

Optimum process parameters for activated carbon production from rice husk for phenol adsorption

ABSTRACT (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)

Aim: The determination of optimum process parameters in the production of activated carbon from rice husk for the uptake of phenol from aqueous solution was the focus of this work.

Study design: The optimization was designed using response surface methodology.

Methodology: Central composite design (CCD) was used to generate the design matrix and analyze the result obtained. Carbonization temperature, percentage acid concentration and carbonization time were the factors considered. Tetraoxophosphoric acid (H₃PO₄) was employed in the activation process. The surface area was determined using the Brunauer-Emmet-Teller (BET) nitrogen adsorption method.

Results: The result indicated the optimum process conditions as carbonization temperature of 575 °C, time of 240 minutes and 45 percentage acid concentration. This gave 96.5% adsorption efficiency of phenol from aqueous solution. There was good agreement between the experimental values and the predicted values. The BET surface area of the activated carbon was 471.1 m²/g.

Conclusion: This work has optimized the process conditions for activated carbon production from rice husk for effective adsorption of phenol from wastewater.

12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27

Keywords: Rice husk, activated carbon, optimization, response surface methodology, phenol

1. INTRODUCTION (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)

One of the major drawback of the oil industry in Nigeria is the release of untreated wastewater containing pollutants into water bodies which the host communities use for drinking, cooking, washing, irrigation etc. This practice has not only endangered the immediate environment of the host communities but is also one of the reasons for the constant clashes between the host communities on one hand and the oil companies and the government on another hand.

Phenol is a major pollutant in the wastewater from the oil industry. This is because it has serious unpleasant effects especially on man both long term and short term. [1] reported that the consumption of water containing phenol can result in the damage of the capillaries in man which may lead to death. Even at low concentration, phenol is very harmful to human beings hence they are considered as priority pollutants [2-3]. Diarrhea, excretion of dark

28 urine, impaired vision etc are among other dangerous side effects of phenol [4]. Therefore,
29 treatment of phenolic wastewater is one of the priority needs for the protection of the
30 environment and for peace in the oil industry.

31
32 Different techniques that have been reported for treatment of waste water include
33 electrocoagulation, biodegradation, solvent extraction, chemical oxidation, biological
34 treatment, phase transfer catalysis, adsorption, ion exchange etc. Adsorption has proved to
35 be the most effective and widely used method in treatment of waste water [5,4]. Adsorption
36 is the process where the adsorbate (in this case phenol) is attached on the surface of an
37 adsorbent and hence removed from the solution [6]. Adsorption is an effective method due to
38 its high treatment efficiency, low cost and the fact that it does not form harmful by-products
39 [7]. The adsorbent is usually activated carbon. Activated carbons have been employed in
40 water purification when removing both organic and inorganic pollutants from industrial
41 wastewater [8]. The drawback in adsorption method is the high cost of the activated carbon
42 which is used as the adsorbent [9] and most times, is imported in commercial quantities in
43 Nigeria. The over-dependence of our local industries on imported raw materials, activated
44 carbon inclusive, is currently the bane of the economy of developing countries such as ours
45 [10]. To curb this, the effectiveness of cheaper and abundant local materials has being
46 studied for use as adsorbents. They include oil palm fibre, saw dust, bamboo, kola nut shell
47 etc [11-14].

48 Rice husk is the chaff that is obtained from the milling of rice grains. It is relatively abundant
49 and can be obtained as waste from the rice milling industries. Its potential as an adsorbent
50 have been reported in adsorption processes. The ash of rice husk contains approximately
51 high silica, which has porous structure and is lightweight, with high specific surface area.
52 Annual rice production in Nigeria was estimated at over 5.8 million tons in 2017 [15].
53 Therefore, huge quantity of rice husk from the rice milling factories is usually discarded as
54 waste annually. Activated carbon adsorbents produced from rice husk has been used in the
55 adsorption of gasoline [16] and in the adsorption of lead (Pb) from car battery wastewater
56 [17]. The adsorptive capacity of the adsorbent can be increased by carbonization. Different
57 conditions of the process parameters used in the carbonization process affect the adsorptive
58 potential of the activated carbon produced.

59 Optimization using response surface methodology can be used to determine the optimum
60 conditions involved in a process [18]. It is different from the method of one factor at a time
61 (OFAT) which involves keeping all other parameters constant while varying one factor.
62 OFAT method uses a large number of experiments in determining the optimum condition. It
63 is time consuming and does not show the interactive effects of the independent factors
64 unlike optimization using response surface methodology (RSM). Design of experiment using
65 RSM is an enhanced systematic experimentation that takes into consideration all the
66 process parameters involved simultaneously [18].

67 Hence, the aim of this work is to use response surface methodology to optimize the
68 carbonization process parameters for optimum production of activated carbon from rice husk
69 that will be used in the treatment of wastewater containing phenol.

