## Original Research Article

# Phytochemical and Pharmacological Potential of *Enhydra fluctuans* available in Bangladesh

#### ABSTRACT

Objectives: The possible phytochemical natureconstituents, thrombolytic and membrane stabilizing nativity nactivities of the crude ethanolic extract of Enhydra fluctuans (CE) were investigated along with the anti-microbial, antioxidant and cytotoxic potentials of its petroleum ether (PESF), carbon tetrachloride (CTCSF), chloroform (CSF) and aqueous (AQSF) soluble fractions.

Materials & Methods: The coarse leaf powder was extracted at room temperature with ethanol. Solvent-solvent partitioning was done to obtain the four soluble fractions by the modified Kupchan method. Anticoagulant potential was determined by the *in\_vitro\_thrombolytic* model, membrane stabilization method was used to assess *in vitro\_anti-inflammatory* activity, the disc diffusion method was used for anti-microbial screening, antioxidant potential was determined by 2,2-diphenyl-1-picrylhydrazyl (DPPH)\_radical scavenging method and brine shrimp lethality bioassay method was used for cytotoxic activity determination.

**Results:** Chemical screening of the crude extract\_evidences the presence of alkaloids, saponins, tannins, flavonoids, reducing sugars and gums. It showed significant clot lysis property of 49.29%. It also significantly inhibited heat and hypotonic solution induced lysis of the human red blood cell membrane with values of 71.67% and 47.93%, respectively. CTCSF and PESF showed mild antimicrobial activity. AQSF showed most prominent antioxidant activity with IC50 value of 12.27  $\mu$ g/mL. CTCSF showed LC50 value of 0.84  $\mu$ g/mL, with most potent cytotoxic activity.

**Conclusion:** Significant thrombolytic, membrane stabilizing, antioxidant and cytotoxic activities of the plant was found from this study. *In vivo* activities and isolation of active compound(s) from this extract are yet to be investigated.

**Keywords:** *Enhydra fluctuans*, thrombolytic—activity, membrane stabilizing—activity, antioxidant activity—and cytotoxic—activity.

#### INTRODUCTION

Medicinal plants have played an important role in treating various diseases since ancient times.

Increased drug resistance and side effects of existing pharmaceutical drugs have escalated the
research based study on traditionally available plants. Myocardial or cerebral infraction and other
atherothrombotic diseases are consequences of thrombus formed in blood vessel<sup>1,2</sup> Fibrinolytic
drugs like-such as tissue plasminogen activator (t-PA), urokinase, streptokinase ete.and others

Formatted: Adjust space between Latin and Asian text, Adjust space between Asian text and numbers

Formatted: Don't adjust space between Latin and Asian text, Don't adjust space between Asian text and numbers

