

1           **Antibacterial activity of some nano particles on antibiotic resistant**  
2                           **bacterial pathogens from air of operation theatre**

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4   **Abstract:**

5   The current research work was carried out to find antibacterial activity of some nano particles  
6   against bacterial pathogens isolated from air of operation theatre of Mayo hospital. Three  
7   pathogenic bacterial strains were isolated. These strains were named as A1, A2, A3.  
8   Molecular characterization, optimum growth conditions and antibiotic resistance of bacterial  
9   isolates was checked. The antibiotics used in this study were Amoxycillin, Cefepime and  
10   Ampicillin. Nano particles were used in methanolic solutions (mg/ml). Nano particles  
11   included ferric oxide, Zinc oxide and Silver oxide. Results showed A3 was resistant to all  
12   antibiotics. Other strains showed sensitivity and resistance to these three antibiotics. All nano  
13   particles showed antibacterial activity against pathogenic bacterial isolates. Maximum zone  
14   of inhibition of 1cm was formed when used Ferric oxide against A1 bacterial pathogen.  
15   Optimum temperature was 37°C while the optimum pH was 7. These bacterial pathogens  
16   were identified by ribotyping as *Staphylococcus aureus* (A1), *Pseudomonas aeruginosa* (A2)  
17   and *Streptococcus pyogenes* (A3).

18   **Key words:** bacterial pathogens; nano particles; antibacterial activity; ribotyping

19   **Introduction:**

20   Both pathogenic and non-pathogenic bacteria are present in air. This contamination is  
21   increasing day by day due to increase in human population. Human population increase  
22   results in increased waste production, improper sanitary conditions and waste disposal  
23   problems (Hanif *et al.*, 1995). Hospital indoor air contains a diverse group of micro-  
24   organisms. Here the significance of these microbes is put to argument, whereas these may be  
25   considered significant in any other sphere. Farzana (1988) studied the airborne pathogenic  
26   bacterial isolates from various wards of Ganga Ram Hospital, Lahore. The work showed that  
27   the *Staphylococcus sp.*, *Streptococcus pyogenes* and *Enterobacter sp.*, were frequent in  
28   hospital air. Airborne bacterial contamination in the operating theatre is one of the reasons for  
29   infections in connection with surgery. Because of overuse and misuse of antibiotics the  
30   bacterial pathogens have become resistant and this resistance is increasing. So there is need

31 of additional therapies for infection control (Jaffal *et al.*, 1997). Nano particles are being used  
32 in research to study their antibacterial activity against these common pathogens. Nano  
33 particles range from 1 to 100 nm in size. Recent studies have proved that nano particles are  
34 not only effective in treatment of cancer cells but also show significant antibacterial activity  
35 against common pathogens.

### 36 **Materials and Methods:**

37 Bacterial pathogens were isolated from air of operation theatre. Sampling was done at  
38 specific selected points in operation theatre. Random sampling was done to get better results.  
39 Sampling was conducted by exposing nutrient agar plates in operation theatre for three  
40 minutes. These plates were exposed at different points in operation theatre (Benson, 2002).  
41 After sampling, plates are placed in an incubator for overnight at 37° C. Isolated bacterial  
42 colonies were streaked on fresh agar plated to obtain pure culture. These pure cultures were  
43 subjected to blood agar test, antibiotic resistance/sensitivity test, nano particles  
44 resistance/sensitivity test, optimum growth conditions and molecular characterization  
45 (Cheesebrough, 1993).

### 46 **Determination of Optimum Growth Conditions:**

47 Optimum growth conditions for each bacterial isolate were determined. The optimum  
48 temperature of three strains was observed. The temperature range was 25°C, 30°C, 37°C and  
49 40°C. The optimum pH of strains was also observed. The pH studied was 6.5, 7.0, 7.5 and  
50 8.0.

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### 52 **Antibiotic resistance of bacterial pathogens:**

53 Assessment of antibiotic resistance of bacterial pathogens was checked against broad-  
54 spectrum antibiotics by performing disc diffusion method. For the test, nutrient agar plates  
55 were prepared for three strains. Bacteria were spread on the plates by spreading plate  
56 method. Antibiotics discs of known concentration were placed on the plates with the help of  
57 sterilized forcep and were incubated at 37 °C for 24 hours. Growth inhibited zones appeared  
58 as clear area near the disc. Growth inhibited zones were measured. Clear zone indicated the  
59 sensitivity of tested bacterial strain against that antibiotic and no zone showed resistance.

