

**Antidiarrhoeal effects of hydromethanolic
leaves extract of *Ipomea asarifolia* in albino rat
model**

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ABSTRACT

Aim: To evaluate the antidiarrhoea effect of hydromethanolic leave extract of *I. asarifolia* (HLEIA) on castor oil-induced diarrhoea

Place and Duration of Study: Department of Biochemistry, Faculty of Life sciences, Kebbi State University of Science and Technology, Aliero, Kebbi state, Nigeria. P.M.B.1144. Kebbi State. Nigeria, between February 2015 and September 2016.

Methodology: In a continuous effort to search for bioactive agents from medicinal plants, the antidiarrhoea activity of *I. asarifolia* was investigated. The effect of hydromethanolic leave extract of *I. asarifolia* (HLEIA) on castor oil-induced diarrhoea, gastrointestinal transit and intestinal fluid accumulation (enteropooling) were assessed in albino rats. Qualitative phytochemical analysis was carried out using standard procedures while acute oral toxicity studies was determined using the staircase method.

Results: The phytochemical analysis showed the presence of alkaloid, terpenoid, tannin, saponin, phenols. The LD₅₀ was estimated to be greater than 3000mg/kg since there was no mortality recorded after 14 days of acute oral toxicity studies. Sub-chronic administration of graded doses (150 – 600mg/kg) of HLEIA significantly (p<0.05) reduced diarrhoea episodes, decreased gastro intestinal movement and inhibited intestinal fluid accumulation compared to the control. The antidiarrhoea effect of treated group (600mg/kg) was comparable to that of the standard drug Loperamide

Conclusion: The findings of the present study scientifically validate the use of *I. asarifolia* in the treatment of diarrhoea.

Keywords: Gastro-intestinal transit, Castor oil, enteropooling, loperamide, diarrhoea episodes.

1. INTRODUCTION

The use of plants for medicinal purposes is an age old tradition in Africa, Asia and Latin America [1, 2]. Medicinal plants are plants containing inherent active ingredients used to cure disease or relieve pain [3]. The striking coincidence between indigenous medicinal plants uses and scientifically-proved phytochemical and pharmacological properties shows that the traditional remedies are an important and effective part of indigenous healthcare systems which is totally dependent on traditional healers [4]. Growing interest on the use of medicinal plants for primary health care is greatly influenced by the rising cost and side effects associated with most modern drugs. Modern pharmacopoeia still contains at least 25% of drugs derived from plants and many others, which are synthetic analogues, built on prototype compounds isolated from plants [5].

29 *Ipomoea asarifolia* (Convolvulaceae) is a glabrous succulent perennial plant trailing on the
30 ground. It is found throughout West Africa and is a common weed of hydromorphic soils, low
31 lying and inland valleys, streams and river banks. In Nigeria, the traditional names include
32 "Duman kada" in Hausa and "Gboro ayaba" in Yoruba [6]. Various parts of the plant are used
33 by traditional medicine practitioners in Nigeria for the management and treatment of several
34 disorders which include ophthalmia, neuralgia, headache, arthritic pains and stomach ache. In
35 Kebbi (North- West Nigeria), *Ipomoea asarifolia* has been widely used for the treatment of
36 various stomach disorders including diarrhoea.

