

## Short communication

### The **In vivo** Antioxidant Protective Activity of *Mangifera indica* Cold Aqueous Leaf Extract in *Drosophila melanogaster*.

#### ABSTRACT

Objective: To evaluate **in vivo** antioxidant activity of *Mangifera indica* cold aqueous leaf extract

Methods: a number of 50 adult flies were exposed to graded concentrations of *Mangifera indica* cold aqueous leaf extract, 2.5 mg/10 g diet, 5 mg/10 g diet and 10 mg/10 g diet for 5 days. Each concentration was prepared in 200µl of distilled water and replicated five times. 10g diet with 200µl distilled water served as control. Mortality reading was taken at 24 hours interval. The flies were homogenized, centrifuged and the supernatant was used to assay for Glutathione-S-transferase (GST), Catalase (CAT) and Total thiol content.

Results: The % mortality of flies after 5 days showed 32.5 %, 0 %, 15.5 % and 37 % in the control (**10g diet with 200µl of distilled water**), 2.5 mg/10 g diet, 5 mg/10 g diet and 10 mg/ 10g diet respectively. There was elevation in total thiol content and high GST and CAT activity in 2.5 mg/10 g diet and 5 mg/10 g diet treated flies.

Conclusion: the 100% and 85% survival of 2.5 mg/10 g and 5 mg/10 g diet-treated flies respectively and increase of fly' s antioxidant system after 5days exposure at these concentrations may suggest protective activity of *Mangifera indica* in *D. melanogaster*.

#### Keywords

In vivo, Antioxidant activity, Cold aqueous extract, *Drosophila melanogaste*, *Mangifera indica*, Catalase ,Total thiol, Glutathione-S-transferase

#### 1 Introduction

Antioxidants act as a defense mechanism that protect against deleterious effects of oxidative reaction produced by reactive oxygen species (ROS) in a biological system <sup>1</sup>.

Overproduction of ROS and/or inadequate antioxidants has been implicated in the pathogenesis and complications of some disease conditions like diabetes, Alzheimer's disease, cancer, atherosclerosis, arthritis, neurodegenerative disease, and aging process <sup>2,3</sup>.

Antioxidants have been reported to prevent oxidative damage caused by ROS by reacting with

32 free radicals, chelating, and catalytic metals and also by acting as oxygen scavengers  
33 <sup>4,5</sup>. Oxidative stress is characterized by imbalance between oxidant-producing systems and  
34 antioxidant defense mechanisms, resulting in excessive formation of reactive oxygen species  
35 (ROS). Excessive accumulation of ROS can damage bio-molecules, including lipids, proteins and  
36 nucleic acids <sup>6</sup>. Thiol groups are important members of the antioxidant team and have been  
37 shown to destroy ROS and other free radicals by enzymatic and non-enzymatic mechanisms <sup>7</sup>.  
38 Total thiol groups of proteins are mainly responsible for their antioxidant response, and they  
39 can serve as a sensitive indicator of oxidative stress <sup>8</sup>.

40 Mango (*Mangifera indica* L.) is a juicy stone fruit belonging to the family of Anacardiaceae  
41 in the order of Sapindales and is grown in many parts of the world, particularly in tropical  
42 countries; Mango is now commercially grown in more than 87 countries <sup>9</sup>. It has been well  
43 documented that mango fruits are an important source of micronutrients, vitamins and other  
44 phytochemicals. Moreover mango fruits provide energy, dietary fiber, carbohydrates, proteins,  
45 fats and phenolic compounds <sup>9</sup>, which are vital to normal human growth, development and  
46 health <sup>10</sup>. Various parts of mango are used for more than thousands of years as wide variety of  
47 ethno medicinal use <sup>11</sup>. **Mango extracts from leaves, fruit, seed kernel, fruit pulp, roots, bark  
48 and stem bark have been used extensively for medicinal purposes in many countries<sup>12</sup>.** The  
49 ethno-medical use of mango stem bark aqueous extract in Cuba has been documented widely <sup>5</sup>.  
50 It has been extensively used in cancer, diabetes, asthma, infertility, lupus, prostatitis, prostatic  
51 hyperplasia, gastric disorders, arthralgias, mouth sores and tooth pain <sup>12</sup>