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**62 Antibacterial activity test of Nano particles:**

63 Antibacterial activity of various Nano particles was tested by well diffusion method. The  
64 solution of Nano particles were made in the organic solvent *i.e.* Methanol The medium used  
65 was nutrient agar; it was prepared by dissolving 28 grams of prepared nutrient agar in 1 liter  
66 (1000ml) of distilled water in a flask. The pH of the medium was maintained at 7.4, the  
67 medium was sterilized by autoclaving for 20 minutes at 121<sup>0</sup>C temperature and 15 lb  
68 pressure. After medium was autoclaved, it was poured in the petri plates under sterile  
69 conditions, a drop of autoclaved water was poured in the center of the plate on which  
70 bacterial isolate was inoculated and it was then evenly spread on the entire plate with the help  
71 of sterilized spreader. After that, wells were made in the plates. Solutions (1mg/ml) of three  
72 Nano particles *i.e.* Ferric oxide, Silver oxide and Zinc oxide were used. 50 micro liters  
73 solution of Nano particles were poured separately in the wells and 50 micro liters of methanol  
74 was also poured in a separate well as a control. Petri plates were covered with lids and  
75 incubated at 37<sup>0</sup>C for 24 hours. After incubation, zone of inhibition around the wells showed  
76 the sensitivity of the isolate against particular particle whereas growth around the well  
77 indicated that the bacterial isolate was resistant against the particular particle.

**78 Molecular characterization**

79 Ribotyping or molecular characterization of 16s rRNA gene was done. Genomic DNA was  
80 isolated by phenol:chloroform extraction method. PCR was done using universal primers;  
81 27f and 1495r. After pcr gene clean was done and then sequencing from molecular  
82 laboratory, Malaysia.

**83 Results:**

84 From air sample taken from operation theatre (Mayo hospital). Three bacterial pathogens A1,  
85 A2, A3 were identified as *Staphylococcus aureus*, *Pseudomonas aeruginosa* and  
86 *streptococcus pyogenes* by ribotyping. Bacterial pathogens showed resistance against  
87 antibiotics used. Bacterial strain A3 was most resistant against Amoxycillin, Cefepime and  
88 Ampicillin (Table 1). The sensitivity/resistance was checked by measuring Zone of  
89 inhibition. The zone of inhibition was measured in centimeter (cm).

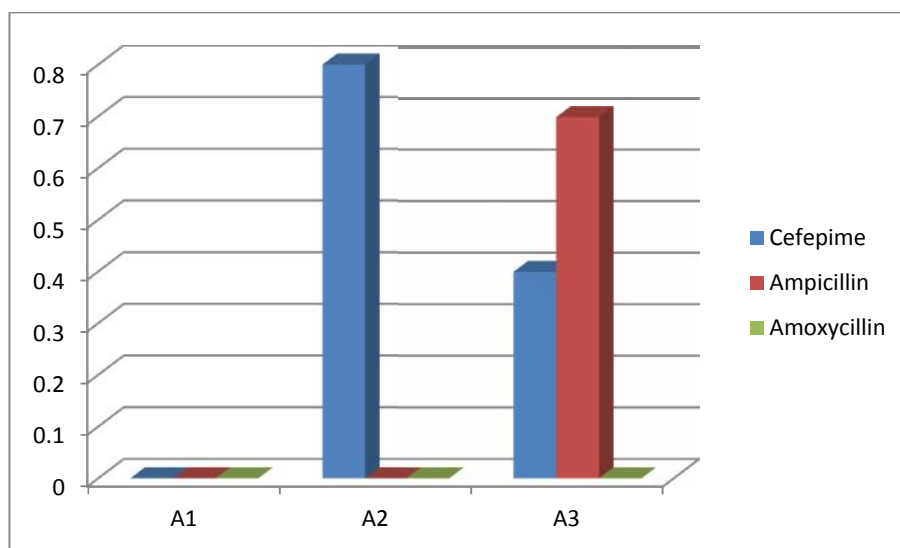
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92 **Table 1: Antibiotic resistance/sensitivity of bacterial pathogens**

strain	Amoxycillin (AMC 30ug)	Ampicillin (AMP 30ug)	Cefepime (CF 30ug)
	cm	cm	cm
<b>A1</b>	R	S (0.8)	S (0.4)
<b>A2</b>	R	R	S (0.7)
<b>A3</b>	R	R	R

