

## **Fertilization With Silicon in Sweet Pepper Improved Plants Grown Under Salt Stress**

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### **ABSTRACT**

**Aims:** The objective of this research was to investigate the effect of calcium silicate on gaseous exchanges and production factors in the sweet pepper, cultivated under conditions of soil salinity induced by potassium fertilization, in protected cultivation.

**Study design:** The experiment was arranged in a randomized complete block design in a 2 × 5 factorial scheme with five replications.

**Place and Duration of Study:** The experiment was conducted in the sector of Olericultura and Experimentation of the course of Agronomy from October 12, 2018 to February 2019.

**Methodology:** The experiment was arranged in a randomized complete block design in a 2 × 5 factorial scheme (two sources of correction: calcareous and silicon and five increasing rates of KCl equivalent to 150, 300, 450, 600 and 700 kg ha<sup>-1</sup> of K<sub>2</sub>O). Ten treatments with five replicates where each experimental unit consisted of a polyethylene pot, with a volume of 19 dm<sup>3</sup>. The electrical conductivity, the determination and quantification of silicon in soil and plant, liquid photosynthesis, stomatal conductance, intercellular CO<sub>2</sub> concentration, transpiration, water use efficiency and instantaneous carboxylation efficiency were analyzed.

**Results:** With the increase of K<sub>2</sub>O in the soil there was a tendency of reduction in liquid photosynthesis, transpiration, stomatal conductance, intercellular CO<sub>2</sub> concentration, water use efficiency and instantaneous carboxylation efficiency in the presence and absence of calcium silicate. Higher rates of K<sub>2</sub>O (300, 450 and 600 kg ha<sup>-1</sup>) reduced the length and yield of sweet pepper fruits in the presence and absence of calcium silicate. The rate of 150 Kg K<sub>2</sub>O favored the growth of sweet pepper plants in the presence of calcium silicate.

**Conclusion:** It is concluded from the research that the dose of 300 kg ha<sup>-1</sup> of K<sub>2</sub>O, in the presence of calcium silicate, provided the best results for the electrical conductivity of 2.76 dS m<sup>-1</sup>, silicon content in the soil of 5.70 mg kg<sup>-1</sup>, 14% silicon leaf content, improving photosynthetic rates, transpiration, water use efficiency and fruit production. The increase in salinity reduced fruit yield, in the presence and absence of Si.

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**Keywords:** *Abiotic stress; photosynthesis; Capsicum annuum; salinization; calcium silicate.*

### **1. INTRODUCTION**

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Brazil is among the main sweet pepper producing countries. The main sweet pepper producing states in Brazil are Minas Gerais, São Paulo, Ceará, Rio de Janeiro, Espírito Santo and Pernambuco (87% of the total). It is possible to produce sweet peppers all year round, but it develops better in the summer. Currently, sweet pepper producers

26 have preferred to cultivate this crop in a protected environment, which allows a  
27 continuous supply and harvesting in periods of low supply of the product in the market,  
28 thus achieving more competitive prices [1].

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30 In the State of São Paulo, in 2018, about 65.800 tons of sweet pepper were produced in  
31 2.560 ha [2]. In the production of vegetables in protected cultivation, it was verified that,  
32 after three years of cultivation, many producers do not obtain adequate productivities  
33 and quality of the fruits, because there are many problems related to excessive  
34 fertilization, leading the soil to an accumulation of salts. The losses suffered by the  
35 producers are generated by improper practices of the incorrect management of the  
36 fertilization in greenhouse [3]. Therefore, the symptoms of these anomalies in plants  
37 under conditions of nutritional imbalance are common, due to the saline stress of the soil  
38 solution. Although irrigation water in protected crops is of good quality, the addition of  
39 fertilizers, when using the fertigation technique, makes it saline, increasing the risk of soil  
40 salinization [4].

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42 Potassium (K) is a nutrient demanded in great quantity by the culture of the sweet  
43 pepper, being the main source used by the producers is potassium chloride that has high  
44 saline index, being one of the main sources of salinization of the soil in cultivation.  
45 Potassium sulfate has a salt content equivalent to half of the salt content of potassium  
46 chloride, which makes it more suitable for soils with tendency to salinization [5].

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48 The exogenous application of silicon (Si) significantly improves the development of  
49 plants under conditions of salt stress [6]. Calcium silicate can be used as a corrective of  
50 soil acidity, neutralizing exchangeable aluminum, providing nutrients to the plant and  
51 increasing soil base saturation [7]. When saline stress occurs, there is a decrease in the  
52 relative water content in the leaf, indicating that the plants are exposed to osmotic stress  
53 [8]. Studies have shown that Si increases the relative water content in plants under  
54 conditions of salt stress [9], decreasing the toxicity of the salts to the plant and improving  
55 its growth, increasing the thickness of the leaves, due to deposition of Si, which reduces  
56 transpiration and decreases water loss [10].

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58 Due to the condition of soil salinization, nutritional disorders may occur, inducing  
59 antagonistic relationships between nutrients in the plant, which significantly reduces crop  
60 yields [11]. Elevation of K content in soil can induce nutritional imbalance for plants [12].  
61 However, it is necessary to know the effects of the interactions between saline stress  
62 and the use of silicon in the culture of sweet pepper that has been cultivated in protected  
63 culture.