70

71

72

73 **2.0 MATERIALS AND METHODS**

74 **2.1 Preparation of raw materials**

75 The rice husk was sourced from rice mills in Anambra State, Nigeria. It was washed with
76 distilled water and sun dried. The phenol, tetraoxophosphoric acid (H₃PO₄), distilled water
77 and other reagents were sourced from Chemical Engineering Laboratory in Nnamdi Azikiwe
78 University, Awka, Anambra State.

79

80 **2.2 Design of experiment**

81 The experimental runs for the carbonization process were designed using central composite
 82 design of the RSM. Design Expert software version 10.0.7 was used to carry out the RSM
 83 analysis. This method uses a minimum number of experiments to optimize a process while
 84 analyzing the interaction between the parameters. The independent variables were
 85 percentage concentration of the tetraoxophosphoric acid, carbonization temperature and
 86 carbonization time. The dependent variable or the response was the percentage of phenol
 87 adsorbed. Table 1 shows the different levels of the independent variables that were used in
 88 the experiment. The distance of the star like points from the core point, that is, the alpha
 89 value was 1.68. The actual experimental design (in Table 3) consist of 20 runs made up of 8
 90 core points, 6 star like points and 6 null points.
 91 Statistical analysis of the model including the analysis of variance (ANOVA) was evaluated
 92 using the Design Expert software.
 93
 94
 95

Table 1: Factor levels of the independent variables

Independent variable	- α	-1	0	1	α
Percentage concentration of acid (%)	10	20	35	50	60
Carbonization temperature ($^{\circ}$ C)	19	60	120	180	221
Carbonization time (minutes)	298	400	550	700	802

96
 97

98 2.3 Carbonization process

99 The carbonization of the rice husk was carried out based on the design of experiment. The
 100 rice husks were broken into small pieces and dried in sunlight. This helps to reduce the
 101 moisture content of the sample. The dried sample was activated by mixing it with the
 102 required percentage concentration of the tetraoxophosphoric acid, H_3PO_4 and kept in an
 103 oven at 383K for 24 hours. Thereafter, the activated sample was washed severally with
 104 deionized water. The activated rice husk was placed in a furnace at the appropriate
 105 temperature and time (based on the experimental design) to undergo the carbonization
 106 process. The sample was cooled, ground using mortar and pestle and sieved using a mesh
 107 size of 75 μ m. The experiment was repeated with different percentage concentration of acid,
 108 carbonization temperature and time according to the experimental design. All produced
 109 activated carbons were properly labeled and used for the actual adsorption experiment.
 110 Response surface was used to determine the individual and interactive effects of the
 111 independent variables on the percentage of the phenol adsorbed.
 112

113 2.4 Adsorption process

114 Stock solution of phenol with concentration 100mg/l was prepared. One hundred milliliters
 115 (100ml) of the phenol solution was placed on a magnetic stirrer set at 50 $^{\circ}$ C. Activated rice
 116 husk adsorbent of mass 0.5g was introduced and the mixture allowed for about 60 minutes.
 117 Thereafter, the mixture was cooled and separated using centrifugation at 1,000 rpm for 20
 118 minutes. The absorbance of the phenol was estimated using UV spectrophotometer at a
 119 wavelength of 250nm and then converted to concentration. The percentage adsorbed (%)
 120 was determined as follows;

$$121 \%Adsorbed = \frac{C_o - C_e}{C_o} \quad (1)$$

122 where C_o is the initial concentration of phenol solution (mg/l), C_e is the equilibrium
 123 concentration of the phenol solution (mg/l).
 124

125 2.5 Physical properties of the activated carbon

126 Some of the physical properties of the activated rice husk were determined using standard
 127 methods. A pH meter (Elico model L1 -120) was used to determine the pH. Fixed carbon
 128 iodine number, moisture content, volatile matter, ash content and porosity were determined

129 using the method reported by [14]. Water displacement method was used to determine the
130 bulk density [19].

131

132 **2.6 Surface area and Pore size distribution analysis**

133 The BET nitrogen (N₂) adsorption-desorption isotherms was used to determine the surface
134 area and micro pore volumes. The Quantachrome NOVA Win version 11.03 was used at
135 77K using N₂ gas sorption analyzer. The total pore volume estimated using liquid volume of
136 adsorbate (N₂) at a relative pressure of 0.99 while the surface area was calculated from the
137 nitrogen adsorption isotherms by assuming the area of a nitrogen molecule was 0.162 nm².

138

139 **2.7 Instrumental characterization of the activated carbon**

140 A JOEL scanning electron microscope model JSM 6400 was used to carried out the
141 scanning electron microscope (SEM) analysis while a Shimadzu Fourier Transform Infrared
142 Spectrophotometer (FTIR) 8400S was used to identify the functional groups present in the
143 activated rice husk.

144

145

146 **3.0 RESULTS AND DISCUSSION**

147

148 **3.1 BET surface area and pore size distribution**

149 The surface area was obtained using N₂ adsorption isotherm of the carbonized at 77K. The
150 multipoint BET surface area was 471.7m²/s while the single point BET surface area was
151 286.8m²/s as seen in Table 2. The surface area was high as a result of the presence of
152 excess pores that developed during the activation and carbonization process. The higher the
153 surface area, the better the adsorption potentials of the adsorbent. The micropore volume
154 was 0.179cm³/g. These values are similar to those reported by [19,20]. The pore radius was
155 16.20Å while the average pore width was 5.55 nm. The values obtained provide qualitative
156 information on the adsorption mechanism and the pore structure of the carbon.