- 40 dissolve thrombin in acutely occluded coronary arteries and restore blood supply to ischemic
- 41 myocardium, to limit necrosis and improve prognosis<sup>3</sup>. Yet all the available thrombolytic agents
- 42 have significant deficiencies, including the necessity of large doses to be maximally effective,
- 43 limited fibrin specificity and a significant associated bleeding tendency. Therefore, studies are
- 44 going on to develop improved thrombolytic drugs in order to minimize deficiencies of the
- 45 available drugs.
- 46 Inflammation is a pathological disorder, in which inflammatory cells produce a complex mixture
- 47 of growth and differentiation cytokines as well as physiologically active arachidonate
- 48 metabolites, also generate reactive oxygen species (ROS) that can damage cellular biomolecules
- 49 which in turn augment the state of inflammation<sup>4</sup>. Compounds that possess radical scavenging
- ability may therefore expect to have the therapeutic potentials against inflammation Moreover,
- 51 stabilization of lysosomal membrane limits inflammatory response by inhibiting the release of
- 52 lysosomal constituents of activated neutrophil such as bactericidal enzymes and proteases, which
- are responsible for further tissue inflammation and damage<sup>6</sup>.
- Bacteria are responsible for many infectious diseases in 21<sup>st</sup> century<sup>7</sup>. Antibiotic resistance has
- become a major clinical and public health problem for most people now-a-days<sup>8,9</sup>. This
- 56 Multidrug multidrug Resistance resistance (MDR) is clearly related to the misusage of different
- 57 antibiotics 10,11. The increasing clinical importance of drug resistant bacterial pathogens
- 58 | necessitates emergence of additional antibacterial therapy. The antibacterial screening which is
- 59 the first stage of antibacterial research is performed to ascertain the susceptibility of various
- 60 bacteria to any agent.
- Free radicals are highly toxic and reactive; and cause diseases like aging, atherosclerosis, cancer,
- 62 diabetes, liver cirrhosis, cardiovascular disorders, etc. by attacking different macro-molecules
- 63 including lipids, proteins and DNA resulting in the cellular damage<sup>12,13</sup>. Antioxidants protect
- 64 body by delaying or reducing the oxidation of the substrate in turn neutralize or terminate the
- 65 chain reaction before vital molecules within the body are damaged 14. Currently available
- 66 synthetic antioxidants like butylated hydroxyl anisole (BHA), butylatedhydroxy toluene (BHT),
- 67 tertiary butylated hydroquinone and gallic acid esters show low solubility and moderate
- antioxidant activity with suspected negative health effects<sup>15</sup>. BHA and BHT are suspected of
- 69 being responsible for liver toxicity and carcinogenesis 16,17. Investigations to screen out naturally
- 70 occurring antioxidants are now mostly demanding.
- 71 Bioactive compounds are always toxic to living body at some higher doses and it justifies the
- 72 statement that 'Pharmacology is simply toxicology at higher doses and toxicology is simply
- 73 pharmacology at lower doses'. Brine shrimp lethality bioassay implies cytotoxicity as well as a
- 74 wide range of pharmacological activities such as antimicrobial, antiviral, pesticidal pesticides and
- 75 anti-tumor etc.and others of the compounds 18,19.
- 76 Enhydra fluctuans Lour (Family: Asteraceae) is a small genus of marsh herb, available in
- 77 | tropical and subtropical regions; likenamely Bangladesh, India, Malaysia, China and the rest of
- South East Asia and Tropical Africa 20,21. The herb is relatively glabrous sometimes pubescent
- 79 glandular. Stems are usually 0.3-to 0.6 m, elongated simple or divaricating rooting at the nodes<sup>22</sup>.
- 80 Traditional uses indicate that the leaves of the plant are good as laxative. The expressed juice of
- 81 the leaves is an excellent demulcent in gonorrhoeagonorrhea and also used in the treatment of
- 82 skin and nervous system<sup>23</sup>. These activities can be attributed mainly to the presence of
- 83 phytochemicals such as flavonoids, alkaloids, saponins, tannins, phenols and carbohydrates. But

there are insufficient records in literature of this plant, regarding its pharmacological activities 84 85

and phytochemical characteristics. Thus the present study focuses on screening of the ethanolic

extract of Enhydra fluctuans Lour for its phytochemical and pharmacological potential. 86

#### MATERIALS AND METHODS

89 Instrumentation

87 88

90 The extract was condensed concentrated in rotary vacuum evaporator (Heidolph Instruments

- 91 GmbH & Co. KG, Germany). Samples were centrifuged on a bench top centrifuge machine
- 92 (HERMLE Labortechnik GmbH, Germany). The absorbance was recorded by a UV-Visible
- 93 visible spectrophotometer (Analytic Jena AG, Germany). Ketoconazole was obtained from
- Beximco Pharmaceuticals Ltd., Bangladesh. Streptokinase vial (Beacon Pharmaceutical Ltd) was 94
- 95 purchased from local pharmacy. Trolox and BHA were obtained from Calbiochem, Merck,
- 96 Germany and VWR International Ltd., England, respectively. Nutrient agar and potato dextrose
- 97 agar and Kanamycin-kanamycin 30 µg standard disc were purchased from HiMedia Laboratories
- Pvt. Ltd., India. All other reagents were purchased from Sigma-Aldrich, USA and all other 98
- 99 solvents used were of analytical grade and purchased from Active Fine Chemicals Ltd.,
- 100 Bangladesh.
- 101 Collection and Identification
- 102 For this investigation, Enhydra fluctuans leaves were collected from Narayangoni, Bangladesh
- 103 on February 2015. Plant was identified by experts. After collection The leaves were thoroughly
- washed with clean water, then shade dried in the shade for several days and oven dried for 24 104
- hours at not more than 40-°C for better grinding. The dried leaves were then ground to a coarse 105
- 106 powder. The powder (140 g) was subjected to cold maceration by soaking in 1200 mL of ethanol
- 107 in a clean, amber colored reagent bottle. The container was kept for a period of 10 days with
- 108 occasional shaking and stirring. The whole mixtures were then filtered through a fresh cotton
- 109 plug and finally with a Whatman No.1 filter paper. The filtrate was concentrated at 40°C under
- reduced pressure to evaporate approximately 70% of the solvent to obtain the crude extract, CE. 110
- 5.0 g of CE was dissolved in 10% aqueous ethanol. Solvent-solvent partitioning of this solution 111
- 112 was done using the protocol designed by Kupchan and modified by Van Wagenenet al., (1993)
- 113 with petroleum ether, then with carbon tetrachloride and finally with Chloroform to obtain three
- 114 fractions (PESF, CTCSF and CSF) and the remaining part was named as aqueous soluble
- fraction (AQSF)<sup>24</sup>. 115
- 116 Phytochemical Screening
- Approximately 5 mg of CE was diluted in ethanol to perform the following\_tests for 117
- identification of different chemical groups<sup>25,26</sup>. 118
- 119 Tests for tannins
- 120 Ferric Chloride Chloride Test: About 5 mL of the solution was taken in a test tube., followed by
- 121 addition of 1 mL of 5% ferric chloride solution was added to it. Greenish black precipitate will
- 122 indicates the presence of tannins.
- 123 Potassium Dichromate dichromate Test: About 1 mL of 10% potassium dichromate solution was
- 124 added with 5 mL of the extract solution in a test tube. Yellow precipitate will suggest the
- 125 presence of tannins.