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94

95 **Figure 1: Antibiotic resistance/sensitivity of bacterial pathogens A1, A2 and A3**

96 Antibacterial activity of nano particles was also studied. All bacterial pathogens were  
 97 resistant against control solution of nano particles i.e, methanol. But nano particles showed  
 98 clear antibacterial activity against all antibiotic resistant bacterial pathogens (Table 2). Ferric  
 99 oxide solution showed maximum antibacterial activity against A1(*Staphylococcus aureus*) by  
 100 forming Zone of inhibition of 1cm while zinc oxide formed zone of inhibition of 0.3cm  
 101 against A3(*Streptococcus pyogenes*).

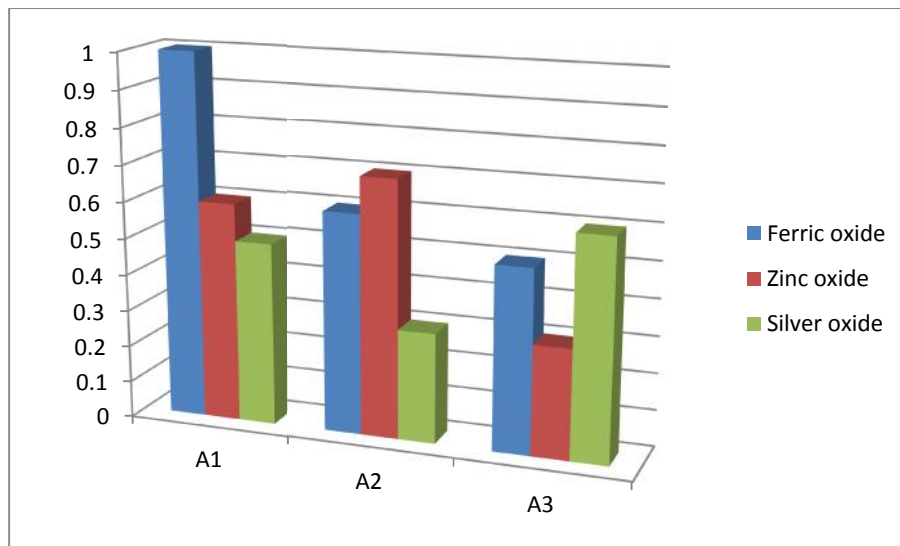
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103 **Table 2: Antibacterial activity test of Nano particles**

Nano particles solutions	Strain A1	Strain A2	Strain A3
Ferric oxide (1mg/ml)	1.0cm	0.6cm	0.5cm
Zinc oxide (1mg/ml)	0.6cm	0.7cm	0.3cm
Silver oxide (1mg/ml)	0.9cm	0.9cm	0.6cm
Methanol (control)	R	R	R

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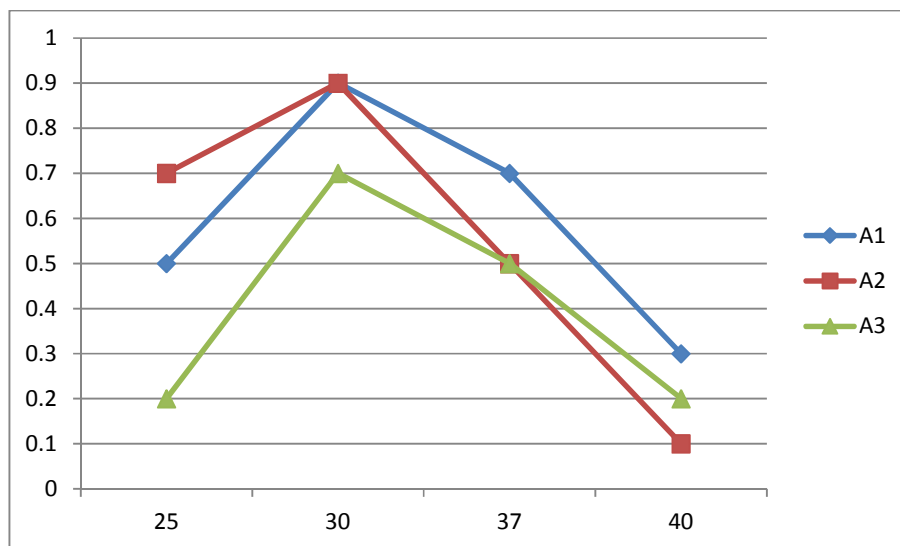
R= RESISTANT



