

Phytochemical and Pharmacological Potential of *Enhydra fluctuans* available in Bangladesh

ABSTRACT

Objectives: The possible phytochemical constituents, thrombolytic and membrane stabilizing activities of the crude ethanolic extract of *Enhydra fluctuans* (CE) were investigated along with the antimicrobial, antioxidant and cytotoxic potentials of its petroleum ether (PESF), carbon tetrachloride (CTCSF), chloroform (CSF) and aqueous (AQSF) soluble fractions.

Materials & Methods: The coarse leaf powder was extracted at room temperature with ethanol. Solvent-solvent partitioning was done to obtain the four soluble fractions. Anticoagulant potential was determined by the *in vitro* thrombolytic model, membrane stabilization method was used to assess *in vitro* anti-inflammatory activity, the disc diffusion method was used for anti-microbial screening, antioxidant potential was determined by 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging method and brine shrimp lethality bioassay method was used for cytotoxic activity determination.

Results: Chemical screening of the crude extract evidenced the presence of alkaloids, saponins, tannins, flavonoids, reducing sugars and gums. It showed significant clot lysis property of 46.91%. It also significantly inhibited heat and hypotonic solution induced lysis of the human red blood cell membrane with values of 71.80% and 47.60%, respectively. CTCSF and PESF showed mild antimicrobial activity. AQSF showed most prominent antioxidant activity with IC₅₀ value of 12.27 µg/mL. CTCSF showed LC₅₀ value of 0.84 µg/mL, with most potent cytotoxic activity.

Conclusion: Significant thrombolytic, membrane stabilizing, antioxidant and *in vitro* cytotoxic activities of the ethanolic plant extract were observed in this study. *In vivo* activities and isolation of active compound(s) from this extract are yet to be investigated.

Keywords: *Enhydra fluctuans*, thrombolytic, membrane stabilizing, antioxidant and cytotoxic.

INTRODUCTION

Medicinal plants have played an important role in treating various diseases since ancient times. Increased drug resistance and side effects of existing drugs have escalated the research on traditionally available medicinal plants.

Myocardial or cerebral infraction and other atherothrombotic diseases are consequences of thrombus formed in blood vessel^{1,2}. Fibrinolytic drugs such as tissue plasminogen activator (t-PA), urokinase, streptokinase and others dissolve thrombin in acutely occluded coronary arteries

40 and restore blood supply to ischemic myocardium, to limit necrosis and improve prognosis³. Yet
41 all the available thrombolytic agents have significant deficiencies, including the necessity of
42 large doses to be maximally effective, limited fibrin specificity and a significant associated
43 bleeding tendency. Therefore, studies are going on to develop improved thrombolytic drugs in
44 order to minimize deficiencies of the available drugs.

45 Inflammation is a pathological disorder, in which inflammatory cells produce a complex mixture
46 of growth and differentiation cytokines as well as physiologically active arachidonate
47 metabolites, also generate reactive oxygen species (ROS) that can damage cellular biomolecules
48 which in turn augment the state of inflammation⁴. Compounds that possess radical scavenging
49 ability may therefore expect to have the therapeutic potentials against inflammation⁵. Moreover,
50 stabilization of lysosomal membrane limits inflammatory response by inhibiting the release of
51 lysosomal constituents of activated neutrophil such as bactericidal enzymes and proteases, which
52 are responsible for further tissue inflammation and damage⁶.

53 Currently antibiotic resistance has become a major clinical and public health problem for most
54 people^{7,8}. This multidrug resistance (MDR) is clearly related to the misuse of different
55 antibiotics^{9,10}. The increasing clinical importance of drug resistant bacterial pathogens
56 necessitates of the search for additional antibacterial therapy. The antibacterial screening which is
57 the first stage of antibacterial research is performed to ascertain the susceptibility of various
58 bacteria to any agent.

