Original Research Article

Association between Non-Secretion of ABH Antigens and Sickle Cell Anaemia

ABSTRACT

Aim: To determine whether non-secretion of ABH blood group antigens was associated with Sickle Cell Anaemia.

Materials and Methods: Haemaglutination inhibition test was carried out on saliva samples from 300 individuals; 100 of whom had haemoglobin (Hb) genotype AA, 100 HbAS, 50 HbAC and 50 HbSS. ABO blood grouping was carried out by standard methods and Haemoglobin genotype test was performed by cellulose acetate electrophoresis technique. **Results:** Eighteen percent (18%) of HbAA, 23% of HbAS, 18% of HbAC and 42% of HbSS individuals were non-secretors of ABH antigens (p = 0.007). Non-secretion of ABH substances was more associated with HbSS persons than HbAA (p = 0.002), HbAS (p = 0.016) and HbAC (p = 0.009) individuals.

Conclusion: Non-secretion of ABH blood group substances is associated with Sickle Cell Anaemia.

Keywords: Haemoglobin genotype, ABO blood group, Sickle cell anaemia, ABH antigens, Secretor status

1. INTRODUCTION

In Southwestern Nigeria, in addition to normal haemoglobin A, haemoglobins S and C exist bringing about variants HbAA, HbAS, HbAC, HbSS, HbCC and HbSC among the people in the region [1]. Haemoglobinopathies especially sickle cell anaemia poses a lot of health challenges in Nigeria [2]. Sickle cell anaemia (SCA) is an inherited disorder caused by mutation resulting in replacement of amino-acid glutamic acid with valine at the 6th base position of the beta globin chain. It is a genetic blood disorder characterized by the presence of 2 alleles of the abnormal haemoglobin S (HbSS) in the red cell instead of HbAA with high morbidity and mortality rates. It is recognized by the United Nations as a global public health concern and the World Health Organisation has recommended that by 2020, half of its members should have set up Sickle Cell Anaemia (SCA) control programmes [3].

21 Worldwide, SCA is estimated to affect 20-25 million people and annually about 300,000 22 children are born with the disorder [4]; approximately 250,000 of whom are in sub-Saharan 23 Africa [5] with 50-80% of affected children dying before the age of 5 years [6]. 24 The ABO blood group and secretor status of a person are inherited independently. While the 25 ABH (FUT 1) gene codes for the ABO blood group, the secretor (FUT 2) gene interacts with 26 the ABH (FUT 1) gene to determine the secretor status of an individual [7]. Individuals can 27 be homozygous (SeSe) or heterozygous (Sese) secretors or non-secretors (sese). Non-28 secretion of ABH antigens has been associated with a number of non-communicable 29 diseases and disorders such as autoimmune diseases [8, 9], blood clotting and thrombotic 30 diseases [10, 11], immunological disorders [12], myocardial infarction [13, 14], rheumatic 31 heart disease [15, 16], duodenal ulcers [17]. Apart from one study [18] which investigated 32 frequency distribution of secretors and non-secretors in HbAA and HbSS individuals in Zaria, 33 Northwestern Nigeria, we are not aware of any other investigation that has related secretor status with haemoglobin variants. In this study, we hypothesized that secretor status varied 34 35 significantly with haemoglobin variants and that non-secretion of ABH antigens was 36 associated with sickle cell anaemia.

37 38

2. METHODOLOGY

39 40

41

45

46

47

48

49

2.1 Study Area and Population

42 This study was carried out in Osogbo, Southwestern Nigeria. It is the capital of Osun State.

43 Osogbo city seats the Headquarters of both Osogbo Local Government Area (situated at

44 Oke Baale Area of the city) and Olorunda Local Government Area (situated at Igbonna Area

of the city). It is some 88 kilometers by road northeast of Ibadan with coordinates Latitude

7.767-7.770°N and Longitude 4.557-4.567°E. A total of 300 participants were screened for

this study: 100 HbAA individuals, 100 HbAS, 50 HbAC and 50 HbSS. They were drawn from

apparently healthy staff, students and patients of LAUTECH Teaching Hospital visiting the

General Out Patient Department for routine examination.