157

158

Table 2: BET surface area analysis of the activated rice husk

Property	Quantitative value
Multipoint BET surface area (m ² /s)	471.67
Single point BET surface area (m ² /s)	286.8
Average pore width (nm)	6.247
Micropore volume (cm ³ /g)	0.179
Adsorption energy (KJ/mol)	4.162
Pore radius (Å)	16.20

159

160

160 **3.2 Physical properties of the activated carbon**

161 Table 3 contains the physical properties of the adsorbent. The fixed carbon analysis gave a
162 value of 10.14% which is not very high suggesting that the carbon content of rice husk is
163 low. The moisture content was equally low (6.5%) as expected while the porosity index
164 indicated 0.339. The iodine number was high at 461.84 mg/g which indicated high surface
165 area. Iodine number is used as an index to investigate the internal structure and surface
166 area of the activated carbon [21]. The high volatile matter (18.01%) and ash content
167 (57.49%) suggested good properties of the activated carbon.

168

169

Table 3: Physical properties of the adsorbent

Property	Quantitative value
Bulk density (g/ml)	0.448
pH	6.8 ± 0.2
Ash content (%)	57.49

Iodine Number (mg/g)	461.84
Moisture content (%)	6.5
Porosity(η)	0.339
Volatile matter (%)	30.82
Fixed Carbon (%)	10.14

170
171
172
173
174
175
176
177
178
179
180
181
182
183
184
185
186
187

3.3 FTIR and SEM analysis

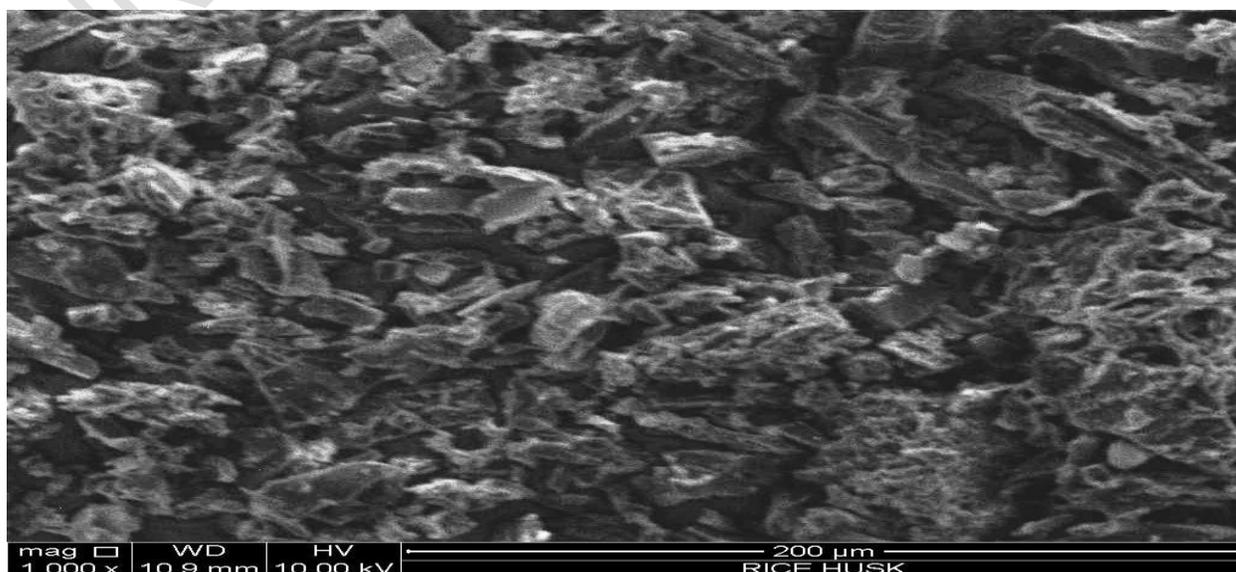
The FTIR analysis revealed the functional groups present in the rice husk as shown in Table 4. The chemical structure of the adsorbent is of vital importance in understanding the adsorption process. The wave number ranged from 3693.8 to 670 cm^{-1} with peaks from 96.68 to 83.45 cm^{-2} . They are instrumental in the adsorption of aromatic compounds. The – C=C- stretch indicates the presence of alkenes while the C-Cl stretch and vibration suggests the presence of alkyl halides. The coupled vibrations are appreciable due to the availability of various constituents [22]. This shows that the rice husk can be a good source of some hydrocarbons such as alcohols, alkenes, alkyl halides.

The SEM in figure 1 was obtained using the activated rice husk sieved at 200 μm . The result was at a magnification of 1000x. It indicated that the texture and surface morphology of the activated carbon were characterized by rough surfaces. Interspatial pores were seen within the matrix of the adsorbent indicating good adsorption properties. The large pores observed is due to the fact the activating agents promote the contact area between the carbon and the activating agent.