59 Free radicals are highly toxic and reactive; and cause diseases like aging, atherosclerosis, cancer,
60 diabetes, liver cirrhosis, cardiovascular disorders, etc. by attacking different macro-molecules
61 including lipids, proteins and DNA resulting in the cellular damage¹¹⁻¹³. Currently available
62 synthetic antioxidants like butylatedhydroxy anisole (BHA), butylatedhydroxy toluene (BHT),
63 tertiary butylated hydroquinone and gallic acid esters show low solubility and moderate
64 antioxidant activity with suspected negative health effects¹⁴⁻¹⁶. Investigations to screen out
65 naturally occurring antioxidants are now mostly demanding.

66 Bioactive compounds are always toxic to living body at some higher doses. Brine shrimp
67 lethality bioassay implies cytotoxicity as well as a wide range of pharmacological activities such
68 as antimicrobial, antiviral, pesticidal and anti-tumor activities^{17,18}.

69 *Enhydra fluctuans* Lour (Family: Asteraceae) is a small genus of marsh herb, available in
70 tropical and subtropical regions namely Bangladesh, India, Malaysia, China and the rest of South
71 East Asia and Tropical Africa^{19,20}. The herb is relatively glabrous sometimes pubescent
72 glandular. Stems are usually 0.3 to 0.6m, elongated simple or divaricating rooting at the nodes²¹.
73 Traditional uses indicate that the leaves of the plant are good as laxative. The expressed juice of
74 the leaves is an excellent demulcent in gonorrhoea and also used in the treatment of skin and
75 nervous system²². Different phytoconstituents such as flavonoids, isoflavone glycosides,
76 terpenoids like sesquiterpene lactones, saponins etc have already been isolated from *E.*
77 *fluctuans*²³⁻²⁶. There are reports on antioxidant property of the methanolic extract, antimicrobial
78 property of the toluene, chloroform and methanolic extract of *E. fluctuans*, moreover CNS
79 depressant activity, anti-inflammatory activity of *E. fluctuans* have also been reported²⁷⁻³⁰. But
80 there are insufficient records in literature, regarding pharmacological activities and
81 phytochemical characteristics of *E. fluctuans* Lour available in Bangladesh which are known to
82 differ with plant's geographical location. Thus the present study focuses on screening of the

83 ethanolic extract of *Enhydra fluctuans* Lour to identify its phytochemical and pharmacological
84 potential.

85 MATERIALS AND METHODS

86 *Instrumentation*

87 The extract was concentrated in rotary vacuum evaporator (Heidolph Instruments GmbH & Co.
88 KG, Germany). Samples were centrifuged on a bench top centrifuge machine (HERMLE
89 Labortechnik GmbH, Germany). The absorbance was recorded by a UV-Visible
90 spectrophotometer (Analytic Jena AG, Germany). Ketoconazole was obtained from Beximco
91 Pharmaceuticals Ltd., Bangladesh. Streptokinase vial (Beacon Pharmaceutical Ltd) was
92 purchased from local pharmacy. Trolox and BHA were obtained from Calbiochem, Merck,
93 Germany and VWR International Ltd., England, respectively. Nutrient agar and potato dextrose
94 agar and kanamycin 30 µg standard disc were purchased from HiMedia Laboratories Pvt. Ltd.,
95 India. All other reagents were purchased from Sigma-Aldrich, USA and all other solvents used
96 were of analytical grade and purchased from Active Fine Chemicals Ltd., Bangladesh.