Comment [SaA1]: Is this true?

Comment [SaA2]: To state the name and designation

- 126 Lead Acetate acetate Test: About 1 mL of 10% lead acetate solution was added to 5 mL of
- extract solution. The Ppresence of tannins will bewere indicated by yellow precipitate.
- 128 Test for Flavonoids
- 129 A few drops of concentrated hydrochloric acid were added to a small amount of CE. Immediate
- red color formation is caused by flavonoids.
- 131 Test for Saponins
- 132 About 1 mL of the solution was diluted with distilled water to 20 mL distilled water and shaken
- 133 in a graduated cylinder for 15 minutes. One-centimeter layer of foam formation will indicate the
- presence of saponins.
- 135 Tests for Gums
- 136 About 5 mL solution of the extract was taken and then added to molisch reagent and sulfuric acid
- were added. Red violet ring at the junction of the two liquids evidences presence of gums.
- 138 Tests for Steroids
- 139 About 1 mL concentrated sulphurie sulfuric acid was added to 1 mL solution of the chloroform
- extract. Red color in the lower layer will indicate the presence of steroids.
- 141 Tests for alkaloids
- 142 | Mayer's test: About 2 mL solution and 0.2 mL of dilute hydrochloric acid were taken in a test
- tube. Then 1 mL of Mayer's reagent was added. Yellowish buff colored precipitate is indicative
- 144 <u>indicates of the presence of alkaloids.</u>
- 145 Dragendroff's test: About 2 mL solution and 0.2 mL of dilute hydrochloric acid were taken in a
- 146 test tube. Then 1 mL of Dragendroff's reagent was added. Orange brown precipitate evidences
- the presence of alkaloids.
- 148 | Hager's test: About 2 mL solution of the extract and 0.2 mL of dilute hydrochloric acid were
- taken in a test tube. Then 1 mL of picric acid solution or Hager's reagent was added. Yellowish
- precipitate evidences presence of alkaloids.
- 151 Test for Reducing reducing Sugar sugar
- 152 About 2 mL of aqueous extract of the plant material—was added to 1\_mL of a mixture of equal
- volumes of Fehling's solutions A and B, boiled for few minutes. Brick red colored precipitate
- will indicate presence of reducing sugar.
- 155 Thrombolytic activity
- 156 As a part of exploration of cardio protective drugs from natural resources the eThe extract of E.
- 157 fluctuans was assessed for thrombolytic activity using in vitro thrombolytic model<sup>27</sup>. Taking
- 158 account of all ethical considerations, and aseptic precautions, 20 mL of venous blood was drawn
- 159 from healthy human volunteer without a history of oral contraceptive or anticoagulant therapy.
- 160 This withdrawn blood was\_then distributed in pre-weighed sterile vials (1 mL/tube) and
- 161 incubated at 37-°C for 45 minutes. After clot formation, the serum was completely removed
- 162 without disturbing the clot and clot weight was determined (clot weight = weight of clot
- 163 containing vial weight of vial alone). About 100 mg of CE was dissolved in 10 mL of ethanol
- and kept overnight. Then the The soluble supernatant was decanted and filtered. Approximately
- 165 | 100 µL of the solution was added to a vial containing pre-weighed clot. Lyophilized

