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2 **A comparative Evaluation Of Selected Medicinal Plants on Male fertility indices (reproductive**

3 **hormones and sperm profile) of albino wistar rats. An Animal Case study**

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5

6 **Abstract:** The study aims to investigate the effects of the medicinal plants{seeds} on the
7 reproductive hormones and sperm profile of male albino rats to ascertain their possible usefulness
8 as fertility agent. Walnuts [Tetracarpidium conophorum] ,Sesame (Sesamine indicum),and Velvet
9 bean(Mucuna pruriens) seeds were obtained and taken to the Department of plant Science and
10 Biotechnology, University Of Nigeria , Nsukka(UNN). The seeds were milled into fine powder. One
11 hundred and ten sexually matured aibino rats of about twelve weeks weighing 130-180g were
12 divided into eleven groups (1-11) using completely randomized design. There were two different
13 control groups and rats in group 1 served as the Control 1 and were fed with normal commercial
14 feed. Rats in group 2 were administered with a drug (Ketoconazole) to induce infertility. The
15 **Infertility Induced groups** were treated with low dose (groups 3-5), medium dose (groups 6-8) and
16 high dose (9-11) for the period of nine (9) weeks. At the end and testes and Epididymides were
17 surgically removed and weighed. Blood sample analysis revealed that the concentration of sex
18 hormones measured in the male rats fed with medium plants (seeds) showed that the testosterone
19 concentration significantly increased ($p<0.05$) in animal control group 1(normal rats) Therefore,
20 these medicinal plants walnut seeds showed significant increase in their testosterone concentration,
21 luteinizing hormone (LH) and follicle stimulating hormone (FSH) which significantly enhanced the
22 production of reproductive hormones which enriched the fertility status of these animals.

23 **Keywords:** albino rats, medicinal plants, reproductive hormones, sperm profile, testis histology

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26 1. INTRODUCTION

27 Medicinal plants are plants that contain properties or compounds that can be used for
28 therapeutic purposes or those that synthesize metabolites to produce useful drugs (WHO 2008). The
29 pharmacological evaluation of substances from plants is an established method for the identification
30 of lead compounds which can lead to the development of novel and safe medicinal agents (Golla et
31 al., 2011). Historically, all medicinal preparations were derived from plants whether in the simple
32 form of raw plants materials or in the refined form of crude extracts, mixtures (Krishnaraju et al.,
33 2005). The rich knowledge base of countries like India and China in medicinal plants and health care
34 has led to the keen interest by pharmaceutical companies to use this knowledge as a resource for
35 research and development programs in the pursuit of discovering novel drugs (Mukherjee & Wahile,
36 2006). Nigeria is also among countries blessed with a wide range of such medicinal plants which
37 have not been thoroughly evaluated for their bioactive/ therapeutic agents. Primary among this list
38 of properties is their antioxidants, antibacterial and antifungal activities (Ajaiyeoba and Fadara ,
39 2006). However, some of such botanicals are Tetracarpidium conophorum, Sesame indicum L and

40 Mucuna pruriens. These plants have antimicrobial activity which may cure some sexually transmitted
41 infections that could be responsible for male fertility (Ajaiyeoba and Fadara 2006). Researchers have
42 also shown that these plants possess some essential antioxidants and Vitamins capable of fighting
43 off reactive oxygen species, thereby assisting seminal plasma exhibiting a strong capacity to maintain
44 a relatively neutral and protective environment for sperm function (Orth et al; 1993; Foresta et al,
45 2004). There are reports that abnormalities in sex hormone biosynthesis may impair
46 spermatogenesis. The failure of the pituitary to maintain proportionate levels of FSH,LH and PRL
47 may lead to disruption of testicular function, leading to infertility (Dashistani and Dayem, 2007).The
48 seeds have also been found to contain major essential fatty acids (Omega-3DHA and Omega-6)
49 which include oleil, linolenic and linolenic acids (Zwarts et al; 1999; savage, 2001; Ozkan and
50 koyuncu, 2005, Ahmed et al, 2008,). Essential fatty acids act as hormone regulators and its
51 purported that sperm contain high concentrations of Omeg-3's.