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65 Therefore, the present research was conducted to evaluate the effect of calcium silicate  
66 on gaseous exchanges and production factors in the sweet pepper under conditions of  
67 soil salinity induced by potassium fertilization, in protected cultivation.

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## 69 **2. MATERIAL AND METHODS**

70

71 The experiment was conducted in the sector of Olericultura and Experimentation of the  
72 course of Agronomy from October 12, 2018 to February 2019 in greenhouse. A  
73 protective structure model was used, with 225 meters each (9 meters wide by 25 meters  
74 long) and right foot of 4 meters. The structure was covered with agrofilm, of blue color.  
75 The sweet pepper cultivar Magali R. was used. The seedlings were produced in trays  
76 with 128 cells, 6.0 to 6.2 cm high, with substrate composed of inert material and free of  
77 pathogens. Transplanting was carried out on November 20, 2018 using a seedling per  
78 pot, when they had three to four definitive leaves, which occurred around 35 days after  
79 sowing.

80

81 The experiment was arranged in a randomized complete block design in a 2 × 5 factorial  
 82 scheme (two sources of correction: calcareous and silicon and five increasing rates of  
 83 KCl equivalent to 150, 300, 450, 600 and 700 kg ha<sup>-1</sup> of K<sub>2</sub>O. It was applied 1.62 ha<sup>-1</sup> Mg  
 84 of calcareous with 80 % total neutralizing power (45 % CaO and 10% MgO)  
 85 corresponding to 15.39 g pot and 1.87 Mg ha<sup>-1</sup> of calcium silicate with total neutralizing  
 86 power 86% (40.7% SiO<sub>2</sub> and 10% CaO) corresponding to 17.85 g by pot, the source  
 87 CaSiO<sub>3</sub> used was reagent pure for analysis. Whose treatments and potency  
 88 equivalence are described in Table 1. Each experimental unit consisted of a 19 dm<sup>-3</sup>  
 89 polyethylene pot filled with Oxisol [13], after incubation of calcareous and calcium  
 90 silicate, fertilization was per-formed for the macro and micronutrients following the  
 91 recommendation of [14] and [15] adapted for experiments conducted in pots and for the  
 92 corn crop.

93  
 94 The soil was classified as Oxisol [16] and samples were collected at a depth of 0-20 cm.  
 95 The samples were placed to dry, crushed through a 5 mm sieve and mixed to describe  
 96 the chemical and physical compositions. Chemical and physical compositions of the soil  
 97 used in this study, according to [17], were: pH in water (1:2.5)= 5.2; level of organic  
 98 matter (OM)= 1.42 (dag kg<sup>-1</sup>); P and K by Mehlich I extraction = 3.69 and 30.41 (mg dm<sup>-3</sup>);  
 99 Mg, Ca and Al extractable by 1 M KCl solution= 7.59, 1.12 and 0.20 (cmol dm<sup>-3</sup>); Si=  
 100 3.29 (mg dm<sup>-3</sup>); Zn= 1.05 (mg dm<sup>-3</sup>); Cu= 1.38 (mg dm<sup>-3</sup>); S= 13.24 (mg dm<sup>-3</sup>); B= 0.07  
 101 (mg dm<sup>-3</sup>); Fe= 53.62 (mg dm<sup>-3</sup>); T = cation exchange capacity at pH 7.0 (3.62 %); t=  
 102 cation exchange capacity effective (5.02 %); m = aluminum saturation index (12.50 %); V  
 103 = Base saturation index (27.85 %). Soil granulometry was the soil physical composition  
 104 used in this study, determined by the pipette method (sand, silt and Clay = 60 %, 11 %  
 105 and 29 %). After incubation of calcareous and calcium silicate, fertilization was  
 106 performed for macro and micro-nutrients following the recommendation of [14] and [15]  
 107 adapted for experiments conducted in pots for sweet pepper crops. The soil chemical  
 108 analysis was done at the soil science laboratory of the Federal University of Lavras,  
 109 Brazil. The pots had holes in the bottom where a layer of 0.30 m of folded sombrite was  
 110 placed to avoid soil loss and to allow drainage of excess water, if it occurred.

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 112 Before the transplanting of the crop, 300 mg dm<sup>-3</sup> of urea (45 % N), 300 mg dm<sup>-3</sup>  
 113 of simple superphosphate (18 % P<sub>2</sub>O<sub>5</sub>) was applied and incorporated into the soil, pure  
 114 reagent was used for analysis for both fertilizers. The calculations for soil correction were  
 115 based on recommendations [18]. For N, the equivalent of 12.22 g of urea per pot was  
 116 divided into three applications and, for P<sub>2</sub>O<sub>5</sub>, 72.52 g of simple superphosphate per pot  
 117 applied at planting was used. Coating fertilizations started at 15 days after transplant  
 118 (DAT) and were performed biweekly. The basic fertilization for K<sub>2</sub>O was made with KCl  
 119 using pure reagent source for analysis (60 % K<sub>2</sub>O), as described in Table 1. After the  
 120 application of the fertilizer, the soil was moistened for 35 days to favor the chemical  
 121 reaction of the corrective and fertilizer. The pots were distributed at spacing of 0.63 m  
 122 between plants and 1.0 m between rows.