2.2 Collection of Blood Samples

Blood samples were collected for Haemoglobin geneotype test and ABO blood grouping. A sample of 3 ml of venous blood was collected from each participant into ethylenediaminetetraacetic acid (EDTA) bottle. Haemogobin genotype test was performed using cellulose acetate electrophoresis method as described elsewhere [1]. In an alkaline pH (8.2-8.6), Hb is a negatively charged molecule and will migrate towards the anode. Different Hbs move at different rates depending on their net charge which is controlled by the amino acid composition of their globin chain. The ABO grouping system is based on agglutination reaction. When a red blood cell carrying an antigen is exposed to its corresponding antibody, they react with each other to form agglutination or clumping. ABO blood group tests were performed by standard techniques as described elsewhere [19].

2.3 Collection of Saliva Samples

Saliva samples were collected from participants for the determination of their secretor status; 2 ml of saliva was collected from each participant for determination of secretor status using haemagglutination inhibition test as described elsewhere [20]. In haemagglutination inhibition technique, processed saliva is mixed with antiserum (anti A, anti B or anti H) and allowed to incubate briefly. If the saliva is from a secretor, the soluble blood antigens in it react and neutralize the antibodies in the antiserum. So when red blood cells of appropriate blood group are added to the test mixture of the saliva and antiserum, there will be no free antibody to agglutinate them because the antibodies have already been neutralized by the antigens in the saliva. Therefore the reaction will be negative for agglutination. However, if the saliva is from a non-secretor, there will be no blood group antigens in it and so the antibodies in the antiserum will not be neutralized but free to react with appropriate test cells when added to produced agglutination. Laboratory investigations were carried out on samples collected in the Research Laboratory of the Department of Medical Laboratory Science, College of Health Sciences, Ladoke Akintola University of Technology, Osogbo, Nigeria.

2.4 Data Analysis

Data were analysed using proportions and percentages. Differences in proportions or percentages were tested by Chi-square test. A p-value of < 0.05 was considered significant.

3. RESULTS

A total of 300 persons comprising 100 HbAA, 100 HbAS, 50 HbAC and 50 HbSS individuals participated in this study. Table 1 shows the age and sex distributions of the study population. There were no significant differences in the age (p = 0.998) and sex (p = 0.718) distributions among the four groups of haemoglobin variants. The distributions of the haemoglobin variants of the study participants in relation to secretor status are given in Table 2. Of the 100 individuals with HbAA, 18% were non-secretors; 23%, 18% and 42% of the HbAS, HbAC and HbSS individuals respectively were non-secretors. Non-secretion of ABH antigens varied significantly with haemoglobin variants ($\chi^2 = 11.99$, df = 3, p = 0.007). Further Chi-square tests showed that non-secretion of ABH antigens was more associated with HbSS individuals than HbAA individuals ($\chi^2 = 9.978$, df = 1, p = 0.002), HbAS individuals ($\chi^2 = 5.805$, df = 1, p = 0.016) and HbAC individuals ($\chi^2 = 6.857$, df = 1, p = 0.009). There was no significant variation in secretion of ABH antigens among HbAA, HbAS and HbAC individuals ($\chi^2 = 0.938$, df = 2, p = 0.626). Altogether, non-secretors in the HbSS group (42.0%) were significantly higher than those in the non-SS (HbAA, AS and AC) group (20.0%) ($\chi^2 = 11.163$, df = 1, p < 0.001).

The distributions of the haemoglobin variants of the study participants in relation to ABO blood group are given in Table 3. Group AB was excluded from the analysis due to its small number across the haemoglobin variants. There was no significant association in the

distributions of haemoglobin variants in relation to ABO blood group ($\chi^2 = 5.69$, df = 6, p = 0.458).

The distributions of the non-secretors of the study participants with respect to haemoglobin variants and ABO blood group is given in Table 4. Of the 100 AA individuals, 18 were non-secretors (10 non-group O and 8 group O); of the 100 AS individuals, 23 were non-secretors (14 non-group O and 9 group O); of the 50 AC individuals, 9 were non-secretors (6 non-group O and 3 group O) while 21 of the 50 SS individuals were non-secretors (14 non-group O and 7 group O). Altogether, of the 150 non-O blood group participants, 44 (29.3%) were non-secretors while 27 of the 150 (18.0%) group O participants were non-secretors ($\chi^2 = 5.332$, df = 1, p = 0.021).