52 *Drosophila melanogaster*, known colloquially as the fruit fly, remains one of the most  
53 commonly used model organisms for biomedical science. For more than one hundred years, the  
54 low cost, rapid generation time, and excellent genetic tools have made the fly indispensable for  
55 basic research. The addition of numerous molecular tools has allowed the model system to  
56 keep up with the latest advances. In this issue, various authors provide examples of how  
57 *Drosophila* is currently being used, and what directions they think the system is moving in.  
58 From human disease modeling to the dissection of cellular morphogenesis and to behavior and  
59 aging.<sup>13</sup>

60 The **in vitro** antioxidant activity of **Mangifera indica** plant extract has been established but  
61 with no or little information on its **in vivo** antioxidant activities. The specific objective of this  
62 work was to evaluate the **in vivo** antioxidant protective activities of *Mangifera indica* in  
63 *Drosophila melanogaster*.

64

## 65 **2 Material and methods**

## 66 2.1 Chemicals

67 All chemicals used were of analytical grade. Distilled water purchased from Africa Centre of  
68 Excellence in Phytomedicine Research and Development, Jos, plateau State. Radox Protein kit  
69 was purchased from Medicom, Jos Plateau State. 1-chloro-2,4-dinitrobenzene, (CDNB) and 5,5'-  
70 dithiobis(2-nitro-benzoic acid) (DTNB) were purchased from Sigma Aldrich (St Louis, MO).

## 71 2.2 Plant Collections

72 *Mangifera indica* leaf was collected from University of Jos Senior staff quarter, Jos North,  
73 Plateau State, Nigeria. The leaves were air dried using room temperature for 7 days, and then  
74 pulverized to powder using a commercial grinding machine. It was kept in an air tight container  
75 before extraction. The extraction was carried out by maceration method using 1:10 of plant  
76 material to distilled water for 72hrs. Filtered and the filtrate was concentrated to dryness using  
77 freeze dryer.

## 78 2.3 *In vivo* Antioxidant Assay

79 *In vivo* antioxidant assay was carried out by exposing (ingestion) 50 flies to graded  
80 concentration (2.5mg, 5mg, and 10mg) of cold aqueous leave extracts for 5 days. Each  
81 concentration was prepared in 200µl of distilled water and replicated 5 times. 10g diet with  
82 200µl of distilled water served as control. At the end of the exposure period (5 days), the flies  
83 (50) from each group of control and cold leaves extract-treated flies were anaesthetized in ice,  
84 weighed, and homogenized in 0.1 M phosphate buffer, pH 7.0 (1 mg: 10 µL), and centrifuge for  
85 10 min at 4000 rpm (temperature, 4 °C). The supernatants obtained were used to determine the  
86 activities of Catalase (CAT), Glutathione-S-transferase (GST) and Total thiol content.

87 **2.3.1 Total thiol determination:** Total thiol content was determined using the method of Ellman  
88 <sup>14</sup>. The reaction mixture contained 510uL potassium phosphate buffer (0.1 M, PH 7.4), 25 uL of  
89 sample as well as 30uL of DTNB (10 mM). After incubation for 30 min at room temperature, the  
90 absorbance was measured at 412 nm and used to calculate the sample total thiol levels (in  
91 mmol/mg protein) using 35ul of GSH as standard.

