

**ASSESSMENT OF PESTICIDE RESIDUES IN FRESH VEGETABLES FROM THREE MAJOR MARKETS IN LAGOS USING QuEChERS METHOD AND GC-MS**

**ABSTRACT**

The study evaluates pesticide residues in fresh vegetables from three major markets in Lagos and verified compliance of these fresh vegetables with the maximum residue levels (MRLs) as specified by Codex Alimentarius Commission. The residues were extracted by means of multi residue method based on the Quick, Easy, Cheap, Effective, Rugged and Safe (QuEChERS) method, Gas Chromatography - Mass Spectrometry (GC-MS) was used to determine two organophosphates (dichlorvos and chlorpyrifos), two organochlorines (endosulfanII and chlorothalonil), two carbamates(carbaryl and carbofuran) and two pyrethroids(alpha-cypermethrin and lambda-cyhalothrin). Fifteen samples of five common vegetables (cabbage, lettuce, fluted pumpkin, bitter leaf and African spinach) were analyzed. The linear ranged used were 0.005, 0.1, 0.25, 0.5 and 1.0mg/l, resulting to  $r^2$  of  $\geq 0.996$ . While the mean recoveries obtained for two fortification levels (0.1 and 0.25 ppm) in three replicates for lettuce control sample and spinach control sample were 96.7 to 104.3 % , 96.3 to 101.3 %,and 92.0 to 114.3 % ,92.1 to102.1 % respectively. A satisfactory precision of  $RSD < 20\%$  was recorded. The limit of detection (LOD) and limit of Quantitation (LOQ) were between 0.005 -0.050mg/kg and 0.015-0.150 mg/kg respectively. Pesticide residues detected ranges from 0.025-0.529 mg/kg. EndosulfanII, Chlorothalonil, carbaryl and Carbofuran were not detected during the entire study. However, pesticide residues detected were below the MRLs specified by Codex Alimentarius Commission.

Keyword: Gas Chromatography, Mass Spectrometry, Organophosphates, Organochlorines Carbamates, Pyrethroids

**INTRODUCTION**

Vegetables are important components of the human diet since they provide essential nutrients and antioxidants that are required for most of the reactions occurring in the body. A high intake of vegetables has been encouraged not only to prevent consequences due to vitamin deficiency but also to reduce the incidence of major diseases such as cancer, cardiovascular diseases and obesity. Like other crops, vegetables are attacked by pests and diseases during production and storage leading to damages that reduce the quality and the yield. In order to reduce the loss and maintain the quality vegetables harvest, pesticides are used to destroy pests and prevent diseases. The use of pesticides has increased because they have rapid action; decrease toxins produced by food infecting organisms and are less labour intensive than other pest control methods. However, the use of pesticides during production often leads to the presence of pesticide residues in vegetables after harvest. Shrestha *et al*,(2010) reported that commercial vegetable growers opt for the application of a variety of pesticides that belong to diverse chemical class, pesticides such as Parathion-Methyl, Dichlorvos, Carbofuran , Chlorpyrifos, Cypermethrin, Deltamethrin, Dimethoate, Endosulfan, Lambda-Cyhalothrin, Profenofos, and fungicides such as Carbendazim,

