

## Original Research Article

### Phytochemical and Pharmacological Potential of Bangladesh *Enhydra fluctuans* available in Bangladesh ethanolic extract

#### ABSTRACT

##### Objective:

The possible phytochemical nature, thrombolytic and membrane stabilizing activity of the crude ethanolic extract of *Enhydra fluctuans*(CE) were investigated along with the anti-microbial, antioxidant and cytotoxic potentials of its petroleum ether (PESF), carbon tetrachloride (CTCSF), chloroform (CSF) and aqueous (AQSF) soluble fractions.

##### Materials & Methods:

The coarse leaf powder was extracted at room temperature with-using ethanol. Solvent-solvent partitioning was done to obtain the four soluble fractions by ~~the~~ modified Kupchan method. Anticoagulant potential was determined by the *invitro* thrombolytic model, membrane stabilization method was used to assess *in vitro* anti-inflammatory activity, the disc diffusion method was used for anti-microbial screening, antioxidant potential was determined by 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging method and brine shrimp lethality bioassay method was used for cytotoxic activity determination.

##### Results:

Chemical screening of the crude extract evidenced the presence of alkaloids, saponins, tannins, flavonoids, reducing sugars and gums. It showed significant clot lysis property of 49.29%. It also significantly inhibited heat and hypotonic solution induced lysis of the human red blood cell membrane with values of 71.67% and 47.93%, respectively. CTCSF and PESF showed mild antimicrobial activity. AQSF showed most prominent antioxidant activity with IC<sub>50</sub> value of 12.27 µg/mL. CTCSF showed LC<sub>50</sub> value of 0.84 µg/mL, with most potent cytotoxic activity.

##### Conclusion:

Significant thrombolytic, membrane stabilizing, antioxidant and cytotoxic *in vitro* activities of the plant extract was ~~found from~~ observed in this study. *In vivo* activities and isolation of active compound(s) from this extract are yet to be investigated.

**Keywords:** *Enhydra fluctuans*, thrombolytic activity, membrane stabilizing activity, antioxidant activity and cytotoxic activity.

#### INTRODUCTION

37 Medicinal plants have played an important role in treating various diseases since ancient times.  
38 Increased drug resistance and side effects of existing pharmaceutical drugs have escalated the  
39 research based study on traditionally available medicinal plants.

40 Myocardial or cerebral infraction and other atherothrombotic diseases are consequences of  
41 thrombus formed in blood vessels<sup>1,2</sup>. Fibrinolytic drugs like tissue plasminogen activator (t-PA),  
42 urokinase, streptokinase etc. dissolve thrombin in acutely occluded coronary arteries and restore  
43 blood supply to ischemic myocardium, to limit necrosis and improve prognosis<sup>3</sup>. Yet all the  
44 available thrombolytic agents have significant deficiencies, including the necessity of large doses  
45 to be maximally effective, limited fibrin specificity and a significant associated bleeding  
46 tendency. Therefore, studies are going on to develop improved thrombolytic drugs in order to  
47 minimize deficiencies of the available drugs.

48 Inflammation is a pathological disorder, in which inflammatory cells produce a complex  
49 mixture of growth and differentiation cytokines as well as physiologically active arachidonate  
50 metabolites, also generate reactive oxygen species (ROS) that can damage cellular biomolecules  
51 which in turn augment the state of inflammation<sup>4</sup>. Compounds that possess radical scavenging  
52 ability may therefore expect to have the therapeutic potentials against inflammation<sup>5</sup>. Moreover,  
53 stabilization of lysosomal membrane limits inflammatory response by inhibiting the release of  
54 lysosomal constituents of activated neutrophils such as bactericidal enzymes and proteases,  
55 which are responsible for further tissue inflammation and damage<sup>6</sup>.

56 Bacteria are responsible for many infectious diseases in the 21<sup>st</sup> century<sup>7</sup>. Antibiotic resistance  
57 has become a major clinical and public health problem for most people now-a-days currently<sup>8,9</sup>.  
58 This Multidrug Resistance (MDR) is clearly related to the misuse of different antibiotics<sup>10,11</sup>.  
59 The increasing clinical importance of drug resistant bacterial pathogens necessitates emergence  
60 of the search for additional antibacterial therapy. The antibacterial screening which is the first  
61 stage of antibacterial research is performed to ascertain the susceptibility of various bacteria to  
62 any agent.

63 Free radicals are highly toxic and reactive; and cause diseases like aging, atherosclerosis, cancer,  
64 diabetes, liver cirrhosis, cardiovascular disorders, etc. by attacking different macro-molecules  
65 including lipids, proteins and DNA resulting in the cellular damage<sup>12,13</sup>. Antioxidants protect  
66 body by delaying or reducing the oxidation of the substrate in turn neutralize or terminate the  
67 chain reaction before vital molecules within the body are damaged<sup>14</sup>. Currently available  
68 synthetic antioxidants like butylated hydroxyl anisole (BHA), butylated hydroxy toluene (BHT),  
69 tertiary butylated hydroquinone and gallic acid esters show low solubility and moderate  
70 antioxidant activity with suspected negative health effects<sup>15</sup>. BHA and BHT are suspected of  
71 being responsible for liver toxicity and carcinogenesis<sup>16,17</sup>. Investigations to screen out naturally  
72 occurring antioxidants are now mostly demanding.