97 *Collection and Identification*

98 *Enhydra fluctuans* leaves were collected from Narayangonj, Bangladesh on February 2015. Plant
99 was identified by taxonomist. The leaves were thoroughly washed with clean water, dried in the
100 shade for several days and oven dried for 24 hours at not more than 40°C. The dried leaves were
101 then ground to a coarse powder. The powder (140 g) was subjected to cold maceration by
102 soaking in 1200 mL of ethanol in a clean, amber colored reagent bottle. The container was kept
103 for a period of 10 days with occasional shaking and stirring. The whole mixtures were then
104 filtered through a fresh cotton plug and finally with a Whatman No.1 filter paper. The filtrate
105 was concentrated at 40°C under reduced pressure to evaporate approximately 70% of the solvent
106 to obtain the crude extract, CE. 5.0 g of CE was dissolved in 10% aqueous ethanol. Solvent-
107 solvent partitioning of this solution was done using the protocol designed by Kupchan and
108 modified by Van Wagenen *et al.*, (1993) with petroleum ether, then with carbon tetrachloride and
109 finally with Chloroform to obtain three fractions (PESF, CTCSF and CSF) and the remaining
110 part was named as aqueous soluble fraction (AQSF)³¹. Before carbon tetrachloride and
111 chloroform extraction, water was added to the remaining aqueous extract to increase the polarity
112 difference for separation.

113 *Phytochemical Screening*

114 Approximately 5 mg of CE was diluted in ethanol to perform the following tests for identification
115 of different chemical groups^{32,33}.

116 *Tests for tannins*

117 Ferric chloride test: About 5 mL of the extract solution was taken in a test tube, followed by
118 addition of 1 mL of 5% ferric chloride solution. Greenish black precipitate indicates the presence
119 of tannins.

120 Potassium dichromate test: About 1 mL of 10% potassium dichromate solution was added with 5
121 mL of the extract solution in a test tube. Yellow precipitate suggests the presence of tannins.

122 Lead acetate test: About 1 mL of 10% lead acetate solution was added to 5 mL of extract
123 solution. Yellow precipitate formation is indicative of presence of tannins.

124 ***Test for Flavonoids***

125 A few drops of concentrated hydrochloric acid were added to a small amount of CE. Immediate
126 red color formation is caused by flavonoids.

127 ***Test for Saponins***

128 About 1 mL of the solution was diluted with 20 mL distilled water and shaken in a graduated
129 cylinder for 15 minutes. One-centimeter layer of foam formation is indicative of the presence of
130 saponins.

131 ***Tests for Gums***

132 About 5 mL solution of the extract was added to a test tube containing molisch reagent and
133 sulfuric acid. Red violet ring at the junction of the two liquids evidences presence of gums.

134 ***Tests for Steroids***

135 About 1 mL concentrated sulfuric acid was added to 1 mL of chloroform extract. Red color in the
136 lower layer indicates the presence of steroids.

137 ***Tests for alkaloids***

138 Mayer's test: About 2 mL solution and 0.2 mL of dilute hydrochloric acid were taken in a test
139 tube followed by addition of 1 mL of Mayer's reagent. Yellowish buff colored precipitate
140 indicates the presence of alkaloids.

141 Dragendroff's test: About 2 mL solution and 0.2 mL of dilute hydrochloric acid were taken in a
142 test tube followed by addition of 1 mL of Dragendroff's reagent. Orange brown precipitate
143 evidences the presence of alkaloids.

144 Hager's test: About 2 mL of the extract and 0.2 mL of dilute hydrochloric acid were taken in a
145 test tube. Then 1 mL of picric acid solution or Hager's reagent was added. Yellowish precipitate
146 evidences the presence of alkaloids.

147 ***Test for reducing sugar***

148 About 2 mL of aqueous extract was added to 1 mL of a mixture of equal volumes of Fehling's
149 solutions A and B followed by boiling for few minutes. Brick red colored precipitate is indicative
150 of the presence of reducing sugar.