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Figure 2: Antibacterial activity of nano particles against bacterial pathogens A1, A2 and A3

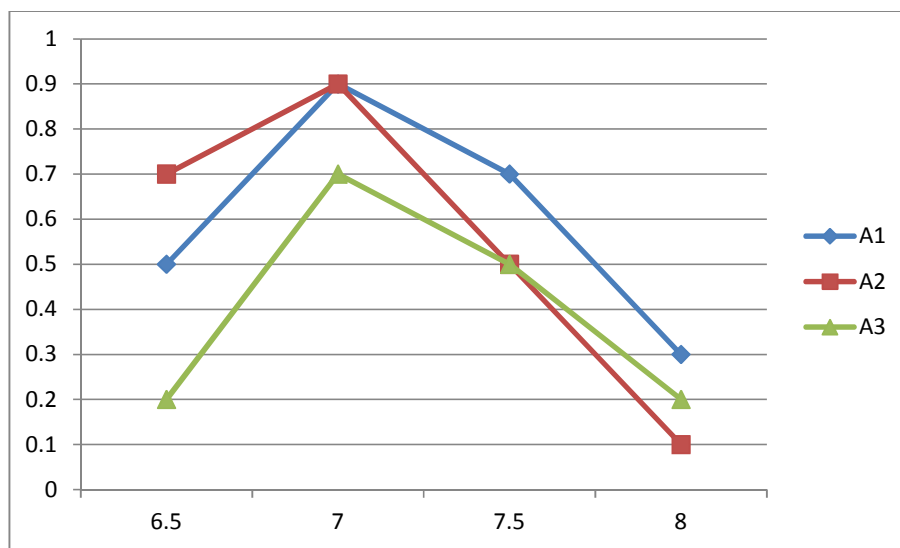
114 Optimum growth conditions were also observed. Optimum temperature for all strains was  
 115 37°C and that optimum pH was 7.



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117 Figure 3: Optimum Temperature (°C) of bacterial pathogens

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120 Figure 4: Optimum pH of bacterial pathogens

121 For molecular characterization sequences obtained were blast on NCBI website.

122 ***Staphylococcus aureus* ( partial sequence 16s rRNA gene)**

123 TTTATGGAGAGTTTGATCCTGGCTCAGGATGAACGCTGGCGGCGTGCCTAATACATGCAAGTCGAG  
 124 CGAACGGACGAGAGCTTGCTTCTATGATGTTAGCGGCGGACGGGTGAGTAACACGTGGATAACCT

125 ACCTATAAGACTGGGATAACTTCGGGAACCGGAGCTAATACCGGATAATATTTTGAACCGCATGG  
 126 TTCAAAAGTGAAAGACGGTCTTGCTGTCACTTATAGATGGATCCGCGCTGCATTAGCTAGTTGGTA  
 127 AGGTAAGTTACCAAGGCAACGGATGCATAGCCGACCTGAGAGGGTGATCGGCCACACTGGAACCTGA  
 128 GACACGGTCCAGACTCCTACGGGAGGCAGCAGTAGGGTCTTCCGCAATGGGCGAAAGCCTGACGG  
 129 CCGAGCAACGCCGCGTGAGTGATGAAGGTCTTCGGATCGTAAACTCTGTTATTAGGGAAGAACA  
 130 TATGTGTAAGTAACTGTGCACATCTCGCGGTACCTAATCAGAAAG

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132 ***Streptococcus pyogenes* ( partial sequence 16s rRNA gene)**

133 GAGAGTTTGATCCTCCGCTCAGGACGAACGCTGGCGGGCGTGCCTAATACATGCAAGTAGAACGCT  
 134 GAGAAGTGGACTGCACCGGTTCAAGGAGTTGCGAACGGGTGAGTAACGCGTAGGTAACCTACCT  
 135 CATAACGGGGGATAACTATTGGAAACGATAGCTAATACCGCATAAGAGAGACTAACGCATGTTAG  
 136 TAATTATAAAAGGGGCAATTGCTCCACTATGAGATGGACCTGCGTTGTATTAGCTAGTTGGTGAGG  
 137 TAAAGGCTCACCAAGGCGACGATACATAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGA  
 138 GACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCGGCAATGGGGGCAACCCTGAC  
 139 CGAGCAACGCCGCGTGAGTGAAGAAGGTTTTTCGGATCGTAAAGCTCTGTTGTTAGAGAAGAATAG  
 140 GTGGGAGTGAAAATCCACCAAGTGACGGTAACTAACCAGAAAGGGACG

