


# ***In Vitro* Analysis of Antimicrobial Susceptibility of *Xanthomonas axonopodis* pv. *punicae* and Comparative Study of Protein Profiling under Biotic Stress**

## **Abstract**

Bacterial blight of pomegranate caused by *X. axonopodis* pv. *Punicae* (*XAP*) assumed epidemic form and resulted in economic burden on farmers. In the current study the pathogen infected samples were collected and the isolated *XAP* was identity and confirmed through the morphological, biochemical characterization and Pathogenicity test. Bacterium was reisolated from infected plant to prove Koch's postulates. Efficacy of different chemicals and oils were tested by disc diffusion assay and turbidometrically. Bronopol 3000 ppm ( $25.6 \pm 1.6$  mm) and Clove oil ( $18.0 \pm 0.7$  mm) formed highest zone of inhibition Turbidometri showed the highest O.D. (0.908 nm) by Copper oxy chloride and Neem oil showed maximum inhibition of growth with O.D. (0.842 nm). Biotic stress (pathogen) induced protein response was studies by using SDS-PAGE method after protein extraction from *XAP*, healthy *P. granatum* L. and infected *P. granatum* L. The protein banding pattern showed the unique band no. 2 (Mol.Wt.66000 Da) in infected *P. granatum* L. as compared to the banding pattern of *XAP* and healthy *P. granatum* L. The over expressed protein due to biotic stress could be useful as a marker for detection of the disease at the early stage and for control of the diseases after knowing the biochemical significance of the protein.

**Keywords:** Pomegranate, bacterial blight, *Xanthomonas axonopodis* pv. *punicae*, *In vitro* efficacy, Antimicrobial activity, SDS-PAGE.

## **Introduction**

Pomegranate (*Punica granatum* L.) belongs to the family Lythraceae, is an ancient, medical fruit and cash crop of Maharashtra. It is also grown in commercially in other state of india viz. Karnataka, Andhra Pradesh, Rajasthan, Gujarat, and Tamil Nadu (Manjula et al., 2009). India is one of the major producers of pomegranate in the world with average total production of 8 lakh tons per annum. 

Many diseases occurred on the pomegranate plant out of them bacterial disease caused by the *Xanthomonas axonopodis* pv. *punicae* (Xap) strongly damaged the pomegranate production, including large economic losses to the Indian growers (Mondal et al. 2009). Bacterial disease cause substantial loss to the productivity of major crop plants. Unlike fungal disease, bacterial disease not control effectively through chemical methods.

The symptoms of the bacterial blight were observed on the leaves stems and fruits as small (2 to 5 mm) in size, irregular, prominent water soaked spots, which later become black often leads to the breaking of branches and cracking of fruits, reducing to the market value of the fruit (Rangaswamy, 1962; Manjula and khan 2002).

It is gram negative, aerobic rods belonging to the family *Pseudomonodaceae*. The pathogen causes disease actively at high temperature (up to 32°C) and high rainfall and humidity and p<sup>H</sup> at 7.0. The pathogen causes destruction of seedlings and adult plants and thus results in yield reduction. The aim of the study was to screen *in vitro* a range of antimicrobial agents and their efficacy against Xap and protein profiling; in laboratory conditions.

## **Material and methods**

### **Collection and isolation of disease sample:**

The samples were collected from two different districts of Maharashtra viz. Ahmednagar and pune as per typical symptoms showed by plant externally. The fresh infected leaves samples were used for isolation employing tissue isolation method by using Nutrient Agar medium (Antre et.al. (2015)).

### **Purification and Maintenance of bacterial culture:**

The respective bacterial cultures were maintained on NA medium at room temperature by adopting subsequent subculturing at periodical, regular intervals. Three days old cultures were used for further studies (Kanwar et.al. (1976)).

### **Identification of the pathogen:**

The identification of the pathogen involved in causing of oily spot in pomegranate was determined by conducting studies on its morphological (Ryan *et.al.* (2011); Kishun R *et al.* (1993)) and biochemical (Duche *et.al.*(2015); Giri *et.al.*(2011)) features of the pathogen as per standard microbiological procedures.

### **Pathogenicity test:**

Veinlets of some leaves were injected with 48 h old bacterial suspension with the help of sterile Dispo Van syringe. Some plant leaves were inoculated by injuring the leaves with sharp needle and by cotton swabbing the bacterial suspension on injured leaves. The plant was kept for incubation. After 10 – 12 days observations made for typical symptoms of bacterial blight on leaves, the organism was reisolated from artificially inoculated leaves of Pomegranate plants showing typical symptoms of disease. The re-isolation was carried out on NAS medium and the growth of organism was observed for next 2-3 days (Popovic *et al.* (2013); Abdo-Hasan *et al.* (2008)).

