

Antimicrobial Activities of *Teucrium creticum* Against Reference Microbial Strains and Multi-Drug Resistant Bacteria Isolated at an Oncology Ward

Brief title: Antimicrobial Activities of *Teucrium creticum*

ABSTRACT

Aim: To determine antimicrobial activity of *Teucrium creticum* (*T. creticum*) leaves extract against bacterial and fungal reference strains and multidrug resistant bacteria isolated at an oncology ward.

Study design: Cross-sectional study.

Place and Duration of Study: The study was carried out in department of biology and biotechnology in An-Najah National University in cooperation with the laboratory of the hospital of the university. An-Najah National University is in West Bank in Palestine. The research was performed from 8th of February to the 15th of April 2017.

Methodology: *Teucrium creticum* plant leaves were collected in Palestine, from which aqueous and methanolic extracts were prepared. Antimicrobial activities of *T. creticum* extracts were determined against reference bacterial and fungal strains as well as against 8 multidrug resistant bacteria isolated at an oncology ward. Antibacterial and anti-yeast activities were determined by Micro broth dilution method, while anti-mold activities were determined by agar dilution method. *Teucrium creticum* methanolic extract strongly inhibited the growth of the studied reference bacterial strains, which were *Staphylococcus aureus* (MIC= 1.56 mg/ml) and *Shigella sonnie* (MIC=3.125 mg/ml). In addition, most of the 8 multi-drug resistant bacterial strains isolated from patients with cancer (*Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Escherichia Coli* and *Enterobacter cloacae*) were also highly susceptible to methanolic extract (MIC=3.125 mg/ml). Both the *Staphylococcus aureus* and *Shigella sonnie* reference strains were inhibited at lower level by the aqueous extract (MIC=12.5 mg/ml). All the bacterial strains isolated from patients with cancer were susceptible to aqueous extract at different levels (3.125 – 25 mg/ml). *Epidermophyton floccosum* mold and *Candida albicans* yeast were strongly inhibited by aqueous extract, where the MIC values were 1.56 and 3.125 mg/ml, respectively.

Conclusion: *T. creticum* plant extracts showed promising antimicrobial activities against multidrug resistant bacterial isolates as well as against reference bacterial and fungal strains.

Keywords: *Teucrium creticum*, antimicrobial activities, multidrug resistance, oncology

1. INTRODUCTION

A major public health problem worldwide is the spread of antimicrobial resistance. This may be attributed mostly to the widespread use of antimicrobials. Infections due to multi-drug resistant (MDR) pathogens have become a therapeutic challenge and a cause of significant morbidity and mortality [1, 2, 3].

Complementary and alternative medicine (CAM) are widely used and still increasing in the Western world [4, 5]. Genus of *Teucrium* L. is a large and polymorphic one. This genus is common in mild parts of Europe, North Africa and Asia. In Turkey, *Teucrium* species have been used for years in order to treat diabetes and diseases of the digestive system as traditional drugs for the purpose of choler expectorant, urine expectorant, antidiabetic, treatment of inflammation, antiseptic, worm removal, flavour, appetizing and breath opener [6, 7]. In Cyprus, the genus *Teucrium* was found to includes four species; *T. cyprium* ssp., *T. micmpodioides*, *T. divaricatum* ssp. *canescens* and *T. kotschyianum* [8]. In a previous study in Palestine, 3 species of *Teucrium* were detected, which included *Teucrium capitatum*, *Teucrium creticum* and *Teucrium divaricatum* [9].

48 In Palestine, *Teucrium creticum* L. (Lamiaceae) (*T. creticum*) is locally known under the common name Ja'adh. The
49 plant is used traditionally to treat diabetes [10]. Due to its limited distribution in Palestine [11], few phytochemical
50 and antimicrobial studies were conducted on this plant.

51 *Teucrium creticum* plant's aerial parts have been reported for its astringent, vulnerary, antipyretic and depurative
52 properties. Furthermore, aerial parts of *T. creticum* is known for its attractiveness to bees [12].

53 The genus *Teucrium* is considered to be one of the most abundant natural source of neoclerodane and 19-nor-neo-
54 clerodane diterpenoids. These compounds were reported to possess useful antifeedant activity [13]. In *Cyprus*, *T.*
55 *creticum* is also known to contain clerodane diterpenoids [14]. In a study carried by Omar et al [15], *Teucrium*
56 *creticum* was not shown to significantly affect human coagulation cascade. It was demonstrated in a previous study
57 that *T. creticum* is an efficient scavenger of free radical [16]. On the other hand, ethanolic *T. creticum* extract
58 possessed no or very low antimicrobial activities against *Pseudomonas aeruginosa*, *Proteus vulgaris*, *Klebsiella*
59 *pneumonia*, *Escherichia coli*, *Salmonella typhi*, *Candida albicans*, *Microsporium canis*, and *Trichophyton rubrum*
60 [17].