124 **Table 1. Treatments and equivalence in pots based on the two correctives**  
 125 **(calcium silicate and calcareous) and rates of K<sub>2</sub>O.**

Treatments	Corrective		K <sub>2</sub> O rates kg ha <sup>-1</sup> of K <sub>2</sub> O	
	T1	Calcium silicate	-	150
T2	Calcium silicate	-	300	-
T3	Calcium silicate	-	450	-
T4	Calcium silicate	-	600	-
T5	Calcium silicate	-	700	-
T6	-	Calcareous	-	150
T7	-	Calcareous	-	300
T8	-	Calcareous	-	450

T9	-	Calcareous	-	600
T10	-	Calcareous	-	700

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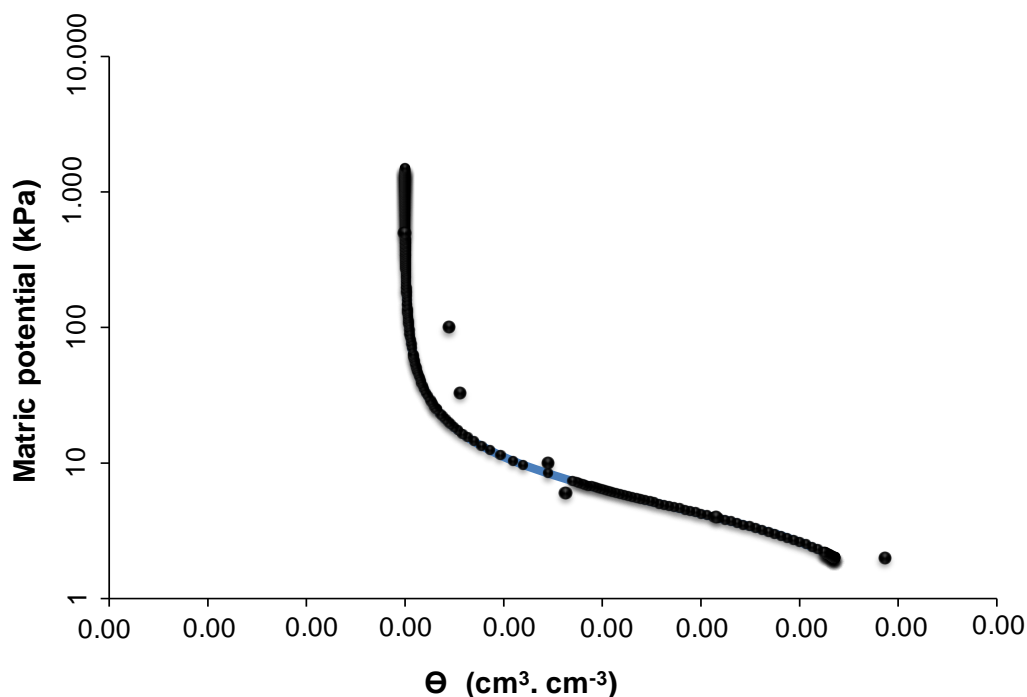
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The water characterization of the soil was determined by its water retention characteristic curve (Fig.1). The parameters of the soil water retention curve used in irrigation and irrigation management were obtained based on the model proposed by [18], with the aid of the Solver application of Microsoft Office Excel<sup>®</sup> software ( $\theta = 0.4215 \times [1 + (0.2040 \times |\Psi_m|)^{1.8757}]^{-0.4669} + 0.2670$ ), where:  $\theta$  = current moisture  $\text{cm}^3.\text{cm}^{-3}$  and  $\Psi_m$  = stress, kPa. The field capacity was estimated to be equivalent to the voltage and humidity at the inflection point of the retention curve, as proposed by [19]:  $\Psi_m = 1 / \alpha [1 / m]^{1/n}$ , where:  $\Psi_m$  = tension at the inflection point of the curve, kPa;  $\alpha$ ,  $m$  and  $n$  = adjustment parameters of the model equation proposed by [18]. The moisture value in the field capacity found was  $0.3458 \text{ cm}^3.\text{cm}^{-3}$  for a voltage of 4.25. Soil moisture was determined through tensiometers, using the water potential of -35 kPa, considered as adequate for the development of the crop [20].

The irrigation was done by drip irrigation; the self-compensating emitters being manually inserted in polyethylene hoses. The calculation of the operating time of the irrigation system was made based on the humidity sensors (tensiometers) installed in the depth of 0.15 m. With the observed stresses, the corresponding moisture values were estimated from the water retention curve in the soil.

With these moistures and the one corresponding to -30 kPa [15] and, considering the effective depth of the root system (0.15 m), the net and gross replacement slides were calculated for the treatments. Aiming at the replacement of soil water, two readings were performed daily in the tensiometers, one in the morning (8:00 am) and one in the afternoon (14:00 pm).