37 Diarrhoea is a leading cause of malnutrition and globally, there are nearly 1.7 billion cases of
38 childhood diarrhoeal disease every year [7]. It is a very common ailment and national
39 problem in many tropical countries and the cause of 4-5 million deaths throughout the world
40 annually [8]. Diarrhoea remains the second leading cause of death among children under
41 five globally [9]. Nigeria was estimated to have a total number of annual child deaths due to
42 diarrhoea to be 151,700 [9]. Diarrhoea may be caused by a wide array of agents such as
43 entero-pathogenic microorganisms (*Shigella flexneri* and *Shigella dysenteriae*,
44 *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi* and *Candida albicans*), alcohol,
45 irritable bowel syndrome, bile salts, hormones, secretory tumors and intoxication [10,11].
46 Dependency on plants as medicine in controlling diseases is common among rural populace
47 in Nigeria because of its relative safety and affordability compared with the cost of
48 conventional medicines. Therefore, there is need to provide scientific bases of justification
49 on the therapeutic uses of medicinal plants against infectious diseases. *Ipomoea asarifolia*
50 has been used in traditional medicine for treating various ailments, including diarrheal,
51 without scientific verification of its effects. The present study was therefore designed to
52 validate this claim of *Ipomoea asarifolia* in the treatment of diarrhoea by the communities in
53 Kebbi State, Northwest Nigeria.

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55 **2. MATERIAL AND METHODS**

56 **2.1 Plant collection**

57 The fresh leaves of *Ipomoea asarifolia* were collected in the month of March, 2015 at Kebbi
58 State University of Science and Technology, Aliero (KSUSTA) main campus. The plant was
59 identified taxonomically and authenticated at the Department of Biological Science, Kebbi
60 State University of Science and Technology Aliero, Nigeria with a voucher specimen no 001.

61 **2.2 Plant extraction**

62 The collected leaves of *I. asarifolia* were air-dried and then grounded into powder. 200g of
63 the powdered leave was macerated in 2000mL of methanol:water (70:30) for 72 hours,
64 filtered using muslin cloth and dried in an oven at 45 °C. The percentage yield of the
65 hydromethanolic extract of *I. asarifolia* was 32.95%

66 **2.3 Animals**

67 Albino rats were used for the study. They were purchased at the animal house of Usmanu
68 Danfodio University Sokoto, Sokoto State. All the animals were kept in the cage and allow
69 acclimatizing for one week in Biochemistry Laboratory of Kebbi State University of Science
70 and Technology Aliero, Kebbi State, before the experiment started. The animals were fed
71 with standard pellet diet and water. The container for the food and water were washed and
72 cleaned daily as food and water were renewed every day to ensure hygiene and maximum
73 comfort for the animal.

74 **2.4 Phytochemical screening**

75 The presence of various phytochemical constituents in the extract was determined using the
76 standard screening tests [12].

77 **2.5 Lethal dose determination (LD₅₀)**

78 The up and down procedure as described by Dixon [12] was used to evaluate the oral acute
79 toxicity of hydromethanolic leaves extract of *I. asarifolia*. Five non-pregnant adult albino rats
80 randomly selected from the pull of acclimatized rats were used for this experiment. The

81 animals were weighed, marked and individually housed in cages prior to treatment. The rats
82 to be treated were fasted overnight but allowed free access to water. Freshly prepared
83 hydromethanolic leaves extract of *I. asarifolia* was administered orally at a limited dose of
84 3000mg/kg. The first animal was dosed and observed for sign of toxicity such as
85 hyperactivity, hypoactivity, spasm, ruffled fur, emesis, inappetance, scratching of mouth part,
86 coma or death. If the animal survived, the same procedure was adopted until all the five rats
87 were dosed and observed for 48 hours for signs of acute toxicity, morbidity and mortality for
88 the first 48 hours and up to 14 days. The behavioral changes and other changes observed in
89 animals were recorded according to Organization for Economic and Cultural Development
90 (OECD) 425 guidelines [13].