73 Bioactive compounds are always toxic to living body at some higher doses and it justifies the  
74 statement that 'Pharmacology is simply toxicology at higher doses and toxicology is simply  
75 pharmacology at lower doses'. Brine shrimp lethality bioassay implies cytotoxicity as well as a  
76 wide range of pharmacological activities such as antimicrobial, antiviral, pesticidal and anti-  
77 tumor etc. of the compounds<sup>18,19</sup>.

78 *Enhydra fluctuans* Lour (Family: Asteraceae) is a small genus of marsh herb, available in  
79 tropical and subtropical regions; like Bangladesh, India, Malaysia, China and the rest of South  
80 East Asia and Tropical Africa<sup>20,21</sup>. The herb is relatively glabrous sometimes pubescent

81 glandular. Stems are usually 0.3-0.6m, elongated simple or divaricating rooting at the nodes<sup>22</sup>.  
82 Traditional uses indicate that the leaves of the plant are good as laxative. The expressed juice of  
83 the leaves is an excellent demulcent in gonorrhoea and also used in the treatment of skin and  
84 nervous system<sup>23</sup>. These activities can be attributed mainly to the presence of phytochemicals  
85 such as flavonoids, alkaloids, saponins, tannins, phenols and carbohydrates. But there are  
86 insufficient records in literature of this plant, regarding its pharmacological activities and  
87 phytochemical characteristics. Thus the present study focuses on screening of the ethanolic  
88 extract of *Enhydra fluctuans* Lour for its phytochemical and pharmacological potential.

89

90

91

## 92 **MATERIALS AND METHODS**

### 93 *Instrumentation*

94 The extract was condensed in rotary vacuum evaporator (Heidolph Instruments GmbH & Co.  
95 KG, Germany). Samples were centrifuged on a bench top centrifuge machine (HERMLE  
96 Labortechnik GmbH, Germany). The absorbance was recorded by a UV-Visible  
97 spectrophotometer (Analytic Jena AG, Germany). Ketoconazole was obtained from Beximco  
98 Pharmaceuticals Ltd., Bangladesh. Streptokinase vial (Beacon Pharmaceutical Ltd) was  
99 purchased from local pharmacy. Trolox and BHA were obtained from Calbiochem, Merck,  
100 Germany and VWR International Ltd., England, respectively. Nutrient agar and potato dextrose  
101 agar and Kanamycin 30 µg standard disc were purchased from HiMedia Laboratories Pvt. Ltd.,  
102 India. All other reagents were purchased from Sigma-Aldrich, USA and all other solvents used  
103 were of analytical grade and purchased from Active Fine Chemicals Ltd., Bangladesh.

### 104 *Collection and Identification*

105 For this investigation, *Enhydra fluctuans* leaves were collected from Narayangonj, Bangladesh  
106 | on February 2015. Plant was identified by expert [taxonomists](#). After collection, leaves were  
107 | thoroughly washed with clean water then shade dried for several days and oven dried for 24  
108 | hours at not more than 40 °C for better grinding. The dried leaves were then ground to a coarse  
109 | powder. The powder (140 g) was subjected to cold maceration by soaking in 1200 mL of ethanol  
110 | in a clean, amber colored reagent bottle. The container was kept for a period of 10 days with  
111 | occasional shaking and stirring. The whole mixtures were then filtered through a fresh cotton  
112 | plug and finally with a Whatman No.1 filter paper. The filtrate was concentrated at 40°C under  
113 | reduced pressure to evaporate approximately 70% of the solvent to obtain the crude extract, CE.  
114 | 5.0 g of CE was dissolved in 10% aqueous ethanol. Solvent-solvent partitioning of this solution  
115 | was done using the protocol designed by Kupchan and modified by Van Wagenet al.,  
116 | (1993) with petroleum ether, then with carbon tetrachloride and finally with Chloroform to obtain  
117 | three fractions (PESF, CTCSF and CSF) and the remaining part was named as aqueous soluble  
118 | fraction (AQSF)<sup>24</sup>.

### 119 *Phytochemical Screening*

120 | Approximately 5 mg of CE was diluted in ethanol to perform the following tests for  
121 | identification of different chemical groups<sup>25,26</sup>.

### 122 *Tests for tannins*

123 Ferric Chloride Test: 5 mL of the extract solution was taken in a test tube. 1 mL of 5% ferric  
124 chloride solution was added to it. Greenish black precipitate ~~will~~ indicated the presence of  
125 tannins.