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**Fig. 1. Water retention characteristic curve of the Oxisol used in the experiment (-  $\Psi_m$ = matric potential).**

158 At the end of the experiment (120 days after plant transplantation), the electrical  
159 conductivity (EC) was determined in the saturated paste extract [21], which is the  
160 method used as reference for EC determination and adopted in various regions of the  
161 world. To do so, the soil passed through the 2 mm sieve and allowed to stand for 24 h to  
162 air dry. Afterwards, 800 g of soil were added in plastic containers, with capacity for 1200  
163 mL, with 500 mL of distilled water added. After the mixture turned into a paste, the  
164 container was covered with foil remaining for 24 h. After this time, the slurry was again  
165 stirred, standing for 1 h. By means of the vacuum filtration of the saturation paste, the  
166 solution of the soil was extracted, after which the EC reading was measured. The  
167 electrical conductivity of the saturated pulp was corrected considering the soil water  
168 retention characteristic using a digital conductivity meter (Lutron, model CD-4303).

169

170 For the quantification of the silicon in the soil, soil samples were taken from the pots  
171 grown with sweet pepper and prepared for analysis. The samples were dried at room  
172 temperature (TFSA) and subsequently sieved (<2,0 mm). The extraction procedure was  
173 performed to maintain the same soil: solution ratio, that is, for each 10 g of soil, 100 mL  
174 of extractor was added. The extractors used were: Acetic acid 0,5 mol L<sup>-1</sup> [22]: 100 mL of  
175 0.5 mol L<sup>-1</sup> acetic acid was added to a 150 mL plastic flask containing 10 g de soil. The  
176 plastic bottle was capped and shaken horizontally for one hour. After 30 minutes, the  
177 extract was filtered (plastic funnel), using filter paper number 42; Buffer pH 4.0: 100 mL  
178 of a buffered solution at pH 4.0 acetic acid plus sodium acetate (49.2 mL of concentrated  
179 acetic acid and 14.800 g of anhydrous sodium acetate were dissolved in 1,0 liter of  
180 distilled water, and the pH adjusted to 4.0 with the addition of acetic acid) were added in  
181 a 150 mL plastic flask with 10 g soil and shaken horizontally for one hour. The vials were  
182 then held for 30 minutes and then the plastic funnel extract and filter paper number 42  
183 filtered; Calcium chloride 0.0025 mol L<sup>-1</sup> [23]: 100 mL of a 0.0025 mol L<sup>-1</sup> calcium  
184 chloride solution was added in a plastic flask containing 10 g of soil. Thereafter, it was  
185 shaken horizontally for 15 minutes and then decanted from overnight. The following day,  
186 the extracts were filtered (plastic funnel and filter paper number 42); Water: 100 mL of  
187 distilled and demineralized water were added in 150 mL plastic bottles with 10 g of soil.  
188 Henceforth, the procedure was the same as for acetic acid.

189

190 The determination of Si in the extract was made by mixing 10 mL of the extract (filtrate /  
191 decanting) in 1 mL of sulfo-molybdenum 7.5% solution (7.5 g ammonium molybdate in  
192 10 mL + ac. sulfuric 9 mol L<sup>-1</sup> in 100 mL). After 10 minutes 2 mL of the 20% tartaric acid  
193 solution was added and after 5 minutes 10 mL of the 0.3% ascorbic acid solution was  
194 added. After one hour, the Si was read in a spectrophotometer and at the wavelength of  
195 660nm. The quantification of silicon in the leaves was performed by the colorimetric  
196 method of molybdenum blue in the laboratory of mineral nutrition of plants in the  
197 Laboratory of Mineral Nutrition of Plants of the Federal University of Uberlandia, Brazil  
198 [24].

199

200 The shoot dry matter (leaves + stem) was collected at 120 days after transplanting  
201 (DAT), to determine dry shoot mass (MMSPA). To dry the material an oven was used at  
202 70 °C with forced ventilation until constant mass was reached. The shoot + stem were  
203 processed together. The heights of the plants (m) were evaluated with the help of a  
204 scale, measuring the distance between the base of the plant collar to the end of the main  
205 stem, the production, which was determined throughout the reproductive stage of the  
206 plants, and also the diameter, length, weight and diameter of commercial fruits.

