

# Original Research Article

## PHYTOCHEMICAL SCREENING AND COMPARATIVE ANTIOXIDANT ACTIVITIES OF ~~DIFFERENT~~ FRACTIONS OF ~~ISOLATED FROM~~ SONNERATIA CASEOLARIS (LINN.) BARKS EXTRACTS.

### ABSTRACT

Aims: Our study was carried out to appraise the phytochemical screening and antioxidant potentials of ~~different~~ fractions of *Sonneratia caseolaris* (Linn.) barks extracts.

Study design: For the purpose of this experiment the extracts ~~were subjected~~ ~~were~~ ~~subjected~~ for an *in-vitro* study.

Place and Duration of Study: The study was carried out in August 2014 in the Department of Pharmacy, Southeast University, Dhaka, Bangladesh.

METHODOLOGY : The various fractions of *Sonneratia caseolaris* (Linn.) barks ~~extracts~~ as Ethanolic ~~fractions~~ (ETF), ethyl acetate (EAF), chloroform (CLF) and pet ether (PTF) fractions ~~—were~~ obtained after extraction were subjected to preliminary phytochemical screening. The antioxidant capacity of ~~Ethanolic fractions (ETF), ethyl acetate (EAF), chloroform (CLF) and pet ether (PTF) extracts of barks of S. caseolaris~~ ~~these fractions~~ were evaluated using 1, 1-diphenyl-2-picryl-hydrazyl (DPPH) free radical scavenging assay .Total antioxidant activity and ~~Total total~~ phenolic content of Ethanolic fractions (ETF), ethyl acetate (EAF), chloroform (CLF)

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and pet ether (PTF) extracts of *S. caseolaris* were determined.

RESULTS: The phytochemical screening showed the presence of ~~Flavonoid~~flavonoid, ~~Steroid~~steroid, ~~Tannin~~tannin compounds in large amounts. In DPPH scavenging assay, among the extracts, ~~Ethanolie~~ethanolic fractions (ETF) exhibited the highest ~~DPPH-radical~~scavenging activity with IC<sub>50</sub> of 4.57 µg /ml .The highest phenolic content was found in EAF extracts (63.00 mg of GAE/-g~~m~~. of dried extract) followed by CLF (36.25 mg of GAE/-g~~m~~. of dried extract) and PTF (26.28 mg of GAE/-g~~m~~. of dried extract). The highest total antioxidant activity was also found in ETF fraction (185 GAE/g~~m~~ of dried sample followed by EAF fraction (99.00GAE/g~~m~~ of dried sample), PTF (84.00 GAE/g~~m~~ of dried sample) and Chloroform (49.00 GAE/gm of dried sample).

CONCLUSION: Our result demonstrates that all the extractives of *S. caseolaris* have appreciable antioxidant ~~activities~~.

11  
12 **KEYWORDS:** *Sonneratia caseolaris* , DPPH, Total antioxidant activity, Total phenolic  
13 content.

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## 16 1. INTRODUCTION

17 *Sonneratia caseolaris* (L.) (Sonneratiaceae) is ~~such~~ a mangrove plant found widespread in  
18 tropical and subtropical tideland. *S. caseolaris* is a medium-size plant (2-to\_20\_m  
19 ~~high~~height), evergreen tree with elliptic-oblong leaves (5-to\_9.5cm long) [1-2]. *S. caseolaris*  
20 is reported to have 24 compounds such as ~~nine triterpenoids, eight~~nine triterpenoids, eight  
21 steroids, three flavonoids and four benzene carboxylic derivatives have been isolated from  
22 stems and twigs of medicinal mangrove plant *S. caseolaris* [3]. This plant contains phenolic  
23 compound like gallic acid and flavonoids e.g. luteolin and luteolin-7-O-glucoside [4]. It  
24 contains alkaloid, tannin, flavonoid, saponin, phytosterol, and carbohydrate[5-6].*S.*  
25 *caseolaris* ~~te-beis~~ used in traditional medicine systems in several countries, it is used for  
26 sprains, swelling helminthiasis, poultices, coughs, hematuria, small pox, astringent,  
27 antiseptic, arresting hemorrhage, piles, and also used as remedy to stop blood bleeding [7].  
28 *S. caseolaris* possessed intestinal  $\alpha$ -glucosidase inhibitory property [8] and it has also been  
29 reported to be toxic against mosquito larvae [7].

