with
166 t(8;21)(q22;q22.1) seems to have more dysplastic changes than *de novo* AML [10]. These
167 dysplastic characteristics were observed in the patient's granulocytic cells (Figure 2A-B).
168 Based on morphologic, phenotypic and molecular analysis and on the previous historic of
169 chemotherapy and RT, this case was finally diagnosed as t-AML.

170 Studies demonstrated that patients with t-AML with t(8;21)(q22;q22.1), are usually
171 older, have lower white blood cells (WBC) counts and an inferior overall survival than their
172 *de novo* counterparts [2-10]. The patient in this study was 52-years old and had a poor
173 response after treatment with (7+3) cytarabine (100 mg/m²) and daunorubicin (60 mg/m²).
174 He presented persistent blasts in the PB and passed away few months later.

175 In t-AML with t(8;21)(q22;q22.1) is observed a short latent period, without a previous
176 myelodysplastic phase, and it is associated with prior TPI therapy or RT alone [1]. The
177 mechanisms responsible for such mutations remain unknown, but may involve several
178 chromatin structural elements such as topoisomerase II cleavage sites, which are found to
179 colocalize with preferential breakage sites after exposure to damage, such as TPIs [10]. The
180 patient's t-AML was diagnosed four months after the administration of doxorubicin, a TPI, for
181 MCL treatment; and no sign of myelodysplasia was observed before the t-AML diagnosis,
182 which is compatible with the literature description.

183

184 **4. CONCLUSION**

185

186 t-AML with t(8;21)(q22;q22.1) is rare and few cases are described in the literature. It is
187 a fatal complication of cancer treatment and its incidence is expected to rise due to the
188 increasing number of cancer survivors. It shares the same morphological, molecular and
189 immunophenotypic features than *de novo* AML with t(8;21)(q22;q22.1), though presenting a
190 worse prognosis. Thus, the correct diagnosis of this disease is crucial due to its severity and
191 low overall survival rates. For that reason, more reports and studies are necessary to better
192 elucidate the mechanisms involved in the development of t-AMLs in order to define better
193 treatment strategies, preventing these events and improving the poor outcomes presented in
194 such cases.

195

196

197

198

199 **COMPETING INTERESTS**

200

201 Authors have declared that no competing interests exist.

202

203

204 **CONSENT**

205

206 All authors declare that written informed consent was obtained for publication of this case
207 report and accompanying images.

208

209 **ETHICAL APPROVAL (WHERE EVER APPLICABLE)**

210

211 "All authors hereby declare that all experiments have been examined and approved by the
212 appropriate ethics committee and have therefore been performed in accordance with the
213 ethical standards laid down in the 1964 Declaration of Helsinki. The Research Ethics
214 Committee of the Federal University of Santa Catarina approved this study (CAAE:
215 61598816.7.0000.0121).

216

217

218 **REFERENCES**

219

220 1. Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H *et al.* WHO
221 classification of tumours of haematopoietic and lymphoid tissues. Revised 4th ed.
222 IARC: Lyon, 201. 585 p.

223 2. Zhang L, Wang SA. A focused review of hematopoietic neoplasms occurring in
224 the therapy-related setting. *Int J Clin Exp Pathol.* 2014;7(7):3512-3523.

- 225 3. Bueso-Ramos CE, Kanagal-Shamanna R, Routbort MJ, Hanson CA. Therapy-
226 related myeloid neoplasms. *Am J Clin Pathol.* 2015;144:207-218.
- 227 4. Morton LM, Dores GM, Tucker MA, Kim CJ, Onel K, Gilbert ES *et al.* Evolving
228 risk of therapy-related acute myeloid leukemia following cancer chemotherapy
229 among adults in the United States, 1975-2008. *Blood.* 2013; 121(15): 2996–3004.
- 230 5. Xu Y1, Wang H, Zhou S, Yu M, Wang X, Fu K *et al.* Risk of second malignant
231 neoplasms after cyclophosphamide-based chemotherapy with or without
232 radiotherapy for non-Hodgkin lymphoma. *Leuk Lymphoma.* 2013;54(7):1396-1404.
- 233 6. Mudie NY, Swerdlow AJ, Higgins CD *et al.* Risk of second malignancy after non-
234 Hodgkin's lymphoma: a british cohort study. *J Clin Onco.* 2006; 24: 1568–1574.
- 235 7. Tward JD, Wendland MM, Shrieve DC *et al.* The risk of secondary malignancies
236 over 30 years after the treatment of non-Hodgkin lymphoma. *Cancer.* 2006;107:
237 108–115.
- 238 8. Malik UU, Zarina S, Pennington SR. Oral squamous cell carcinoma: Key clinical
239 questions, biomarker discovery, and the role of proteomics. *Arch Oral Biol.* 2016;63:
240 53-65.
- 241 9. Liang F, Zhang S, Xue H, Chen Q. Risk of second primary cancers in cancer
242 patients treated with cisplatin: a systematic review and meta-analysis of randomized
243 studies. *BMC Cancer.* 2017;17(1):871.
- 244 10. Duployez N, Willekens C, Marceau-Renaut A, Boudry-Labis E, Preudhomme C.
245 Prognosis and monitoring of core-binding factor acute myeloid leukemia: current
246 and emerging factors. *Expert Rev Hematol.* 2015;8(1):43-56.