61 This study aimed to evaluate antimicrobial activity of *T. creticum* leaves extract against bacterial and fungal
62 reference strains and multidrug resistant bacteria isolated at oncology ward of An-Najah National university.
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2. MATERIALS AND METHODS

2.1 Bacterial isolates

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67 Reference microbial strains were obtained from American Type Culture Collection. Bacterial strains were
68 *Staphylococcus aureus* ATCC 25923 and *Shigella sonnie* ATCC 25931, while fungal isolates were *Candida*
69 *albicans* ATCC 90028 and *Epidermophyton floccosum* ATCC 52066. In addition, 38 isolates recovered from
70 different wards at An-Najah national hospital were provided with their identification. These isolates were collected
71 from 8th of February to the 15th of April 2017. These strains were isolated from various clinical samples (blood,
72 wound swabs, sputum, urine, etc.). The bacterial isolates were *Escherichia coli*, *Pseudomonas aeruginosa*,
73 *Staphylococcus aureus* and *Enterobacter cloacae*.

2.2 Antibiotic susceptibility test

75 In order to select representative clinical multidrug-resistant isolates recovered at oncology ward and to examine
76 their susceptibility to *T. creticum*, antibiotic susceptibility test was performed. Disc diffusion technique according
77 to CLSI [18], was used for antibiotic susceptibility testing. The used antibiotic discs were Cephalothin (30 µg),
78 Cefoxitin (30 µg), Nalidixic acid (30 µg), Amoxicillin (20 µg), Ciprofloxacin (5 µg), Imipenem (10 µg), Amikacin
79 (30 µg), Aztreonam (30 µg), Ceftazidime (30 µg) and Amoxycillin-Clavulanic Acid (20/10 µg). In addition, *S.*
80 *aures* isolates were examined against Vancomycin (30µg). All antibiotics were obtained from OXOID (UK).

2.3 Determination of antimicrobial activities of *T. creticum*

2.3.1 Plant collection

83 The plant was classified and kindly provided by Dr. Ghadeer Omar. The plant was collected from Palestine and
84 included leaves, which was ground into powder.
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3.3.2 Plant extracts preparation

88 Plant powder (20g) was separated into two equal parts then 100 ml of 100% methanol and 100 ml of deionized
89 water were added separately. The powder was soaked with the solvents with continues shaking for 7 days. Then the
90 extracts were centrifuged for 5 minutes at 1000 g and the supernatant was aspirated. The organic extract was dried
91 using rotary evaporator, and the aqueous extract was dried using lyophilizer. Aqueous extract was dissolved in water
92 and organic extract was dissolved in 100% DMSO (Dimethyl sulfoxide). The concentration of both extracts was 50
93 mg/ml. The extracts were sterilized by syringe filtration method.

2.3.3 Micro broth dilution method

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95 A 100 µl volume of Mueller Hinton broth (Becton dickinson and company, France) was pipetted in each well of
96 a 96 well plates (Greiner bio-one, China). Plant extract (100 µl) was pipetted in the first well and mixed. Then 100
97 µl was transferred to next well. This was repeated to well number 11, from which 100 µl were discharged after
98 mixing. Well number 12 (positive control of microbial growth) was free from plant extract. Bacterial suspension
99 was prepared equivalently to 0.5 McFarland standards, and 2 ml from suspended bacteria was diluted with 4 ml
100 Mueller Hinton broth, this suspension was inoculated in each well (1µl) except well number 11 (negative control of
101 microbial growth). The plates were incubated at 35°C for 24 hours for bacterial species [19, 20]. The yeast species
102 (*Candida albicans*) suspension was prepared equivalently to 0.5 McFarland standards, and diluted 1:20 then 1:50
103 with Mueller Hinton broth, this suspension was inoculated in each well (100 µl) except well number 11 (negative
104 control of microbial growth). Plate was incubated at 35°C for 48 hours. MIC value was defined as the lowest
105 concentration that inhibited any visible microbial growth [19, 20, 21].