151 ***Thrombolytic activity***

152 The extract of *E. fluctuans* was assessed for thrombolytic activity using *in vitro* thrombolytic
153 model³⁴. Taking account of all ethical considerations, and aseptic precautions, 20 mL of venous
154 blood was drawn from healthy human volunteer without a history of oral contraceptive or
155 anticoagulant therapy. This withdrawn blood was then distributed in pre-weighed sterile vials (1
156 mL/tube) and incubated at 37°C for 45 minutes. After clot formation, the serum was completely
157 removed without disturbing the clot and clot weight was determined (clot weight = weight of clot
158 containing vial – weight of vial alone). About 100 mg of CE was dissolved in 10 mL of ethanol
159 and kept overnight. The soluble supernatant was decanted and filtered. Approximately 100 µL of
160 the solution was added to a vial containing pre-weighed blood clot. Lyophilized streptokinase
161 (SK) vial of 15,00,000 I.U. was reconstituted with 5 mL sterile distilled water. About 100 µL
162 (30,000 I.U) of the solution was added as positive control and 100 µL of distilled water was
163 added as a negative non thrombolytic control to the vial containing pre-weighed blood clot

164 separately. All the three vials were then incubated at 37°C for 90 minutes and observed for clot
165 lysis. After incubation, the released fluid was removed and vials were again weighed to observe
166 the weight difference after clot disruption. This difference of weight before and after clot lysis
167 was expressed as the percentage of clot lysis as the following³⁴:

168 **Percentage of clot lysis = (wt. of released clot/clot wt.) × 100**

169 *Membrane stabilization activity*

170 To assess the anti-inflammatory activity, membrane stabilization potential of CE was evaluated
171 by measuring the heat and hypotonic solution induced hemolysis of erythrocyte following
172 standard protocol³⁵.

173 ***Preparation of erythrocyte suspension***

174 Fresh whole blood (20 mL) was collected intravenously in syringe containing 3.1% sodium
175 citrate solution as anti-coagulant from healthy human volunteer without history of non steroidal
176 anti-inflammatory drugs (NSAIDs) therapy for 2 weeks prior the experiment. The blood cells
177 were washed three times with isotonic buffer solution, 154 mM NaCl in 10 mM sodium
178 phosphate buffer (pH 7.4) through centrifugation at 3000 g for 10 min.

179 ***Heat induced hemolysis***

180 CE was dissolved in isotonic solution. Isotonic buffer containing aliquots (5 mL) of the extract at
181 1.0 mg/mL was taken in six centrifuge tubes in three sets of two. Two sets of control tubes
182 containing 5 mL of the vehicle control and 5 mL of 0.1 mg/mL of **acetyl salicylic acid (Aspirin)**,
183 respectively. Erythrocyte suspension was added to each tube and mixed gently by inversion. One
184 pair of the tubes was incubated at 56°C for 30 min in a water bath, while the other pair was
185 maintained at 0 to 5°C in an ice bath. The reaction mixture was centrifuged for 5 min at 2500g
186 and the absorbance of the supernatant was measured at 560 nm. The percentage inhibition or
187 acceleration of hemolysis was calculated according to the equation³⁵:

188 Percentage of inhibition of hemolysis = $100 \times [1 - (OD_2 - OD_1) / (OD_3 - OD_1)]$

189 Where,

190 OD₁ = optical density of unheated test sample; OD₂ = optical density of heated test sample; OD₃ =
191 optical density of heated control sample.

192 ***Hypotonic solution induced hemolysis***

193 About 5 mL hypotonic solution (distilled water) containing 1.0 mg/mL of CE were put in
194 centrifuge tube in triplicates. About 5 mL of hypotonic solution and 5 mL of acetyl salicylic acid
195 **(Aspirin)** at 0.1 mg/mL were taken as negative and positive controls respectively in separate
196 centrifuge tubes. Erythrocyte suspension (0.5 mL) was added to each of the tube and mixed
197 gently. The mixture was incubated for 10 min at room temperature and then centrifuged for 10
198 min at 3000 g. The absorbance of the supernatant was measured at 540 nm. The percentage
199 inhibition of hemolysis or membrane stabilization was calculated using the following equation³⁵:

200 Percentage of inhibition of hemolysis = $100 [1 - \{(OD_2 - OD_1) / (OD_3 - OD_1)\}]$

201 Where OD₁ = absorbance of test sample in isotonic solution; OD₂ = absorbance of test sample in
202 hypotonic solution; OD₃ = absorbance of control hypotonic sample.