Streptokinase streptokinase (SK) vial of 15,00,000 I.U. was reconstituted with 5 ml-mL sterile 167 distilled water. About 100 µL (30,000 I.U) of the solution was added as positive control and 100 168 μL of distilled water was added as a negative non thrombolytic control to the vial containing pre-169 weighed clot separately. All the three vials were then incubated at 37-°C for 90 minutes and 170 observed for clot lysis. After incubation, the released fluid was removed and vials were again 171 weighed to observe the weight difference after clot disruption. This difference of weight before and after clot lysis was expressed as the percentage of clot lysis as shown belowthe following: 172 Comment [SaA3]: Please include reference for 173 %-Percentage of clot lysis = (wt. of released clot/clot wt.)  $\times$  100 174 Membrane stabilization activity 175 To assess the anti-inflammatory activity, membrane stabilization potential of CE was evaluated 176 by measuring the heat and hypotonic solution induced haemolysis hemolysis of erythrocyte following standard protocol<sup>28</sup>. 177 178 Preparation of erythrocyte suspension 179 Fresh whole blood (20 mL) was collected intravenously in syringe containing 3.1% sodium 180 citrate solution as anti-coagulant from healthy human volunteer without history of non steroidal 181 anti-inflammatory drugs (NSAIDs) therapy for 2 weeks prior the experiment. The blood cells 182 were washed three times with isotonic buffer solution, 154 mM NaCl in 10 mM sodium phosphate buffer (pH 7.4) through centrifugation at 3000 g for 10 min. 183 184 Heat induced haemolysishemolysis CE was dissolved in isotonic solution. Isotonic buffer containing aliquots (5 mL) of the extract at 185 186 1.0 mg/mL was taken in 6-six centrifuge tubes in three sets of two. Two sets of control tubes were taken-containing 5 mL of the vehicle control and 5 mL of 0.1 mg/mL of acetyl salicylic 187 188 acid (ASA), respectively. Erythrocyte suspension was added to each tube and mixed gently by 189 inversion. One pair of the tubes was incubated at 56°C for 30 min in a water bath, while the other 190 pair was maintained at (0-to 5)-°C in an ice bath. The reaction mixture was centrifuged for 5 min at 2500 g and the absorbance of the supernatant was measured at 560 nm. The percentage 191 192 inhibition or acceleration of hemolysis was calculated according to the equation: Comment [SaA4]: Please include the reference! %—Percentage of Inhibition of hemolysis =  $100 \times [1 - (OD_2 - OD_1)/(OD_3 - OD_1)]$ 193 194 Where. 195  $OD_1$ = optical density of unheated test sample;  $OD_2$ = optical density of heated test sample;  $OD_3$ = 196 optical density of heated control sample 197 Hypotonic solution induced haemolysishemolysis 198 About 5 mL hypotonic solution (distilled water) containing 1.0 mg/mL of CE were put in 199 centrifuge tube in triplicates. About 5 mL of hypotonic solution and 5 mL of acetyl salicylic acid 200 (ASA) at 0.1 mg/mL concentration was taken as negative and positive controls respectively in 201 separate centrifuge tubes. Erythrocyte suspension (0.5 mL) was added to each of the tubes and 202 mixed gently. The mixture was incubated for 10 min at room temperature and then centrifuged 203 for 10 min at 3000 g. The absorbance of the supernatant was measured at 540 nm. The percentage inhibition of haemolysishemolysis or membrane stabilization was calculated using

**Comment [SaA5]:** Please include the reference!

166

204 205

206

the following equation:

Inhibition of haemolysis ( $\frac{\%}{}$ ) = 100 [1-{(OD<sub>2</sub>-OD<sub>1</sub>)/(OD<sub>3</sub>-OD<sub>1</sub>)}]

- 207 Where  $OD_1$  = absorbance of test sample in isotonic solution;  $OD_2$  = absorbance of test sample in 208 hypotonic solution;  $OD_3$  = absorbance of control hypotonic sample
- 209 Antimicrobial assay
- The disc diffusion technique<sup>29</sup> was used for preliminary screening of antimicrobial activity. Two 210
- 211 Gram Positive bacterial strains (Bacillus megaterium, Staphylococcus aureus), two Gram
- 212 Negative bacterial strains (Escherichia coli, Pseudomonas aeruginosa) and two fungal strains
- 213 (Aspergillus niger and Aspergillus flavus) were collected as pure cultures. PESF, CTCSF, CSF,
- 214 AQSF fractions of the crude extracts were dissolved separately in specific volume of
- 215 dichloromethane or methanol depending on their solubility. The diluted samples were applied on
- 216 the sterile discs at a concentration of 500 µg/disc. Kanamycin (30 µg/disc) and Ketoconazole 217 ketoconazole (30 μg/disc) were used as standard antibiotics for antibacterial and antifungal
- 218 screening, respectively. Solvent was used as negative control. The antimicrobial activity of the
- 219 test agent was determined by measuring the diameter of zone of inhibition expressed in
- 220 millimetermm.
- 221 Antioxidant scavenging activity
- 222 Antioxidant potential was studied using DPPH radical scavenging activity with slight
- modification of the methodas described by Brand-William et al<sup>30</sup>. Here, solution of varying 223
- concentrations such as 200 µg/mL, 100 µg/mL, 50 µg/mL, 25 µg/mL, 12.5 µg/mL, 6.25 µg/mL, 224
- 225 3.125 µg/mL, 1.5625 µg/mL and 0.78125 µg/mL were obtained by serial dilution technique in
- 226 test tubes, where 2 mL of each of the test sample solution was mixed with 2 mL of a DPPH-
- 227 methanol solution (20 µg/mL) to obtain the above mentioned concentrations. Test tubes were
- allowed to stand for 20 minutes in dark for the reaction to occur. The absorbance was determined 228
- 229 at 517 nm and percentage of inhibition was calculated by using the following equation:
- 230
- 231  $\frac{\text{--Percentage of inhibition of radical scavenging}}{\text{--Inhibition of radical scavenging}} = [1 - (ABS_{sample} / ABS_{control})] \times 100.$
- 232 233
- Then % inhibitions were plotted against respective concentrations used and from the graph IC<sub>50</sub> 234 was calculated. Here, ascorbic acid and BHA were used as the positive controls.
- 235 Cytotoxic activity
- 236 Brine shrimp lethality bioassay technique was applied for the determination of general toxic
- 237 properties of the plant extractives extracts against Artemiasalina<sup>19,31</sup>. The test samples were
- dissolved in dimethyl sulfoxide (DMSO) following serial dilutions to obtain concentrations of 238
- 239 400, 200, 100, 50, 25, 12.5, 6.25 and 3.125 µg/mL in separate test tubes containing 10 shrimps in
- 240 simulated brine water (5 mL). The test tubes were incubated at room temperature for 24 hours.
- 241 The LC<sub>50</sub> of the test samples was determined by a plot of percentage of the shrimp mortality
- 242 against the logarithm of the sample concentrations. Vincristine sulphate sulfate was used as
- 243 positive control in this assay to compare the cytotoxicity of the test samples.
- 244 245
- 246