106 **2.3.4 Agar Dilution Method**

107 The prepared SDA media (Becton dickinson and company, France) were kept in melted state (1ml in each test
108 tube) in water bath at 40 °C. Six test tubes for each plant extract (water and methanolic) were prepared. A volume
109 of 1 ml of plant extract was added into tube number one and mixed. Then 1ml was transferred from tube number 1
110 to next tube. The process was repeated up to tube number 6. Then tubes were put on slant position, Tube number six
111 was used as negative control of fungal growth. One tube was free from plant extract (positive control of fungal
112 growth) [20, 22]. In addition, serial dilution of 100% DMSO was examined.

113 A suspension of mold (*Epidermophyton floccosum*) was prepared to be equivalent to a McFarland standard with
114 90% transmission. A total of 20µl of fungal suspension was placed on surface of each slant in the tubes with the
115 exception of negative control of fungal growth. The tubes were incubated at 25 °C for 14 days [20, 22].

116 Each of plant extract and DMSO were examined by micro-broth and agar dilution methods two times.

117 **3. RESULTS**

118 **3.1 Antimicrobial activities of *T. creticum* extracts**

119 **3.1.1. Inhibitory activity against reference strains**

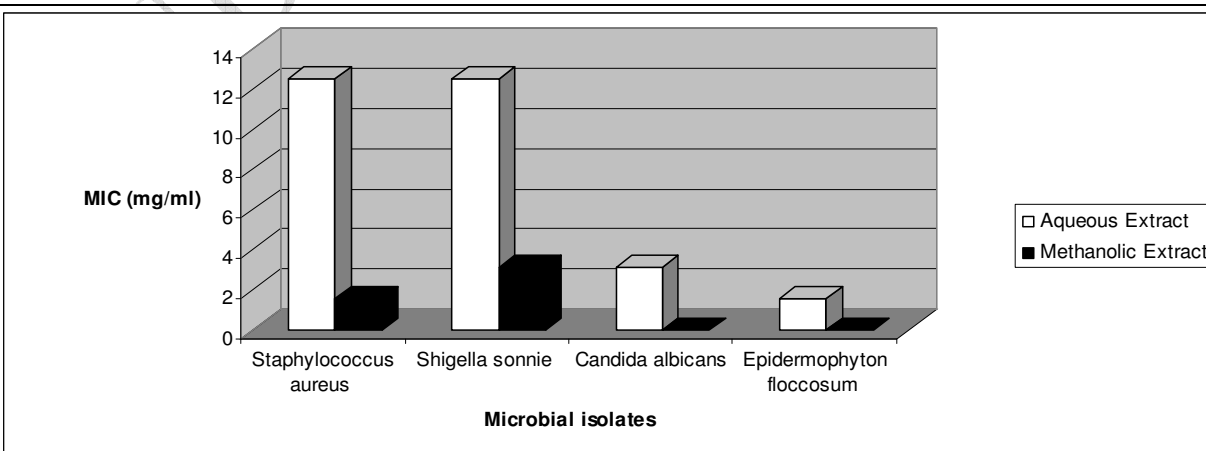
120 Methanolic extract of *T. creticum* was more effective than aqueous one against *S. aureus* and *S. sonnie*. In more
121 detail, growth of *S. aureus* and *S. sonnie* was inhibited by methanolic extract concentration of 1.56 and 3.125 mg/ml,
122 respectively (Table 1, Figure1). On the other hand, higher concentration of aqueous plant extract (MIC= 12.5
123 mg/ml) was required to stop the growth of both bacterial types. Both fungal types were more susceptible than
124 bacteria to aqueous extract of *T. creticum*, where MIC for *E. floccosum* and *C.albicans* were 1.56 and 3.125 mg/ml,
125 respectively. The limited solubility of methanolic *T. creticum* extract caused impediment for the determination of
126 MIC due to the antimicrobial activity of solvent DMSO. Where the container (well or tube) showing no growth of
127 fungi and containing plant extract was also containing an inhibitory concentration of DMSO (determined by MIC
128 value of DMSO alone).