92 **2.3.2 Glutathione-S-transferase (GST) activity:** The activity of glutathione-S-transferase (GST;  
93 EC 2.5.1.18) was determined by the method of Habig and Jacoby <sup>15</sup> using 1-chloro-2,4-  
94 dinitrobenzene (CDNB) as substrate. The assay reaction mixture contained 600uL of solution A  
95 (20 uL of 0.25 M potassium phosphate buffer, pH 7.0 with of 2.5 mM EDTA, and 510 uL of  
96 0.1 M GSH at 25 °C), 60 uL of sample (1:5 dilution) and 30uL of 25 mM CDNB. An increase in  
97 absorbance was measured at 340 nm for 2min at 10 s interval using spectrophotometer (Jenway).

98 The data were expressed in mmol/min/mg of protein using the molar extinction coefficient ( $\epsilon$ ) of  
99 9.6 mM<sup>-1</sup> cm<sup>-1</sup> of the coloured GS–DNB conjugate formed by GST.

100 **2.3.3 Catalase (CAT) activity:** The measurement of catalase (CAT; EC 1.11.1.6) activity was  
101 followed by a procedure described by Aebi <sup>16</sup>. The reaction mixture containing 100 mL of  
102 potassium phosphate buffer, pH 7.0, 194mL of 300 mM H<sub>2</sub>O<sub>2</sub> to form solution A. 10 uL of  
103 sample was reacted with 590ul of solution A and monitoring the clearance of H<sub>2</sub>O<sub>2</sub> at 240 nm at  
104 25 °C. The decrease in H<sub>2</sub>O<sub>2</sub> was monitored for 2 min (10 s intervals), at 240 nm using a  
105 UV– visible spectrophotometer (Jenway) and expressed as mmol of H<sub>2</sub>O<sub>2</sub> consumed/min/mg of  
106 protein.

## 107 **2.4 Statistical analysis**

108 The data was expressed as mean±SEM (standard error of mean), and the statistical analysis was  
109 carried out using one-way analysis of variance (ANOVA) followed by Turkey's post-hoc test.  
110 The results was considered statistically significant at p <0.05.

## 111 **3. Results and discussion**

### 112 **3.1 Five (5) Days Mortality of *Mangifera indica* cold aqueous leaf extract-treated Flies**

113 Mortality result of Flies exposed to *Mangifera indica* cold aqueous leaf extract was high at 10  
114 mg / 10 g diet while the least mortality was recorded in 2.5 mg /10 g diet. 32.5 % mortality was  
115 recorded in the control while 0 %, 15.5 % and 37 % was recorded in 2.5 mg /10 g diet, 5 mg /10  
116 g diet and 10 mg/10 g diet of *Mangifera indica* cold aqueous leaf extract respectively. There  
117 was significant difference (p<0.05) comparing 2.5 mg/10 g diet of *Mangifera indica* cold  
118 aqueous leaf extract to the control group. This suggests that 2.5 mg/10 g diet and 5 mg /10 g  
119 diet of *M. indica* may have protective activity in *D. melanogaster*. The % mortality result is  
120 presented in figure 1

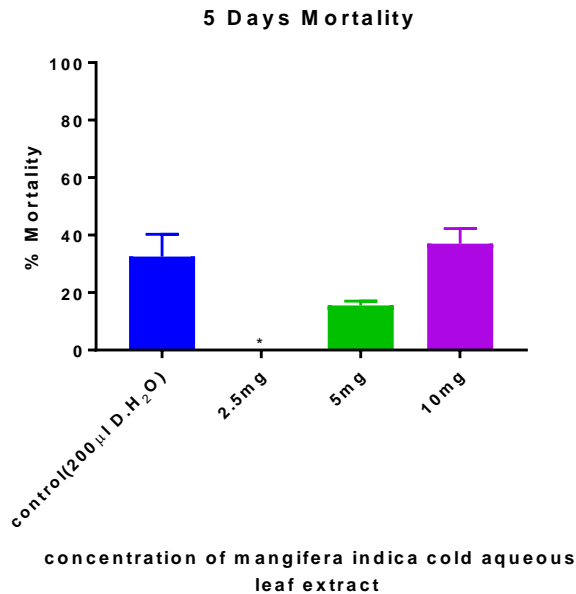


Fig 1: 5 Days %Mortality of *Mangifera indica* cold aqueous leaf extract-treated Flies