Table 1: Age and Sex distributions among the Study Participants

Haemoglobin Variants							
HbAA	HbAS	HbAC	HbSS	Total	р		
n=100(%)	n=100(%)	n=50(%)	n=50(%)	300			
					0.998		
32(32.0)	35(35.0)	17(34.0)	15(30.0)	99(33.0)			
28(28.0)	27(27.0)	14(28.0)	15(30.0)	84(28.0)			
40(40.0)	38(38.0)	19(38.0)	20(40.0)	117(39.0)			
					0.718		
45(45.0)	48(48.0)	26(52.0)	27(54.0)	146(48.7)			
55 (55.0)	52(52.0)	24(48.0)	23(46.0)	154(51.3)			
	n=100(%) 32(32.0) 28(28.0) 40(40.0) 45(45.0)	HbAA	HbAA n=100(%)	HbAA n=100(%) HbAS n=50(%) HbAC n=50(%) HbSS n=50(%) 32(32.0) 35(35.0) 17(34.0) 15(30.0) 28(28.0) 27(27.0) 14(28.0) 15(30.0) 40(40.0) 38(38.0) 19(38.0) 20(40.0) 45(45.0) 48(48.0) 26(52.0) 27(54.0)	HbAA n=100(%) HbAS n=50(%) HbAS n=50(%) Total n=50(%) 32(32.0) 35(35.0) 17(34.0) 15(30.0) 99(33.0) 28(28.0) 27(27.0) 14(28.0) 15(30.0) 84(28.0) 40(40.0) 38(38.0) 19(38.0) 20(40.0) 117(39.0) 45(45.0) 48(48.0) 26(52.0) 27(54.0) 146(48.7)		

Table 2: Distribution of the Haemoglobin Variants of the Study Participants in Relation to Secretor Status

Haemoglobin Variants						
	HbAA	HbAS	HbAC	HbSS	Total	р
	n=100(%)	n=100(%)	n=50(%)	n=50(%)	300	
Secretor status						0.007
Secretor	82(82.0)	77(77.0)	41(82.0)	29(58.0)	229(76.3)	
Non-secretor	18(18.0)	23(23.0)	09(18.0)	21(42.0)	71(23.7)	

Table 3: Distribution of the Haemoglobin Variants of the Study Participants in Relation to ABO Blood Group

	Haemoglobin Variants					
	HbAA	HbAS	HbAC	HbSS	Total	р
	n=100(%)	n=100(%)	n=50(%)	n=50(%)	300(%)	
ABO Blood Group						0.458
Α	22(22.0)	23(23.0)	11(22.0)	16(32.0)	72(24.0)	
В	19(19.0)	26(26.0)	13(26.0)	14(28.0)	72(24.0)	
*AB	02(2.0)	02(2.0)	01(2.0)	01(2.0)	06(2.0)	
0	57(57.0)	49(49.0)	25(50.0)	19(38.0)	150(50.0)	
		1 1 11 1 11 11				

*AB group was not included in the statistical analysis

Table 4: Distribution of the Non-Secretors of the Study Participants with Respect to Haemoglobin Variants and ABO Blood Group

144	
145	

	Haemoglobin Variants						
	HbAA(NS)	HbAS(NS)	HbAC(NS)	HbSS(NS)	Total(NS)		
ABO Blood Group							
Non-O	43(10)	51(14)	25(06)	31(14)	150(44)		
0	57(08)	49(09)	25(03)	19(07)	150(27)		
Total	100(18)	100(23)	50(09)	50(21)	300(71)		