126 Potassium Dichromate Test: 1 mL of 10% potassium dichromate solution was added ~~with to~~ 5  
127 mL of the extract solution in a test tube. Yellow precipitate ~~will~~ suggested the presence of  
128 tannins.

129 Lead Acetate Test: 1 mL of 10% lead acetate solution was added to 5 mL of extract solution.  
130 Presence of tannins ~~will be~~ was indicated by yellow precipitate.

### 131 ***Test for Flavonoids***

132 A few drops of concentrated hydrochloric acid were added to a small amount of CE. Immediate  
133 red color formation the presence of is caused by flavonoids.

134

### 135 ***Test for Saponins***

136 1 mL of the extract solution was diluted with distilled water to 20 mL and shaken in a graduated  
137 cylinder for 15 minutes. One-centimeter layer of foam formation ~~will~~ indicated the presence of  
138 saponins.

### 139 ***Tests for Gums***

140 5 mL extract solution of the extract was taken and then molisch reagent and sulfuric acid were  
141 added. Red violet ring at the junction of the two liquids evidences s presence of gums.

### 142 ***Tests for Steroids***

143 1 mL concentrated sulphuric acid was added to 1mL extract solution of the chloroform extract.  
144 Red color in the lower layer ~~will~~ indicated the presence of steroids.

### 145 ***Tests for alkaloids***

146 Mayer's test: 2 mL extract solution and 0.2 mL of dilute hydrochloric acid were taken in a test  
147 tube. Then 1 mL of Mayer's reagent was added. Yellowish buff colored precipitate ~~is~~ indicated  
148 ~~of the~~ presence of alkaloids.

149 Dragendroff's test: 2 mL extract solution and 0.2 mL of dilute hydrochloric acid were taken in a  
150 test tube. Then 1 mL of Dragendroff's reagent was added. Orange brown precipitate evidences  
151 the presence of alkaloids.

152 Hager's test: 2 mL extract solution of the extract and 0.2 mL of dilute hydrochloric acid were  
153 taken in a test tube. Then 1 mL of picric acid solution or Hager's reagent was added. Yellowish  
154 precipitate evidences presence of alkaloids.

### 155 ***Test for Reducing Sugar***

156 2 mL of aqueous extract of the plant material was added to 1mL of a mixture of equal volumes  
157 of Fehling's solutions A and B, boiled for few minutes. Brick red colored precipitate ~~will~~  
158 indicated presence of reducing sugar.

### 159 ***Thrombolytic activity***

160 As a part of exploration of cardio protective drugs from natural resources the extract of *E.*  
161 *fluctuans* was assessed for thrombolytic activity using *in vitro* thrombolytic model<sup>27</sup>. Taking  
162 account of all ethical considerations, and aseptic precautions, 20 mL of venous blood was drawn  
163 from healthy human volunteer without a history of oral contraceptive or anticoagulant therapy.  
164 This withdrawn blood was then distributed in pre-weighed sterile vials (1 mL/tube) and  
165 incubated at 37 °C for 45 minutes. After clot formation, the serum was completely removed  
166 without disturbing the clot and clot weight was determined (clot weight = weight of clot  
167 containing vial – weight of vial alone). 100 mg of CE was dissolved in 10 mL of ethanol and kept  
168 overnight. Then the soluble supernatant was decanted and filtered. 100 µL of the extract solution  
169 was added to a vial containing pre-weighed blood clot. Lyophilized Streptokinase (SK) vial of  
170 15,00,000 I.U. was reconstituted with 5 ml sterile distilled water. 100µL (30,000 I.U) of the  
171 solution was added as positive control and 100 µL of distilled water was added as a negative non  
172 thrombolytic control to the vial containing pre-weighed blood clot separately. All the three vials  
173 were then incubated at 37 °C for 90 minutes and observed for clot lysis. After incubation, the  
174 released fluid was removed and vials were again weighed to observe the weight difference after  
175 clot disruption. This difference of weight before and after clot lysis was expressed as percentage  
176 of clot lysis as shown below:

177 **% of clot lysis = (wt. of released clot/clot wt.) × 100**

178 *Membrane stabilization activity*

179 To assess the anti-inflammatory activity, membrane stabilization potential of CE was evaluated  
180 by measuring the heat and hypotonic solution induced haemolysis of erythrocyte following  
181 standard protocol<sup>28</sup>.

182 ***Preparation of erythrocyte suspension***

183 Fresh whole blood (20 mL) was collected intravenously in syringe containing 3.1% sodium  
184 citrate solution as anticoagulant from healthy human volunteer without history of non steroidal  
185 anti-inflammatory drugs (NSAIDs) therapy for 2 weeks prior the experiment. The blood cells  
186 were washed three times with isotonic buffer solution, 154 mM NaCl in 10 mM sodium  
187 phosphate buffer (pH 7.4) through centrifugation at 3000 g for 10 min.