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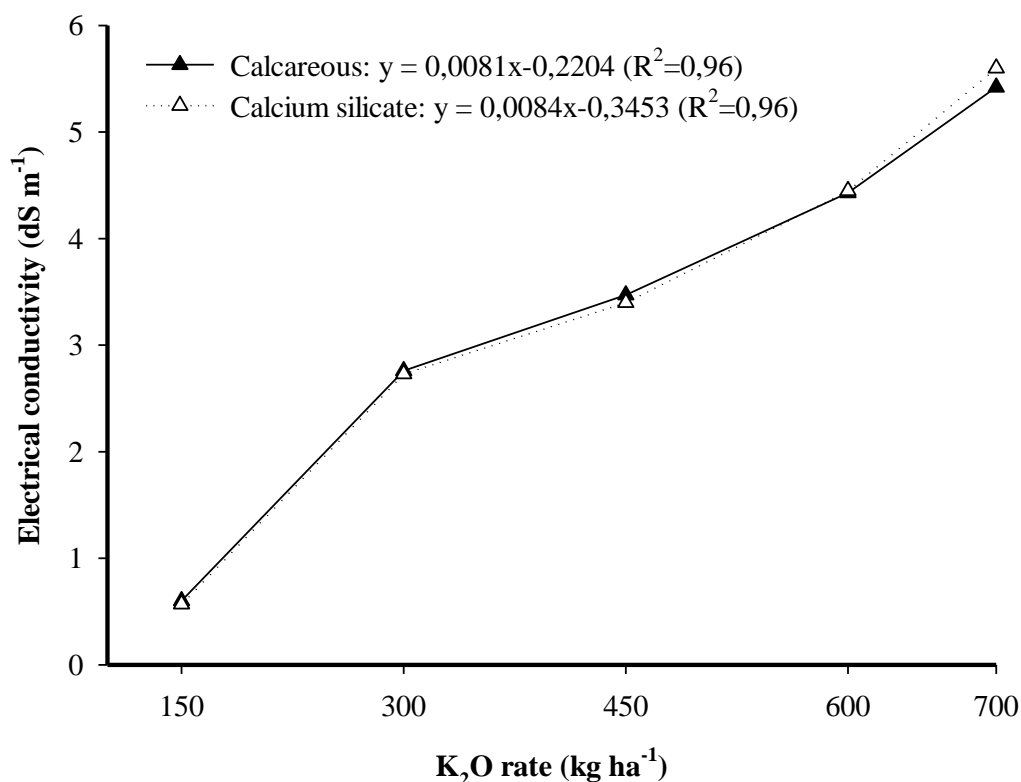
208 For the analysis of liquid photosynthesis, stomatal conductance, intercellular CO<sub>2</sub>  
209 concentration, transpiration, water use efficiency and instantaneous carboxylation  
210 efficiency, the IRGA model LI-6400XT, (Li-Cor, Lincoln, Nebraska, USA) was used. Two  
211 plants of each cultivar were chosen randomly, being defined as the sample unit the sixth  
212 leaf from top to bottom, fully expanded and mature. Because it is a species with a  
213 composite leaf, the first three leaflets of each leaf were used to measure, totaling six  
214 measurements. The value of 850 μmol m<sup>-2</sup> s<sup>-1</sup> of saturation irradiance, defined by the  
215 realization of a light curve, was set using the value of radiation that induced the

216 maximum photosynthesis. Sweet pepper is a C3 plant, where a cyclic mechanism of  
217 enzymatic reactions converts CO<sub>2</sub> into carbohydrates through the reductive  
218 photosynthetic cycle (C3), generating the 3 phosphoglycerate. Therefore, IRGA camera  
219 temperature was controlled at 28 °C, since in C3 plants the maximum rate of  
220 photosynthesis is reached at relatively low radiation intensity, causing no destruction or  
221 damage to the photosynthetic apparatus. Measurements were performed on a 6 cm<sup>2</sup>  
222 sheet area.

223  
224 The results found in the different evaluations were submitted to analysis of variance. For  
225 the evaluation of the means, the Scott-Knott or t-test were applied, according to the  
226 theories recommended by [25]. The standard deviations were calculated and the  
227 correlation estimators (Pearson or Spearman) were used, using SISVAR software [26].  
228

### 229 **3. RESULTS AND DISCUSSION**

230  
231 The electrical conductivity (EC) of the soil (Fig. 2) increased with increasing rates of K<sub>2</sub>O  
232 in both correctives (calcium silicate and calcareous). The EC of 2.76 and 2.16 dS m<sup>-1</sup>  
233 were the ones that provided the greatest vegetative development and production, these  
234 results agree with those found by [27], who studied the influence of EC on eggplant  
235 concluded that the EC of 2.36 dS m<sup>-1</sup> provided the greatest development and fruiting.  
236 The higher dry matter yield of roots, stems, leaves and fruits in eggplant plants was  
237 obtained with EC of nutrient solution of 2.10 dS m<sup>-1</sup> [28]. The use of a rate greater than  
238 60 kg ha<sup>-1</sup> of K<sub>2</sub>O may cause some damage to the legumes due to its saline effect, which  
239 may have occurred in this experiment with rates greater than 100 kg ha<sup>-1</sup> of K<sub>2</sub>O [29]. The  
240 electrical conductivity increased linearly with the increase of the KCl rate applied in two  
241 sources of potassium fertilization, due to the increase of the electrolytic concentration of  
242 the soil solution, which is proportional to the increase in the concentration of ions in the  
243 solution [30].  
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248  
249 **Fig. 2. Electrical conductivity of the soil as a function of the K<sub>2</sub>O rates and sources**  
250 **of correctives (calcium silicate and calcareous).**

251  
252 The concentration of Si in the soil did not vary in the different rates of K<sub>2</sub>O studied when  
253 calcium or calcium silicate was applied (Table 2). However, in the interaction between  
254 the rates of K<sub>2</sub>O x sources of correctives it was observed that the silicon concentration  
255 was higher for the treatment using calcium silicate, due to the fact that it is a soluble  
256 source of Si.

