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3 **CORRELATION OF SMALL DENSE LOW DENSITY LIPOPROTEIN, TUMOUR**

4 **NECROSIS FACTOR-ALPHA WITH LIVER ENZYMES IN CHRONIC HEPATITIS B**

5 **PATIENTS.**

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7 **ABSTRACT**

8 **Aim:** This study investigated the relationship between small dense low density lipoprotein (sdLDL),
9 tumour necrosis factor-alpha (TNF- α), aspartate aminotransferase (AST), alanine aminotransferase (ALT)
10 and alkaline phosphatase (ALP) in chronic hepatitis B patients.

11 **Subjects and methods:** Sixty (60) participants were recruited for this cross sectional study. They
12 comprise thirty (30) clinically diagnosed chronic hepatitis B virus (HBV) infected patients attending clinic
13 at a tertiary hospital in Osogbo, Osun sta, **country name**. Thirty (30) apparently healthy volunteers were
14 recruited as control subjects after fulfilling the inclusion criteria. Anthropometric measurements were
15 performed using standard method. About 6mL of venous blood was collected from each study
16 participants and serum was extracted and kept at -80°C until time of analysis. Small dense LDL, TNF- α ,
17 AST, ALT and ALP were determined using enzyme linked immunosorbent assay and colorimetric method
18 as appropriate. Data analysis was done obtained using Student's t-test for comparison of variables and
19 Pearson's correlation was used to determine the relationship between variables. *P*-value less than 0.05
20 was considered significant.

21 **Results:** SdLDL, TNF- α , AST and ALT were significantly elevated in HBV patients when compared with
22 the control subjects ($P < 0.05$). SdLDL had a significant positive correlation with TNF- α ($P = 0.03$), AST
23 ($P = 0.01$), ALT ($P = 0.00$). TNF- α had a significant positive correlation with AST ($P = 0.02$) and ALT
24 ($P = 0.00$).

25 Conclusion: This study revealed a noteworthy positive relationship between sdLDL, TNF- α and hepatic
26 aminotransferases in chronic hepatitis B patients.

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28 **1. INTRODUCTION**

29 Viral hepatitis is now recognized as a major public health challenge that requires an urgent response (1).
30 In 2015, about 325 million people were living with chronic hepatitis infections worldwide and it was
31 reported that approximately 1.34 million people died of hepatitis globally (2). This global mortality is
32 comparable to deaths caused by tuberculosis and human immunodeficiency virus (HIV). While deaths
33 resulting from tuberculosis and HIV appear to be declining, deaths from hepatitis are on the increase.

34 Hepatitis B viral infection has been described as one of the leading causes of mortality worldwide with
35 about 650,000 annual deaths (2). Hepatitis B viral infection poses a major threat to human health and it is
36 highly prevalent in developing countries (3). The prevalence of Hepatitis B infection is about 12% in
37 Nigeria (4).

38 Hepatitis B virus has the potential to affect the functional integrity of the liver of an infected host. Liver as
39 a homeostatic organ plays a pivotal role in lipid metabolism. Thus, the circulating levels of lipids in plasma
40 depend greatly on the functionality of the liver. In the setting of acute or chronic hepatic dysfunction
41 circulating lipids and lipoproteins are altered with respect to quantity as well as pattern of their
42 electrophoretic mobility (5).

43 Previous studies documented diverse reports about the alterations of serum lipids in patients suffering
44 from acute hepatitis due to the actions of hepatotropic viruses (6-8). Additionally, it was reported that
45 chronic HBV infection is associated with elevated levels of low density lipoprotein (LDL), which is known
46 to be a predictor of atherosclerotic cardiovascular disease risk (9-13).

47 Low-density lipoprotein consists of several subclasses of particles with different sizes and densities and
48 they include the large buoyant, intermediate and small dense (sd) LDL particles. It is well documented
49 that sdLDL cholesterol (sdLDL-C) proportion is a better marker for prediction of cardiovascular disease
50 than total LDL-C (14, 15).

51 Furthermore, activation of the immune response during viral hepatitis leads to the production of many pro-
52 inflammatory cytokines that act as mediators of disease activity (16). These pro-inflammatory cytokines
53 particularly interleukin-6 (IL-6), interleukin-1 (IL-1) and tumour necrosis factor-alpha (TNF- α) appear to
54 accentuate lipogenesis (17).