188 ***Heat induced haemolysis***

189 CE was dissolved in isotonic solution. Isotonic buffer containing aliquots (5 mL) of the extract at  
190 1.0 mg/mL was taken in 6 centrifuge tubes in three sets of two. Two sets of control tubes were  
191 taken containing 5 mL of the vehicle and 5 mL of 0.1 mg/mL of acetyl salicylic acid (ASA),  
192 respectively. Erythrocyte suspension was added to each tube and mixed gently by inversion. One  
193 pair of the tubes was incubated at 56°C for 30 min in a water bath, while the other pair was  
194 maintained at (0-5) °C in an ice bath. The reaction mixture was centrifuged for 5 min at 2500g  
195 and the absorbance of the supernatant was measured at 560 nm. The percentage inhibition or  
196 acceleration of hemolysis was calculated according to the equation:

197 % Inhibition of hemolysis =  $100 \times [1 - (OD_2 - OD_1) / (OD_3 - OD_1)]$

198 Where,

199 OD<sub>1</sub>= optical density of unheated test sample; OD<sub>2</sub>= optical density of heated test sample; OD<sub>3</sub>=  
200 optical density of heated control sample

201 ***Hypotonic solution induced haemolysis***

Formatted: Font: Not Bold, Not Italic

202 5 mL hypotonic solution (distilled water) containing 1.0mg/mL of CE were put in centrifuge  
203 tube in triplicates. 5 mL of hypotonic solution and 5 mL of acetyl salicylic acid (ASA) at 0.1  
204 mg/mL concentration was taken as negative and positive control respectively in separate  
205 centrifuge tubes. Erythrocyte suspension (0.5 mL) was added to each of the tubes and mixed  
206 gently. The mixture was incubated for 10 min at room temperature and then centrifuged for 10  
207 min at 3000 g. The absorbance of the supernatant was measured at 540 nm. The percentage  
208 inhibition of haemolysis or membrane stabilization was calculated using the following equation:

209 Inhibition of haemolysis (%) =  $100 [1 - \{(OD_2 - OD_1) / (OD_3 - OD_1)\}]$

210 Where OD<sub>1</sub> = absorbance of test sample in isotonic solution; OD<sub>2</sub> = absorbance of test sample in  
211 hypotonic solution; OD<sub>3</sub> = absorbance of control hypotonic sample

#### 212 Antimicrobial assay

213 | The disc diffusion technique<sup>29</sup> was used for preliminary screening of antimicrobial activity. Two  
214 Gram Positive bacterial strains (*Bacillus megaterium*, *Staphylococcus aureus*), two Gram  
215 Negative bacterial strains (*Escherichia coli*, *Pseudomonas aeruginosa*) and two fungal strains  
216 (*Aspergillus niger* and *Aspergillus flavus*) were collected as pure cultures. PESF, CTCSF, CSF,  
217 AQSF fractions of the crude extract were dissolved separately in specific volume of  
218 dichloromethane or methanol depending on their solubility. The diluted samples were applied on  
219 the sterile discs at a concentration of 500 µg/disc. Kanamycin (30 µg/disc) and Ketoconazole (30  
220 µg/disc) were used as standard antibiotics for antibacterial and antifungal screening, respectively.  
221 Solvent was used as negative control. The antimicrobial activity of the test agent was determined  
222 | by measuring the diameter of zone of inhibition expressed in millimeters.

#### 223 Antioxidant scavenging activity

224 Antioxidant potential was studied using DPPH radical scavenging activity with slight  
225 modification of the method described by Brand-William *et al*<sup>30</sup>. Here, solution of varying  
226 concentrations such as 200 µg/mL, 100 µg/mL, 50 µg/mL, 25 µg/mL, 12.5 µg/mL, 6.25 µg/mL,  
227 3.125 µg/mL, 1.5625 µg/mL and 0.78125 µg/mL were obtained by serial dilution technique in  
228 test tubes where 2 mL of each of the test sample solution was mixed with 2 mL of a DPPH-  
229 methanol solution (20 µg/mL) to obtain the above mentioned concentrations. Test tubes were  
230 allowed to stand for 20 minutes in dark for the reaction to occur. The absorbance was determined  
231 at 517 nm and percentage of inhibition was calculated by using the following equation:

232

233 % inhibition =  $[1 - (ABS_{\text{sample}} / ABS_{\text{control}})] \times 100$ .

234

235 | Then % inhibitions were plotted against respective concentrations used and from the graph IC<sub>50</sub>  
236 | was calculated. Here, ascorbic acid and BHA were used as the positive control.