52 The main purpose of the research work therefore was to investigate the effects of the
53 medicinal plants (seeds) on the reproductive hormones and sperm profile of male albino rats in
54 order to ascertain possible usefulness as a fertility agent.

55 2. MATERIALS AND METHODS

56 The seeds of walnut (*Tetracarpidium conophorum*), Sesame (*Sesamium indicum*), and
57 Velvet bean (*Mucuna pruriens*) were procured and sent to the department of plant science and
58 Biotechnology, University of Nigeria (UNN) for identification and authentification. **The seeds were**
59 **cut into pieces and dried for fort-eight hours and ground** into fine powder using electric blender.

60 **Experimental Animals and Administration of Seeds.** Ninety sexually matured male albino rats of
61 about twelve weeks weighing between 130-180g were used for this experiment. They were kept in a
62 well ventilated conventional cage (temperatures: 28-31°C, photoperiod: 12hours of natural light and
63 12hours of darkness; humidity: 50-55%) and acclimatized for two weeks with tap water before
64 commencement of treatments. The rats were divided into eleven groups (1-11) using completely
65 randomized design with six male and four female rats in each group. There were two different
66 control groups in this research. Rats in group (1) served as the control 1 and were fed with normal
67 commercial feed only and rats in group (2) served as control ii and were administered with a drug
68 (ketoconazole) reported to induce infertility to rats, including dog and primate (Vawda and Davies,
69 1986; Donald et al , 1990). To evaluate further the potentials of the seeds, the infected groups were
70 treated with low dose (groups 3-5), medium dose (groups 6-8) and high dose (groups 9-11) using
71 dietary inclusion for the period of nine weeks.

72 Preparation of Serum and Semen for fertility Examination.

73 At the end of the treatment regime, the rats were anaesthetized using diethyl ether. The testes and
74 Epididymides were surgically removed and weighed. Blood samples, 15ml per each rat were
75 collected through cardiac puncture into sterile tubes for hormonal analysis.

76 Hormone Assay

77 The blood samples collected were centrifuged at 2500rpm for 5minutes using a centrifuge at 10-
78 20oc to obtain the serum samples which were analyzed for testosterone, follicle stimulating

79 hormone (FSH), luteinizing hormone (LH) and estradiol hormone level using enzyme linked
80 immunoassay (ELISA) technique (Ekaluo et al; 2010)

81

82 **Semen Analysis**

83 (a) Determination of Epididymides and weight.

84 These Epididymides and testes were dissected out and excess blood damped into cotton wool and
85 placed in a clean weighing balance to record the weight.

86 (b) Evaluation of sperm motility

87 Semen samples from the different treatment groups were dropped on a glass slide and viewed
88 under the microscope. A minimum of five microscopic fields were assessed to evaluate sperm
89 motility and were analyzed for progressive motile sperm (PMS), non-progressive motile sperm
90 (NPMS) and non-motile sperm (NMS) distinguished by the movement of the sperm cells (World
91 Health Organization, 1992)

92 (c) Estimation of mean sperm count

93 The mean sperm count was carried out according to the method of (Ekaluo et al; 2009)

94 (d) Estimation of sperm viability

95 Sperm viability was estimated using the improved one step eosin-nigrosin staining technique. A
96 fraction of each suspension of the sperm samples were mixed with equal volume of eosin-nigrosin
97 stain and air dried smears were prepared on glass slides for each samples as described by Bjorn dahl
98 et al, (2003).

99 (e) Estimation of Semen pH

100 The pH of semen was measured using a specially treated calibrated paper blot that changes colour
101 according to the pH of the semen that it is exposed to (Comhaire and Vermeulen, 1995)

102 (f) Sperm head abnormalities

103 A fraction of each of the sperm suspension was examined by placing the solution (10:1) for 30
104 minutes on a glass slide. The slide was examined for percentage abnormalities in every 200
105 spermatozoa on each slide and fine air dried smear prepared on glass slide for each sample
106 according to (Ekaluo et al; 2005).