Table 4: FTIR analysis result of carbonized rice husk

Wave Number (cm^{-1})	Peak area (cm^{-2})	Bond source	Compound
3693.8	96.68	O-H vibration	Alcohols
3272.6	90.66	-C=C- stretch	alkenes
2922.2	88.05	-C=C- stretch	alkenes
2855.1	85.08	O-H bending	Carboxylic acid
2209.9	95.58	O-H vibration	Alcohols
1640.0	889.48	O-H bending	Carboxylic acid
1233.7	90.23	C-H vibration	alkanes
853.6	87.83	C-Cl stretch	Alkyl halides
670.9	83.45	C-Cl stretch	Alkyl halides

188
189
190
191
192



193
 194
 195
 196
 197
 198
 199
 200
 201
 202
 203
 204
 205
 206
 207
 208
 209
 210
 211

Figure 1: SEM image of the activated carbon

3.4 Optimization process

The model summary statistics for the adsorption efficiency of phenol is presented in Tables 5. The model summary values suggested that a quadratic model best fitted the optimization process. The R-squared values for the quadratic and cubic models have the best values of 0.9909 and 0.9936 respectively when compared to that of other models (2FI and linear). The R-Squared is usually a measure of how efficient the variability in the actual response values can be explained by the experimental variables and their interactions. The cubic model is always aliased because the CCD does not contain enough runs to support a full cubic model. Aliases are false signals of any sort present hence the quadratic model was suggested.

Table 5: Model Summary Statistics

Source	Std. Dev.	R-squared	Adjusted R-sq	Predicted R-sq	PRESS	Remark
Linear	20.48	0.1550	-0.0035	-0.4966	11887.43	Not suggested
2FI	22.44	0.1757	-0.2047	-0.8298	14533.60	Not suggested
Quadratic	6.01	0.9546	0.9137	0.6717	2607.29	Suggested
Cubic	6.43	0.9688	0.9012	-5.2957	50005.71	Aliased

212
 213
 214
 215
 216
 217
 218
 219
 220
 221
 222
 223
 224

The ANOVA in table 6 was used to analysis the result and validate the adsorption model. The lack of fit test and the adequacy of the regression models were equally performed. A significance level of 5% was used hence P-values greater than 0.05 are considered insignificant while those at 0.05 or less are significant. Hence, only the interactions of AB, AC and C² are insignificant. The model F-value of 23.35 implies that the model is significant agreeing with the P-value being less than 0.0001. The P values check the significance of the factors and equally help to understand the pattern of the mutual interactions between the test variables [23]. The R² value of 0.9546 is in close agreement with the adjusted R² value of 0.9137.

Table 6: ANOVA of the optimization process

Source	Sum squares	Df	Mean square	F value	p-value (Prob>F)
Model	7582.08	9	842.45	23.35	< 0.0001
A-Acid Concentration	580.95	1	580.95	16.10	0.0025
B-Carbonization time	268.53	1	268.53	7.44	0.0213
C-Carbonization temperature	381.26	1	381.26	10.57	0.0087
AB	1.45	1	1.45	0.040	0.8454
AC	22.44	1	22.44	0.62	0.4485

BC	141.12	1	141.12	3.91	0.0761
A ²	5180.95	1	5180.95	143.62	< 0.0001
B ²	585.03	1	585.03	16.22	0.0024
C ²	0.41	1	0.41	0.011	0.9169
Residual	360.73	10	36.07		
Lack of Fit	339.73	5	67.95	16.18	0.0042
Pure Error	21.00	5	4.20		
Cor Total	7942.81	19			

225 Std. Dev. = 6.01; Mean = 54.26; C.V. = 11.07%; PRESS = 2607.29
 226 R-Squared = 0.9546 Adj R-Sq = 0.9137; Pred R-Sq = 0.6717; Adeq Precision =
 227 21.210
 228
 229
 230
 231

232 3.5 Optimum model equation

233 The generated model equation for the adsorption process in terms of coded factors is
 234 Percentage Adsorbed (%) = +62.97 – 6.52A + 4.43B + 5.28C + 0.43AB – 1.67AC + 4.20BC
 235 – 18.96A² + 6.37B² - 0.17C²
 236 (1)

237 The positive sign of a factor indicates that there will be increase in the response when there
 238 is an increase in the factor while negative sign will lead to decrease in the response [24].
 239 Increase in carbonization temperature will show the most significant increase in the
 240 response on the account that its coefficient is highest.

241 Since a significant level of 5% was used, all factors with P-values greater than 0.05 are
 242 eliminated giving the final model equation as

$$243 \text{ Percentage Adsorbed (\%)} = +62.97 - 6.52A + 4.43B + 5.28C + 4.20BC - 18.96A^2 + 6.37B^2 \quad (2)$$

247 3.6 Comparism of predicted and experimental values

248 A comparism of the actual experimental response and the predicted response are given in
 249 Table 7. The result of the experimental runs in the optimization process indicated that the
 250 best carbonization conditions are at an acid concentration of 45%, carbonization time of
 251 240.9 minutes and carbonization temperature of 575 °C. This gave the highest adsorption
 252 efficiency of 96.6% of phenol adsorbed from the phenol solution. The result equally revealed
 253 that the three factors optimized have great effect on the production of activated carbon.