### 3.2 Total Thiol contents and Glutathione-S-transferase(GST) Activity

The Thiol groups are important members of the antioxidant team and have been shown to destroy ROS and other free radicals by enzymatic and non-enzymatic mechanism<sup>17</sup>. Total thiol groups of proteins are mainly responsible for their antioxidant response, and they can serve as a sensitive indicator of oxidative stress<sup>8</sup>. Total thiols are composed of both intracellular and extracellular thiols. Intracellular thiols such as glutathione and thioredoxin play an important role in maintaining the highly reduced environment inside the cell<sup>17</sup>. Extracellular thiols are protein bound and are mainly disulfide proteins due to the oxidative environment. Total thiol status in the body, especially thiol groups present on protein are considered as major plasma antioxidants *in vivo* and most of them are present over albumin, and they are the major reducing groups present in our body fluids<sup>18</sup>. Total thiol groups are very susceptible to oxidation and considered as one of the most important plasma sacrificial antioxidants. When the cells are exposed to oxidative stress, thiol groups are the first antioxidants that are consumed<sup>8</sup>. We found decreased plasma total thiol levels in oxidative or *nitro oxidative* condition. The total thiol contents of flies treated with *Mangifera indica* cold aqueous leaf extract for 5 days ranged from 9.25±1.8- 24±2.3 Mmol/mgprotein. The highest total thiol contents was recorded in the 5mg-treated flies while the lowest total thiol contents was recorded in the 10mg-treated flies. There was significant difference (p<0.05) comparing only 5 mg-treated flies to the control flies. The total thiol result is presented in figure 2. The GST activity of *Mangifera indica* leaf cold aqueous extract-treated flies ranged from 0.49±0.11- 1.08±0.27 µmol/min/mgprotein. The highest activity was recorded in 2.5 mg-treated flies while

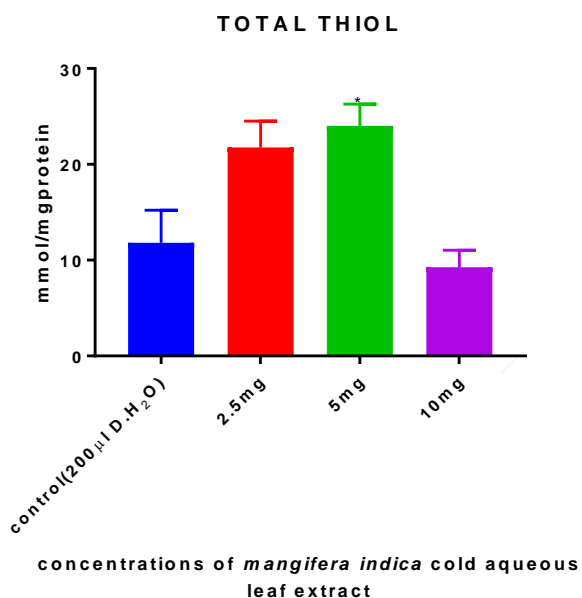
147 the lowest activity was recorded in the controlled flies. There was elevation of GST activities in  
148 all the extract-treated flies but no significant difference ( $p>0.05$ ) comparing to the controlled  
149 flies. The result is presented in figure 3. The result suggests that *Mangifera indica* leaf cold  
150 aqueous extract could improve the total thiol content and GST activity of flies.