257  
258 **Table 2. Soil silicon content in CaCl<sub>2</sub> 0.01 mol L<sup>-1</sup> as a function of K<sub>2</sub>O rates and**  
259 **corrective sources (calcareous and calcium silicate).**

K <sub>2</sub> O rates (kg ha <sup>-1</sup> )	Calcium silicate	Calcareous
	Content tho Si (mg kg <sup>-1</sup> )	
150	6.00 Aa	5.00 Ab
300	5.70 Aa	5.00 Ab
450	5.80 Aa	4.80 Ab
600	5.75 Aa	5.00 Ab
700	6.00 Aa	5.20 Ab

261 *Capital letters equal in the column, do not differ at the level of significance of 5%; Minor letter in*  
262 *the same line, do not differ at the level of significance of 5%.*

263  
264 For the silicon content in the sweet pepper leaf (Table 3) differences were observed  
265 between the rates of K<sub>2</sub>O. When the calcium silicate was applied, the highest levels were  
266 found with 600 and 700 kg ha<sup>-1</sup> K<sub>2</sub>O. As for the interaction between the correctives  
267 (calcium silicate x calcareous), independent of the K<sub>2</sub>O rate, the higher silicon contents  
268 were found when calcium silicate was applied.

269

270 **Table 3. Silicon content in the leaf (%) as a function of K<sub>2</sub>O rates and corrective**  
 271 **sources (calcareous and calcium silicate).**  
 272

K <sub>2</sub> O rates (kg ha <sup>-1</sup> )	Calcium silicate Content tho Si (%)	Calcareous
150	13 Ca	12 Ab
300	14 Ca	12 Ab
450	18 Ba	13 Ab
600	20 Aa	13 Ab
700	20 Aa	14 Ab

273 *Capital letters equal in the column, do not differ at the level of significance of 5%; Minor letter in*  
 274 *the same line, do not differ at the level of significance of 5%.*

275  
 276 With increasing rates of K<sub>2</sub>O in the soil there was a tendency of reduction in the liquid  
 277 photosynthesis (total photosynthesis) (Fig. 3A), transpiration (Fig. 3B), stomatal  
 278 conductance (Fig. 3C), intercellular CO<sub>2</sub> concentration (Fig. 3D), water use efficiency  
 279 (Fig. 3E) and instantaneous carboxylation efficiency (Fig. 3F), in the presence and  
 280 absence of calcium silicate. However, it was observed that with the application of  
 281 calcium silicate all these variables presented higher values. The deposition of silicon in  
 282 plant tissues improves the interception of light and decreases transpiration [31].  
 283 Increased availability of Si favors increased productivity, since Si can act indirectly in  
 284 photosynthetic and biochemical processes, especially when the plant is subjected to  
 285 some type of stress [32]. The translocation of silicon from the roots to the aerial part of  
 286 plants may be related to the increase in photosynthetic capacity, greater resistance to  
 287 possible damage and reduction in the evapotranspiration process, which, consequently,  
 288 improves the use of available water in the soil [33].

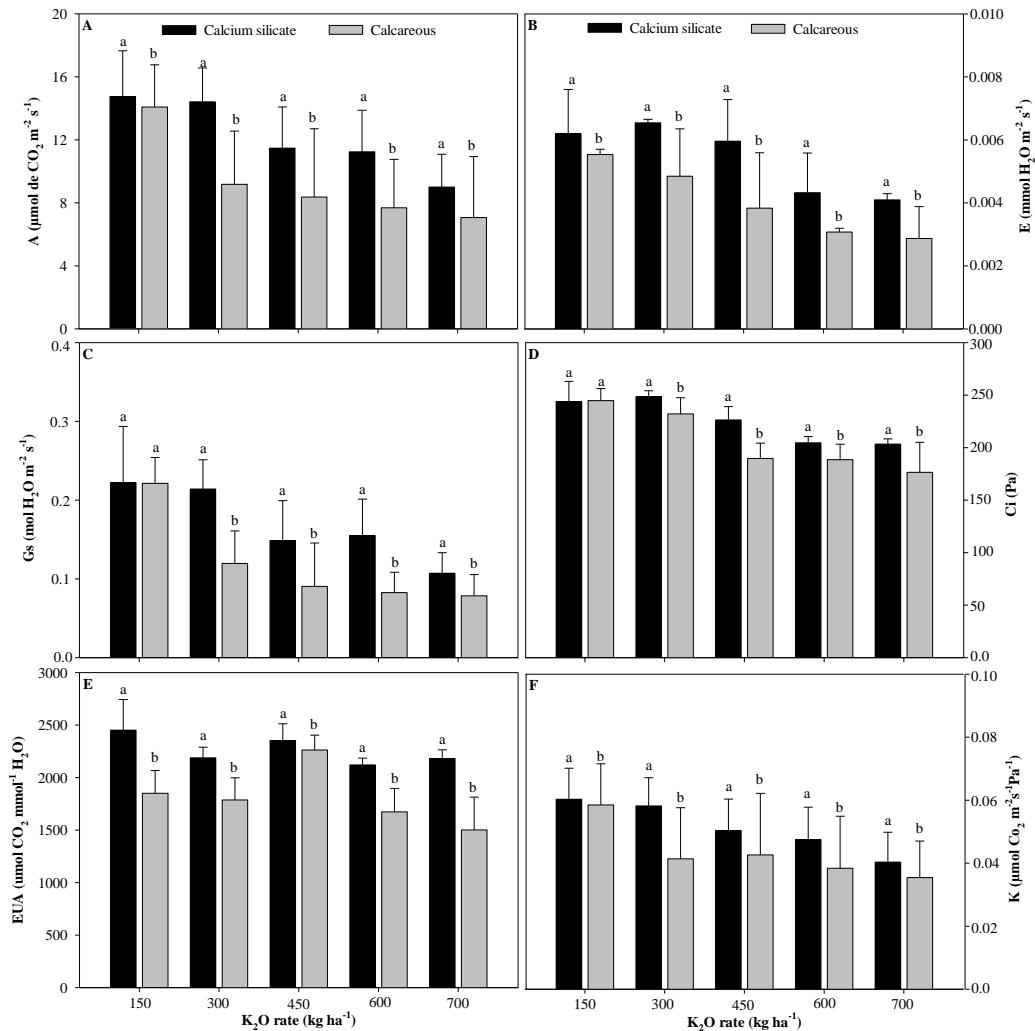
289  
 290 The increase in CO<sub>2</sub> concentration inside leaves promotes the closure of stomata, which  
 291 may occur in response to a biotic stress [34]. This CO<sub>2</sub> concentration may be directly  
 292 related to the increase in transpiration, which was greater than 0,006 mmol H<sub>2</sub>O m<sup>-2</sup>s<sup>-1</sup>  
 293 (Fig. 3B). The increase in transpiration by plants is mainly due to the inability of some  
 294 plants to absorb enough water to replenish that consumed in the transpiration process,  
 295 and the loss of water by plants is regulated by the activity of the guard cells. As  
 296 temperature rises, relative air humidity decreases and responses of metabolic processes  
 297 in plants will reflect the interaction between transpiration and guard cell activities [35].