203 *Antimicrobial assay*

204 The disc diffusion technique³⁶ was used for preliminary screening of antimicrobial activity. Two
 205 Gram Positive bacterial strains (*Bacillus megaterium* ATCC 13578, *Staphylococcus aureus*
 206 ATCC 25923), two Gram Negative bacterial strains (*Escherichia coli* ATCC 25922,
 207 *Pseudomonas aeruginosa* ATCC 27833) and two fungal strains (*Aspergillus niger* and
 208 *Aspergillus flavus*) were freshly cultured in nutrient agar media for bacteria and potato dextrose
 209 agar media for fungi. PESF, CTCSF, CSF, AQSF fractions of the crude extracts were dissolved
 210 separately in specific volume of dichloromethane or methanol depending on their solubility. The
 211 diluted samples were applied on the sterile discs at a concentration of 500 µg/disc. Kanamycin
 212 (30 µg/disc) and ketoconazole (30 µg/disc) were used as standard antibiotics for antibacterial and
 213 antifungal screening, respectively. Solvent was used as negative control. The antimicrobial
 214 activity of the test agent was checked after 18 hrs of incubation for bacteria at 37 °C and 48 hrs
 215 of incubation for fungi at 28 °C. The result was determined by measuring the diameter of zone of
 216 inhibition expressed in mm.

217 *Antioxidant scavenging activity*

218 Antioxidant potential was studied using DPPH radical scavenging activity with slight
 219 modification of the method described by Brand-William *et al*³⁷. Here, sample solution of varying
 220 concentrations such as 200 µg/mL, 100 µg/mL, 50 µg/mL, 25 µg/mL, 12.5 µg/mL, 6.25 µg/mL,
 221 3.125 µg/mL, 1.5625 µg/mL and 0.78125 µg/mL were obtained by serial dilution technique in
 222 test tubes, where 2 mL of each of the test sample solution was mixed with 2 mL of a DPPH-
 223 methanol solution (20 µg/mL) to obtain the above mentioned concentrations. Test tubes were
 224 allowed to stand for 20 minutes in dark for the reaction to occur. The absorbance was determined
 225 at 517 nm and percentage of inhibition was calculated by using the following equation³⁷:

226 Percentage of radical inhibition = $[1 - (\text{ABS}_{\text{sample}} / \text{ABS}_{\text{control}})] \times 100$.

227 Then % inhibitions were plotted against respective concentrations used and from the graph IC₅₀
 228 was calculated. Here, ascorbic acid and BHA were used as the positive controls.

229 *Cytotoxic activity*

230 Brine shrimp lethality bioassay technique was applied for the determination of general toxic
 231 properties of the plant extracts against *Artemia salina*^{18,38}. The test samples were dissolved in
 232 dimethyl sulfoxide (DMSO). Serial dilution technique is used to obtain sample concentrations of
 233 400, 200, 100, 50, 25, 12.5, 6.25 and 3.125 µg/mL in separate test tubes containing 10 shrimps in
 234 simulated brine water (5 mL). The test tubes were incubated at room temperature for 24 hours.
 235 The LC₅₀ of the test samples was determined by a plot of percentage of the shrimp mortality
 236 against the logarithm of the sample concentrations.

237 **RESULTS AND DISCUSSION**

238 *Phytochemical screening*

239 Preliminary phytochemical screening evidenced the presence of alkaloids, saponins, tannins,
 240 flavonoids, reducing sugars and gums (**Table 1**).

241 **Table 1:** Results of chemical group tests.