130 **3.2.2 Inhibitory activity against bacteria isolated from patients with cancer**

131 In general ,the frequencies of resistance to different antibiotics were higher among isolates collected at oncology
132 ward in comparison to other sources. All the 8 isolates of bacterial strains isolated at the oncology wards were
133 multidrug resistant (resistant to 4 or more antibiotics). Table 1 and Figure 2 show the antimicrobial activities of *T.*
134 *creticum* extract against bacteria isolated from patients suffering from cancer. Among the extract for which MIC was
135 determined, both methanolic and aqueous *T. creticum* extract showed inhibitory activity against all bacteria isolated
136 from cancer patients. The 4 *P. aeruginosa* isolates were strongly inhibited by methanolic extract (MIC=3.125
137 mg/ml) and inhibited mostly at lower level (3.125-12.5 mg/ml) by aqueous extract. Out of the 2 *E. coli* isolates, one
138 was similar to *P. aeruginosa* isolates, where the MIC values were 3.125 and 12.5 mg/ml for methanolic and aqueous
139 extracts, respectively. On other hand, the second *E. coli* isolate was more resistant to activity of *T. creticum* extract,
140 where the MIC of aqueous extract was 25 mg/ml and MIC of methalonic extract was not determined. *S. aureus* and
141 *E. cloacae* isolated at oncology wards expressed more resistance to aqueous extract (MIC= 25mg/ml) and were more
142 susceptible to organic extract, where MIC was 3.125 mg/ml for both.

143 **Table 1** *Teucrium creticum* antimicrobial activities against bacteria isolated from cancer patients and
 144 reference microorganism strains

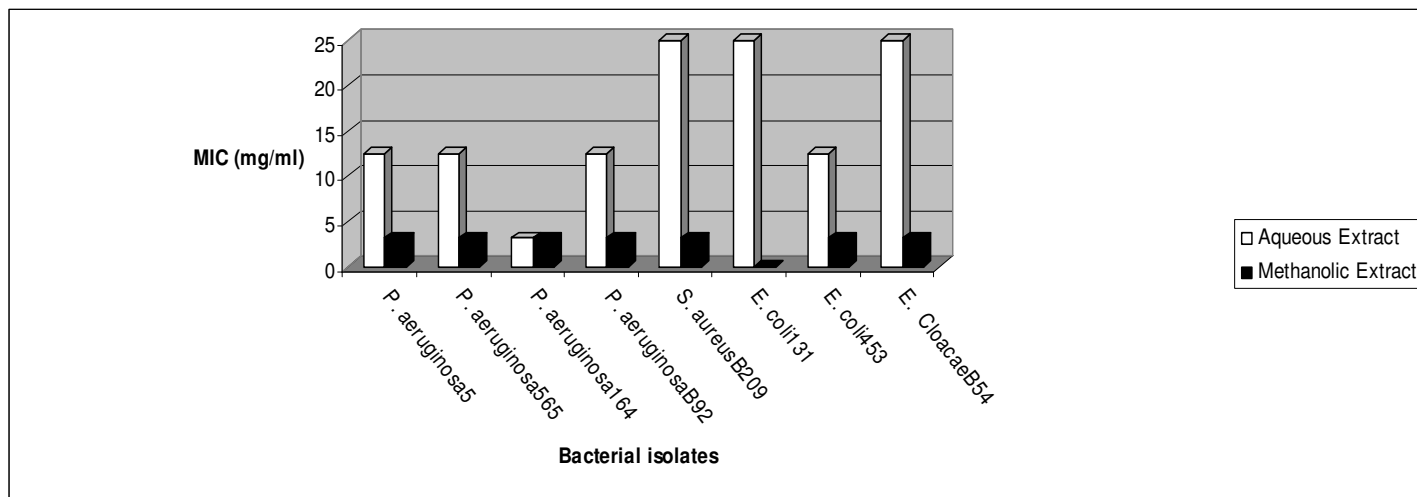
Microbial Isolates	MIC (mg/ml)	
	Aqueous Extract	Methanolic Extract
<i>Staphylococcus aureus</i> ATCC 25923	12.5	1.56
<i>Shigella sonnie</i> ATCC 25931	12.5	3.125
<i>Candida albicans</i> ATCC 90028	3.125	Undetermined
<i>Epidermophyton floccosum</i> ATCC 52066	1.56	Undetermined
Strains isolated from patients with Cancer		
<i>Pseudomonas aeruginosa</i> 5	12.5	3.125
<i>Pseudomonas aeruginosa</i> 565	12.5	3.125
<i>Pseudomonas aeruginosa</i> 164	3.125	3.125
<i>Pseudomonas aeruginosa</i> B92	12.5	3.125
<i>Staphylococcus aureus</i> B209	25	3.125
<i>Escherichia coli</i> 131	25	Undetermined
<i>Escherichia coli</i> 453	12.5	3.125
<i>Enterobacter cloacae</i> B54	25	3.125



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Figure 1. Antimicrobial activities of methanolic and aqueous *T. creticum* extracts against reference strains



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Figure 2. Antimicrobial activities of methanolic and aqueous *T. creticum* extracts against multidrug resistant strains isolated at oncology wards