298  
 299 The efficiency in the use of water by sweet pepper plants demonstrates a relationship  
 300 between photosynthesis and transpiration in which the observed values are directly  
 301 related to the amount of carbon that the plant fixes for each unit of water it loses [36]. In  
 302 this sense, decreases observed in water use efficiency (Fig. 3E) are reflective of  
 303 increases in the rate of carbon dioxide assimilation and transpiration of plants. As for the  
 304 instantaneous efficiency of carboxylation (Fig. 3F).

305  
 306 The results obtained in this work indicate that the increase in the instantaneous  
 307 efficiency of carboxylation is related to the increase in the concentration of CO<sub>2</sub> and to  
 308 the gains related to the rate of assimilation of CO<sub>2</sub> [37]. Point out that this efficiency is  
 309 related to the intercellular CO<sub>2</sub> concentration and the rate of assimilation of CO<sub>2</sub>. The  
 310 CO<sub>2</sub> assimilation from the external environment promotes water loss, which restricts CO<sub>2</sub>  
 311 entry [35]. The gas exchanges, are influenced by climatic conditions, so the reduction in  
 312 the efficiency of water use may be related to the increase of solar radiation, temperature  
 313 and relative humidity.

314  
 315 It is noteworthy that the stomatal behavior determines the transpiratory requirement of  
 316 the plants, thus controlling the loss of water in the form of vapor. Although Si is not  
 317 considered an essential element for plants, studies show that its application to the soil  
 318 contributes to the growth and increase of productivity [38], as can be observed in this  
 319 work (Table 3). In saline stress conditions, the plant growth is compromised due to the

320 reduction of the osmotic potential of the soil solution, which reduces the water potential of  
 321 the plants [39]. According to [40], this reduction of the water potential of the plants can  
 322 be mitigated by the application of Si, which reduces the toxicity caused by excess  
 323 sodium chloride in the soil solution.  
 324



325 **Fig. 3. Liquid photosynthesis (A), transpiration (B), stomatal conductance (C),**  
 326 **intercellular CO<sub>2</sub> concentration (D), water use efficiency (E) and instantaneous**  
 327 **efficiency of carboxylation (F) as a function of presence and absence of calcium**  
 328 **silicate and rate of K<sub>2</sub>O.**  
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330  
 331 The production and weight of sweet pepper fruits were higher when the 150 kg K<sub>2</sub>O rate was applied in the presence and absence of calcium silicate (Fig. 4A and 4B). Higher  
 332 rates of K<sub>2</sub>O reduced sweet pepper production (Fig. 4A) and caused a significant  
 333 decrease in plant height (Fig. 4E). There was a reduction in the length of the chili fruits  
 334 when the K<sub>2</sub>O rates increased, in the presence and absence of calcium silicate (Fig. 4C).  
 335 The application of calcium silicate favored the increase of the diameter of the fruits in the  
 336 rates of K<sub>2</sub>O studied (Fig. 4D). The beneficial effects of Si on the growth have been  
 337 reported in a wide of plant species, which are characterized by protecting the plant from  
 338 various biotic and a biotic stresses [41]. Transporters responsible for Si unloading from  
 339 xylem in leaves also have been identified in many plant species [42]. The aerial plant  
 340 parts accumulate more Si than roots [43]. Deposition of Si takes place in different parts  
 341 of plant such as epidermis of shoots but can also occur in the cell wall of root  
 342