91 2.6 Antidiarrhoea studies

92 2.6.1 Gastrointestinal motility test

93 Rats were fasted for 18 h and divided into five groups of five animals each. Group I received
94 5 mL/kg normal saline orally, group II received Loperimide (5 mg/kg), group III - V received
95 hydromethanolic leave extract of *Ipomea asarifolia* (150, 300 and 600 mg/kg) respectively.
96 After 1h of administration, 1mL of deactivated charcoal meal was administered to all the rats.
97 Thirty minutes later, each rat was sacrificed and the small intestine removed. The total
98 length of the intestine and the distance moved by the charcoal meal from the pylorus to the
99 caecum was measured (cm). The intestinal charcoal transit was expressed as a percentage
100 of the distance moved by charcoal to the length between pylorus and the caecum [14] and
101 was calculated according to the following formula:

$$\% \text{ Inhibition of intestinal transit} = \frac{\text{distance travelled by charcoal meal in control group} - \text{treated group}}{\text{Distance travelled by charcoal meal in control group}} \times 100$$

107 2.6.2 Castor oil induced diarrhea

108 Twenty rats were fasted for 18 h and divided into five groups of five animals each. Castor oil
109 (1 mL) was orally given to all groups of animals for the induction of diarrhoea. Thirty minutes
110 after castor oil administration various treatments were given. Group I (control) animals were
111 treated with normal saline (5 mL/kg), Group II animals were treated with Loperamide (5
112 mg/kg), a positive control. Group III-V was treated with hydromethanolic extract of *Ipomea*
113 *asarifolia* (150, 300 and 600 mg/kg) orally administered. Animals were placed separately in
114 individual cages lined with filter paper. The filter papers were changed every hour and the
115 severity of diarrhoea was assessed hourly for 6 hours [15]. The total score of diarrhoea
116 faeces for the control group was considered as 100% and percentage inhibition of diarrhoea
117 was calculated using the following formula:

$$\% \text{ Inhibition of diarrhoea} = \frac{\text{Total no. of diarrheal faeces in control group} - \text{Total no. of diarrheal faeces in treated group}}{\text{Total no. of diarrheal faeces in control group}} \times 100$$

122 2.6.3 Castor oil induced enteropooling

123 Intraluminal fluid accumulation was determined by the method described by Robert *et al* [16].
124 Rats were divided into five groups of five animals each. One hour before oral administration
125 of castor oil (2 mL/rat.), group I orally received normal saline (5 mg/kg) and served as
126 control, group II animals received Loperamide (5 mg/kg) while groups III – V through oral
127 intubation, respectively received the plant extract at 150, 300 and 600 mg/kg body weight
128 respectively. Two hours later, the rats were sacrificed and the small intestine from the
129 pylorus to the caecum was isolated. The intestinal contents were collected by milking into a
130 graduated tube and their volume measured.

131 2.7 Statistical analysis

132 The results were expressed in as mean standard error of mean (SEM) and statistical
133 analysis were carried out employing one way analysis of variance (ANOVA) followed by

134 Dunnett multiple comparisons test at $p < 0.05$ significance level using Graphpad software,
135 San Diego California USA, (www.graphpad.com).

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137 3. RESULTS AND DISCUSSION

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139 The percentage yield of Hydromethanolic leaves extract of *Ipomea asarifolia* (HLEIA) was
140 found to be 32.95%. The high percentage yield of HLEIA suggests that the plant is a good
141 source of extract since it contains sufficient amount which could be subjected further for
142 isolation studies.

143 In the acute oral toxicity studies, it was observed that oral administration of HLEIA to the rats
144 at 3000 mg/kg neither caused no mortality nor signs (hyperactivity, spasm, ruffled fur,
145 emesis, inappetance etc.) of toxicity in the animals within the first 24 hours and up to 14
146 days after its administration. This indicates that the lethal median dose (LD_{50}) of the extract
147 is greater than 3000 mg/kg suggesting the plant extract may be considered safe for
148 consumption as herbal formulation [17].

149 One hour after castor oil administration, all the rats in the control group produced copious
150 diarrhoea. HLEIA produced a marked anti-diarrhoea effect in the rats, as shown in Table 1.
151 At 150 mg/kg, the extract significantly ($p < 0.01$) decreased the total number of wet faeces
152 produced upon administration of castor oil compared with control group. Highest inhibition
153 percentage of defecation was observed with the extract at 150 mg/kg (40.00%) and with
154 Loperamide (64.62%).