107

108 **3. RESULTS AND DISCUSSION**

109 Table 1 shows the results of the impacts of medicinal plants on the sperm quality of the albino rats.
110 The result revealed no significant increase ($p>0.5$) in the sperm motility between the animal groups
111 1,6,7,9 and 11 and these groups showed a significant increase ($p<0.5$) when compared with that of
112 other experimental groups. The sperm motility was significantly lower ($p<0.05$) in the animal groups

113 2,3,4,5,8 and 10. The result in respect of sperm count showed a significant increase ($p<0.05$) in the
 114 group 1 compared across all the induced animal groups 2-11. However, there was no observed
 115 significant difference ($p>0.05$) between the group 2 and the animal groups treated with the
 116 medicinal plant samples. Sperm viability was observed to be significantly high($p<0.05$) in the groups
 117 1,5,6,8,10, and 11 compared to other treated groups. Also, there were no significant difference
 118 ($p>0.05$) in the sperm viability between the animals in groups 2,3, and these groups showed
 119 significant decrease($p<0.05$) in the sperm pH of the rats in all the groups. Meanwhile, Testes and
 120 Epididymides weight of the group 6 was significantly higher($p<0.05$) than that of groups 1,4,5, and
 121 8. However, the results revealed no significant increase ($p>0.05$) of the Testes and Epididymides
 122 weight between the groups 2,3,7,9,10 and 11.

123 All values are expressed as Mean +Standard Deviation (SD). Mean values with the same letters as
 124 superscripts along the column are considered non-significant difference ($p>0.05$) while mean values
 125 with different letters as superscripts are considered significant($p<0.05$).

126 **Table1: Effects of the medicinal plants (Tetracarpidium conophorum,Sesame indicum L, and**
 127 **Mucuna pruriens) seed on the sperm profile of the albino rats (mean \pm SD)**

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GROUP S	Sperm Motility	Sperm Count	Sperm Viability	Sperm Head Abnormality	Sperm pH	Testes and Epididymides weight (g/kg)
Group 1	61.67 ± 7.64^a b	53.33 ± 2.89^a	58.33 ± 3.51^a	38.33 ± 7.63^b	7.50 ± 0.49^a	0.93 ± 0.2^b
Group 2	26.67 ± 5.77^d	26.67 ± 7.64^b	34.00 ± 8.72^c	71.67 ± 6.36^a	6.63 ± 0.72^a	0.97 ± 0.06^{ab}
Group 3	33.33 ± 5.77^c d	26.67 ± 7.64^b	29.67 ± 7.50^c	60.00 ± 1.50^{ab}	6.90 ± 0.42^a	1.13 ± 0.58^{ab}
Group 4	31.67 ± 6.07^d	21.66 ± 7.60^b	41.33 ± 5.51^{bc}	60.00 ± 2.42^{ab}	6.77 ± 0.40^a	0.73 ± 0.12^b
Group 5	33.33 ± 2.58^c d	20.00 ± 5.00^b	44.33 ± 6.03^{ab} c	63.32 ± 3.64^{ab}	7.03 ± 0.49^a	0.90 ± 0.17^b
Group 6	55.00 ± 3.23^a b	26.66 ± 7.63^b	42.00 ± 1.16^{ab} c	56.67 ± 2.89^{ab}	7.00 ± 1.20^a	1.53 ± 0.23^a
Group 7	65.00 ± 5.00^a	28.33 ± 2.89	27.00 ± 2.65^c	61.66 ± 4.64^{ab}	7.07 ± 0.57	1.00 ± 0.35^{ab}

		b			a	
Group 8	45.00 ± 1.00^c d	26.66 ± 2.88 b	43.66 ± 5.13^{ab} c	63.33 ± 1.41^{ab}	7.03 ± 0.32 a	0.93 ± 0.12^b
Group 9	48.33 ± 1.41^b c	28.33 ± 2.80 b	36.67 ± 2.58^{bc}	66.66 ± 2.88^{ab}	7.00 ± 0.72 a	1.10 ± 0.10^{ab}
Group 10	35.00 ± 8.03^c d	35.00 ± 1.54 b	42.60 ± 4.93^{ab} c	61.24 ± 2.08^{ab}	7.10 ± 0.61 a	1.07 ± 0.31^{ab}
Group 11	58.33 ± 1.93^a b	30.00 ± 5.00 b	49.32 ± 5.13^{ab}	58.33 ± 2.16^{ab}	7.33 ± 1.10 a	1.07 ± 0.32^{ab}

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(*Tetracarpidium conophorum*, *Sesame indicum* L, and *Mucuna pruriens*) seeds.