#### 237 Cytotoxic activity

238 Brine shrimp lethality bioassay technique was applied for the determination of general toxic  
239 | properties of the plant extractives against *Artemia salina*<sup>19,31</sup>. The test samples were dissolved in  
240 dimethyl sulfoxide (DMSO) following serial dilutions to obtain concentrations of 400, 200, 100,  
241 | 50, 25, 12.5, 6.25 and 3.125 µg/mL in separate test tubes containing 10 shrimps in simulated  
242 brine water (5 mL). The test tubes were incubated at room temperature for 24 hours. The LC<sub>50</sub> of

243 the test samples was determined by a plot of percentage of the shrimp mortality against the  
244 logarithm of the sample concentrations. Vincristine sulphate was used as positive control in this  
245 assay to compare the cytotoxicity of the test samples.

246  
247  
248  
249  
250  
251  
252  
253

## 254 RESULT AND DISCUSSION

### 255 *Phytochemical screening*

256 Preliminary phytochemical screening evidenced the presence of alkaloids, saponin, tannins,  
257 flavonoids, reducing sugars and gums (Table1).

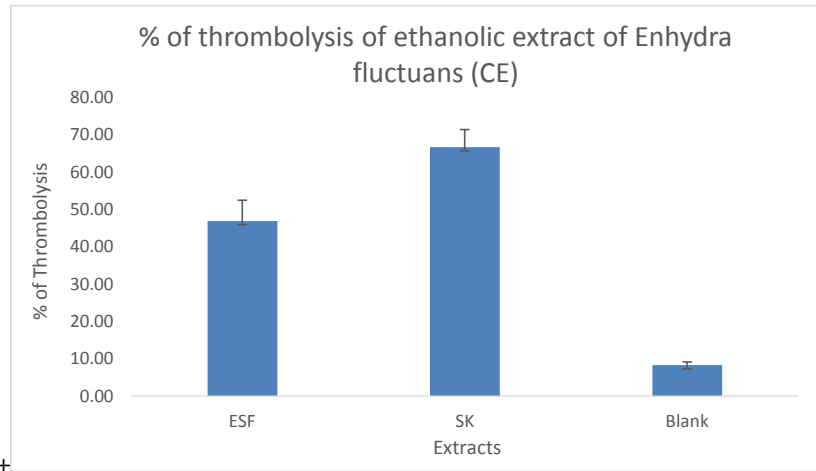
258 Table 1: Results of chemical group tests

Tested groups	Alkaloids	Steroids	Saponin	Tannins	Flavonoids	Reducing Sugars	Gums
Ethanollic Extract of <i>Enhydra fluctuans</i>	+	-	+	+	+	+	+

259 Note: + =Indicates the presence of the tested group, - = Indicates the absence of the tested  
260 group.

### 261 *Thrombolytic activity*

262 The erythrocyte membrane resembles to lysosomal membrane and as such, the effect of drugs on  
263 the stabilization of erythrocyte could be extrapolated to the stabilization of lysosomal  
264 membrane<sup>32,33</sup>. Addition of 100µL SK solution, a positive control to the blood clots and  
265 subsequent incubation resulted in 66.67% lysis of blood clot. On the other hand, negative control  
266 exhibited a negligible lysis of blood clot (8.33%). When blood clots were treated with the test  
267 sample, significant blood clot lysis activity (46.91%) was observed. When compared with the  
268 negative control (water) the mean ~~of~~ percentage (%) of blood clot lysis was significant ( $p <$   
269 0.001).



270 +  
 271 | Figure 1:Thrombolytic activity (in terms of % of **blood** clot lysis) of CE with standard deviation  
 272 error bar where n=3

273 *Membrane stabilizing activity*

274 Flavonoid rich ethyl acetate extract of *E. fluctuans* collected from West Bengal, India showed  
 275 significant anti-inflammatory activity in carrageenan and histamine induced paw edema in rats<sup>34</sup>.  
 276 The ethanol extracts of *E. fluctuans* at concentration 1.0mg/mL significantly protected the lysis  
 277 of human erythrocyte membrane by hypotonic solution and heat induced haemolysis compared  
 278 | to the standard acetyl salicylic acid (0.10mg/ml). For heat induced condition CE demonstrated  
 279 71.80% inhibition of haemolysis of RBCs, whereas acetyl salicylic acid inhibited 77.20%.On the  
 280 other hand, during hypotonic solution induced haemolysis, CE inhibited 47.60% haemolysis of  
 281 RBCs as compared to 71.90% produced by acetyl salicylic acid.