- - Comment [SaA8]: To state the culture method and medium used for the assay

Comment [SaA7]: Please include the reference!

Comment [SaA6]: To state culture agar and

conditions for assay?

## RESULTS AND DISCUSSION

248 Phytochemical screening

247

251

255

256

257258

259

260

261

262

263

264

249 Preliminary phytochemical screening evidences the presence of alkaloids, saponins, tannins,

250 flavonoids, reducing sugar and gums (**Table1**).

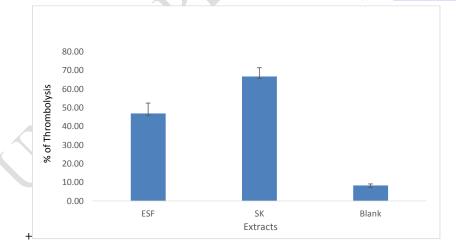
Table 1: Results of chemical group tests.

Tested groups	Alkaloids	Steroids	Saponin	Tannins	Flavonoids	Reducing Sugars	Gums
Ethanolic Extract of Enhydra fluctuans	+	-	+	+	+	+	+

Note: + = Indicates the presence of the tested group, - = Indicates the absence of the tested group.

254 Thrombolytic activity

The erythrocyte membrane resembles to lysosomal membrane and as such, the effect of drugs on the stabilization of erythrocyte could be extrapolated to the stabilization of lysosomal membrane  $^{32,33}$ . Addition of  $100\mu L$  SK solution, a positive control to the clots and subsequent incubation resulted in 66.67% lysis of clot. On the other hand, negative control exhibited a negligible lysis of clot (8.33%). When clots were treated with the test sample significant clot lysis activity (46.91%) was observed. When compared\_with the negative control (water) the mean of percentage (%) of clot lysis was significant (p < 0.001). Figure 1 shows ??????????



**Figure 1**: Thrombolytic activity (in terms of % of clot lysis) of CE with standard deviation error-bar where n=3.

**Comment [SaA9]:** State the amount: low, intermediate or highly presence!

Formatted: Font: Bold
Formatted: Font: Bold

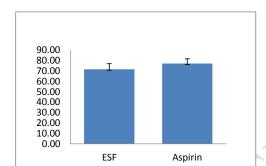
**Comment [SaA10]:** Please describe the figure in detail!

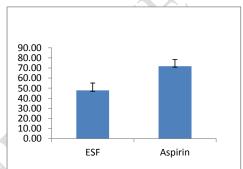
Formatted: Font: Bold

Formatted: Justified

### Membrane stabilizing activity

Flavonoid rich ethyl acetate extract of *E. fluctuans* collected from West Bengal, India showed significant anti-inflammatory activity in carrageenan and histamine induced paw edema in rats<sup>34</sup>. The ethanol extracts of *E. fluctuans* at concentration 1.0\_mg/mL significantly protected the lysis of human erythrocyte membrane by hypotonic solution and heat induced haemolysishemolysis compared to the standard acetyl salicylic acid (0.10\_mg/ml). For heat induced condition CE demonstrated 71.80% inhibition of haemolysishemolysis of RBCs, whereas acetyl salicylic acid inhibited 77.20%.On the other hand, during hypotonic solution induced haemolysishemolysis, CE inhibited 47.60% haemolysishemolysis of RBCs as compared to 71.90% produced by acetyl salicylic acid.