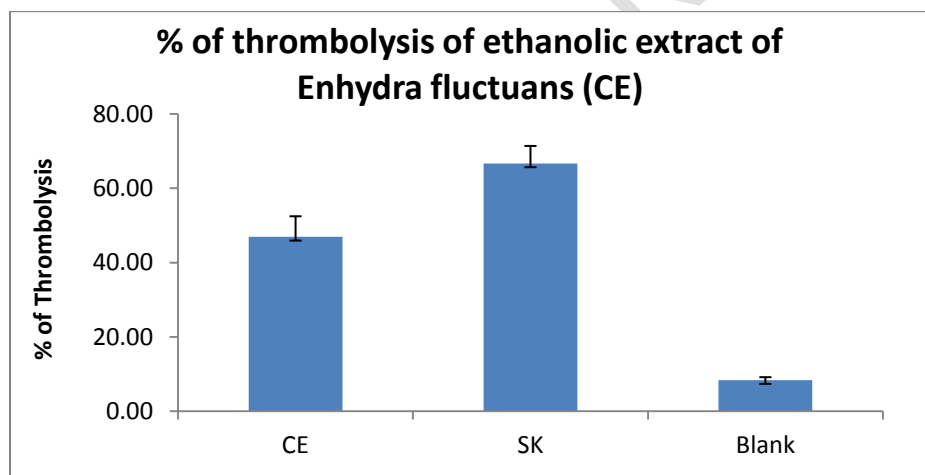
Tested groups	Alkaloids	Steroids	Saponin	Tannins	Flavonoids	Reducing Sugars	Gums
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Ethanollic Extract of <i>Enhydra fluctuans</i>	+	-	+	+	+	+	+
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242 Note: + = Indicates the presence of the tested group, - = Indicates the absence of the tested
 243 group.

244 *Thrombolytic activity*

245 Addition of 100 μ L SK solution, a positive control to the blood clots and subsequent incubation
 246 resulted in 66.67% lysis of clot. On the other hand, negative control exhibited a negligible lysis
 247 of blood clot (8.33%). When blood clots were treated with the test sample significant clot lysis
 248 activity (46.91%) was observed. When compared with the negative control (water) the mean
 249 percentage (%) of blood clot lysis was significant ($p < 0.001$) as shown in **Figure 1**. Blood clot
 250 or fibrin lysis in thrombosis occurs by plasmin produced from plasminogen which in turn
 251 activated by plasminogen activator³⁹. The plant extract thus may act as plasminogen activator to
 252 produce clot lysis activity almost similar to standard which must be proved through more
 253 specific test.

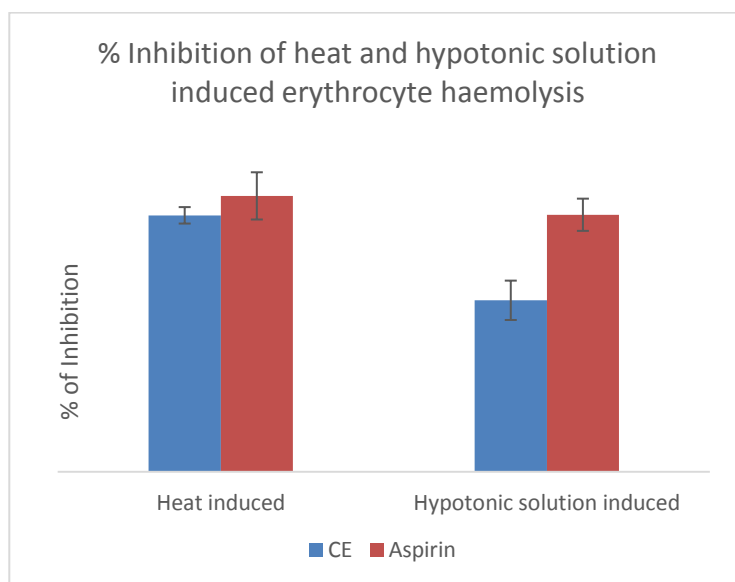


254
 255 **Figure 1:** Thrombolytic activity (in terms of % of clot lysis) with standard deviation error bar
 256 where n=3. “CE”-Ethanolic crude extract of *E. fluctuans*, “SK”-Positive control.