### ***In vitro* efficacy of different chemicals and plant derived oils against Xap by using paper disc and turbidimetric method:**

In vitro efficacy of different chemicals and oils was checked against *Xanthomonas axonopodis* pv. *Punicae* by using paper disc and turbidimetric method (Copper oxy chloride: 2500ppm, 1000ppm, Bronopol: 3000ppm, 1000ppm, Bordeaux mixture: 3000ppm, 1000ppm, Bactomycin: 1200ppm, 1000ppm, Clove oil: 100%, Nilgiri oil: 100%, Neem oil: 100%). (Kizil et al.(2005), Pawar et al. (2014)).

#### **Isolation of protein from healthy and infected pomegranate leaves:**

Kumar et al. (2015) standardized the protein isolation protocol from plant leaves.

#### **Isolation of protein from *Xanthomonas axonopodis* pv. *punicae*:**

Bioera's teaching kit (2017) described the protein extraction from bacteria and clear result was observed.



#### **Results and discussion:**

##### **Collection and isolation of diseased sample:**

Bacterial blight infected samples were collected from Ahmednagar and Pune districts of Maharashtra, associated bacterium *Xanthomonas axonopodis* pv.*punicae* was isolated. Purified culture was maintained on NSA slants and coded as Xap I, Xap II respectively. (Bradbury et al. (1970)).

##### **Identification of the pathogen:**

Morphological and biochemical characteristic viz., shape, colony colour, Gram reaction, starch hydrolysis, gelatin liquefaction, indole production, acid and gas production, KOH, catalase test were performed and confirmed to identity the bacterium as *Xanthomonas axonopodis* pv. *punicae* and following table 1 showed the morphological and biochemical test. (Gamangatti et al. (2013),Bhardwaj et al. (2014)).

##### **Pathogenicity test:**

Pathogenicity test of isolates was confirmed by injecting bacterial solution to disease free pomegranate plant. Occurrence symptoms of bacterial blight symptoms on plant leaf confirmed pathogenicity test of isolates (Fig. 1). Bacterium was reisolated from infected plant to prove Koch's postulates. (Yenjerappa et al. (2009)).

96 ***In vitro* efficacy of different chemicals and plant derived oils against Xap by using paper**  
 97 **disc method:**

98 In order to assess the efficacy of chemicals and oils against two isolates of  
 99 *Xanthomonas axonopodis* pv. *Punicae* an experiment was conducted and the  
 100 evaluation was made by paper disc method. The data presented in (Table 2) revealed  
 101 the significant differences among the different treatment. In the chemical treatment  
 102 Bronopol were significantly superior in inhibiting the growth of bacteria ( $18.6 \pm 0.7$   
 103 mm,  $25.6 \pm 1.6$  mm) against *Xap*. Among the oils Clove oil showed maximum zone of  
 104 inhibition  $18.3 \pm 0.7$  mm (Fig. 2). (Meerts and Fadavi et al. (2009)).

105 ***In vitro* efficacy of different chemicals and plant derived oils against Xap by using**  
 106 **turbidimetric method:**

107 Efficacy different chemicals and oils was tested turbidometrically. After 24 hours of  
 108 incubation list O.D. 0.713 nm was exhibited by Bronopol(3000 ppm), followed by Copper  
 109 oxy chloride (3000 ppm) with O.D. 0.908 nm and Bordeaux (3000 ppm) 0.834 nm  
 110 respectively whereas Bronopol (1000 ppm) with O.D 0.219 nm showed minimum inhibition  
 111 of growth and Among the oils tested Neem oil showed maximum inhibition of growth with  
 112 O.D. 0.842 nm, followed by Clove oil with O.D. 0.810 nm, Nilgiri oil showed minimum  
 113 inhibition of growth among oils showed in following table 3.

114 **Protein isolation from Healthy pomegranate leaves, infected pomegranate leaves and**  
 115 ***Xap I, Xap II* Bacteria:**

116 **Data analysis of Healthy pomegranate leaves, infected pomegranate leaves and**  
 117 **Bacteria:**

118 Following data analysis table shows the actual position of bands on the gel as  
 119 compared to the standard marker on the basis of molecular weights provided by BioEra's  
 120 Teaching Kit.