4. DISCUSSION

In the present study, although the number of isolates is limited, the bacteria isolated from patients with cancer were more resistant to the examined antibiotics than that of bacteria isolated from patients without cancer. Cancer patients provide different and new environment for bacteria in comparison with other patients without cancer, because cancer patients are exposed to different types of treatments (chemotherapy and radiotherapy). In addition, the immune system of cancer patients is weak, which will give bacteria more time for the development of resistant. This elevated frequency of resistance of bacteria isolated from oncology ward provoked us to include antibiotic resistant bacteria isolated from patient with cancer in the evaluation of antibacterial activities of *T. creticum*. This may provide help for treatment of multidrug resistant bacteria strains isolated from patients with cancer.

In Algeria, *Teucrium polium* L. essential oil collected at Beni Aziz region has a high antibacterial activity against *Escherichia coli*, *Staphylococcus aureus* and the yeast *Saccharomyces cerevisiae*. The population of Boutaleb region has significant activity against *Bacillus cereus* and no action against *S. aureus* and the yeast *S. cerevisiae* [23].

In a previous research [17], ethanolic *T. creticum* extract possessed no or very low antimicrobial activities against *Pseudomonas aeruginosa*, *Proteus vulgaris*, *Klebsiella pneumonia*, *Escherichia coli*, *Salmonella typhi*, *Candida albicans*, *Microsporium canis*, and *Trichophyton rubrum*. In our study, aqueous plant extract of *T. creticum* showed antimicrobial activities against *Staphylococcus aureus*, *Shigella sonnie*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Enterobacter cloacae*, *Candida albicans* and *Epidermophyton floccosum*. In addition, methanolic extract of *T. creticum* possessed stronger antimicrobial for most of isolates. The differences of the results may be due to use of different solvents in the extraction procedures. Husein *et al.* [17] in the extraction procedure used 75% ethanol on the other hand in present study, water and 100% methanol solvents were used. In addition, collection of plants from different regions and differences in extractions methods may attribute to these variations in the results. For our knowledge, we are the first to report *T. creticum* antimicrobial activities against *Shigella sonnie*, *Enterobacter cloacae*, *Epidermophyton floccosum* and multidrug resistant bacterial strains isolated from patients with cancer. Further work is needed to isolate and identify the active compounds from this plant.

It's recommended to repeat the study on larger number of patients to confirm the results of the present study.

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184 **5. CONCLUSION**

185 In conclusion, although methanolic extract of *T. creticum* possessed stronger antimicrobial activities both
186 aqueous and methanolic extracts inhibited the growth of most examined microbial isolates including the multidrug
187 resistant ones isolated from oncology ward, thus *T. creticum* may represent a candidate for the development of new
188 antimicrobial agent that will be helpful for the treatment of multidrug resistant bacterial infections.