254 The close correlation between the actual experimental response and the predicted response
 255 confirms the suitability of the quadratic model used the analysis.
 256

257 Table 7: Experimental and predicted responses of the optimization process

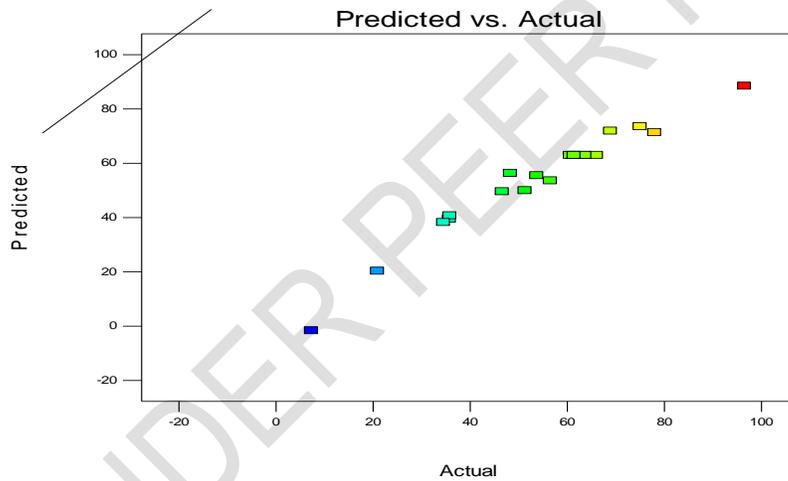
Standard order	Acid Conc	Carbonization time (mins)	Carbonization temperature	Experimental Response	Predicted Response
1	30	80	400	51.3	49.9
2	60	80	400	35.7	39.4
3	30	200	400	46.6	49.6
4	60	200	400	35.8	40.7
5	30	80	750	53.7	55.5
6	60	80	750	34.5	38.2
7	30	200	750	68.9	71.9
8	60	200	750	48.3	56.4
9	19.7731	140	575	20.9	20.3

10	70.2269	140	575	7.3	-1.6
11	45	39.0924	575	75	73.5
12	45	240.908	575	96.5	88.5
13	45	140	280.686	56.5	53.6
14	45	140	869.314	78	71.4
15	45	140	575	61.1	62.9
16	45	140	575	63.5	62.9
17	45	140	575	62.6	62.9
18	45	140	575	63.6	62.9
19	45	140	575	60.6	62.9
20	45	140	575	61.4	62.9