121 . The protein banding pattern showed the unique band no. 2 (Mol.Wt.66000 Da) in  
 122 infected *P. granatum L.* as compared to the banding pattern of *X. axonopodis* pv.*punicae* and  
 123 healthy *P. granatum L.* (Fig. 3) (Walker J.M. (2005)).

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## 126 Conclusion:

127 In the current study the pathogen infected samples identified as infected with *XAP*  
 128 was through morphological, biochemical characterization and Pathogenicity test. Efficacy of  
 129 different chemicals and oils were tested by disc diffusion assay and turbidometrically.  
 130 Bronopol 3000 ppm ( $25.6 \pm 1.6$  mm) formed highest zone of inhibition and Clove oil formed  
 131 highest zone of inhibition ( $18.0 \pm 0.7$  mm). Turbidometri showed the highest O.D. (0.908 nm)  
 132 by Copper oxy chloride and Neem oil showed maximum inhibition of growth with O.D.  
 133 (0.842 nm).

134 Biotic stress (pathogen) induced protein response was studies by using SDS-PAGE  
 135 method after protein extraction from *XAP*, healthy *P. granatum L.* and infected *P. granatum*  
 136 *L.* The protein banding pattern showed the unique band no. 2 (Mol.Wt.66000 Da) in infected  
 137 *P. granatum L.* as compared to the banding pattern of *X. axonopodis pv. punicae* and healthy  
 138 *P. granatum L.* The overexpressed protein due to biotic stress could be useful as a marker for  
 139 detection of the disease at the early stage and for control of the diseases after knowing the  
 140 biochemical significance of the protein.

141 **Table1. Morphological and Biochemical characteristics of *Xanthomonas axonopodis***  
 142 ***pv. punicae* isolates.**

Sr. No.	Morphological and Biochemical test	<i>Xap I</i>	<i>Xap II</i>
1.	Colony color	Yellow	Yellow
2.	Gram's reaction	-ve	-ve
3.	Starch hydrolysis	+++	++
4.	Indole production	++	+
5.	Catalase test	+++	+++
6.	KOH test	++	+
7.	Gelatin liquefaction	++	++
8.	Acid production	++	+

144 - Negative, + Positive, Varying degree of + ve reaction (+ poor, ++ moderate, +++ strong)

145 **Table 2 revealed the significant differences among the different treatment by paper disc**  
 146 **method.**

Sr. No.	Treatment	Concentration	Zone of inhibition produced by Xap I	Zone of inhibition produced by Xap II
T <sub>0</sub>	Control(+ve) water	10µl	00	00
T <sub>1</sub>	Control(-ve) Ampicillin	15µl	17.9±0.6	27.7±0.7
T <sub>2</sub>	Bronopol	1000 ppm	10.8±0.4	18.6±0.7
T <sub>3</sub>	Bordeaux	1000 ppm	10.2±0.2	9.3±0.1
T <sub>4</sub>	COC	1000 ppm	12.1±0.3	11.9±1.5
T <sub>5</sub>	Bactomycin	1000 ppm	10.1±0.3	13.2±0.9
T <sub>6</sub>	Bronopol	3000 ppm	11.6±0.4	25.6±1.6
T <sub>7</sub>	Bordeaux	3000 ppm	9.7±0.7	8.1±.02
T <sub>8</sub>	COC	2500 ppm	7.1±1.0	9.2±0.4
T <sub>9</sub>	Bactomycin	1200 ppm	11.2±0.4	15.3±0.2
T <sub>10</sub>	Clove oil	100%	15.9±0.9	18.3±0.7
T <sub>11</sub>	Neem oil	100%	13.4±1.6	7.3±1.1
T <sub>12</sub>	Nilgiri oil	100%	8.3±0.9	10.6±0.4

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148 **Table 3 Efficacy of chemicals and oils against *Xanthomonas axonopodis pv***  
 149 ***punicae* by turbidometric method:**

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Sr.No.	Treatment	Concentration	O.D. of culture	O.D. after 24 hrs at 620 nm
T <sub>0</sub>	Control(+ve)Nutrient Broth	1 ml		00
T <sub>1</sub>	Control (-ve) antibiotic	500 µl		0.324
T <sub>2</sub>	Bronopol	1000 ppm		0.219
T <sub>3</sub>	Bordeaux	1000 ppm		0.337
T <sub>4</sub>	COC	1000 ppm		0.410

T <sub>5</sub>	Bronopol	3000 ppm	0.998	0.713
T <sub>6</sub>	Bordeaux	3000 ppm		0.834
T <sub>7</sub>	COC	3000 ppm		0.908
T <sub>8</sub>	Clove oil	100%		0.810
T <sub>9</sub>	Neem oil	100%		0.842
T <sub>10</sub>	Nilgiri oil	100%		0.392