55 Even though previous studies have documented that viral hepatitis might interfere with lipid metabolism,
56 however, the link between sdLDL, TNF- α and liver enzymes in individuals with hepatitis B patients has
57 not been fully elucidated. The present study therefore aimed to determine the association between
58 sdLDL, TNF- α , AST, ALT and ALP in chronic hepatitis B patients visiting a tertiary hospital in the south-
59 western part of Nigeria.

60 **2. MATERIALS AND METHODS**

61 **2.1 Subject selection**

62 A total of sixty (60) subjects were recruited for this cross-sectional study. The test group comprise thirty
63 (30) clinically diagnosed chronic hepatitis B virus (HBV) infected patients attending clinic at the
64 Department of Gastroenterology, Ladoke Akintola University of Technology Teaching, Osogbo, Osun
65 state, **country name**. These patients continuously tested positive for HBsAg for more than one year during
66 their periodic visit to the clinic and they had one or more of these features; pallor, jaundice and liver
67 enlargement. The control group comprise thirty (30) age matched apparently healthy HBV seronegative
68 individuals.

69 A short structured questionnaire was administered to each study participant to obtain information on
70 demography, alcohol use, drug use, smoking habits, medications and established diseases. Persons
71 diagnosed with dyslipidemia and other metabolic conditions, record of alcoholism, smoking, usage of
72 medications that affect lipid status and pregnant women were excluded from this study.

73 **2.2 Ethical consideration**

74 All participants were recruited for this study after ethical clearance was obtained from the ethics
75 committee of Ladoke Akintola University of Teaching, Osogbo, Osun state. Written informed consent was
76 obtained from each participants.

77 **2.3 Blood pressure and anthropometric measurement**

78 The blood pressure was measured using mercury sphygmomanometer with appropriate cuff size.
79 Korotkoff phases 1 and 5 were used. Body weight in kilogram (kg) was measured using a standard
80 weighing scale and height (m) was measured using standiometer. Body mass index (BMI) was calculated
81 as the ratio of body weight (kg) to the square of height (m²).

82 **2.4 Sample collection and assay methodology**

83 About 6 milliliters (mL) of venous blood was collected from each participant and dispensed into plain
84 bottles to obtain serum which was aliquoted into a small vial and stored at -80°C until time of analysis for
85 the determination of small dense low density lipoprotein (sdLDL), tumour necrosis factor-alpha (TNF-α),
86 aspartate aminotransferase (AST), alanine aminotransferase (ALT) and alkaline phosphatase (ALP).

87 **2.4.1 Detection of Hepatitis B surface antigen (HBsAg).**

88 The serum samples of subjects were screened to detect the presence of hepatitis B surface antigen. Fifty
89 microliter (50µL) of serum was added vertically into the hole on the cassette and the result was read after
90 15 minutes (Melsin Medical Co., Limited, China). Appearance of two distinct red lines; one line at the
91 control region (C) and the other at the test region (T) indicated positive test. Whereas appearance of only
92 one red line, at the control region (C) indicated negative test. The result was invalid when the line at the
93 test region appeared but the control region failed to appear.

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95 **2.4.2 Determination of liver enzymes**

96 The serum activities of ALT and AST were determined colorimetrically using Randox reagents (Randox
97 Laboratories, UK) as described by Reitman and Frankel (18). The serum activities of ALP were
98 determined colorimetrically using Randox reagents (Randox Laboratories, UK) as described by
99 Gesellschaft (19).

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104 **2.4.3 Determination of sdLDL**

105 Small dense LDL (sdLDL) was analyzed based on the principle of solid phase enzyme linked
106 immunosorbent assay (ELISA) using a unique monoclonal antibody directed against distinct antigenic
107 determinant on sdLDL molecule immobilized on microtitre wells with kits supplied by ElabScience,
108 Biotech, Ltd (USA). The standard working solution, biotinylated detection ab working solution and HRP
109 conjugate working solution were prepared according to the manufacturer's instruction.