258
259
260
261
262
263
264
265
266
267
268

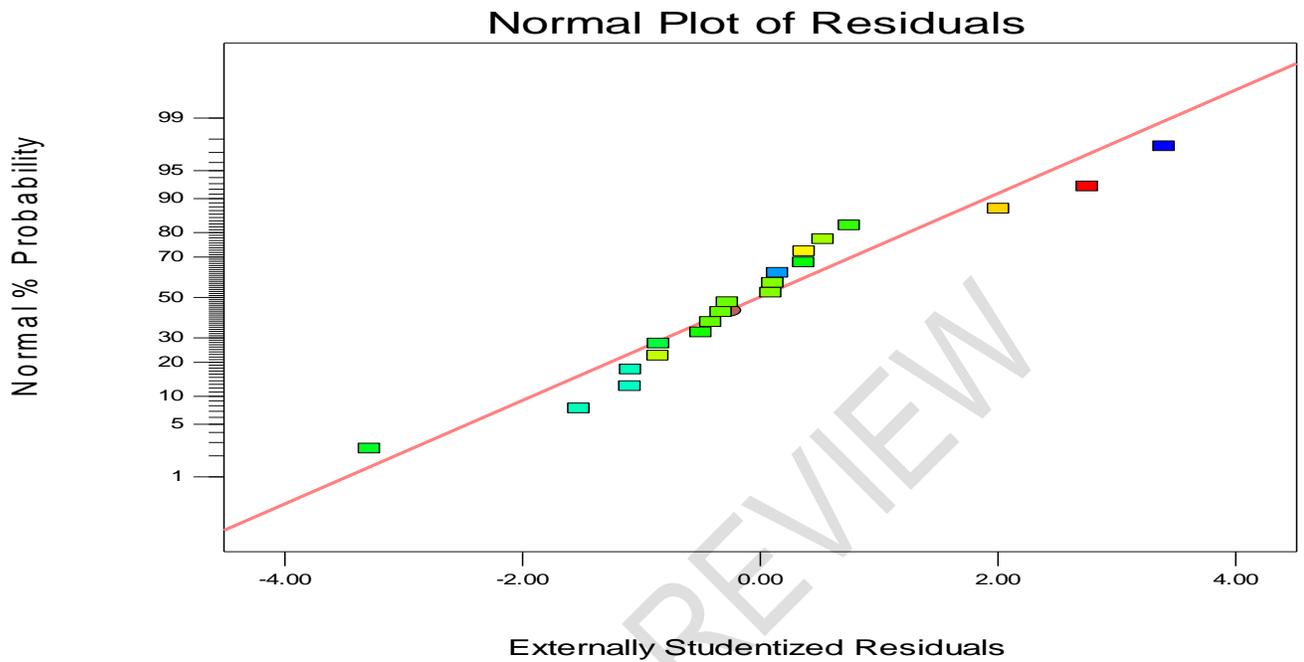
3.7 Error graphs

The Predicted vs Actual plot in figure 2 and the Normal plot of Residuals in figure 3 were used to determine if the residuals follow a normal distribution. It is assumed to have followed a normal distribution as the points closely aligned to the straight line of the plot thereby confirming the good relationship between the experimental values and the predicted values of the response and the adequacy of the suggested model in predicting the response variables in the experimental values.



269
270
271
272
273

Figure 2: The Predicted vs Actual plot of the optimization process



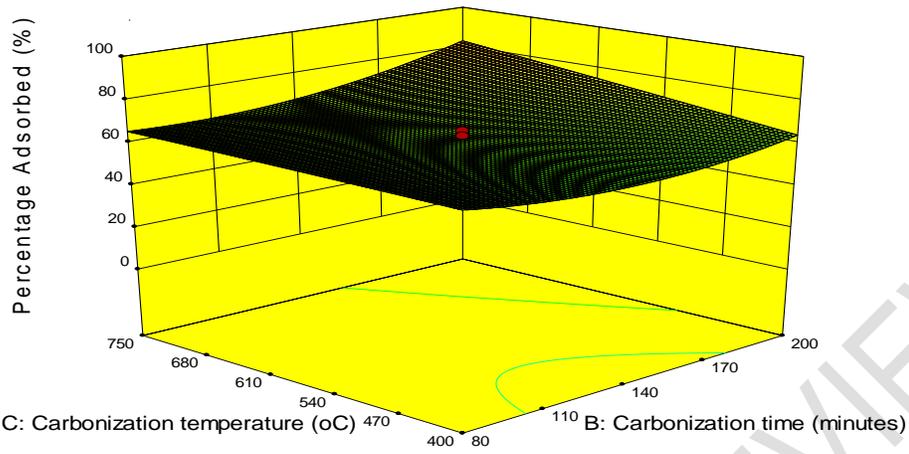
274
 275
 276
 277
 278
 279
 280
 281
 282
 283
 284
 285
 286
 287
 288
 289
 290
 291
 292
 293
 294
 295
 296
 297
 298
 299

Figure 3: Normal plot of Residuals of the optimization process

3.8 3-D response surface plots

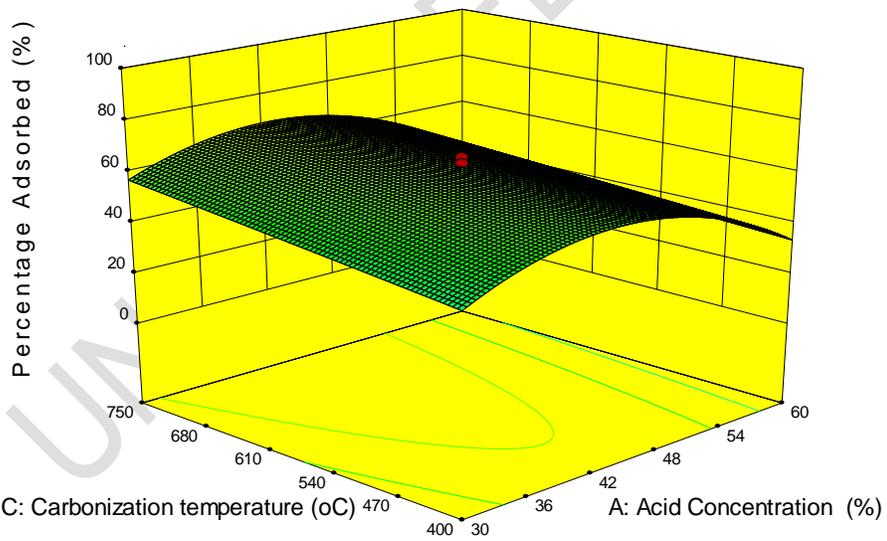
The 3-D response surface plots are graphical representation of the interactive effects of any two variables factors. Response surface estimation serves as a function of two factors at a time, maintaining other factors at fixed levels. This is more helpful in understanding both the main and the interaction effects of those two factors. These plots can be easily obtained by calculating from the model, the values taken by one factor where the second varies with constraint of a given response value. The response surface curves were plotted to understand the interaction of the variables and to determine the optimum levels of each variable for maximum response.

The nature of the response surface curves shows the interaction between the variables. The elliptical shape of the curve indicates good interaction of the two variables and circular shape indicates no interaction between the variables. There was a relative significant interaction between every two variables, and there was a maximum predicted efficiency as indicated by the surface confined in the smallest ellipse in the contour diagrams.