155

156 Table 1: Effect of HLEIA on castor oil induced diarrhoea in albino rats

Treatment	Total number of faeces	Number of diarrhoea faeces	% Inhibition of diarrhoea
Normal saline(5mg/kg)+ castor oil (2mL)	22.25 ± 2.66	16.25 ± 2.18	-
Loperamide (5mg/kg)+ castor oil (2mL)	13.00 ± 0.91	5.75 ± 0.63**	64.62
HEIA (150 mg/kg)+ castor oil (2mL)	20.50 ± 0.87	9.75 ± 0.63**	40.00
HEIA (300 mg/kg)+ castor oil (2mL)	24.00 ± 0.91	11.25 ± 0.63*	30.77
HEIA (600 mg/kg)+ castor oil (2mL)	22.50 ± 0.87	11.00 ± 0.41*	32.31

157 Values are expressed as mean ± S.E.M; (n=5) in each group. Data were analyzed by one way ANOVA followed by Turkey-
158 Kramer multiple comparisons test. * $P < 0.05$ and ** $P < 0.01$ when compared to the control. HEIA=Hydromethanolic extract of
159 *Ipomoea asarifolia*.

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161 Several studies have shown that prior administration with some plant extract had protective
162 effect on the intestinal tract. These studies have validated the use of antidiarrhoea medicinal
163 plants by investigating the biological activity of extracts of such plants which have
164 antispasmodic effects, delayed intestinal transit, reduced gut motility, stimulate water
165 adsorption, or reduce the intraluminal fluid accumulation [18, 19, 20, 21].

166 In antimotility test, sub-chronic administration of graded doses of HLEIA showed significant
167 effect in treated animals receiving plant extract at 300mg/kg ($P < 0.01$) and at 600mg/kg
168 ($P < 0.05$) respectively compared with the control (Table 2). There was also a significant
169 increase ($P < 0.01$) in percentage intestinal transit in the drug-treated group when compared
170 with the control. The highest anti-diarrhoea effect was produced at 300 mg/kg of the extract,
171 which was comparable to the effect of the standard drug Loperamide.

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Table 2: Gastro intestinal motility effect of HLEIA in albino rats

Treatment	Length of intestine (cm)	Distance moved by charcoal meal(cm)	% Inhibition
Normal saline(5mg/kg) + castor oil (2mL)	86.03± 2.78	45.45 ± 2.56	0.00
Loperamide (5mg/kg) + castor oil (2mL)	90.00 ± 4.44	9.50 ± 3.43**	79.10
HEIA (150mg/kg) + castor oil (2mL)	82.25 ± 2.75	37.75 ± 8.41	16.94
HEIA (300mg/kg) + castor oil (2mL)	87.35 ± 3.65	14.63 ± 1.55**	68.10
HEIA (600mg/kg) + castor oil (2mL)	84.88 ± 3.33	21.80 ± 5.29*	52.04

174 Values are expressed as mean ± SEM from the experiment. Data analyzed by one way ANOVA, using Dunnett's comparison test.
175 *(P<0.05) and ** (P <0.01) significantly difference when compared with control group.

176 Gastrointestinal motility test with activated charcoal was carried out to find the effect of the
177 hydromethanolic extract of *I. asarifolia* on peristalsis movement. The result shows that
178 HLEIA (300mg/kg) was found to be comparable with the standard drug Loperamide, a drug
179 which is widely used for the treatment of diarrhoea. Loperamide is known to exert its
180 antidiarrhoea activity by changing the motor function of the intestine, which results in
181 increased capacitance of the gut and a delay in the passage of fluid through the intestine
182 [22].