The results of the concentration of sex hormones measured in the male rats fed with medicinal plants (seeds) shown in the table 2. From the result, the testosterone concentration showed significant increase ($p<0.05$) in animal control group 1 (normal rats) while the concentration of the hormone in group 2 (ketoconazole induced animals with no treatment) significantly decreased ($p<0.05$) when compared across the treated groups 3-10 (Induced rats fed with 4g, 8g and 12g/kg between day of the test samples). Group 11 (Induced rats fed with 12g/kg between day of walnut sample) also revealed a significant increase ($p<0.05$) when compared with the rest of the treatment groups 3-10. The result of the present study also revealed a significant increase ($p<0.05$) in the concentration of FSH in the control group 1 (normal rats) when compared with that of groups 2-11 (animals fed with 4,8 and 12kg/g test samples). However, there was also an observed significant increase ($p<0.05$) in the concentration of Luteinizing hormone (LH) in the group 1 compared to that of groups 2-10 but the result revealed no significant increase ($p>0.05$) when compared group 1 (normal rats) and that of group 11 (animals fed with 12kg/g walnut sample). Group 5 (animals fed with 4kg/g walnut sample) and 6 (animals fed with 8kg/g velvet bean sample) showed significant decrease ($p<0.05$) when compared across other treatment groups. Moreover, the results of the estradiol in table 2 indicate a significant decrease ($p<0.05$) in group 1 (normal rats) compared to that group 2 (ketoconazole induced animals with no treatment) and that of the treatment groups 3-10 (animals fed with 4, 8 and 12kg/g test samples) but no significant difference ($p>0.05$)was observed when compared with that of group 11 (animals fed with 12kg/g walnut sample).

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153 Table 2: The concentration of sex hormones measured in the male rats fed with medicinal
 154 plants (mean \pm SD)

GROUPS	Testosterone (μ l/ml)	FSH (μ l/ml)	LH (μ l/ml)	Estradiol (μ l/ml)
Group 1	1.63 \pm 0.15 ^a	2.57 \pm 0.86 ^a	2.60 \pm 0.31 ^a	32.90 \pm 3.24 ^c
Group 2	0.33 \pm 0.06 ^d	1.37 \pm 0.12 ^b	1.90 \pm 0.10 ^{bc}	50.96 \pm 4.94 ^a
Group 3	0.40 \pm 0.20 ^{cd}	1.60 \pm 0.17 ^b	1.93 \pm 0.12 ^{bc}	42.77 \pm 11.01 ^{ab}
Group 4	0.43 \pm 0.25 ^{cd}	1.63 \pm 0.15 ^b	1.83 \pm 0.06 ^{bc}	43.80 \pm 8.58 ^{ab}
Group 5	0.53 \pm 0.23 ^{cd}	1.80 \pm 0.10 ^b	1.07 \pm 0.06 ^c	37.46 \pm 3.04 ^b
Group 6	0.50 \pm 0.26 ^{cd}	1.40 \pm 0.10 ^b	1.07 \pm 0.05 ^c	44.20 \pm 8.80 ^{ab}
Group 7	0.50 \pm 0.35 ^{cd}	1.53 \pm 0.06 ^b	1.10 \pm 0.10 ^{bc}	43.47 \pm 7.05 ^{ab}
Group 8	0.63 \pm 0.32 ^{cd}	1.43 \pm 0.06 ^b	1.10 \pm 0.01 ^{bc}	39.13 \pm 7.98 ^b
Group 9	0.47 \pm 0.11 ^{cd}	1.53 \pm 0.05 ^b	1.53 \pm 0.11 ^{bc}	50.23.6.44 ^{ab}
Group 10	0.67 \pm 0.06 ^{bc}	1.37 \pm 0.06 ^b	1.43 \pm 0.15 ^{bc}	48.70 \pm 8.55 ^{ab}
Group 11	0.77 \pm 0.05 ^b	1.63 \pm 0.28 ^b	1.47 \pm 0.05 ^{ab}	38.77 \pm 4.54 ^{bc}

4. CONCLUSION

159 The concentration of sex hormones measured in the male rat fed with medicinal plants
 160 (seeds) showed that the testosterone concentration significantly increased ($p<0.05$). It was also
 161 deduced that these medicinal plants including Walnut seeds significantly increased their Luteinizing
 162 hormone(LH) and Follicle Stimulating Hormone (FSH). This eventually enhanced the production of the
 163 reproductive hormones which enriched the fertility status of these animals.