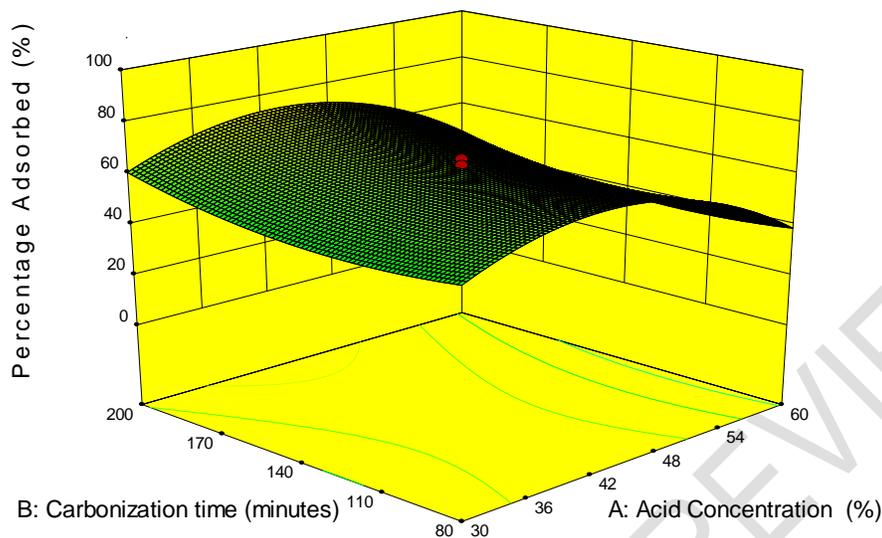
300
301
302
303
304
305

Figure 4: Interactive effects of temperature and time



306
307
308
309
310
311
312

Figure 5: Interactive effect of temperature and concentration



313
314 Figure 6: Interactive effects of time and concentration
315

316 An adsorption capacity of 38.2 mg/g was obtained in this work for phenol adsorption using
317 the activated rice husk produced. This is more 26.26 mg/g reported in phenol removal using
318 sewage sludge based adsorbent modified with H₂SO₄ [25] and 25.0 mg/g obtained in phenol
319 adsorption using multiwalled carbon nanotubes [26]. However, it is much less than 91 to
320 112.5 mg/g reported in phenol adsorption using *Latana camara* [27].
321

322 323 324 **4. CONCLUSION**

325 The process parameters for activated carbon production from rice husk were optimized
326 using response surface methodology for the treatment of phenolic wastewater. BET multi-
327 point surface area of the activated carbon was high indicating favourable applicability of the
328 adsorbent in the adsorption processes. Maximum adsorption efficiency of 96.5% was
329 obtained at carbonization time of 240 minutes, carbonization temperature of 575 °C and at
330 acid concentration of 45%. A quadratic model with a high correlation coefficient was
331 suggested in describing the interactive effects of the process parameters. This study has
332 shown that activated carbon can be produced from rice husk at optimum process conditions
333 for the uptake of phenol from wastewater.
334
335
336
337

338
339
340
341
342
343
344
345
346
347
348
349
350
351
352
353
354
355
356
357
358
359
360
361
362
363
364
365
366
367
368
369
370
371
372
373
374
375
376
377
378
379
380
381
382
383
384
385
386
387
388
389
390

COMPETING INTERESTS

The authors have declared that no competing interest exists.