151

152

153

154



155

156 Fig2: Total Thiol contents of *Mangifera indica* cold aqueous leaf extract-treated Flies

157

158

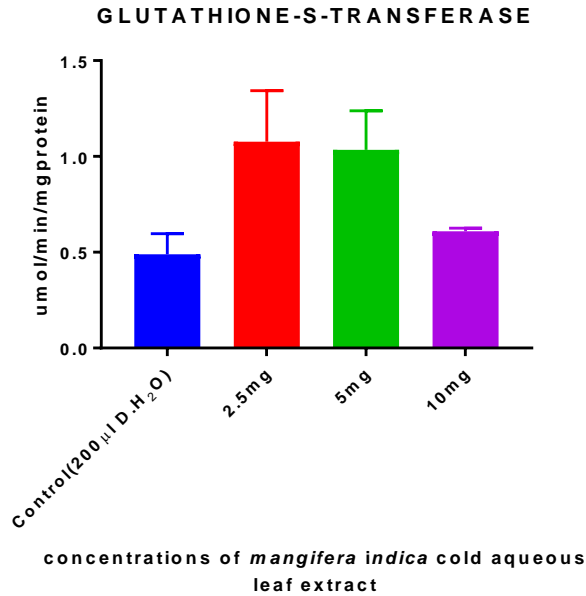
159

160

161

162

163



164  
165  
166  
167

Fig3: GST activity of *Mangifera indica* cold aqueous leaf extract-treated Flies

### 3.2 Catalase (CAT) activity

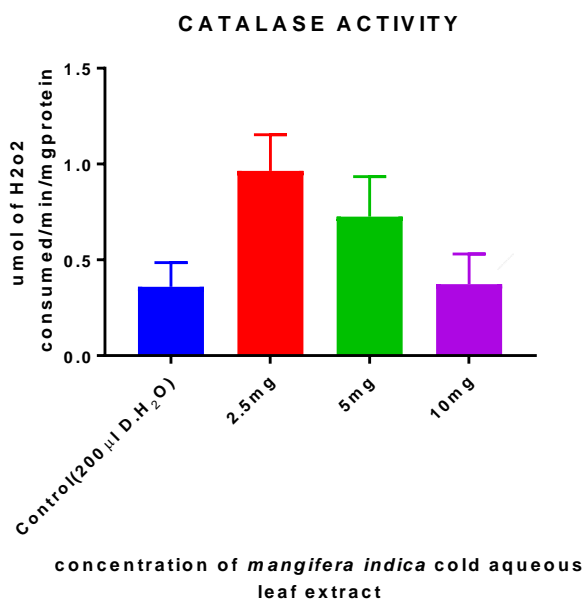
To scavenge ROS (Reactive Oxygen Species), SOD (superoxide dismutase) is the first and most important enzyme of the antioxidant system, catalyzing the dismutation of superoxide anions to hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and water. In the second step, CAT catalyzes the decomposition of H<sub>2</sub>O<sub>2</sub> to water and oxygen<sup>19</sup>

The CAT activity of *Mangifera indica* leaf cold aqueous extract-treated flies ranged from 0.36±0.13-0.97±0.19 µmol/min/mgprotein. The highest activity was recorded in 2.5 mg-treated flies while the lowest activity was recorded in the controlled flies. There was elevation of CAT activities in all the extract-treated flies but no significant difference (p>0.05) comparing to the controlled flies. The result is presented in figure 3. This result suggests that *Mangifera indica* leaf cold aqueous extract may improve the production of CAT to scavenge free radicals. Similar result was reported by Karuppanan *et al.*,<sup>20</sup> who evaluated the *in vivo* antioxidant properties of *Mangifera indica* leaf in mice. Their result showed similar high level of CAT activity against free radical. This high activity of catalase in both *Drosophila melanogaster* and Mice may indicate high antioxidant activity of *Mangifera indica* leaf extract. Zhang, *et al.*<sup>21</sup>, carried out a work on the effects of Rosemary extract on the lifespan and Antioxidant System of *Drosophila melanogaster*; their result showed that the CAT activity in each dosage group is higher than that of control group compared with control group. They concluded that a certain amount of extract of Rosemary can increase endogenous antioxidant activity(SOD, CAT) in *Drosophila*. Lu and Yeap Foo<sup>22</sup> studied *Salvia officinalis* (L.) for its antioxidant activity and polyphenol content and reported that rosmarinic acid and various catechols were responsible for the radical scavenging activity and caffeic acid was responsible for the xanthine oxidase (EC 1.17.3.2) inhibition. Zhao *et al.*<sup>23</sup> investigated the