41 Mancozeb and Metalaxyl in order to overcome pest resistance and pest resurgence[1]. According  
42 to Petsas A.S *et al.*,(2017), vegetables are capable of retaining large quantities of pesticides[2].  
43 The presence of pesticide residues is a concern for consumers because pesticides are known to  
44 have potential harmful effects to other non-targeted organisms. The major concerns are their  
45 toxic effects such as interfering with the reproductive systems and fetal development as well as  
46 their capacity to cause cancer and asthma [3]. Some of the pesticides are persistent and therefore  
47 remain in the body causing long term exposure. Use of pesticides in Nigeria is not well  
48 controlled as compare to the developed countries due to ineffective legislation, lack of awareness  
49 and inappropriate pesticide management. Global scientific concerns have been raised regarding  
50 the potential toxicity of pesticides that have promoted their strict regulation in order to protect  
51 consumers, environment and also the users of pesticides. To ensure the safety of food for  
52 consumers, numerous legislations such as codex directives (CODEX Committee on Pesticide  
53 Residues, 2003) have established maximum residue limits (MRLs) for pesticides in foodstuffs  
54 [4]. Maximum residue limits (MRLs) values defined as the highest levels of pesticide residues  
55 that are legally tolerated in or on food or feed when pesticides are applied correctly (adoption of  
56 Good Agricultural Practices, GAPs) were established internationally by Codex Alimentarius  
57 Commission. Different extraction and quantification methods are used by various researchers for  
58 estimation of pesticide residues in several vegetables. The main criteria for opting any  
59 methodology is that analytical method should be fast, easy, inexpensive and applicable to  
60 different matrices. In recent years, gas chromatography-mass spectrometry (GC-MS) and gas  
61 chromatography-tandem mass spectrometry (GC-MS/MS) have been a versatile tool use in  
62 pesticide analysis. The ability to perform multi-analysis of analytes using GC-MS is another  
63 attribute that makes the technique unique in its applications. Analysis of several pesticides in  
64 vegetable samples from countries like Kenya, India, Pakistan, Kuwait by GC-MS in one run has  
65 been reported[5, 6, 7, 8]. The aim of this study was to evaluate presence of some pesticide  
66 residues in fresh vegetable from three major markets in Lagos and to verify compliance of these  
67 fresh vegetables with the maximum residue levels' (MRLs) as specified by Codex Alimentarius  
68 regulations for products present in the market.

## 69 **EXPERIMENTAL**

### 70 **Sampling Collection and Storage**

71 A total number of 75 vegetables were sampled from the 3 major vegetable markets. Five samples  
72 of each vegetable in each market were collected. The vegetable species are Lettuce, Cabbage,  
73 Fluted pumpkin, Bitter leave and spinach. The sampling was done according to guideline in  
74 China (SAC,2008) on sampling for official control of pesticide residues[9]. Samples were  
75 packed in separated polythene bags, sealed and labeled with a unique sample identity and placed  
76 in an ice chest box. All samples were stored at 4°C.The composite samples were prepared by  
77 systematic mixing of the five samples of each vegetable in each market. After the vegetable  
78 samples were mixed and blended using Stephan blender, the composite sample were then  
79 extracted within 24 hours from the time of their collection. Control sample of lettuce and spinach  
80 were also collected.

### 81 **Sample Extraction and Cleanup**

82 The QuEChERS sample preparation method for pesticides was applied to all the samples [10]. A  
83 10 g portion of the homogenized sample was weighed into a 50 ml polytetrafluoroethylene  
84 (PTFE) tube added 10 ml of acetonitrile. Then, 4 g magnesiumsulphate (MgSO<sub>4</sub>), 1 g sodium  
85 chloride (NaCl), 1g sodium citrate tribasic dehydrate(Na<sub>3</sub> C<sub>6</sub>H<sub>5</sub>O<sub>7</sub>.2H<sub>2</sub>O) and 0.5g sodium citrate  
86 dibasic sesqihydrate (C<sub>6</sub>H<sub>5</sub>Na<sub>2</sub> O<sub>7</sub>.1.5H<sub>2</sub>O ) were added, and the sample was shaken vigorously  
87 for 1 min on a vortex .The samples were then centrifuged at 3000 rpm for 5 min. The supernatant  
88 (6 ml) were transferred to a 15 ml PTFE tube to which 900 mg MgSO<sub>4</sub>, 150 mg PSA and 150  
89 mg GCB were added. The extract was shaken using a vortex mixer for 30 second and centrifuged  
90 at 3000 rpm again for 5 min. 2ml of the supernatant were taken into a graduated test tube and 20  
91 microliter of formic acid (HCOOH) was added to adjust the <sup>pH</sup>. These extracts were evaporated  
92 to dryness under a stream of nitrogen and reconstituted with 2ml of Hexane: Acetone (4:1) and  
93 transferred to 2 ml vial and sealed for quantification using gas chromatograph equipped with  
94 mass spectrometry (GC-MS).

### 95 **Preparation of Pesticide Standard Solution**

96 The pesticide reference standards were sourced from Dr Ehrenstorfer, Augsburg, Germany and  
97 the purity of standards ranged from 96% to99%. Individual stock standard solution of the studied  
98 pesticide were prepared by weighing accurately 10mg of each pesticide into 10ml volumetric  
99 flask and dissolved with acetone, yielding a concentration of 1mg/ml. A mixed stock standard  
100 solution of pesticides were prepared 5µg/mL in hexane: acetone (4:1).The standard mixture of  
101 pesticides was prepared at 0.005, 0.1, 0.25, 0.5 and1.0 µg/mL concentrations by serial dilution  
102 technique for preparing the calibration curve. The entire standard was stored at -20°C.