282

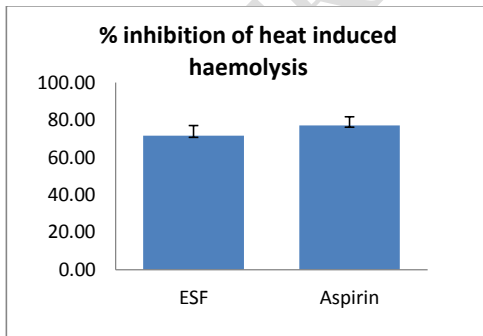


Figure 2:Heat induced haemolysis with standard deviation error bar

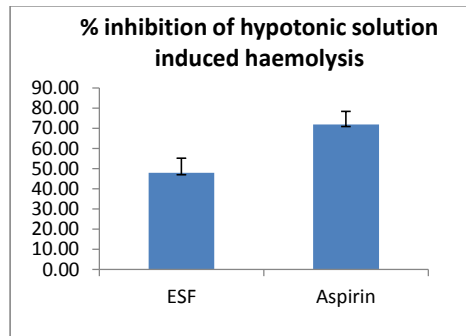


Figure 3:Hypotonic solution induced haemolysis with standard deviation error bar

283 *Antimicrobial assay*



284 | Table-2 shows antimicrobial activity of the tested samples. PESF showed mild activity against  
 285 | gram negative bacteria *Escherichia coli* and *Pseudomonas aeruginosa*. CTCSF exhibited mild  
 286 | activity against both gram positive and gram negative bacteria. All the fractions were inactive  
 287 | against fungal strains. This result is consistent with some antimicrobial screening reports on the  
 288 | plant<sup>35</sup>.

289 | Table 2: Antimicrobial activity of test samples of *E. fluctuans*

Test organisms	Diameter of zone of inhibition(mm)				
	PESF 100µg/dis c	CTSF 100µg/di sc	CSF 100µg/di sc	AQSF 100µg/di sc	Ketoconazole 30µg/disc
Gram positive Bacteria					
<i>Bacillus megaterium</i>	–	7 mm	–	–	40 mm
<i>Staphylococcus aureus</i>	–	8 mm	–	–	28 mm
Gram negative Bacteria					
<i>Escherichia coli</i>	7 mm	8 mm	–	–	26 mm
<i>Pseudomonas aeruginosa</i>	7 mm	7 mm	–	–	27 mm
Fungal Strain					
<i>Aspergillus niger</i>	–	–	–	–	26 mm
<i>Aspergillus flavus</i>	–	–	–	–	36 mm

290

291 | *Antioxidant assay*

292 | The antioxidant activity was expressed from-by the IC<sub>50</sub> values of all samples. Comparison of  
 293 | IC<sub>50</sub> values of different fractions and extracts with standard was-is shown in figure 4. AQSF was  
 294 | found to be most potent with lowest IC<sub>50</sub> value. IC<sub>50</sub> values of PESF, CTCSF, CSF, AQSF,  
 295 | trolox, BHA, Vitamin C are 59.59 µg/mL, 29.28 µg/mL, 26.62 µg/mL, 12.27 µg/mL, 2.04  
 296 | µg/mL, 2.21 µg/mL and 0.17 µg/mL respectively. Previous report of *E. fluctuans* collected  
 297 | from West Bengal, India suggested significant antioxidant property of ethyl acetate extract<sup>36</sup>.  
 298 | Also there has been report of isolation of antioxidative carbohydrate polymer from *E. fluctuans*  
 299 | which is most consistent with our finding<sup>37</sup>.

300

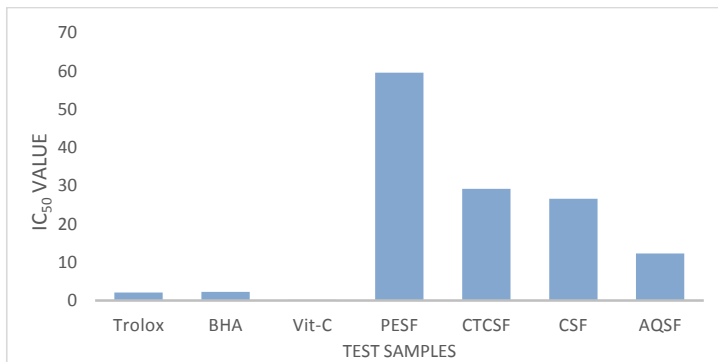


Figure 4: Comparison of  $IC_{50}$  value of different extracts of *E. fluctuans* & standards.

#### Cytotoxicity assay

In case of brine shrimp lethality bioassay, the lethality of the PESF, CTCSF, CSF and AQSF fractions against *A. salina* were shown in Table 3. Lethality assay is proved to be suggestive of different pharmacological properties by isolation of cytotoxic, antimalarial or insecticidal compounds from the cytotoxic plant extracts<sup>38</sup>. Flavonoids isolated from *E. fluctuans* exhibited anticancer activity tested *in vitro* in mice<sup>39</sup>. There is also report of *E. fluctuans* leaves stimulating cell-mediated immune system by increasing neutrophil's phagocytic activity<sup>40</sup>. Here, CTCSF fraction was most potent with the lowest  $LC_{50}$  value (Table 3) which is consistent with the existing reports on *E. fluctuans*.