189

190 antioxidant activity of *Salvia miltiorrhiza* and *Panax notogensing* . The results showed that *Salvia*  
 191 *miltiorrhiza* had a higher reducing power and scavenging activities against free radicals, including  
 192 superoxide and hydroxyl radicals, although it showed weak hydrogen peroxide scavenging . Furthermore,  
 193 Javanmardi *et al.*<sup>24</sup> tested the Iranian *Ocimum sp.* accessions to determine the antioxidant activities and  
 194 total phenolic contents and demonstrated that the antioxidant activity increased in parallel with the  
 195 total phenolic content .Evaluation of the pomegranate peel extracts to discover its antioxidant and  
 196 antimutagenic activities using different solvents such as ethyl acetate, acetone, methanol and water has  
 197 been carried out <sup>24</sup> . The results showed the highest anti-mutagenic and the lowest antioxidant activity  
 198 in the water extract. Catalase activity result of *Mangifera indica* cold aqueous leaf extract-treated flies is  
 199 presented in figure 4.

200  
 201



202  
 203 Fig 4: CAT activity of *Mangifera indica* cold aqueous leaf extract-treated Flies

204  
 205

## 206 Conclusion and Recommendations

207

### 208 Conclusion

209 From the results, 2.5 mg/ 10 g diet and 5 mg/10 g diet of *Mangifera indica* aqueous leaf  
 210 extract-treated flies showed low % mortality and high level of total thiol content, GST activity and  
 211 Catalase activity compared to the control and this may be due to its scavenging power. Therefore, it can  
 212 be concluded that *Mangifera indica* cold aqueous leaf extract at these concentrations may have high  
 213 activity against free radicals in *D. melanogaster*.

214

### 215 Recommendation

216 **Characterization** and **In vivo** antioxidant screening of *Mangifera indica* cold aqueous leaf extract  
 217 Fractions should be carried out in *Drosophila melanogaster* to determine the phyto-chemicals and  
 218 bioactive components of the extract responsible for its antioxidant activity.