**\_Figure 2**: Heat--induced haemolysishemolysis with standard deviation error bar.

**Figure 3**: Hypotonic solution induced haemolysishemolysis with standard deviation error bar.

Antimicrobial assay

**Table-2** shows antimicrobial activity of the tested samples. PESF showed\_mild activity against gram\_Gram\_negative bacteria *Escherichia coli* and *Pseudomonas aeruginosa*. CTCSF exhibited mild activity against both gram\_Gram\_positive and gram\_Gram\_negative bacteria.\_All the fractions were inactive against fungal strains.\_This result is consistent with some antimicrobial screening reports on the plant<sup>35</sup>.

**Comment [SaA11]:** Please explain Figures 2 and 3 in the paragraph!

Comment [SaA12]: The figures y-axis and x-axis are not labelled. Standardize the fonts and

Formatted: Font: Bold
Formatted: Font: Bold

Formatted: Font: Bold

**Comment [SaA13]:** Please state the ATCC number for the bacteria?

	Diameter of zone of inhibition(mm)						
Test organisms	PESF	CTSF	CSF	AQSF	Ketoconazole		
Č	100μg/dis	100μg/di	100µg/di	100μg/di	30µg/disc		
	c	sc	sc	sc	30µg/disc		
Gram positive Bacteria							
Bacillus megaterium	_	7 <del>mm</del>	_	_	40 <del>mm</del>		
Staphylococcus aureus	_	8 <del>mm</del>	_	_	28 <del>mm</del>		
Gram negative Bacteria							
Escherichia coli	7 <del>mm</del>	8 <del>mm</del>	_	-	26 <del>mm</del>		
Pseudomonas aeruginosa	7 <del>mm</del>	7 <del>mm</del>	_	4-	27 <del>mm</del>		
Fungal Strain							
Aspergillus_niger	_	_	- /	-	26 <del>mm</del>		
Aspergillus flavus	_	_	-	<u> </u>	36 <del>mm</del>		

Formatted: Centered

Formatted: Font: Bold

Formatted: Centered

Formatted: Font: Bold

Formatted: Font: Bold

# 291292

293

294

295

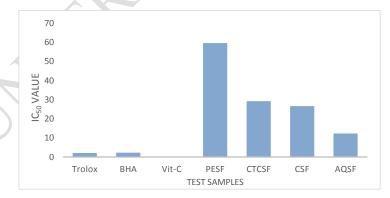
296 297

298 299

## Antioxidant assay

The antioxidant activity was expressed from the IC<sub>50</sub> value of all samples. Comparison of IC<sub>50</sub> values of different fractions and extracts with standard was shown in figure Figure 4. AQSF was found to be most potent with lowest IC<sub>50</sub> value. IC<sub>50</sub> values of PESF, CTCSF, CSF, AQSF, trolox, BHA, Vitamin C are 59.59  $\mu$ g/mL, 29.28  $\mu$ g/mL, 26.62  $\mu$ g/mL, 12.27  $\mu$ g/mL, 2.04  $\mu$ g/mL, 2.21  $\mu$ g/mL and 0.17  $\mu$ g/mL respectively. Previous report of *E. fluctuans*\_collected from West Bengal, India suggest\_significant antioxidant property of ethyl acetate extract<sup>36</sup>. Also there has been report of isolation of antioxidative carbohydrate polymer from *E fluctuans*\_which is the most consistent with our finding<sup>37</sup>.





302303

304

**Figure 4**: Comparison of LC<sub>50</sub> value of different extracts of *E. fluctuans* & standards.

#### Cytotoxicity assay

In case of brine shrimp lethality bioassay, the The lethality of the PESF, CTCSF, CSF and AQSF fractions against A. salina were shown in Table 3. Lethality assay is proved to be suggestive of different pharmacological properties by isolation of cytotoxic, antimalarial or insecticidal compounds from the cytotoxic plant extracts 18. Flavonoids isolated from E. fluctuans exhibited anticancer activity tested in vitro in mice 19. There is also report of E. fluctuans leaves stimulating cell-mediated immune system by increasing neutrophil's phagocytic activity 10. Here, CTCSF fraction was most potent with the lowest LC50 value which is consistent with the existing reports on E. fluctuans.