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190 **COMPETING INTERESTS**

191 There is no competing interest to mention

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193 **REFERENCES**

- 194 1. Prestinaci F, Pezzotti P, Pantosti A. Antimicrobial resistance: a global multifaceted
195 phenomenon. *Pathogens and Global Health*. 2015;109(7):309-318.
196 doi:10.1179/2047773215Y.0000000030.
- 197 2. Centres for Disease Control and Prevention, US Department of Health and Human Services.
198 Antibiotic resistance threats in the United States. Atlanta: CDC; 2013. Available
199 from: <http://www.cdc.gov/drugresistance/pdf/ar-threats-2013-508.pdf>
- 200 3. Giske CG, Monnet DL, Cars O, Carmeli Y. Clinical and economic impact of common multidrug-
201 resistant Gram-negative bacilli. *Antimicrob Agents Chemother*. 2008;52(3):813-821.
- 202 4. Chang HA, Wallis M, Tiralongo E. Use of Complementary and Alternative Medicine among People
203 with Type 2 Diabetes in Taiwan: A Cross-Sectional Survey. *Evidence-Based Complementary and*
204 *Alternative Medicine*. 2011, Article ID 983792, 8 pages doi:10.1155/2011/983792
- 205 5. Ventola CL. Current Issues Regarding Complementary and Alternative Medicine (CAM) in the
206 United States. *Complementary and Alternative Medicine*. 2010; 35(8): 461-468.
- 207 6. Bagcı Y. Aladag'lar (Yahyalı, Kayseri) ve çevresinin etnobotanik özellikleri. *Ot Sistematik Bot.*
208 *Dergisi*. 2000; 7: 8994. Turkish.
- 209 7. Öztürk M, Dinc, M. Nizip Bölgesi'nin (Aksaray) etnobotanik özellikleri. *Ot Sistematik Bot.*
210 *Dergisi*. 2005;12: 93-102. Turkish.
- 211 8. Arnold N. *Ethnobotanique - Ethnopharmacologie de la Flore de Chypre et de l'Île Méditerranéenne*.
212 Bailleul, France. 1991; 3, 1198-1199. French.
- 213 9. Al Sheikh B, Mahassneh M. Flora of Wadi Al-Quff Protected Area, Hebron Governorate, Palestine.
214 *Jordan Journal of Natural History*. 2017;3: 47-57.
- 215 10. Shtayeh MS, Jamous RM. Complementary and alternative medicine use amongst Palestinian
216 diabetic patients. *Complement. Ther. Clin. Pract*. 2012; 18(1):16-21.
- 217 11. Saad B, Azaizeh H, Said O. Tradition and perspectives of Arab herbal medicine: A review. *Evid.*
218 *Based Complement. Alternat. Med*. 2005; 2 (4):475-479.
- 219 12. Arnold N, Bellomaria B, Valentini G, Rafaiiani SM. Comparative study on essential oil of some
220 *Teucrium* species from Cyprus. *J. Ethnopharmacol* 1991; 35: 105-113.
- 221 13. Simmonds MSJ, Blaney WM, Ley, SV, Savona G, Bruno M, Rodriguez B. The antifungal activity of
222 clerodane ditetpenoids from *Teucrium*. *Phytochemistry* 1989; 28, 1067-1071.
- 223 14. Savona G, Piozzi F, Bruno M, Dominguez G, Servetaz O, Teucretol. A new clerodane ditetpenoid
224 from *Teucrium creticum*. *Phytochemistry* 1987; 26: 3285.
- 225 15. Omar G, Abdallah L, Rahim A, Othman R, Barakat A. Selected Wild Plants Ethanol Extracts
226 Bioactivity on the Coagulation Cascade. *Journal of Scientific Research and Reports*. 2017; 13(6), 1-
227 10. <https://doi.org/10.9734/JSRR/2017/32989>
- 228 16. Husein AI, Shtayeh MS, Jondi WJ, Zatar NAA, Abu-Reidah IM, Jamous RM. In vitro antioxidant
229 and antitumor activities of six selected plants used in the Traditional Arabic Palestinian herbal
230 medicine. *Pharmaceutical Biology*. 2014;52:10, 1249-1255, DOI: 10.3109/13880209.2014.886274.
- 231 17. Husein AI, Shtayeh MS, Jamous RM, Abu Zaitoun SY, Jondi WJ, Zatar NAA. Antimicrobial
232 activities of six plants used in Traditional Arabic Palestinian Herbal Medicine. *African Journal of*
233 *Microbiology Research*. 2014; 8(38):3501-3507.
- 234 18. Clinical and Laboratory Standard Institute. Performance Standards for Antimicrobial Susceptibility

- 235 Testing. Seventeenth Informational Supplement. M100-S17. Wayne, PA, USA: CLSI; 2007.
236 19. Wikler MA. Performance Standards for Antimicrobial Susceptibility Testing; Twenty-First
237 Informational Supplement, Clinical and Laboratory Standards Institute, USA. 2011; 1-165.
238 20. Forbes BA, Sahm DF, Weissfeld AS. Bailey and Scott's the Diagnostic Microbiology. 12 ed. USA:
239 Elsevier -Mosby; 2007.
240 21. Klepser ME, Wolfe EJ, Jones RN, Nightingale CH, Pfaller MA. Antifungal pharmacodynamic
241 characteristics of fluconazole and amphotericin B tested against *Candida albicans*, *Antimicrob.*
242 *Agents Chemother.* 1997;41 (6):1392–1395.
243 22. Falahati M, Tabrizi NO, Jahaniani F. Anti Dermatophyte Activities of *Eucalyptus camaldulensis* in
244 Comparison with Griseofulvin. *Iranian Journal of Pharmacology & Therapeutics (IJPT)*. 2005; 4(2):
245 p. 80-83.
246 23. Lograda T, Ramdani M, Chalard P, Figueredo G, Deghar A. Chemical Analysis and Antimicrobial
247 Activity of *Teucrium polium* L. Essential Oil from Eastern Algeria. *American Journal of Advanced*
248 *Drug Delivery.* 2014/ www.ajadd.co.uk/ISSN 2321-547X.