### 103 **Instrumental method**

104 Chromatographic instrumentation and quantification were carried out by Gas chromatograph-  
105 mass spectrometer GC-MS (Shimadzu QP2010 Ultra) with a GC column HP-5MS 5 % phenyl-  
106 95 % methyl siloxane, 30x0.25 mm id x 0.25 film thickness. The GC operating conditions: split  
107 less injection, injector temperature 250 °C, helium carrier gas (99.9999 purity) at flow rate 1.20  
108 mL min<sup>-1</sup> with column head pressure 89.4 kpa, oven temperature from 100°C (1 min hold), then  
109 raised to 200 °C at the rate of 10 °C (2min hold) afterwards raised to 300 °C at the rate of10 °C  
110 (5 min hold). The sample was injected in split less modes. The MS system was routinely set in  
111 selective ion monitoring (SIM) mode and each compound was quantitated based on peak area  
112 using one target and one or two qualifier ion. Mass spectrometer parameter was set as follows:  
113 electron impact ionization mode with 70 Ev electron energy. Ion source temperature 200 °C, MS  
114 interface temperature 250 °C.

### 115 **RESULT AND DISCUSSION**

116 Table 1 show the LOD and LOQ obtained for each pesticide. The LOD and LOQ values  
117 obtained ranged from 0.005 to 0.05mg/kg and 0.015 to 0.150mg/kg respectively. The linearity  
118 was assessed by the correlation coefficient (R<sup>2</sup>) resulted from the five-point calibration curve.  
119 The linearity was observed in the range 0.005 -1.0mg/kg and all correlation coefficient were R<sup>2</sup>  
120 ≥0.996.

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Table1: List of Pesticides with retention time (Rt), LOD, LOQ and Correlation coefficient (R<sup>2</sup>)

<b>Pesticide Name</b>	<b>Retention time (Rt)min</b>	<b>LOD(mg/kg )</b>	<b>LOQ(mg/kg)</b>	<b>Correlation Coefficient(R<sup>2</sup>)</b>
<b>Dichlorvos</b>	6.570	0.005	0.015	0.995860
<b>Chlorpyrifos</b>	13.429	0.010	0.030	0.997184
<b>Endosulfan II</b>	15.557	0.010	0.030	0.998576
<b>Chlorothanolin</b>	11.176	0.050	0.150	0.998796
<b>Carbaryl</b>	10.040	0.050	0.150	0.998460
<b>Carbofuran</b>	10.861	0.010	0.030	0.998063
<b>α-Cypermethrin</b>	21.919	0.005	0.015	0.999423
<b>λ-Cyhalothrin</b>	19.981	0.010	0.030	0.999671

Recovery study was carried out to determine the method accuracy and precision .For each blank matrix (Lettuce and spinach), two concentration levels of 0.1 ug/ml and 0.25 ug/ml at replicate (n=3) were determined. All the studied pesticides, the recoveries of these two spiking levels ranged between 92.0 to114.3 percent. The method was proved to be repeatable with RSD in range of 1.2 to 14.0 % at all spiking levels. Result obtained complied with SANCO guidelines [11]. (Document No. SANCO/12571/2013). Recovery within the range70-120 % and RSD ≤ 20 %.

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Table 2: Average recovery (n=3) and Relative standard deviation (%RSD)

Spiking level(ug/ml)	Lettuce				African Spinach			
	0.1		0.25		0.1		0.25	
	Recov ery(%)	RSD (%)	Recov ery(%)	RSD (%)	Recov ery(%)	RSD (%)	Recover y(%)	RSD (%)
<b>Dichlorvos</b>	100.7	1.4	98.7	1.6	114.3	11.8	92.7	3.4
<b>Chlorpyrifos</b>	102.3	6.7	99.6	1.2	95.7	7.9	100.3	14.0
<b>EndosulfanII</b>	100.7	2.1	101.1	2.0	102.5	6.4	99.9	7.1
<b>Chlorothanon il</b>	100.7	6.5	96.3	4.0	92.0	13.3	100.3	3.0
<b>Carbaryl</b>	101.3	4.1	98.9	1.5	104.7	8.3	98.0	6.0
<b>Carbofuran</b>	104.3	4.8	101.1	1.4	110.3	9.2	102.1	6.2
<b>α- Cypermethri n</b>	98.0	11.4	99.3	1.3	82.0	1.4	99.2	9.0
<b>λ- Cyhalothrin</b>	96.7	6.9	97.7	2.3	94.9	4.0	101.4	8.5