**Table 3:** Lethal concentration for 50% mortality ( $LC_{50}$ ) of different extracts of *E. fluctuans*.

Sample	PESF	CTCSF	CSF	AQSF
$LC_{50}$ value ( $\mu\text{g/mL}$ )	0.97	0.84	1.43	1.16

#### Conclusion

Preliminary chemical screening and *in-vitro* bioactivity study data suggests *Enhydra fluctuans* potential pharmacological activity. It showed significant cytotoxic activity with good thrombolytic and anti-inflammatory potential. Further higher and detailed experiments for isolation of bioactive compounds from *Enhydra fluctuans* are needed.

#### Conflict of Interest

There is no conflict of interest to declare by the authors.

#### References

- Davies MJ, Thomas AC. Plaque fissuring--the cause of acute myocardial infarction, sudden ischaemic death, and crescendo angina. *British Heart Journal* 1985; 53:363-373.
- DeWood MA, Spores J, Notske R, Mouser LT, Burroughs R, Golden MS, Lang HT. Prevalence of total coronary occlusion during the early hours of transmural myocardial infarction. *The New England Journal of Medicine* 1980; 303:897-902.

Formatted: Font: Italic

- 327 3. Laurence DR, Bennett PN. *Clinical Pharmacology*. Ed 7, Churchill Livingstone, New York,  
328 1992.
- 329 4. Cochrane CG. Cellular injury by oxidants. *American Journal of Medicine* 1991; 91: 23 – 30.
- 330 5. Lipinski B. Hydroxyl radical and its scavengers in health and disease. *Oxidative Medicine*  
331 *and Cellular Longevity* 2011; 2011: Article ID 809696.
- 332 6. Murugasan N, Vember S, Damodharan C. Studies on erythrocyte membrane IV: in vitro  
333 haemolytic activity of oleander extract. *Toxicology Letters* 1981; 8: 33-38.
- 334 7. Morris AK, Masterton RG. Antibiotic resistance surveillance: action for international studies.  
335 *Journal of Antimicrobial Chemotherapy* 2002; 49: 7-10.
- 336 8. Levy SB, Marshall B. Antibacterial resistance worldwide: causes, challenges and responses.  
337 *Nature Medicine* 2004; 10: 122-129.
- 338 9. Taubes G. The bacteria fight back. *Science* 2008; 321: 356-361.
- 339 10. Levy SB. *The antibiotic paradox-How miracle drugs are destroying the miracle*. Plenum Press,  
340 New York, 1992.
- 341 11. Levy SB. Antibiotic resistance: an ecological imbalance. In: *Antibiotic Resistance: Origins,*  
342 *Evolution, Selection and Spread*, editors: Chadwick D, Goode J, John Wiley and Sons,  
343 Chichester, 1997, 1-14.
- 344 12. Gutteridge JM. Free radicals in disease processes: A complication of cause and  
345 consequence. *Free Radical Research Communications* 1993; 19(3): 141-158.
- 346 13. Aruoma OI. Free radicals, oxidative stress, and antioxidants in human health and diseases.  
347 *Journal of the American Oil Chemists' Society* 1998; 75: 199-212.
- 348 14. Halliwell B. The antioxidant paradox. *Lancet* 2000; 355: 1179-1180.
- 349 15. Branen AL. Toxicology and biochemistry of butylated hydroxyanisole and  
350 butylated hydroxytoluene. *Journal of the American Oil Chemists' Society* 1975; 5: 59-63.
- 351 16. Grice HP. Enhanced tumour development by butylated hydroxyanisole (BHA) from the  
352 prospective of effect on fore-stomach and oesophageal squamous epithelium. *Food and*  
353 *Chemical Toxicology* 1988; 26: 717-723.
- 354 17. Wichi HC. Safety evaluation of butylated hydroxytoluene (BHT) in the liver, lung and  
355 gastrointestinal tract. *Food and Chemical Toxicology* 1986; 24: 1127-1130.
- 356 18. Persoone G. *Proceeding of the International Symposium on Brine Shrimp, Artemiasalina.*  
357 *Universa Press, Witteren, 1980, 1-3.*
- 358 19. Meyer BN, Putnam JE, Jacobsen LB, Nichols DE, and McLaughlin JL. *Planta Medica* 1982;  
359 45: 31-32.
- 360 20. Bora P, Kumar Y. *Floristic diversity of Assam. Study of pabitorawildlife sanctuary*. Daya  
361 *Publishing House, Delhi, 2003.*
- 362 21. Nadkarni AK. *The Indian materia medica*. Vol. 2, Popular Prakashan, Bombay, 1999.
- 363 22. Kirtikar KR, Basu BD. *Indian Medicinal Plants*. Vol. 2, International Book Distributors, Dehra  
364 *Dun, 1999.*
- 365 23. Chatterjee A, Pakrashi S. *The treatise on Indian medicinal plants*. Vol. 5, Publication and  
366 *Information Directorate, CSIR, Delhi, 1997.*
- 367 24. Van Wagenen BC, Larsen R, Cardellina JH III, Ran dazzo D, Lidert ZC, Swithenbank C.  
368 Ulosantoin a potent insecticide from the sponge Ulosaruetzleri. *Journal of Organic*  
369 *Chemistry* 1993; 58: 335-337.
- 370 25. Tiwari P, Kumar B, Kaur M, Kaur G, Kaur H. Phytochemical screening and extraction: a  
371 review. *Internationale Pharmaceutica Scientia* 2011; 1: 103-104.