151 **Table4. Data analysis of Healthy pomegranate leaves, infected pomegranate leaves and**  
152 **bacteria**

Sr. No.	Standard Marker (Band no)	Molecular weight	Healthy plant (Band no)	Infected plant (Band no)	<i>Xap I</i> proteins (Band no)	<i>Xap II</i> proteins (Band no)
1.	1	97,400	Absent	Absent	Absent	Absent
2.	2	66,000	Absent	<b>Present</b>	Absent	Absent
3.	3	43,000	Present	Present	Absent	Absent
4.	4	29,000	Present	Present	Absent	Absent
5.	5	20,100	Present	Present	Present	Present
6.	6	14,300	Present	Present	Present	Present

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154 **Fig. 1 (a) Symptoms of bacterial blight observed after 10-12 days of inoculation (b)**  
155 **Reisolated *Xanthomonas axonopodis* pv. *punicae* after pathogenicity test**

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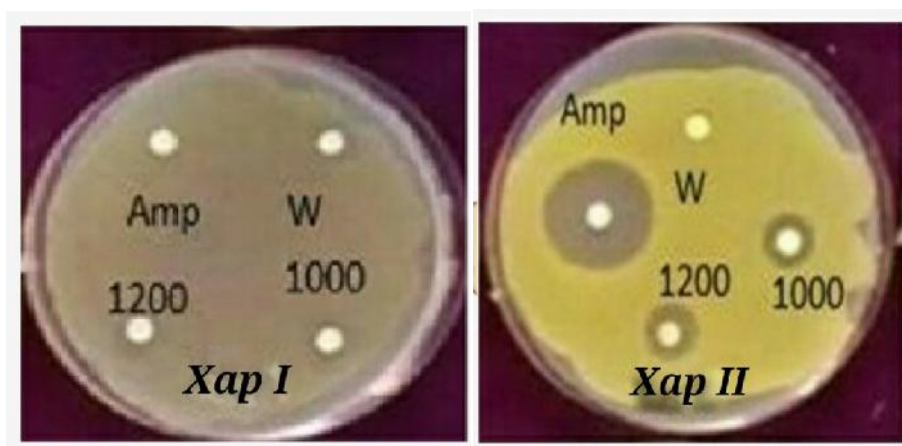
(a)



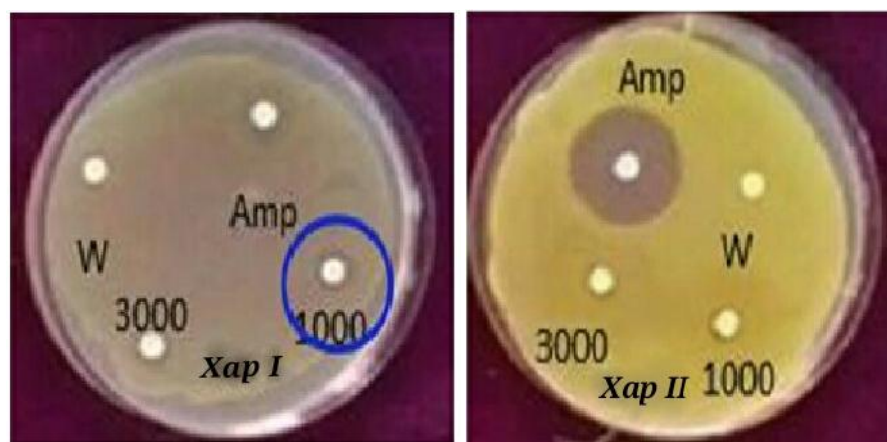


(b)