164 Ethical Approval:

166 As per international standard or university standard written ethical approval has been collected and
 167 preserved by the author(s).

168 Consent: NA

169 **Disclaimer:**

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171 This paper is based on preliminary dataset. Readers are requested to consider this paper as
172 preliminary research article, as authors wanted to publish the initial data as early as possible.
173 Authors are aware that detailed statistical analysis is required to get a scientifically
174 established conclusion. Readers are requested to use the conclusion of this paper judiciously
175 as statistical analysis is absent. Authors also recommend detailed statistical analysis for
176 similar future studies.

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179 **REFERENCES**

180

181 Ajaiyeoba, E. O. and Fadare,D.A.(2006) Antimicrobial potential of extracts and fractions of the
182 African Walnut *Tetracarpidium conophorium*. **African Journal of Biotechnology**,5
183 (22):2322-2325.

184 BJORNDALH, L., SODOURLUND, I., KVIST, U.(2003).Evaluation of the one –step eosin-nigrosin staining
185 technique for human sperm vitality assessment.Human Reproduction.18:813-816.

186 COMHAIR , F., and VERMEULEM,(1995). Human semen analysis .Human Reproductive Updates.
187 1(4):343 362.

188 DASHISTANI , H .I and DAYEM, M .A.(2007). Hyperprolactinemia and hyper gonadotro-pins in infertile
189 men with severe oligospermia and azospermia. **International Journal of Endocrinology**,3:3-8.

190 EKALUO,U.B., IKPEME,E.V., UDENSI,O., MARKSON,A.A., MADUNAGU,B.E., OMOSUN,G. and UMANA,E.
191 (2010)Effect of aqueous ieaf extract of neem(*Azadirachta indica*) on the hormonal milieu of
192 male rats. **International Journal of Current Research**.

193 EKALUO ,U.B., UDOKPOH, A.E.,UDOFIA . U.U. AJANG , R.O.(2005).Comparative Toxicity of five Commonly
194 Used Analysis ,on Rat Sperm Count and Morphology. **Global Journal of Pure and Applied**
195 **Sciences**. 2(1):81-82.

196 ORTH, A .B., ROYSE, D.J.,and TIEN,M.(1993).Ubiquity of lignin –degrading peroxidases among various
197 wood-degrading fungi.Applied Environmental Microbiology,59 :4017-4023.

198 VAWDA ,A.I. and DAVIES, A .G.(1986). An Investigation into the effects of Ketoconazole on testicular
199 function in Wistar rats. Acta Endocrinology. 111(2): 246 -251.

200 World Health Organization .(1992).WHO laboratory manual for the Examination of human Semen
201 and Sperm.Cervical Mucus Interaction Cambridge University Press.

202 ZWARTS,L., SAVAGE ,G.P. and MC NEIL, B.L.(1999) Fatty Acid Content of New Zealand-Grown
203 Walnuts(*Juglans regia L*), **International Journal of Food Sciences and Nutrition**,50: 189

204 Golla, U.R. Gajam, P.K. Mohammad, A. R. Kumar. A. K. Raj, B. S. S. ASSESSMENT OF
205 BIOACTIVITY OF Desmostachya bipinnata (L.) Stapf USING BRINE SHRIMP (ARTEMIA
206 SALINA) LETHALITY ASSAY. Pharmacologyonline 3: 982-990 (2011).

207
208 Krishnaraju, A. V., Rao, T. V., Sundararaju, D., Vanisree, M., Tsay, H. S., & Subbaraju, G. V.
209 (2005). Assessment of bioactivity of Indian medicinal plants using brine shrimp (Artemia
210 salina) lethality assay. *Int J Appl Sci Eng*, 3(2), 125-34.

211 Mukherjee, P. K., & Wahile, A. (2006). Integrated approaches towards drug development
212 from Ayurveda and other Indian system of medicines. *Journal of ethnopharmacology*,
213 103(1), 25-35.

214