NS: Non-Secretor

150 151

4. DISCUSSION

152 153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

Previous studies in this study area had shown that secretor status was independent of sex [20]. A similar finding was reported in Calabar, South south Nigeria[21]. Similarly, in this study locality, the distribution of ABO blood group and haemoglobin variants had been reported to be sex independent [1, 22] which were in line with ABO studies carried out in the same region by other researchers [23, 24]. In this study, we tested the hypothesis that non-secretors were more associated with HbSS compared to the other haemoglobin variants. The frequency of non-secretors in SCA (HbSS) individuals was significantly higher than the frequency of non-secretors in the other haemoglobin variants (HbAA, HbAS and HbAC) showing that secretor status varied significantly with haemoglobin variants. A study in Northwestern Nigeria reported a higher frequency of non-secretor in HbSS individuals compared to HbAA individuals [18]. Also in this study, non-secretion of ABH substances was more associated with persons of non-O group compared to those of O group. Previous studies in the study area and elsewhere had reported lower proportion of group O non-secretors compared to non-O group non-secretors [7, 20]. Another study in the area showed that malaria was less associated with group O secretors than non-group O secretors [22]. These studies showed that with respect to ABO blood group system, more group O persons were secretors compared to the other groups. The protective effect offered by group O individuals had been linked to higher incidence of secretor compared to non-O group [7]. Non-secretion of ABH antigens has been associated with many non-communicable diseases and disorders as stated earlier on. Similarly, sickle cell anaemia individuals are known to have several complications including chronic pain, intermittent painful episodes, musculoskeletal problems, stroke, pulmonary hypertension and septicaemia [3, 25]. It is not unlikely that the complications exhibited by majority of the persons with sickle cell disorder might largely be due to their inability to secrete ABH substances. Also, the observed association might be linked with the Le^a antigens which are present in greater amounts on the epithelial surface of non-secretors [26]. The positive interaction observed between HbSS and inability to secrete ABH antigens could be suggestive of the fact that the sickle cell gene and the secretor gene might directly or indirectly interact to confer susceptibility on persons with sickle cell anaemia. We opine that the severity of symptoms and complications observed in HbSS patients could be due to their inability to secrete ABH antigens. Further studies can be carried out to confirm or disprove this view.

4. CONCLUSION

We conclude that secretor status varies significantly with haemoglohin variants and that inability to secrete ABH antigens is associated sickle cell anaemia. The increased risk of symptoms and complications of sickle cell anaemia.

CONSENT

Informed consent was obtained from all the participants. Ethical approval for this study was obtained from the Ethical Committee of the College of Health Sciences, Ladoke Akintola University of Technology, Osogbo, Osun State, Nigeria.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Igbeneghu C, Olisekodiaka MJ, Akinola FFS, Odaibo AB. Impact of haemoglobin variants AS and AC on Asymptomatic falciparum malaria among adults in Iwo, Southwestern Nigeria. SJAMS. 2015a;3(1A):17-20.

2. Emechebe GO, Onyire NB, Orji ML, Achigbu KI. Sickle cell disease in Nigeria - A review. IOSR-JDMS. 2017;16(1):87-94.

3. Mulumba LL, Wilson L. Sickle cell disease among children in Africa: An integrative literature review and global recommendations. Intern J Afr Nursing Sci. 2015;3:56-64.