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Table 3: Showing Codex Maximum Residue Limits (MRLs) of studied vegetables mg/kg [23]

<b>Pesticide</b>	<b>Lettuce</b>	<b>Cabbage</b>	<b>Fluted Pumpkin</b>	<b>Bitter leaf</b>	<b>African Spinach</b>
<b>Dichlorvos</b>	0.5	0.5	0.5	0.5	0.5
<b>Chlorpyrifos</b>	0.1	0.05	0.05	0.05	0.05
<b>Carbaryl</b>	10	5.0	3.0	10	10
<b>Carbofuran</b>	0.1	0.5	0.2	0.2	0.2
<b>Endosulfan II</b>	1.0	2.0	2.0	2.0	2.0
<b>Chlorothanolin</b>	1.0	1.0	1.0	1.0	1.0
<b><math>\alpha</math>- Cypermethrin</b>	2.0	1.0	1.0	1.0	1.0
<b><math>\lambda</math>-Cyhalothrin</b>	0.2	0.2	0.2	0.2	0.2

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Table 4: Pesticide Residue Concentrations (mg/kg ) in fresh vegetable samples collected from Mile12 market.

<b>Pesticide</b>	<b>Lettuce</b>	<b>Cabbage</b>	<b>Fluted Pumpkin</b>	<b>Bitter Leaf</b>	<b>African Spinach</b>
<b>Dichlorvos</b>	0.178	0.268	0.048	0.025	ND
<b>Chlorpyrifos</b>	0.082	0.049	0.044	ND	0.038
<b>Carbaryl</b>	ND	ND	ND	ND	ND
<b>Carbofuran</b>	ND	ND	ND	ND	ND
<b>Endosulfan II</b>	ND	ND	ND	ND	ND
<b>Chlorothanolin</b>	ND	ND	ND	ND	ND
<b><math>\alpha</math>- Cypermethrin</b>	0.168	0.529	0.094	0.096	0.065
<b><math>\lambda</math>-Cyhalothrin</b>	0.118	0.171	0.113	0.075	0.064

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184 Table 5: Pesticide Residue Concentrations (mg/kg) in fresh vegetable samples collected from  
 185 Oyingbo market.

<b>Pesticide</b>	<b>Lettuce</b>	<b>Cabbage</b>	<b>Fluted Pumpkin</b>	<b>Bitter Leaf</b>	<b>African Spinach</b>
<b>Dichlorvos</b>	0.088	0.058	0.153	0.080	0.077
<b>Chlorpyrifos</b>	0.060	0.044	0.039	0.038	ND
<b>Carbaryl</b>	ND	ND	ND	ND	ND
<b>Carbofuran</b>	ND	ND	ND	ND	ND
<b>Endosulfan II</b>	ND	ND	ND	ND	ND
<b>Chlorothanolin</b>	ND	ND	ND	ND	ND
<b><math>\alpha</math>- Cypermethrin</b>	0.045	0.072	0.028	0.090	ND
<b><math>\lambda</math>-Cyhalothrin</b>	0.129	0.200	0.071	0.150	0.101

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188 Table 6: Pesticide Residue Concentrations (mg/kg ) in fresh vegetable samples collected from  
 189 Oshodi market.

<b>Pesticide</b>	<b>Lettuce</b>	<b>Cabbage</b>	<b>Fluted Pumpkin</b>	<b>Bitter Leaf</b>	<b>African Spinach</b>
<b>Dichlorvos</b>	0.045	0.400	0.058	0.121	0.027
<b>Chlorpyrifos</b>	0.036	0.048	0.038	0.047	0.040
<b>Carbaryl</b>	ND	ND	ND	ND	ND
<b>Carbofuran</b>	ND	ND	ND	ND	ND
<b>Endosulfan II</b>	ND	ND	ND	ND	ND
<b>Chlorothanolin</b>	ND	ND	ND	ND	ND