30 | Oxidative stress ~~follow-on~~cause due to imbalance of oxidizing agents and natural  
31 | antioxidants in the body induces the brutality of a number of diseases like atherosclerosis,  
32 | cancer, cardiovascular ailments, neurodegenerative disorders and diabetes [9]. As self-  
33 | protective measure against such oxidative damages, biological systems have evolved a  
34 | range of enzymatic machineries and scavengers. These include dietary antioxidants ( $\alpha$ -  
35 | tocopherol,  $\beta$ -carotene, ascorbic acid, glutathione, uric acid), hormones (estrogen,  
36 | angiotensin), enzyme systems (superoxide dismutase, glutathione peroxidase, catalase).  
37 | ~~etc.~~[10-11]. A large ~~number~~ of antioxidative agents, both natural (e.g.  $\alpha$ -tocopherol) and  
38 | synthetic (e.g. butylated hydroxyanisole, butylated hydroxytoluene, tert-butyl hydroquinone  
39 | and propyl gallate) are broadly used in the food industry to lengthen shelf life as they inhibit  
40 | lipid oxidation [12]. However, the use of these synthetic antioxidants is increasingly getting  
41 | restricted because of their toxicity and health risks [13]. Therefore, discovery of novel  
42 | antioxidative of natural origin is the urgent need of the hour and plants can be a good source  
43 | for the purpose[12]. Earlier research focused on methanolic bark extracts to illustrate the  
44 | antioxidant activity of *S. caseolaris*. However, here we focus on comparative antioxidant  
45 | activities of different ~~fractions-offractions of~~ *Sonneratia caseolaris* (Linn.) barks extracts.

46 |

## 47 | 2. METHODS

### 48 | 2.1 Collection, identification and preparation of plant material

49 | The stems were harvested after identification by an expert taxonomist from ~~the plant growing~~  
50 | at Barisal on August 5, 2014. The stems were dried under shade at room temperature for a  
51 | period of two weeks in order to avoid solar radiations from altering the API. These stems  
52 | were spread on plastic bags while avoiding their stacking. ~~And Every every~~ day ~~we turned~~  
53 | these stems were mixed upside down so that to favor a homogenous drying process. The  
54 | dried leaves were ground in a clean electric grinding machine in such a way to obtain a fined  
55 | powder, which was stored in an airtight container. The total dried powder material was  
56 | obtained 600 gm. It was divided equally into four portions and was refluxed with ~~ethanol~~

57 | ~~ethylethanol, ethyl~~ acetate, pet ether and chloroform solvent for three times. The extract  
58 | was filtered with Whiteman No. 1. filtered paper and the collected filtrate was evaporated in  
59 | an oven at 50°C. This extract was weighed so that to determine the yield obtained from the  
60 | initial powder quantity and then stored in an air-tight container for subsequent experimental  
61 | tests.

62 |

### 63 | **2.2.1 Phytochemical screening**

64 | Phytochemical screening of the stems extracts of *S.caseolaris* ~~were tested~~ were tested for  
65 | the presence of active principles such as alkaloids, flavonoids, tannins, reducing sugar ~~etc.~~  
66 | using the standard procedures.

67 |

#### 68 | **Test for saponin:**

69 | About 2 g of the powdered sample was boiled in 20 ml of distilled water in a water bath and  
70 | filtered. About 10 ml of the filtrate was mixed with 5 ml of distilled water and shaken  
71 | vigorously for a stable persistent froth. The frothing was mixed with 3 drops of olive oil and  
72 | shaken vigorously, then observed for the formation of emulsion.

73 |

#### 74 | **Test for saponins (Kokate, 1999):-**

75 | The extract was diluted with distilled water and made up to 20 ml. The suspension was  
76 | shaken in a graduated cylinder for 15 min. ~~and a~~ 2 cm layer of foam indicates the presence  
77 | of saponins.

78 |

#### 79 | **Test for Tannins:**

80 | About 2.5 g of the plant extract was dissolved in 5 ml of distilled water, filtered and ferric  
81 | chloride reagent added to the filtrate. A blue-black, green, or blue-green precipitate was  
82 | taken as evidence for the presence of tannins (Trease and Evans, 1989).

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84 **Test for Flavonoid**

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85 The presence of flavonoids in the samples was determined using the methods [14]. About 10  
86 ml ~~Ethyl-ethyl~~ acetate ~~was added~~was added to 0.2 g of the powdered sample and heated in  
87 a water bath for 5 min. The mixture was ~~cooled~~,cooled, filtered and the filtrates used for the  
88 test.