110 One hundred microliter (100 μ L) of sdLDL standards, samples and controls were added to appropriate
111 wells. One hundred microliter (100 μ L) of biotinylated detection ab, solution was added to each well, they
112 were mixed thoroughly and then incubated at 37 $^{\circ}$ C for 60 minutes, after which the wells were washed 3
113 times. One hundred microliter (100 μ L) of HRP conjugate was then added to each well and was incubated
114 at 37 $^{\circ}$ C for 30 minutes after which the wells were washed 5 times. Ninety microliter (90 μ L) of substrate
115 reagent was added to each well and was incubated at 37 $^{\circ}$ C for 15 minutes after which 50 μ L of stop
116 solution was added to each well to stop the reaction. Absorbance was read at 450nm with a microtitre
117 well reader. The grades of standard were used to plot a curve of absorbance against concentration for the
118 calculation of sdLDL concentration.

119 **2.4.4 Determination of TNF-alpha**

120 Tumour necrosis factor-alpha(TNF-alpha) was analyzed based on the principle of solid phase enzyme
121 linked immunosorbent assay (ELISA) using a unique monoclonal antibody directed against distinct
122 antigenic determinant on TNF-alpha molecule immobilized on microtitre wells with kits supplied by
123 ElabScience, Biotech, Ltd (USA). The standard working solution, biotinylated detection ab working
124 solution and HRP conjugate working solution were prepared according to the manufacturer's instruction.

125 One hundred microliter (100 μ L) of TNF-alpha standards, samples and controls were added to appropriate
126 wells. One hundred microliter (100 μ L) of biotinylated detection ab. solution was added to each well, they
127 were mixed thoroughly and then incubated at 37 $^{\circ}$ C for 60 minutes, after which the wells were washed 3
128 times. One hundred microliter (100 μ L) of HRP conjugate was then added to each well and was incubated

129 at 37°C for 30 minutes after which the wells were washed 5 times. Ninety microliter (90µL) of substrate
130 reagent was added to each well and was incubated at 37°C for 15 minutes after which 50µL of stop
131 solution was added to each well to stop the reaction. Absorbance was read at 450nm with a microtitre
132 well reader. The grades of standard were used to plot a curve of absorbance against concentration for the
133 calculation of TNF-alpha concentration.

134 **2.5 Statistical analysis**

135 Data analysis was done using SPSS version 21.0. All values were expressed as mean±standard
136 deviation for test and control groups. Comparison of variables was done using Student's t-test and
137 Pearson's correlation was used to determine the relationship between variables. $P < 0.05$ was considered
138 to be statistically significant.

139 **3. RESULTS**

140 The demographic data, anthropometric and biochemical parameters of the study participants are
141 summarized in table 1. The mean age, BMI and blood pressure of the case and control subjects were not
142 statistically significant ($P > 0.05$). The mean levels of sdLDL, TNF-α and mean activities of AST, ALT were
143 significantly elevated in hepatitis B patients compared with control. ($P < 0.05$).

144 Table 2 shows the correlation between sdLDL and other biochemical parameters in HBV patients. Small
145 dense LDL had significant positive correlation with TNF-α ($P=0.03$), AST ($P=0.01$) and ALT ($P=0.00$).
146 There was also positive correlation with ALP but not significant ($P > 0.05$)

147 Table 3 shows the correlation between TNF-α, AST, ALT and ALP. TNF-α had significant positive
148 correlation with AST ($P=0.02$) and ALT ($P=0.00$) but not with ALP ($P > 0.05$)

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154 **Table 1. Demographic, anthropometric and biochemical parameters of the study participants**

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Parameters	HBV	Control	P-value
Age (years)	35.6±8.7	32.3±6.4	0.64
BMI (kg/m ²)	23.7±3.9	24.8±4.3	0.14
SBP (mmHg)	128.4±12.5	125.2±8.4	0.34
DBP (mmHg)	78.2±7.4	75.6±5.9	0.22
AST (IU/L)	56.8±33.5	28.2±12.5	0.00*
ALT (IU/L)	46.2±23.2	21.1±14.3	0.00*
ALP (IU/L)	65.3±23.7	58.2±16.8	0.16
SdLDL(nmol/mL)	67.9±23.8	29.8±15.9	0.00*
TNF-α (pg/mL)	29.2±13.5	15.7±10.5	0.01*

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157 *Statistically significant at $P < 0.05$. Results are expressed as mean±standard deviation. BMI-Body mass
 158 index; SBP-systolic blood pressure; DBP- diastolic blood pressure; AST-aspartate aminotransferase;
 159 ALT- alanine aminotransferase; ALP-alkaline phosphatase; sdLDL-small dense low density lipoprotein;
 160 TNF-α- tumour necrosis factor-alpha.