216

219

224

227

228

229

239

242

252

256

- 217 4. Azar S, Wong TE. Sickle cell disease: a brief update. Med Clin North Am. 2017; 218 101(2):375-393.
- 5. Macharia AW, Mochamah G, Uyoga S, Ndila CM, Nyutu G, Makale J, Tendwa M, Nyatichi E, Ojal J, Shebe M, Awuondo KO, Mturi N, Peshu N, Tsofa B, Scott JAG, Maitland K and Williams TN. The clinical epidemiology of sickle cell anaemia in Africa. Am J Haematol. 2018;93(3):363-370.
- 225 6. Aygun B, Odame I. A global perspective on Sickle cell disease. Pediatr Bld Cancer. 2012; 59(2): 386-390.
 - 7. Jaff MS. Higher frequency of secretor phenotype in O blood group-its benefits in prevention and /or treatment of some diseases. Intern J Nanomed. 2010;5:901-905.
- Shinebaum R, Blackwell CC, Forster PJ, Hurst NP, Weir DM, Nuki G. Non-secretion of ABO blood group antigens as host susceptibility factor in the spondyloarthropathies. BMJ (Clin Res Ed). 1987;294(6566):208-210.
- Shinebaum R. ABO blood group and secretor status in the spondyloarthropathies. FEMS
 Microbiol Immunol. 1989;1(6-7):389-395.
- 237 10. Orstavik, KH, Kornstad L, Reisner H, Berg K. Possible effect of secretor locus on plasma 238 concentration of factor VIII and von Willebrand factor. Blood. 1989;73(4):990-993.
- 11. Orstavik KH. Genetics of plasma concentration of von Willebrand factor. Folia Haematol
 Int Mag Klin morphol Blutforsch. 1990;117(4):527-531.
- 12. Al-Agidi SK, Shukri SM. Association between immunoglobulin levels and known genetic
 markers in an Iraqi population. Ann Hum Biol. 1982;9(6): 565-569.
- 13. Hein HO, Sorensen H, Suadicani P, Gyntelberg F. The Lewis blood group a new
 genetic marker of ischaemic heart disease. J Intern med. 1992;232(6):481-487.
- 14. Ellison RC, Zhang Y, Myers RH, Swanson JL, Higgins M, Eckfeldt J. Lewis blood group phenotype as an independent risk factor for coronary heart disease (the NHLBI Family Heart Study). Am J Cardiol. 1999;83(3):345-348.
- 253 15. Robinson WM, Salzano FM, Achutti AC, Franco MH. Blood groups, salivary secretion and other immunologic variables in rheumatic fever and rheumatic heart disease. Acta Anthropogenet. 1984;8(3-4):217-221.
- 16. Jhingham B, Mehra NK, Reddy KS, Taema V, Valdya MC, Bhatia ML. HLA, Blood groups and secretor status in patients with established rheumatic fever and rheumatic heart disease. Tissue Ag. 1986;3:172-178.
- 261 17. Dickey W, Collins JSA, Watson RGP, Sloan JM, Porter KG. Secretor status and 262 helicobacter pylori infection are independent risk factors for gastroduodenal disease. Gut. 263 1994;34(3):351-353.
- 18. Olorunshola KV, Audu I. ABO (H) secretor status of sickle cell disease patients in Zaria,
 Kaduna State, Nigeria. Niger J Physiol Sci. 2013;28:29-34.

19. Igbeneghu C, Odaibo GN, Olaleye DO, Odaibo AB. Malaria Infection and ABO blood grouping in Iwo community, Southwestern Nigeria. Res J Med Sci. 2012;6(5):247-250.

20. Igbeneghu C, Olisekodiaka JM, Alabi T, Onuegbu JA, Oseni BA, Odaibo A. ABH secretors status in Osogbo, Southwestern Nigeria. Indian J Fund Appl Life Sci. 2015b;5(3):42-47.

21. Emeribe AO, Igweagu CA and Ossim EE. ABH secretor status in saliva of Calabar Municipal residents. East Afr Med J. 1992;69(1):27-30.

22. Igbeneghu C, Olisekodiaka MJ, Okanlawon BM, Onuegbu JA, Odaibo AB. Non-Secretors of ABH Antigens are susceptible to falciparum malaria. 2015c;3(5A):1838-1841.

23. Falusi AG, Ademowo CA, Latunji CA, Okeke AC, Olatunji PO, Onyekwere TO, Jimmy EO; Raji Y. Distribution of ABO and Rh genes in Nigeria. Afr J Med Med Sci. 2000;29:23-26.

24. Bakare AA, Azeez MA, Agbolade JO. Gene frequencies of ABO and rhesus blood groups and haemoglobin variants in Ogbomoso, Southwest Nigeria. Afr J Biotech. 2006;5(3):224-229.

25. Kapoor S, Little JA, Pecker LH. Advances in the treatment of Sickle cell disease. Mayo Clin Proc. 2018;93(12):1810-1824.

26. Raza MW, Backwell CC, James VS, Ogilvie MM, Weir DM, Molyneaux P et al. Association between secretor status and respiratory viral illness. BMJ. 1991;303:815-818.