<b><math>\alpha</math>- Cypermethrin</b>	0.240	0.045	0.072	0.044	0.032
<b><math>\lambda</math>-Cyhalothrin</b>	0.054	0.081	0.075	ND	ND`

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191 The result of GC-MS analysis of pesticide residues in fresh vegetable samples is presented in  
 192 table 4; table 5 and table 6. Eight different pesticides were investigated and quantified with 54  
 193 detections. From table 4, residues of dichlorvos were found in 4 of 5 fresh vegetable samples in  
 194 order of cabbage (0.268mg/kg) > lettuce (0.178mg/kg) > fluted pumpkin (0.048mg/kg) > bitter leaf  
 195 (0.025mg/kg). None of the sample had dichlorvos above the Codex recommended MRL value of  
 196 0.5mg/kg. Chlorpyrifos was another pesticide detected in 4 of 5 fresh vegetable samples  
 197 analysed in mile 12 market, This pesticide was not detected in bitter leaf, however, the  
 198 concentration of chlorpyrifos was found to be in the order of lettuce(0.082mg/kg) > cabbage(  
 199 0.049mg/kg) > fluted pumpkin(0.044mg/kg) > African spinach(0.038mg/kg). None of the  
 200 vegetable samples exceeded the MRL value of 0.05mg/kg for cabbage, fluted pumpkin, African  
 201 spinach and 0.1mg/kg stipulated for lettuce. Alpha cypermethrin was detected in all the five fresh  
 202 vegetable samples from mile 12 market, detected concentration in vegetable samples were in the  
 203 range of cabbage(0.529 mg/kg) > lettuce(0.168mg/kg) > bitter leaf(0.096  
 204 mg/kg) > fluted pumpkin(0.094 mg/kg) > African spinach(0.065 mg/kg) all the vegetable samples  
 205 did not exceed the 1.0mg/kg MRL stated for cabbage, fluted pumpkin, bitter leaf, African  
 206 spinach and 2.0mg/kg stated for lettuce. Lambda-cyhalothrin was detected in all the five  
 207 vegetable samples from mile 12 market in the order of cabbage (0.171 mg/kg) > lettuce (0.118  
 208 mg/kg) > fluted pumpkin(0.113 mg/kg) > bitter leaf(0.075 mg/kg) > African spinach(0.064mg/kg)  
 209 and were all below the Codex MRL of 0.2mg/kg. EndosulfanII, chlorothanolin, carbaryl and  
 210 carbofuran were not detected in all the five fresh vegetable samples from mile 12. A similar  
 211 finding by Adyel, T.M *et al.*, (2013) did not detect any carbofuran residues in vegetable  
 212 samples[12].

213 Table 5 ; Result from Oyingbo market followed similar trend with that of mile 12 market,  
 214 dichlorvos was detected in all the five fresh vegetable samples from Oyingbo market ranging  
 215 from the fluted pumpkin(0.153 mg/kg) > lettuce(0.088 mg/kg) > bitter leaf(0.080 mg/kg) >  
 216 African spinach(0.077 mg/kg) > cabbage(0.058 mg/kg). The residues of dichlorvos detected were  
 217 below the MRL of 0.5 mg/kg stipulated by Codex. Chlorpyrifos residue was detected in 4 of the  
 218 5 fresh vegetable samples from Oyingbo market. The concentrations were in the order of lettuce  
 219 (0.060mg/kg) > cabbage (0.044 mg/kg) > fluted pumpkin (0.039 mg/kg) > bitter  
 220 leaf(0.038mg/kg). Residues in these vegetable samples were not above the MRL of 0.05 mg/kg  
 221 and 0.1mg/kg for lettuce. African spinach was free of chlorpyrifos residues. This study revealed  
 222 that 4 out of the 5 fresh vegetables from Oyingbo market were contaminated with  $\alpha$ -  
 223 cypermethrin in the order of bitter leaf (0.090 mg/kg) > cabbage(0.072 mg/kg) > lettuce(0.04  
 224 5mg/kg) > fluted pumpkin(0.028 mg/kg). None of the detected fresh vegetable samples exceeded  
 225 the MRL of 1.0mg/kg and 2.0 mg/kg for lettuce. The concentrations of  $\lambda$ - cyhalothrin in this  
 226 study were significantly below their MRLs of 0.2 mg/kg. Their contamination followed the order  
 227 of cabbage (0.200 mg/kg) > bitter leaf (0.150 mg/kg) > lettuce(0.129 mg/kg) > African  
 228 spinach(0.101mg/kg) > fluted pumpkin(0.071mg/kg). EndosulfanII, chlorothanolin, carbaryl and

229 carbofuran were not detected. The finding of this study is comparable to the findings of the study  
230 conducted by Mahugija J.A. *et al.*,(2017) [13].