161 **Table 2: Correlation between sdLDL and other biochemical parameters.**

SdLDL	R	P-value
AST	0.929	0.01*
ALT	0.745	0.00*
ALP	0.294	0.162
TNF-α	0.813	0.03*

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163 *Statistically significant at $P<0.05$ (2-tailed)

164 **Table 3: Correlation between TNF- α and liver enzymes.**

TNF- α	R	P-value
AST	0.835	0.02*
ALT	0.665	0.00*
ALP	0.440	0.146

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166 *Statistically significant at $P<0.05$ (2-tailed)

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183 **4. DISCUSSION**

184 Liver is an important homeostatic organ that is mainly responsible for the synthesis of lipids. Moreover the
185 synthesis of key enzymes for lipid metabolism takes place in the liver (20). Liver also regulates the
186 catabolism of various plasma lipoproteins via hepatic cellular surface receptors which help to maintain the
187 levels of lipids and lipoproteins in humans (21). Thus these processes depend upon the integrity of the
188 cellular function of liver. Hepatocellular damage or injury can interfere with these processes thereby
189 leading to alteration of lipids and lipoprotein patterns.

190 Small dense LDL is a major component of LDL-cholesterol and it is believed to be a very promising
191 biomarker for the prediction of cardiovascular event because it possesses more atherogenic potential
192 than other fractions of LDL-cholesterol and it has the profound ability to exhibit prolonged residency in the
193 sub endothelial space (22-24). Additionally, sdLDL particles have reduced affinity to the liver LDL
194 receptor, consequently they stay longer in the circulation (25, 26).

195 Experimental evidence suggests that most proinflammatory cytokines especially TNF- α plays an
196 important role in liver injury induced by hepatitis B virus and TNF-alpha may also be associated with
197 persistent HBV infection and severity (27, 28).

198 The present study revealed that both sdLDL and TNF- alpha are significantly elevated in chronic HBV
199 patients when compared with control subjects ($P < 0.05$). Additionally, we also observed a significant
200 positive correlation between sdLDL and TNF- α in HBV patients. The underlying mechanism for this
201 association is not entirely clear but one possible explanation is that TNF- alpha has the ability to modify
202 the activities of hepatic lipase thereby causing it to increase the lipolysis of triglyceride-rich LDL with
203 consequent increased formation of sdLDL (29-33).

204 The present study also demonstrated significantly higher levels of hepatic aminotransferases (AST, ALT)
205 in HBV patients when compared with control subjects. This is consistent with finding of previous studies
206 (8,13,34) and this has been attributed to a localized autoimmune reaction mediated by major

207 histocompatibility complex-1-hepatitis B surface protein complex which results into the degeneration of
208 hepatic tissue and during this process the cell membranes become more permeable, thereby leading to
209 leakage of the hepatic aminotransferases into the blood stream (6, 35).

210 Our findings also revealed a significant positive correlation with between sdLDL and hepatic
211 aminotransferases. This agrees with findings of previous studies that reported significant rise in AST and
212 ALT in proportion to raised LDL and triglycerides levels in patients with HBV infection (13, 36-38).

213 Furthermore, the present study revealed that there is a significant positive correlation between TNF-
214 alpha and hepatic aminotransferases and this is consistent with reports of previous studies (28, 39, 40).
215 The significant positive association that exists between TNF- alpha and hepatic aminotransferases,
216 indicates the progression of inflammation and severity of injury induced by HBV infection (41).

217 **5. CONCLUSION**

218 The present study demonstrated that there is significant relationship between sdLDL, TNF- alpha and
219 hepatic aminotransferases. Taken together, sdLDL and TNF- alpha can serve as potential predictors of
220 liver damage induced by HBV. Also based on our findings, HBV patients need to be closely monitored for
221 signs of cardiovascular disease.

222 While results from cross sectional study on a larger scale would play significant role in understudying the
223 observations reported in this study, longitudinal studies would also facilitate better understanding of the
224 findings of this study.

225 **Competing interest:** Nil

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UNDER PEER REVIEW