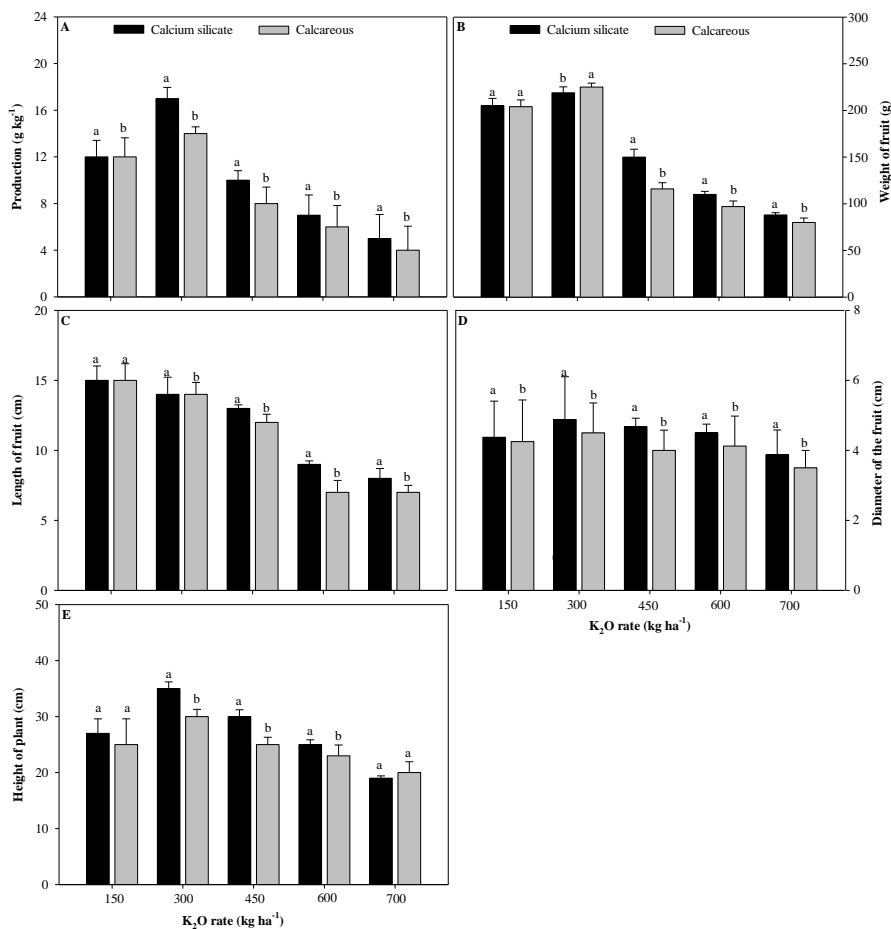
343 endodermis [10]. However, phytoliths formation, composition, and localization vary  
 344 among plant species [44].

345

346 The rate of 150 Kg K<sub>2</sub>O favored the growth of sweet pepper plants in the presence of  
 347 calcium silicate. In Fig. 4C it is observed that, as increasing rates of K<sub>2</sub>O were applied,  
 348 there was reduction in fruit length. Under conditions of higher salinity and osmotic  
 349 pressure of the soil solution the absorption of water from the root cells decreases,  
 350 allowing the occurrence of ionic toxicity. The addition of 16.6 g KCl m<sup>-2</sup> reduced root yield  
 351 and P uptake by sweet pepper plants cultivated on an Oxisol with 24.0 g dm<sup>-3</sup> of organic  
 352 matter [3] in addition, [8] reported that high salinity promotes changes in photosynthesis  
 353 (CO<sub>2</sub> assimilation, stomatal conductance and leaf transpiration), thus inhibiting plant  
 354 growth and reducing its height, as shown in Fig. 4E.

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300



356

357 **Fig. 4. Production (A), fruit weight (B), fruit length (C), fruit diameter (D) and plant**  
 358 **height (E) as a function of the presence and absence of calcium silicate and K<sub>2</sub>O**  
 359 **rates.**

360

#### 361 4. CONCLUSION

362

363 It is concluded from the research that the dose of 300 kg ha<sup>-1</sup> of K<sub>2</sub>O, in the presence of  
 364 calcium silicate, provided the best results for the electrical conductivity of 2.76 dS m<sup>-1</sup>,  
 365 silicon content in the soil of 5.70 mg kg<sup>-1</sup>, 14% silicon leaf content, improving  
 366 photosynthetic rates, transpiration, water use efficiency and fruit production. The  
 367 increase in salinity reduced fruit yield, in the presence and absence of Si.

368

## 369 **COMPETING INTERESTS**

370

371 We declared that no competing interests exist.

372

## 373 **CONSENT**

374

375 It is not applicable.

376

## 377 **ETHICAL APPROVAL**

378

379 It is not applicable.

380

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