89  
90 **Ammonium test:**

91 About 4 ml filtrate ~~is was~~ shaken with 1 ml of dilute ammonia solution. The layer ~~is was~~  
92 allowed to separate and the yellow color in the ammoniacal layer indicates the presence of  
93 flavonoids.

94  
95 **Aluminum chloride solution test:**

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96 Approximately, 1 ml of 1% aluminum chloride solution ~~is was~~ added to 4 ml of the filtrate and  
97 shaken. A yellow coloration indicates the presence of flavonoids.

98  
99 **Test for alkaloids**

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100 Mayer's test (Evans, 1997): To a few ml of the filtrates, a drop of Mayer's reagent was added  
101 by the side of the test tube. A creamy or white precipitate indicates the test is positive.

102  
103 **Test for carbohydrates**

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104 In Benedict's test: ~~To~~, 0.5 ml of the filtrate, 0.5 ml of Benedict's reagent was added. The  
105 mixture was heated on boiling water bath for 2 min. A characteristic red colored precipitate  
106 indicates the presence of sugar. [15]

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## 111 **Test for steroids**

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112 Two ~~ml~~ milliliters of acetic anhydride was added to 0.5 g of extracts of each sample with 2  
113 ml H<sub>2</sub>SO<sub>4</sub>. The colour changed from violet to blue or green in some samples indicating the  
114 presence of steroids. [15]

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## 116 **2.2 Evaluation of Antioxidant ~~activity~~Activity**

### 117 **2.2.1 Estimation of ~~Total total~~ Phenolic-phenolic content**

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118 The Folin-Ciocalteu's reagent was used as oxidizing agent and test-using gallic acid as  
119 standard, the total phenolic content of extractives of *S. caseolaris* was described by Singleton  
120 *et al* [16] with some modifications. The assay mixture consisted of extract (0.5 ml that was  
121 adjusted to 1.0 ml with distilled water) and 2.5 ml of Folin-Ciocalteu's reagent. Furthermore,  
122 after incubation at room temperature for 15 min, 2.5 ml of (w/w) Na<sub>2</sub>CO<sub>3</sub> solution was added  
123 into the test tube and the test tube was incubated at the same temperature for 20 minutes.  
124 Finally, the absorbance was read at 760 nm against reagent blank. However, the methanol  
125 extract and in different fractionates in Gallic acid equivalents (GAE) was calculated by the  
126 according to the formula.

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$$127 C = (c \times V)/m$$

128 Where,

129 C = total content of phenolic compounds, mg/g plant extract, in GAE; c= the concentration of  
130 Gallic acid established from the calibration curve, mg/ml; V = the volume of extract, ml; m =  
131 the weight of different pure plant extracts, gm.

### 133 **2.2.2 Estimation of ~~Total total~~ antioxidant capacity**

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134 Catechin reagent was used as a standard; the total antioxidant capacity of extractives of *S.*  
135 *caseolaris* was determined by the method of ~~of~~ Prieto *et al* [17].with slight modifications. The  
136 ~~experiment~~ mixture consisted of extracts (0.5 ml standard or plant extract solution) was

137 taken in a test tube with 3 ml of reaction mixture containing 0.6 M sulphuric acid, 28 mM  
 138 sodium phosphate and 1% ammonium molybdate was added into the test tube. In addition,  
 139 after incubation at 95°C for 10 minutes, the absorbance of the solution was read at 695 nm  
 140 against reagent bank using a spectrophotometer. The experiment was done three times at  
 141 each concentration.

### 142 2.2.3 DPPH Radical scavenging Assay

143 The 1, 1- diphenyl-2-picrylhydrazyl (DPPH) free radical scavenging activity was evaluated  
 144 with the help of according to the method used by Fresin [18]. using with modified procedure.

145 Test samples were prepared by dissolving 5 mg of dry extracts in 5 ml of methanol. The  
 146 assay mixture contained extract (0.5 ml) and DPPH (1.0 ml) which were mixed well and  
 147 incubated in the dark for 30 minutes. The blank was prepared and made to contain methanol  
 148 (0.5 ml) and DPPH (1.0 ml). The absorbance was measured at 517 nm on a visible  
 149 spectrophotometer. All experiments were performed in triplicate. DPPH radical activity was

150 calculated by the following equation.  $\% \text{Percentage Inhibition} = \{(A_0 - A_1)/A_0\} \times 100$

151 Where,  $A_0$  is the absorbance of the control, and  $A_1$  is the absorbance of the extract/standard.

152 In addition, % inhibitions were plotted against concentration and from the graph  $IC_{50}$  was  
 153 calculated.

### 155 3. RESULTS

156 **Table 1.** Phytochemical test results of different extractives of *S. caseolaris*.