231 Table 6 illustrated the result of pesticide residues obtained from Oshodi market. Dichlorvos was  
232 detected in all the 5 vegetable samples from Oshodi market. The order of contamination were  
233 found to be cabbage (0.400mg/kg) > bitter leaf (0.121mg/kg)>fluted pumpkin (0.058mg/kg)  
234 >lettuce (0.045mg/kg)> African spinach(0.027mg/kg). The concentration of dichlorvos residues  
235 were not above the MRLs 0.5mg/kg set by codex. In same way, chlorpyrifos was detected in all  
236 the 5 vegetable samples ,the order of magnitude of chlorpyrifos residues in vegetable samples  
237 were as follows: cabbage(0.048 mg/kg) >bitter leaf(0.047 mg/kg)>African spinach (0.040  
238 mg/kg)> fluted pumpkin(0.038 mg/kg) > lettuce (0.036 mg/kg). The residues of chlorpyrifos in  
239 all the vegetable samples did not exceed the MRLs of 0.05mg/kg and 0.1mg/kg set for lettuce.  
240 Similarly,  $\alpha$ -cypermethrin was detected in all the 5 vegetable samples analysed. The result  
241 showed that lettuce (0.240 mg/kg)>fluted pumpkin(0.072 mg/kg) >cabbage (0.045  
242 mg/kg)>bitter leaf(0.044 mg/kg) > African spinach(0.032 mg/kg). $\alpha$ -cypermethrin residues in this  
243 study were below the MRLs of 1.0 mg/kg and 2.0 mg/kg set for lettuce. In the same vein,  $\lambda$ -  
244 cyhalothrin was detected in 3 of 5 of the vegetable samples analysed. The findings revealed that  
245 cabbage (0.081mg/kg) >fluted pumpkin (0.075 mg/kg)>lettuce (0.054 mg/kg). None of the  
246 sample detected exceeded the MRLs of 0.2mg/kg set by Codex. Endosulfan II, chlorothanolin,  
247 carbaryl and carbofuran were not detected in all the samples.

248 This study shows the evidence of the presence of pesticide residues in vegetable samples in  
249 Lagos market. The samples analyzed contained pesticide residues below the MRLs. All the five  
250 fresh vegetable samples investigated in this study were free from contamination of endosulfan II  
251 and chlorothanolin. These results were not surprising since most of organochlorine pesticides  
252 were banned or their use is severely restricted in Nigeria. Carbaryl and carbofuran were not  
253 detected in all the vegetable samples analyzed in this study. This suggests that they were not  
254 used for the vegetables studied or there was no significant contamination due to these  
255 compounds. The occurrences of multiple residues in all the samples analyzed were likely to be a  
256 consequence of the application of different types of pesticides to protect vegetables against insect  
257 pests and diseases [1]. The incidence of multi-residue pesticide contamination in different  
258 vegetable samples has also been reported in other studies [1,14]. Pesticide residues detected in  
259 this study were similar to those detected in other studies [15, 16, 17, 18, 19, 20].**Extensive  
260 spraying of various chemicals on vegetables and non-availability of proper guidance about  
261 pesticide application may lead to high or low pesticide residue levels on vegetables [21].**

262 From the results in this study, it is plausible to state that vegetable growers from studied area  
263 applied good agricultural practice (GAP) and being cautious of with-hold period (WHP). A  
264 withholding period is the minimum time for vegetables treated with pesticide must wait before  
265 harvesting, such a period allows enough time for the pesticide to degrade to an acceptable  
266 level.[1,22]The results of this study were interpreted in relation to maximum residue  
267 limits(MRLs) requirements set by Codex Alimentarius Commission for vegetables.

## 268 CONCLUSION

269 The results indicated that all the five vegetable samples were contaminated with two or more  
270 pesticide residues with concentrations below the maximum residues limits(MRLs).From a health  
271 perspective, the observed levels of pesticide residues did not pose a potential health risk to  
272 consumers. Hence, the consumption of vegetables is safe.

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