257 *Membrane stabilizing activity*

258 Flavonoid rich ethyl acetate extract of *E. fluctuans* collected from West Bengal, India showed
 259 significant anti-inflammatory activity in carrageenan and histamine induced paw edema in rats²³.
 260 In our study the ethanolic extracts of *E. fluctuans* at concentration 1.0 mg/mL significantly
 261 protected the lysis of human erythrocyte membrane by hypotonic solution and heat induced
 262 hemolysis compared to the standard aspirin (0.10 mg/mL). For heat induced condition CE
 263 demonstrated 71.80% inhibition of hemolysis of RBCs, whereas aspirin inhibited 77.20%. On
 264 the other hand, during hypotonic solution induced hemolysis, CE inhibited 47.60% hemolysis of
 265 RBCs as compared to 71.90% produced by aspirin as shown in **Figure 2**. The erythrocyte
 266 membrane resembles to lysosomal membrane and as such, the effect of drugs on the stabilization

267 of erythrocyte could be extrapolated to the stabilization of lysosomal membrane to exhibit anti-
 268 inflammatory activity^{40,41}.



269
 270 **Figure 2:** Membrane stabilizing activity (in terms of % inhibition of erythrocyte hemolysis) with
 271 standard deviation error bar where n=3. “CE”-Ethanollic crude extract of *E. fluctuans*, “Aspirin”-
 272 Positive control.

273 *Antimicrobial assay*

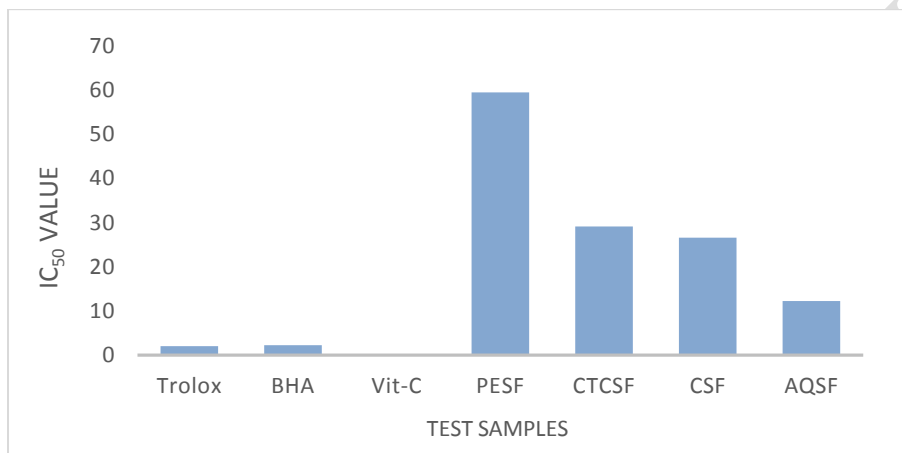
274 **Table 2** shows antimicrobial activity of the tested samples. PESF showed mild activity against
 275 Gram negative bacteria *Escherichia coli* and *Pseudomonas aeruginosa*. CTCSF exhibited mild
 276 activity against both Gram positive and Gram negative bacteria. All the fractions were inactive
 277 against fungal strains. There are reports of mild to moderate antibacterial activity of the
 278 methanolic, ethanolic, acetonic and chloroform extract of the plant against *Staphylococcus*
 279 *aureus*, *Staphylococcus saprophyticus*, *Bacillus cereus*, *Escherichia coli*, *Salmonella typhi*,
 280 *Shigella sonnei*, *Shigella shiga*⁴²⁻⁴⁴. As CTCSF exhibited most prominent activity as compared to
 281 the other fractions, further study is necessary to isolate phytoconstituent with antimicrobial
 282 property from the chloroform extract of *E. fluctuans*.