Phytochemical tests	Crude methanol extract	Ethanol fraction	Chloroform fraction	Petroleum Ether fraction	Ethyl acetate fraction
Saponin	++	+	-	-	+
Tannin	+++	+++	++	++	++
Flavonoid	+++	+++	++	++	++
Alkaloid	++	+	-	-	+
Carbohydrate	++	++	-	+	-
Steroid	+++	++	++	+	+

157 + = Present in mild amount, ++ = Present in moderate amount, +++ = Present in large amount, - = Not  
 158 present  
 159

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160 **3.1 Determination of Total total Phenolicsphenolics**

161 The results were expressed as mg of Galic-gallic Acid-acid Equivalent equivalent (GAE)/gm  
162 of dried extractives. Among the fractions the highest phenolic content was found in EAO  
163 fractions -(63.00 mg of GAE-/gm of dried extract) followed by ETF (60.25 mg of GAE-/gm of  
164 dried extract), CLF (36.25 mg of GAE-/gm of dried extract) and PET (26.28 mg of GAE-/gm  
165 of dried extract). Comparing the phenolic content of different fractions of *S. caseolaris* it was  
166 observed that EAO contains considerable amount of phenolic compounds than the other  
167 extracts. However, phenolic content of the samples were calculated on the basis of the  
168 standard curve for gallic acid as shown in table-Table 2 and in figure-Figure 01.

169

170 **Table 2:** Determination of total phenolic content of different fractions of *S. caseolaris*.

<u>SampleFraction</u>	<u>Conc. (µg/ml)</u>	<u>Absorbance</u>	<u>GAE/gm of dried sample</u>
Ethanol fraction	250	0.296	60.25
Chloroform fraction	250	0.25	36.25
Ethylacetate fraction	250	0.324	63.00
Pet-ether fraction	250	0.174	26.28

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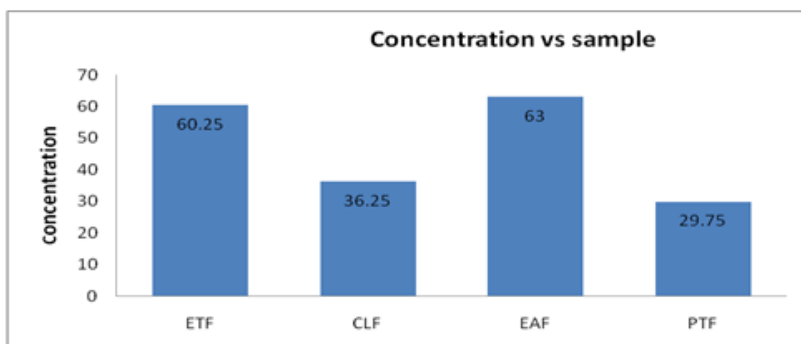
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**Figure 01:** Total phenolic content of different fractions of barks of *S. caseolaris*.

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Here, ETF = Ethanol fraction, CLF = Chloroform fraction, EAF = Ethyl acetate fraction, PTF = pet-ether fraction.

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### 3.2 Determination of total antioxidant activity

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Total antioxidant activity of four different solvents of crude extract such as ethanol (ETF),

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chloroform (CLF), ethyl acetate (EAF) and pet-ether fraction (PTE) were investigated.

183

Among the four different fractions ETF showed the highest total antioxidant activity with

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absorbance at 200 µg/ml concentration followed by EAF (absorbance of 0.388 at 200

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µg/ml), PTF (absorbance of 0.187 at 200 µg/ml) and CLF (absorbance of 0.166 at 200

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µg/ml). Our result demonstrates that all the extractives of *S. caseolaris* have appreciable

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total antioxidant activity. However, total antioxidant activity of plant extracts and (+)-catechin

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(standard) were depicted in [table-Table 03](#) and [04](#) and in [figure-Figures 02](#) and [03](#).