183 Castor oil caused accumulation of water and electrolytes in intestinal loop. HLEIA compare
184 with the control, significantly (P < 0.01) and dose dependently inhibited castor oil-induced
185 enteropooling in rats (Table 3). The inhibition rates for the extract were 24.02, 41.69 and
186 50.53 % respectively at 150, 300 and 600 mg/kg. The intestinal fluid in control animals was
187 2.83 ± 0.48 mL. Inhibitions of intestinal fluid accumulation were 24.02, 41.69 and 50.53%
188 respectively at 150, 300 and 600 mg/kg. The standard drug Loperamide (5 mg/kg), also
189 significantly inhibited intestinal fluid accumulation (60.07 %).

190
191 Table 3: Enteropooling effect of HLEIA in albino rats

Treatment	Volume of intestinal fluid (mL)	% Inhibition
Normal saline(5mg/kg) + castor oil (2mL)	2.83 ± 0.48	--
Loperamide (5mg/kg) + castor oil (2mL)	1.13 ± 0.10**	60.07
HEIA (150mg/kg) + castor oil (2mL)	2.15 ± 0.16	24.02
HEIA (300mg/kg) + castor oil (2mL)	1.65 ± 0.06*	41.69
HEIA (600mg/kg) + castor oil (2mL)	1.40 ± 0.04**	50.53

192 Values are expressed as mean ± S.E.M; (n=5) in each group. Data were analyzed by one way ANOVA followed by Turkey-
193 Kramer multiple comparisons test. *P<0.05 and **P<0.01 when compared to the control. HEIA=Hydromethanolic extract of
194 *Ipomoea asarifolia*.
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196 Castor oil produces diarrhoea due to its most active metabolite, ricinoleic acid by
197 hypersecretory response, which stimulates peristaltic activity in the small intestine, leading to
198 changes in the electrolyte permeability of the intestinal mucosa [23]. Ricinoleic acid causes
199 irritation and inflammation of the intestinal mucosa leading to the release of prostaglandins

200 which stimulate hyper-motility, alteration in the electrolyte permeability of the intestinal
201 mucosa and increase in the volume of intestinal contents by preventing the reabsorption of
202 sodium, potassium and water [24, 25, 26]. In the present study, HLEIA showed a dose-
203 related anti-enteropooling effect via reduced volume of the intestinal contents and also
204 significantly inhibited castor oil-induced diarrhoea in rats by the significant reduction of the
205 number of diarrhoeal episodes and total faeces. This implies that the extract probably
206 enhanced the absorption of electrolytes and water from the intestinal lumen, while reducing
207 the rate of their secretion into the small intestine or has the ability to inhibit the castor oil-
208 induced intestinal accumulation of fluid in a manner similar to the standard anti-diarrhoeal
209 drug (Loperamide) [22].

210 In the phytochemical analysis, HLEIA showed the presence of alkaloids, saponins,
211 terpenoids, tannins, phenols, steroids and resins. The need for phytochemical screening has
212 become imperative since many plants accumulate biologically active complex organic
213 chemicals (secondary metabolites) in their tissues. Previous reports have demonstrated that
214 anti-diarrhoeal properties of medicinal plants were due to tannins, alkaloids, saponins,
215 terpenoids, flavonoids and sterols [27, 28, 29, 30, 31]. It could therefore be suggested that
216 the secondary metabolites present in *I. asarifolia* could be responsible for the
217 pharmacological effects observed.

218 4. CONCLUSION

219 The present study reveals that hydromethanolic leaves extract of *I. asarifolia* contains
220 phytoconstituents such as alkaloids, terpenoids, resins, tannin, saponin, phenols and
221 steroids that are known for their anti-diarrhoeal properties. The result obtained in this
222 research establishes its efficacy and scientifically validate the use of *I. asarifolia* in the
223 treatment of diarrhoea. Further research need to be undertaken to isolate and purify the
224 bioactive components of this plant.

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227 COMPETING INTERESTS

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229 The authors declare that they have no competing interests.

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233 ETHICAL APPROVAL

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235 "All authors hereby declare that "Principles of laboratory animal care" (NIH publication No.
236 85-23, revised 1985) were followed, as well as specific national laws where applicable.

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