313314315

305

306

307

308

309

310

311 312

**Table 3:** Lethal concentration for 50% mortality (LC<sub>50</sub>) of different extracts of *E. fluctuans*.

Sample	PESF	CTCSF	CSF	AQSF	
LC <sub>50</sub> value (µg/mL)	0.97	0.84	1.43	1.16	+

316317

318

319

Preliminary chemical screening and *in-vitro* bioactivity study data suggests *Enhydra fluctuans* potential pharmacological activity. It showed significant cytotoxic activity with good thrombolytic and anti-inflammatory potential. Further higher and detailed experiments for isolation of bioactive compounds from *Enhydra fluctuans*.

320321322

### **Conflict of Interest**

323 There is no conflict of interest to declare by the authors.

324325

326

327

328

329

330

334

335

336

337

## References

- 1. Davies MJ, Thomas\_AC. Plaque fissuring—the cause of acute myocardial infarction, sudden ischaemic schemic death, and crescendo angina. British Heart Journal\_1985; 53:363-373.
- 2. DeWood MA, Spores J, Notske R, Mouser LT, Burroughs R, Golden MS, Lang HT. Prevalence of total coronary occlusion during the early hours of transmural myocardial infarction. The New England Journal of Medicine 1980; 303:897-902.
- 331 | 3. Laurence DR, Bennett PN. Clinical Pharmacology. Ed 7, Churchill Livingstone, New York, 1992.
- 333 4. Cochrane CG. Cellular injury by oxidants. American Journal of Medicine 1991;91: 23 30.
  - Lipinski B. Hydroxyl radical and its scavengers in health and disease. Oxidative Medicine and Cellular Longevity 2011; 2011: <u>Article</u>: <u>Article</u> ID 809696.
  - 6. MurugasanN, Vember S, Damodharan C. Studies on erythrocyte membrane IV: <u>#In vitro haemolytichemoly</u>
- Morris AK, Masterton RG. Antibiotic resistance surveillance: action for international studies.
   Journal of Antimicrobial Chemotherapy 2002; 49:7-10.
- Levy SB, Marshall B. Antibacterial resistance worldwide: causes, challenges and responses.
   Nature Medicine 2004; 10:122-129.

Formatted: Font: Bold

**Comment [SaA14]:** Explanation for your results obtained?

Formatted: Centered, Line spacing: single

Formatted: Line spacing: single

Formatted: Font: Italic

- 342 9. Taubes G. The bacteria fight back. Science 2008; 321: 356-361.
- 343 10. Levy SB. The antibiotic paradox-How miracle drugs are destroying the miracle. Plenum Press,
   344 New York, 1992.
- 11. Levy SB. Antibiotic resistance: an ecological imbalance. In: Antibiotic Resistance: Origins,
   Evolution, Selection and Spread, editors: Chadwick D, Goode J, John Wiley and Sons,
   Chichester, 1997, 1-14.
- 348 12. Gutteridgde JM.Free radicals in disease processes: A complication of cause and consequence. Free Radical Research Communications1993;19\_(3):141-158.
- 350 13. Aruoma OI. Free radicals, oxidative stress, and antioxidants in human health and diseases.
  351 Journal of the American Oil Chemists' Society 1998; 75: 199-212.
- 352 14. Halliwell B. The antioxidant paradox. Lancet2000; 355: 1179-1180.
- 15. Branen AL. Toxicology and biochemistry of butylated\_hydroxyanisol and butylated hydroxytoluene. Journal of the American Oil Chemists' Society1975; 5: 59-63.
- 355 | 16. Grice HP. Enhanced tumour development by butylated\_hydroxyanisole (BHA) from the
   356 | prospective of effect on fore-stomach and oesophageal squamous epithelium.
   357 | Food and Chemical Toxicology1988; 26: 717-723.
- 358 17. Wichi HC. Safety evaluation of butylated\_hydroxytoluene (BHT) in the liver, lung and gastrointestinal tract. Food and Chemical Toxicology1986; 24: 1127-1130.
- 18. Persoone G.\_Proceeding of the International Symposium on Brine Shrimp, *Artemiasalina*.
   Universa Press, Witteren, 1980,1-3.
- 362 | 19. Meyer BN, Putnam JE, Jacobsen LB, Nichols DE, and McLaughlin JL. Planta\_Medica 1982;
   363 | 45: 31-32.
- 20. Bora P, Kumar Y. Floristic diversity of Assam. Study of pabitora\_wildlife sanctuary. Daya
   Publishing House, Delhi, 2003.
- 366 21. Nadkarni AK. The Indian materiamaterial medica. Vol. 2, Popular Prakashan, Bombay, 1999.
- 22. Kirtikar KR, BasuBD. Indian Medicinal\_Plants. Vol. 2, International Book Distributors,
   Dehra Dun. 1999.
- 23. Chatterjee A, Pakrashi S. The treatise on Indian medicinal plants. Vol. 5, Publication and
   Information Directorate, CSIR, Delhi. 1997.
- 24. Van Wagenen BC, Larsen R, Cardellina JHII, Ran dazzo D, Lidert ZC,Swithenbank C.
   Ulosantoin a potent insecticide from the sponge Ulosaruetzleri. Journal of Organic Chemistry1993; 58: 335-337.
- 25. Tiwari P, Kumar B, Kaur M, Kaur G, Kaur H. Phytochemical screening and extraction: a review. Internationale Pharmaceutica Sciencia 2011; 1: 103-104.
- 376 26. Ahmed ZU, Bithi SS, Khan MM, Hossain MM, Suriya S, Rony SR. Phytochemical
   377 screening, antioxidant and cytotoxic activity of fruit extracts of *Calamustenuis*\_Roxb. Journal
   378 of Coastal Life Medicine. 2014;2(8):645-50.
- 27. Prasad S, Kashyap RS, Deopujari JY, Purohit HJ, Taori GM, Daginawala HF. Development
   of an *in vitro* model to study clot lysis activity of thrombolytic drugs. Thrombosis Journal
   2006; 4: 14.
- 382 28. Shinde UA, Phadke AS, Nair AM, Mungantiwar AA, Dikshit VJ, Saraf MN. Membrane
   383 stabilizing activity a possible mechanism of action for the anti-inflammatory activity of
   384 *Cedrus deodara* wood oil. Fitoterapia 1999; 70: 251-257.
- 29. Bauer AW, Kirby WMM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standard
   single disc method. American Journal of Clinical Pathology1966; 45: 493-96.