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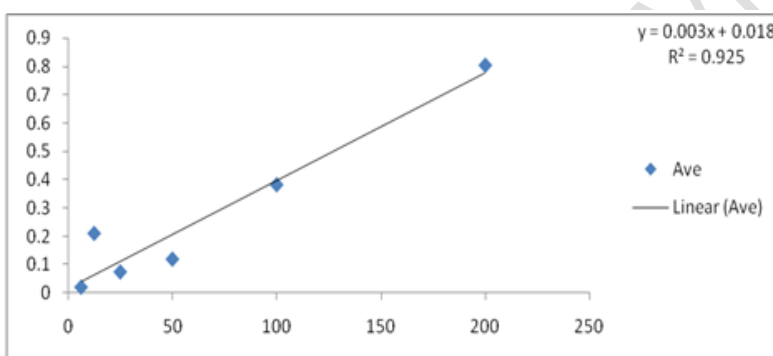
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195 **Table 03:** Absorbance of catechin (standard) at different concentrations for  
 196 determination of total antioxidant activity.

Name of sample	Concentration (µg/ml)	Absorbance			Absorbance Mean ±STD
		a	b	c	
(+) - Catechin	6.25	0.018	0.019	0.021	0.019 ± 0.0015
	12.5	0.207	0.211	0.209	0.209 ± 0.002
	25	0.037	0.039	0.035	0.037 ± 0.002
	50	0.118	0.119	0.116	0.117 ± 0.001
	100	0.380	0.383	0.379	0.381 ± 0.002
	200	0.803	0.801	0.805	0.803 ± 0.002

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199 **Figure 02:** Standard curve of catechin for the determination of total antioxidant capacity.

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201 **Table 04:** Determination of total antioxidant capacity of different solvent fractions of  
 202 crude ethanolic extract of *S. caseolaris*.

Sample Fraction	Conc. (µg/ml)	Absorbance	GAE/gm of dried sample
Ethanol fraction	200	0.388	185
Chloroform fraction	200	0.166	49.00
Ethyl acetate fraction	200	0.216	99.00
Pet-ether fraction	200	0.187	84.00

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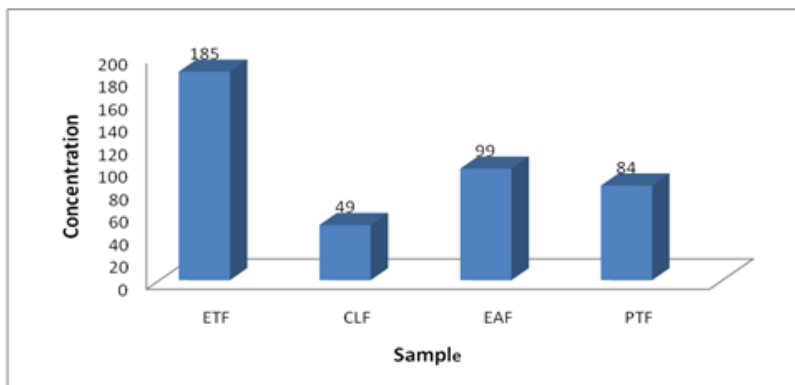
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210 **Figure 03:** Total antioxidant activity of different solvents fractions of the extracts of *S.*  
 211 *caseolaris*.

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### 3.3 Determination of DPPH radical scavenging activity

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DPPH radical scavenging activity of different fractions of solvents of ethanolic (ETF),

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chloroform (CLF), ethyl acetate (EAF) and pet-ether fraction (PTE) were investigated.

217

Among all extracts ethanol fraction (EAF) showed the highest DPPH radical scavenging

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activity with IC<sub>50</sub> value of 4.57 µg/ml and chloroform fraction ethanol fraction (EAF) showed

219

the lowest DPPH radical scavenging activity with IC<sub>50</sub> value of 197.27 µg/ml, respectively.

220

The results of DPPH radical scavenging assays of plant extracts and butylated

221

hydroxytoluene (BHT) (standard) are given in [table-Table 05](#) and in [figure-Figure 04](#).

222

223

224 **Table 05:** DPPH radical scavenging activity of different fractions of extracts of *S. caseolaris*  
 225 and BHT (Standard) at different concentrations.