219  
220  
221

Reference

- 222 1. A. Jayachitra and N. Krithiga, "Study on antioxidant property in selected medicinal plant extract,"  
223 International Journal of Medicinal and Aromatic Plants, vol. 2, no. 3, pp. 495-500, 2010.
- 224 2. N. A. Khalaf, A. K. Shakya, A. Al-Othman, Z. El-Agbar, and H. Farah, "Antioxidant activity of some  
225 common plants," Turkish Journal of Biology, vol. 32, no. 1, pp. 51-55, 2008.
- 226 3. V. R. Patel, P. R. Patel, and S. S. Kajal, "Antioxidant activity of some selected medicinal plants in  
227 western region India," Advances in Biological Research, vol. 4, pp. 23-26, 2010.
- 228 4. F. Shahidi and P. K. Wanasundara, "Phenolic antioxidants," Critical reviews in food science and  
229 nutrition, vol. 32, no. 1, pp. 67-103, 1992.
- 230 5. M. E. Büyükokuroğlu, I. Gülçin, M. Oktay, and Ö. I. Küfrevioğlu, "In vitro antioxidant properties of  
231 dantrolene sodium," Pharmacological Research, vol. 44, no. 6, pp. 491-494, 2001.
- 232 6. H. Sies, Oxidative stress: from basic research to clinical application. Am J Med 1911; 91: 31-9.
- 233 7. D.P.Jones, J.L. Carlson, V.C., Mody, et al. Redox state of glutathione in human plasma. Free  
234 Radic Biol Med 2000; 28: 625-35.
- 235 8. B. Halliwell, J.M. Gutteridge, The antioxidants of human extracellular fluids. Arch Biochem Biophys  
236 1990; 280: 1-8
- 237 9. R.N., Tharanathan, H.M. Yashoda & T.N. Prabha, (2006); Mango (*Mangifera indica* L.), the  
238 king of fruits - A review. Food Reviews International. 22:95-123.
- 239 10. M.H.A, Jahurul, J.S.M, Zaidul, K., Ghafoor, F.A., Al-Juhaimi, K.L., Nyam, N.A.N., Norulaini, F.,  
240 Sahena & A.K.M. Omar (2015); Mango (*Mangifera indica* L.) by-products and their valuable  
241 components: A review. Food Chemistry 183:173-180.
- 242 11. Ethno medicinal use of mango. <http://naturalhomeremedies.co/Mango.html>. 31 March,  
243 2016.
- 244 12. A. J. Nunez-Selles, Antioxidant Therapy; Myth or Reality? J. Braz. Chem. Soc., 16(4), 101-108(2005).
- 245 13. R.E. Kohler, Lords of the Fly: Drosophila Genetics and the Experimental Life. University of Chicago  
246 Press; Chicago, IL, USA: 1994. p. xv.321p
- 247 14. G.L. Ellman, Tissue sulfhydryl groups, Arch. Biochem. Biophys. 82 (1) (1959) 70-77,  
248 [http://dx.doi.org/10.1016/0003-9861](http://dx.doi.org/10.1016/0003-9861(59)90090-6) (59)90090-6 13650640.

249 15. W.H. Habig, W.B. Jakoby, Assays for differentiation of glutathione S-transferases, *Methods Enzymol.*  
250 77 (1981) 398-405 7329316.

251 16. H. Aebi, Catalase in vitro, *Methods Enzymol.* 105 (1984) 121-126 6727660

252 17. D.P., Jones, J.L., Carlson, V.C., Mody, *et al.* Redox state of glutathione in human plasma. *Free*  
253 *Radical Biological Medical* 2000; 28: 625-35.

254 18. F.Q., Schafer GR. Buettner Redox environment of the cell as viewed through the redox state of the  
255 glutathione disulfide/glutathione couple. *Free Radica Biological Medical* 2001; 30: 1191-212.

256 19. P., Chelikani, I. Fita and P.C. Loewen (2004). Diversity of structures and properties among catalases.  
257 *Cell Molecular Life Science.* 61(2), 192-208

258 20. M., Karuppanan, M., Krishnan, P., Padarathi & E.Namasivayam, (2014). Hepatoprotective and  
259 Antioxidant Effect of *Mangifera Indica* Leaf Extracts against Mercuric Chloride-induced Liver Toxicity in  
260 Mice. *Euroasian journal of hepato-gastroenterology*, 4(1), 18.

261 21. Z. S., ZHANG, S. P., WEN, H., WANG, , C., SHAO & S. HU, (2012). Study on Antisenescence Potential of  
262 *Rosmarinus Officinalia* Extract. *Journal of Food Research and Development*, 3

263 22. Y., Lu & F.L. Yeap, Antioxidant activities of polyphenols from sage (*salvia officinalis*). *Food Chem.*  
264 2001, 75, 197-202

265 23. G.R Zhao, Z.J Xiang, , T.X. Ye, Y.J. Yuan, Z.X. Guo , Antioxidant activities of salvia miltiorrhiza and  
266 *panax notoginseng*. *Food Chem.* 2006, 99, 767-774.

267 24. J. Javanmardi, Sf, C. tushnof, E. Locke; J.M. Vivanco, Antioxidant activity and total phenolic content  
268 of iranian *ocimum accessions*. *Food Chem.* 2003, 83, 547-550.

269

270

271

272

273

274

275