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142 ***Pseudomonas aeruginosa* ( partial sequence 16s rRNA gene)**

143 GGTGCACAGCCGTCTGAGCGGTTGCTCAGCTGCTCAAGGACGCTGCCAAGGCAAACGCCTAAGC  
 144 CGTCATGAGTGAAATGCCGACACCCGCCGACGACCTGGTTCGTGATCGGCAAGATCGTTTCGGTGTA  
 145 CGGCATCCGCGGTGAGGTGAAGGTGTATTCTTTACCGACCCGTTGGACAACCTGCTGGACTATCG  
 146 CCGCTGGACGCTCCGGCGCGACGGCGAGATTCCGCGAGCCGAGCTGGTCAGGGGGCGCCTGCATG  
 147 GCAAGGTCTTGCCGCCAAGCTCAAGGGGCTCGACGATCGCGAAGAGGCCCGCACCTTACCCGGT  
 148 TACGAGATCTGCATCCCGCGTAGCGAGTTGCCCTCTCTCGAGGAAGGTGAGTACTACTGGCACCAG  
 149 CTGGAAGGCCTGAAGGTGATCGACCAGGGCAGGCAGTTGCTCGGCGTGATCGACCATCTGCTGGA  
 150 AACCGGTGCCAACGATGTATGGTGGTCAAGCCCTGCGCGGGCAGCCTGGACGACCCGCGAGCGCC  
 151 TGTTGCCCTACACCGGGCAGTGCGTGCTGTCGATCGACCTGGCCGCTGGCGAGATGCGGGTGGACT  
 152 GGGACGCGGACTTCTGATCATCCATGGACAAGCGTTTGTGGGTGGGCGTCGTCAGCATCTTCCGG  
 153 AGATGTTCCGCGGATCAGTGACTATGGCAT

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156 **Discussion:**

157 In recent study bacterial pathogens were isolated from operation theatre (OT) air. Air of OTs  
 158 is supposed to be sterile and bacteria free but countries like Pakistan where hygienic  
 159 conditions are not ideal, contamination of air is an issue. So present work was carried out to  
 160 study these common pathogens not only present outdoor but also in indoor environment even  
 161 places like OTs. The bacterial pathogens isolated are of common occurrence in hospitals yet  
 162 their presence in air of OT is questionable. Airborne bacterial pathogens introduced at  
 163 surgery are an important source of wound contamination and joint sepsis. It has already been  
 164 shown that even in ultraclean-air operating theatres; the surgical sucker forms a reservoir for  
 165 those organisms which have been implicated in septic loosening of the prostheses (Whyte *et*  
 166 *al.*, 1991; Hanif *et al.*, 1995).

167 The bacterial strains isolated were *Staphylococcus aureus*, *Streptococcus pyogenes* and  
 168 *Pseudomonas aeruginosa*. *Staphylococcus aureus* is most common pathogen among all in  
 169 environment and its infections are most common. *S. aureus* is Gram +ve cocci present in

170 form of clusters or bunches. It is coagulase positive which differentiates it from other species.  
171 *Streptococcus* sp. is Gram +ve cocci found in chains. Its infections are most common in  
172 operation wounds or postoperative wounds. *Pseudomonas aeruginosa* is commonly found in  
173 air of hospitals or soil near to the hospitals. It is oxidase positive and is an opportunistic  
174 pathogen (Cheesebrough, 1993).

175 The present study also provided data related to continuous increase in drug resistance against  
176 certain bacterial species. The misuse and overuse of antibiotics against infectious diseases  
177 results in the increase of drug resistance ability of microorganisms including bacteria (Canu  
178 *et al.*, 2002).