283 **Table 2:** Antimicrobial activity of test samples of *E. fluctuans*.

Test organisms	Diameter of zone of inhibition(mm)				
	PESF 100 µg/disc	CTSF 100 µg/disc	CSF 100 µg/disc	AQSF 100 µg/disc	Kanamycin/ Ketoconazole (30 µg/disc)
<i>B. megaterium</i>	–	7	–	–	40
<i>S. aureus</i>	–	8	–	–	28
<i>E. coli</i>	7	8	–	–	26
<i>P. aeruginosa</i>	7	7	–	–	27
<i>A. niger</i>	–	–	–	–	26
<i>A. flavus</i>	–	–	–	–	36

284 *Antioxidant assay*

285 The antioxidant activity was expressed by the IC₅₀ value of the samples. Comparison of IC₅₀
286 values of different fractions and extracts with standard was shown in **Figure 3**. AQSF was found
287 to be most potent with lowest IC₅₀ value. IC₅₀ values of PESF, CTCSF, CSF, AQSF, trolox,
288 BHA, Vitamin C are 59.59 µg/mL, 29.28 µg/mL, 26.62 µg/mL, 12.27 µg/mL, 2.04 µg/mL, 2.21
289 µg/mL and 0.17 µg/mL respectively. Previous report of *E. fluctuans* collected from West Bengal,
290 India suggested significant antioxidant property of ethyl acetate extract⁴⁵. Also there has been
291 report of isolation of antioxidative carbohydrate polymer from aqueous extract of *E fluctuans*
292 which is most consistent with our finding⁴⁶.



293
294 **Figure 3:** Comparison of IC₅₀ value of different fractions of *E. fluctuans* ethanolic extract and
295 standards.

296 *Cytotoxicity assay*

297 The lethality of the PESF, CTCSF, CSF and AQSF fractions against *A. salina* were shown in
298 **Table 3**. Lethality assay is proved to be suggestive of different pharmacological properties by
299 isolation of cytotoxic, antimalarial or insecticidal compounds from the plant extracts⁴⁷. Previous
300 reports had suggested cytotoxic potential of the crude methanolic and ethanolic extracts of *E.*
301 *fluctuans*⁴⁴. Flavonoids isolated from ethyl acetate fraction of *E. fluctuans* exhibited anticancer
302 activity tested *in vitro* in mice⁴⁸. There is report on cell-mediated immune system stimulation by
303 *E. fluctuans* leaves through increasing neutrophil's phagocytic activity⁴⁹. In this study all the
304 fractions exhibited significant cytotoxic activity in the brine shrimp lethality bioassay of which
305 CTCSF fraction was most potent with the lowest LC₅₀ value. Presence of several
306 phytoconstituents such as saponins, tannins, flavonoids in the ethanolic extracts of *E. fluctuans*
307 have been proved in our study which may facilitate these potent activities. So further
308 investigation on its different fractions is necessary to isolate bioactive metabolites and specify
309 their pharmacological activities.

310 **Table 3:** Lethal concentration for 50% mortality (LC₅₀) of different extracts of *E. fluctuans*.

Sample	PESF	CTCSF	CSF	AQSF
LC ₅₀ value (µg/mL)	0.97	0.84	1.43	1.16

311 **CONCLUSION**

312 Preliminary chemical screening and *in-vitro* bioactivity study data suggests *Enhydra fluctuans*'s
313 potential pharmacological activity. It showed significant cytotoxic activity with good
314 thrombolytic and anti-inflammatory potential. The study necessitates further higher and detailed
315 experiments for isolation of bioactive compounds from *Enhydra fluctuans* which may act as lead
316 compounds for new or improved drug development.

317 **Conflict of Interest**

318 There is no conflict of interest to declare by the authors.

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