**Fig. 2 Efficacy of different chemicals and oils against *Xanthomonas axonopodis* pv. *Punicae* by Paper disc method.**

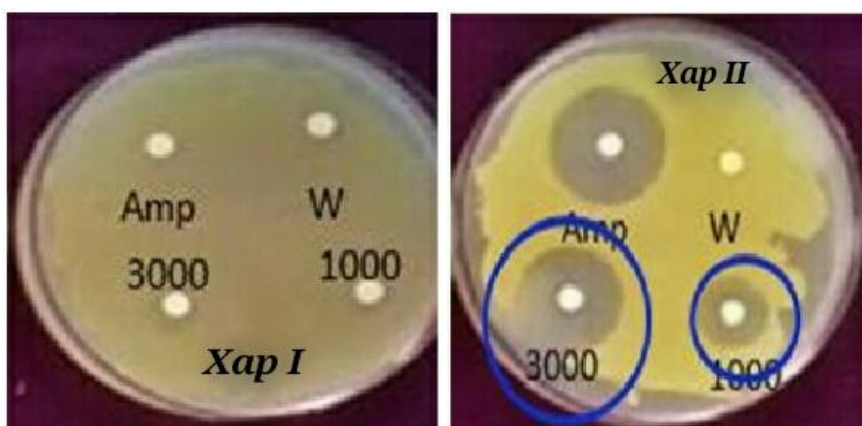


a) Bactomycin



b) Bordeaux

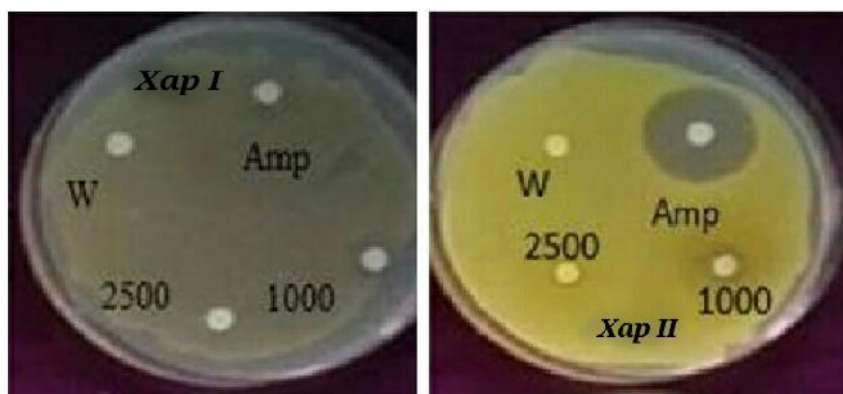




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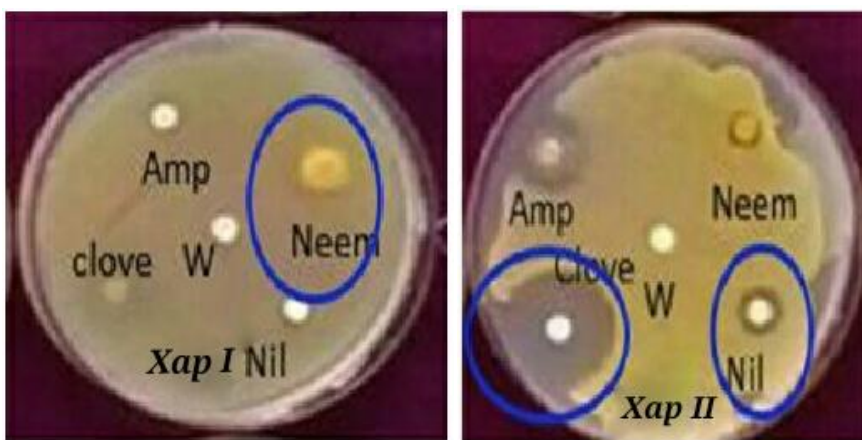
c) Bronopol



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d) Copper oxychloride



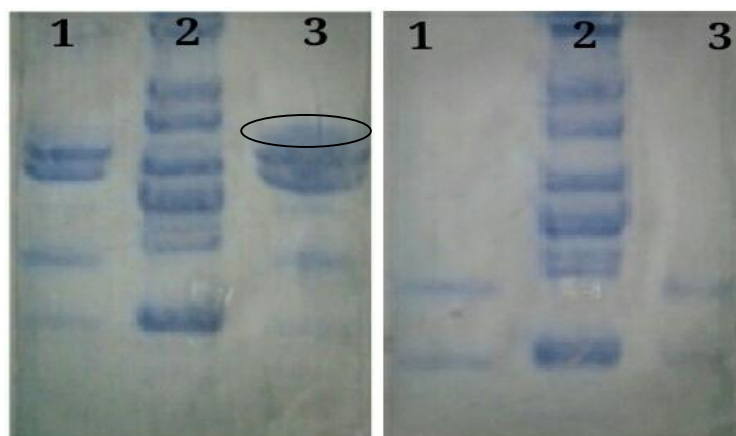
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e) Oils

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175 **Fig 3(a) Isolated proteins of 1) Healthy, 2) Marker, 3) Infected leaves (b) 1) *Xap I*, 2)**  
 176 **Marker and 3) *XapII* Bacteria (*Xanthomonas axonopodis* pv.*punicae*).**



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