- 387 30. William WB, CuvelierME, Berset C. Use of Free Radical Method to Evaluate Antioxidant Activity.LWT-Food Science and Technology 1995; 28(1): 25- 30.
- 389 31. Maclaughlin JL, Anderson JE, Rogers LL. The use of biological assays to evaluate botanicals. Drug Information Journal 1998; 32: 513-524.

391

392

393

394

395

396

397

398

399

400

401

402

403

- 32. Omale J, Okafor PN. Comparative antioxidant capacity, membrane stabilization, polyphenol composition and cytotoxicity of the leaf and stem of *Cissus\_multistriata*. African Journal of Biotechnology 2008; 7:3129-3133.
- 33. Shahriar M, Khair NZ, Sheikh Z, Chowdhury SF, Kamruzzaman M, Bakhtiar MSI *et al*.

  Characterization of phytoconstituents and potential bioactivity of *Annona\_reticulata\_reticulate*L. leaf extract. Journal of Pharmacognosy and Phytochemistry 2016; 5(1): 42-45.
- 34. Sannigrahi S, Mazumder UK, Pal D, Mishra SL, Maity S. Flavonoids of *Enhydra\_Fluctuans* <u>fluctuans</u> exhibits analgesic and anti-inflammatory activity in different animal models. Pakistan journal of Pharmaceutical Sciences 2011; 24(3): 369-375.
- 35. Bhakta J, Majumdar P, Munekage Y. Antimicrobial efficacies of methanol extract of *Asteracantha\_longifolia*, *Ipomoea aquatica* and *Enhydra\_fluctuans* against *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Micrococcus luteus*. Internet Journal of Alternative Medicine 2009;7(2).
- 404 36. Sannigrahi S, Mazuder UK, Pal DK, Parida S, Jain S. Antioxidant potential of crude extract and different fractions of *Enhydra\_fluctuans\_*Lour. Iranian journal of pharmaceutical research2010;9(1):75.
- 407 37. Ghosh D, Ray S, Ghosh K, Micard V, Chatterjee UR, Ghosal PK, Ray B. Antioxidative carbohydrate polymer from *Enhydra fluctuans* and its interaction with bovine serum albumin.
   409 Biomacromolecules 2013; 14(6):1761-1768.
- 38. Krishnaraju AV, Rao TV, Sundararaju D, Vanisree M, Tsay HS, Subbaraju GV. Biological screening of medicinal plants collected from Eastern Ghats of India using *Artemia\_salina* (brine shrimp test). International Journal of Applied Science and Engineering2006; 4(2):115-125.
- 414 39. Sannigrahi S, Mazumder UK, Mondal A, Pal D, Mishra SL, Roy S. Flavonoids of *Enhydra* 415 fluctuans exhibit anticancer activity against Ehrlich's ascites carcinoma in mice. Natural 416 product communications. 2010; 5(8):1239-1242.
- 417 | 40. Patil KS, Majumder P, Wadekar RR. Effect of *Enhydra\_fluctuans\_*Lour. leaf extract on phagocytosis by human neutrophils. Journal of Natural Remedies 2008; 8(1):76-81.

Formatted: Font: Italic