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Name of Sample	Conc. (µg/ml)	Absorbance	% of scavenging	IC <sub>50</sub> (µg/ml)
BHT	200	0.073	94.45	3.25 µg/ml
	100	0.071	94.48	
	50	0.079	94.33	
	25	0.085	93.40	
	12.5	0.098	92.39	

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	6.25	0.147	88.58	
Ethanol fraction	200	0.085	93.40	4.57 µg/ml
	100	0.073	94.33	
	50	0.071	94.48	
	25	0.080	93.78	
	12.5	0.126	90.21	
	6.25	0.409	68.24	
Chloroform fraction	200	0.635	50.69	197.27 µg/ml
	100	1.038	19.40	
	50	0.675	47.59	
	25	0.707	45.10	
	12.5	0.935	27.40	
	6.25	0.689	46.50	
Ethyl acetate fraction	200	0.061	95.26	13.09 µg/ml
	100	0.228	82.29	
	50	0.432	66.45	
	25	0.555	56.90	
	12.5	0.673	47.74	
	6.25	0.697	45.85	
Pet-ether fraction	200	0.749	41.84	12.32 µg/ml
	100	0.637	51.47	
	50	0.698	45.80	
	25	0.742	42.39	
	12.5	0.635	50.69	
	6.25	0.524	59.31	

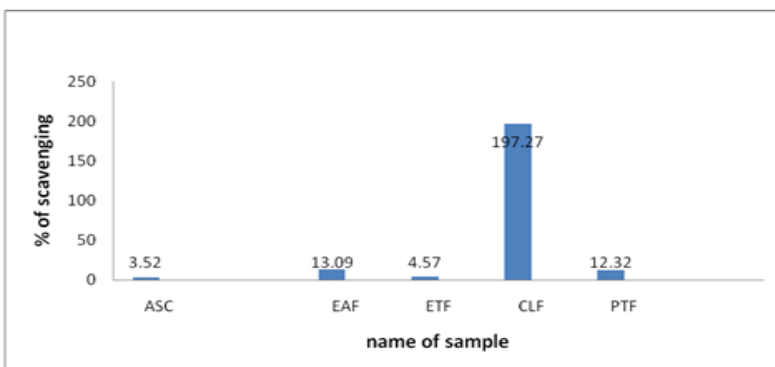
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**Figure 04:**  $IC_{50}$  ( $\mu\text{g/ml}$ ) values of different extractives of *S. caseolaris* for DPPH radical scavenging activity.

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#### 4. DISCUSSION

At maximum wavelength at 517 nm, The DPPH free radical can easily accept an electron or hydrogen from antioxidant molecules to develop into a stable diamagnetic molecule. Due to the DPPH radical's ability to bind hydrogen, it is considered to have a radical scavenging property. Discoloration occurs due to the decreasing quantity of DPPH radicals in the environment. The discoloration of the DPPH therefore reflects the radical scavenging activity of the analyzed extracts [19]. Based on the data obtained from this study, DPPH radical scavenging activity of *S. caseolaris* extract of chloroform fraction ( $IC_{50}$  4.57 $\mu\text{g/ml}$ ) was similar to that standard BHT ( $IC_{50}$  3.25  $\mu\text{g/ml}$ ).

Phenolic compounds have redox properties, which let them to act as antioxidants. [19]. Free radical scavenging ability is facilitated by their hydroxyl groups, the total phenolic concentration could be used as a basis for rapid screening of antioxidant activity. Among the fractions the highest phenolic content was found in EAF\_(63 mg of GAE / gm. of dried extract) and then ETF (60.25 of GAE / gm. of dried extract) ,CLF(36.25 mg of GAE / gm. of dried extract) and PTF(29.75 mg of GAE / gm. of dried extract)—. Comparing the phenolic

248 content of ETF, EAF, CLF and PTF extracts of *S. caseolaris* it was observed that ETF  
249 contains considerable amount of phenolic compounds than the other extracts.

250

251 The total antioxidant capacity (TAC) was based on the reduction of Mo(VI) to Mo(V) by the  
252 extract and subsequent formation of greenphosphate/Mo(V)complex at acid pH . It evaluates  
253 both water-soluble and fat-soluble antioxidants. Among the different extracts, Ethanol  
254 fraction showed the highest total antioxidant activity (185 GAE/gm of dried sample).

255

## 256 5. CONCLUSION

257 Our study investigation brings out the scientific rationale for the folkloric uses of the plant in  
258 the management of oxidative stress associated disorders. Even so, further research is  
259 needed towards isolation and ascertainment of active principles present in the extracts,  
260 which could possibly be explored for pharmaceutical use.

261

## 262 COMPETING INTERESTS

263 There are no competing interests.

## 264 CONSENT: NOT APPLICABLE

## 265 ETHICAL APPROVAL: NOT APPLICABLE

266

## 267 6. REFERENCES

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