179 Nano particles are being extensively used to study antibacterial activity as these are  
180 considered as bactericidal agents. Many studies have shown that nano particles like ferric  
181 oxide, zinc oxide and especially silver oxide are used as bactericidal agents. This property is  
182 because of their small size thus contributing in bactericidal activity. In recent research study  
183 the nano particles have shown significant antibacterial activity against locally isolated common  
184 bacterial pathogens. Almost all bacterial pathogens are antibiotic resistant yet showed  
185 sensitivity against nano particles by forming clear zones. (Taylor and Webster, 2009). So in  
186 future the nano particles are strong candidates of being bactericidal agents against  
187 drug/antibiotic resistant bacterial pathogens

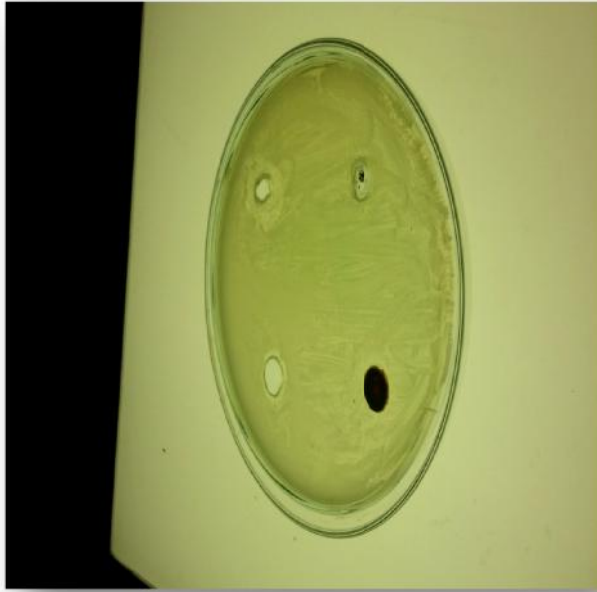
188 Now there is need to minimize or diminish the bacterial pathogens from OTs air as it is life  
189 threatening. There is need to improve sterile techniques and hygienic conditions, so that  
190 chances of operative or post operative infections would be minimized. .



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192 **Figure 5: Antibiotic resistance/sensitivity test**





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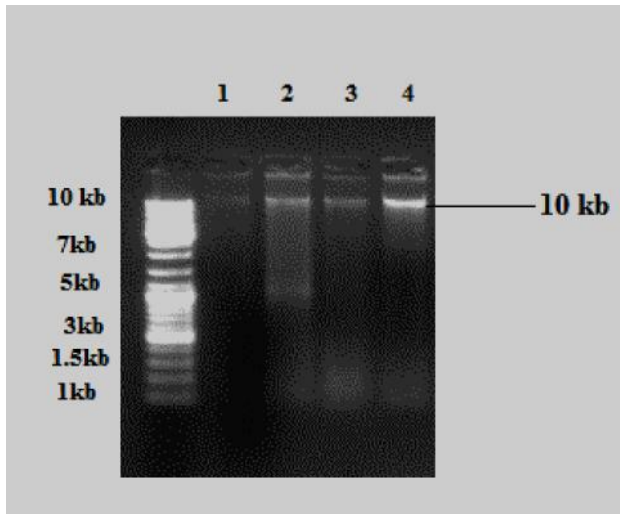
194 **Figure 6: Nano particles antibacterial activity test**

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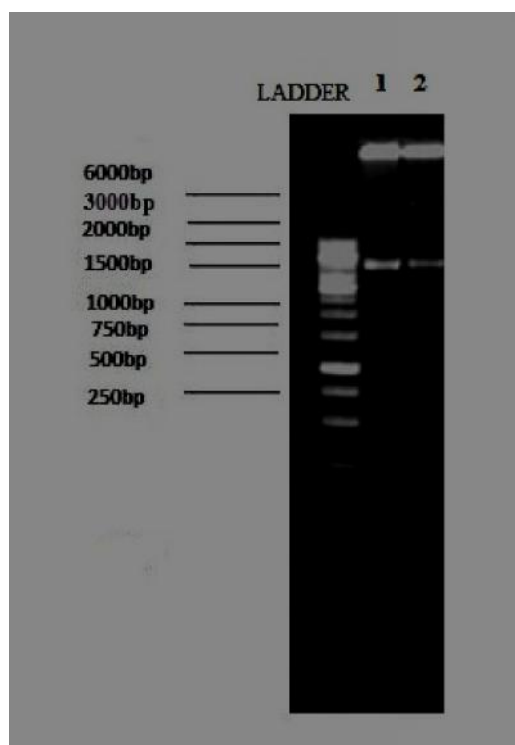
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200 **Figure 7: 1% agarose gel electrophoresis of bacterial genomic DNA**



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203 **Figure 8: PCR product of bacterial strains after agarose gel electrophoresis**

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205 **References:**

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