REFERENCES

1. Uddin M T. Islam MS. Abedin MZ. Adsorption of phenol from aqueous solution by water hyacinth ash. *ARPN Journal of Engineering and Applied Sciences*. 2007; 2 (2): 11 – 18
2. Ahmaruzzaman M. Adsorption of phenolic compounds on low-cost adsorbents: A review. *Advances in Colloid and Interface Science*. 2008; 143: 48–67.
3. Bousba S. Meniai AH. Removal of phenol from water by adsorption onto sewage sludge based adsorbent, *Chemical Engineering Transactions*. 2014; 40, 235-240. DOI: 10.3303/CET1440040
4. Sunil JK. Jayant PK. Review on Research for Removal of Phenol from Wastewater. *International Journal of Scientific and Research Publications*. 2013;3(4): 1 - 7
5. Salim B. Abdeslam HM. Removal of Phenol from Water by Adsorption onto Sewage Sludge Based Adsorbent. *Chemical engineering transactions*. 2014; 40: 235 – 240.
6. Onu CE. Nwabanne JT. Adsorption kinetics for Malachite green removal from aqueous solution using Nteje clay. *Journal of Environment and Human*. 2014; 1(2): 133 – 150.
7. Oguanobi NC, Onu CE Onukwuli OD. Adsorption of a dye (crystal violet) on an acid modified non-conventional adsorbent. *Journal of Chemical Technology and Metallurgy*. 2019; 54 (1): 95-110.
8. Azry B. Ahmad FK. Preparation and characterization of activated carbon from rubber-seed shell by chemical activation. *Journal of applied sciences*. 2012; 12(11):1124 - 1129
9. Rajeshkannan R. Rajasimman M. Rajamohan N. Decolourisation of malachite green using tamarind seed: Optimisation, isotherm and kinetic studies. *Chemical Industry and Chemical Engineering Quarterly*. 2011; 17(1): 67-79.
10. Nwabanne JT. Okpe EC. Igbokwe PK. Asadu CC. Onu CE. Isotherm and kinetic modelling of adsorption of dyestuffs onto kola nut (cola acuminata) shell activated carbon. *Journal of Chemical Technology and Metallurgy*. 2016;51 (2): 188 – 201.
11. Tan IA. Hameed BH. Ahmed AL. Equilibrium and kinetic studies on basic dye adsorption by oil palm fibre activated carbon. *Chemical Engineering Journal*. 2007; 127: 111-119.
12. Nestor T. Natalia M. Fabiana M. Javier P. Carina P. Tomas C. Phenol adsorption onto powdered and granular activated carbon, prepared from eucalyptus wood. *Journal of Colloid and Interface Science*. 2004; 279:357-363.
13. Hameed BH, Din ATM. Ahmad AL. Adsorption of methylene blue onto bamboo-based activated carbon: Kinetics and equilibrium studies. *J. Hazard. Mater*. 2007; 141: 819-825.
14. Nwabanne JT. Okpe EC. Asadu CO. Onu CE. Application of Response Surface Methodology in Phenol Red Adsorption Using Kola Nut (Cola acuminata) Shell Activated Carbon. *International Research Journal of Pure & Applied Chemistry*. 2017; 15 (4): 1 – 14.
15. Udemezue JC. Analysis of Rice Production and Consumption Trends in Nigeria. *Journal of Plant Science and Crop Protection*. 2018; 1(3): 305, 1 – 6.
16. Arunrat C. Sukjit K. Preparation of Activated Carbon Derived from Rice Husk by Simple Carbonization and Chemical Activation for Using as Gasoline Adsorbent.

- 391 International Journal of Environmental Science and Development, 2014; 5 (2) 171 –
392 175., DOI: 10.7763/IJESD.2014.V5.472
- 393 17. Hanum F. Bani O. Wirani LI. (2017) Characterization of Activated Carbon from Rice
394 Husk by HCl Activation and Its Application for Lead (Pb) Removal in Car Battery
395 Wastewater. 1st Annual Applied Science and Engineering Conference. IOP
396 Conference Series: Materials Science and Engineering. 2017; 180, 1 – 11.
- 397 18. Onu CE. Nwabanne JT. Application of Response Surface Methodology in Malachite
398 green adsorption using Nteje clay”. Open Journal of Chemical Engineering and
399 Science. 2014; 1(2): 19 – 33.
- 400 19. Dipa D. Debi PS. Meikap BC. Preparation of Activated Carbon from Green Coconut
401 Shell and its Characterization. Journal of Chemical Engineering & Process
402 Technology. 2015; 6(5):1-7
- 403 20. Adegboyega SO. Olusegun AA. Michael SO. Mku TI. Sam SA. Preparation of
404 phosphoric acid activated carbons from Canarium Schweinfurthii Nutshell and its
405 role in methylene blue adsorption. Journal of Chemical Engineering and Materials
406 Science. 2015; 6(2) : 9-14.
- 407 21. Enaime G. Ennaciri K. Ounas A. Baçaoui A. Seffen M. Yaacoubi A. Preparation and
408 characterization of activated carbons from olive wastes by physical and chemical
409 activation: Application to Indigo carmine adsorption. Journal of Materials and
410 Environmental Sciences. 2017; 8 (11): 4125-4137
- 411 22. Preeti SN. Singh BK. Instrumental characterization of clay by XRD and FTIR, Indian
412 Academy of Sciences. 2007; 30(3): 235-238.
- 413 23. Shrivastava A. Sandagar P. Baja I. Singha R. Media optimization for the production
414 of U-linolenic acid by cunninghamella echinulata varielegans MTCC 522 using
415 response surface methodology. International Journal of food Engineering. 2008;
416 4(2):132-140
- 417 24. Kumar A. Prasad B. Mishra IM. Adsorptive removal of acrylonitrile using powered
418 activated carbon. Journal of Environmental Protection Science. 2008; 2: 54 – 62.
- 419 25. Salim B. Abdeslam HM. Removal of Phenol from Water by Adsorption onto Sewage
420 Sludge Based Adsorbent. Chemical engineering transactions. 2014; 40, 235 – 240.
- 421 26. Nour TA. Ghadir AE. Farag SH. Individual and competitive adsorption of phenol and
422 nickel onto multiwalled carbon nanotubes. Journal of advanced research. 2015; 6(3):
423 405–415.
- 424 27. Girish CR. Ramachandra MV. Adsorption of Phenol from Aqueous Solution Using
425 Lantana camara, Forest Waste: Kinetics, Isotherm, and Thermodynamic Studies.
426 International Scholarly Research Notices. 2014; 1 – 17.