- 372 26. Ahmed ZU, Bithi SS, Khan MM, Hossain MM, Suriya S, Rony SR. Phytochemical  
373 screening, antioxidant and cytotoxic activity of fruit extracts of *Calamustenuis*Roxb. Journal  
374 of Coastal Life Medicine. 2014;2(8):645-50.
- 375 27. Prasad S, Kashyap RS, Deopujari JY, Purohit HJ, Taori GM, Dagainawala HF. Development  
376 of an *in vitro* model to study clot lysis activity of thrombolytic drugs. Thrombosis Journal  
377 2006; 4: 14.
- 378 28. Shinde UA, Phadke AS, Nair AM, Mungantiwar AA, Dikshit VJ, Saraf MN. Membrane  
379 stabilizing activity - a possible mechanism of action for the anti-inflammatory activity of  
380 *Cedrusdeodarawood* oil. Fitoterapia 1999; 70: 251-257.
- 381 29. Bauer AW, Kirby WMM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standard  
382 single disc method. American Journal of Clinical Pathology 1966; 45: 493-96.
- 383 30. William WB, CuvelierME, Berset C. Use of Free Radical Method to Evaluate Antioxidant  
384 Activity. LWT-Food Science and Technology 1995; 28(1): 25- 30.
- 385 31. Maclaughlin JL, Anderson JE, Rogers LL. The use of biological assays to evaluate  
386 botanicals. Drug Information Journal 1998; 32: 513-524.
- 387 32. Omale J, Okafor PN. Comparative antioxidant capacity, membrane stabilization, polyphenol  
388 composition and cytotoxicity of the leaf and stem of *Cissusmultistriata*. African Journal of  
389 Biotechnology 2008; 7:3129-3133.
- 390 33. Shahriar M, Khair NZ, Sheikh Z, Chowdhury SF, Kamruzzaman M, Bakhtiar MSI *et al.*  
391 Characterization of phytoconstituents and potential bioactivity of *Annonareticulata*L. leaf  
392 extract. Journal of Pharmacognosy and Phytochemistry 2016; 5(1): 42-45.
- 393 34. Sannigrahi S, Mazumder UK, Pal D, Mishra SL, Maity S. Flavonoids of *EnhydraFluctuans*  
394 exhibits analgesic and anti-inflammatory activity in different animal models. Pakistan journal  
395 of Pharmaceutical Sciences 2011; 24(3): 369-375.
- 396 35. Bhakta J, Majumdar P, Munekage Y. Antimicrobial efficacies of methanol extract of  
397 *Asteracanthalongifolia*, *Ipomoea aquatica* and *Enhydrafluctuans* against *Escherichia coli*,  
398 *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Micrococcus luteus*. Internet Journal  
399 of Alternative Medicine 2009;7(2).
- 400 36. Sannigrahi S, Mazuder UK, Pal DK, Parida S, Jain S. Antioxidant potential of crude extract  
401 and different fractions of *Enhydrafluctuans*Lour. Iranian journal of pharmaceutical  
402 research 2010;9(1):75.
- 403 37. Ghosh D, Ray S, Ghosh K, Micard V, Chatterjee UR, Ghosal PK, Ray B. Antioxidative  
404 carbohydrate polymer from *Enhydrafluctuans* and its interaction with bovine serum albumin.  
405 Biomacromolecules 2013; 14(6):1761-1768.
- 406 38. Krishnaraju AV, Rao TV, Sundararaju D, Vanisree M, Tsay HS, Subbaraju GV. Biological  
407 screening of medicinal plants collected from Eastern Ghats of India using  
408 *Artemiasalina*(brine shrimp test). International Journal of Applied  
409 Science and Engineering 2006; 4(2):115-125.
- 410 39. Sannigrahi S, Mazumder UK, Mondal A, Pal D, Mishra SL, Roy S. Flavonoids of  
411 *Enhydrafluctuans* exhibit anticancer activity against Ehrlich's ascites carcinoma in mice.  
412 Natural product communications. 2010; 5(8):1239-1242.
- 413 40. Patil KS, Majumder P, Wadekar RR. Effect of *Enhydrafluctuans*Lour. leaf extract on  
414 phagocytosis by human neutrophils. Journal of Natural Remedies 2008; 8(1):76-81.